

Differences in the Inactivation of *Legionella pneumophila* Serogroups Using UV-C LED Technology in Drinking Water

Helen Y Buse

Coauthors: John Hall, Gary Hunter, James Goodrich

International Ultraviolet Association 2021 World Congress

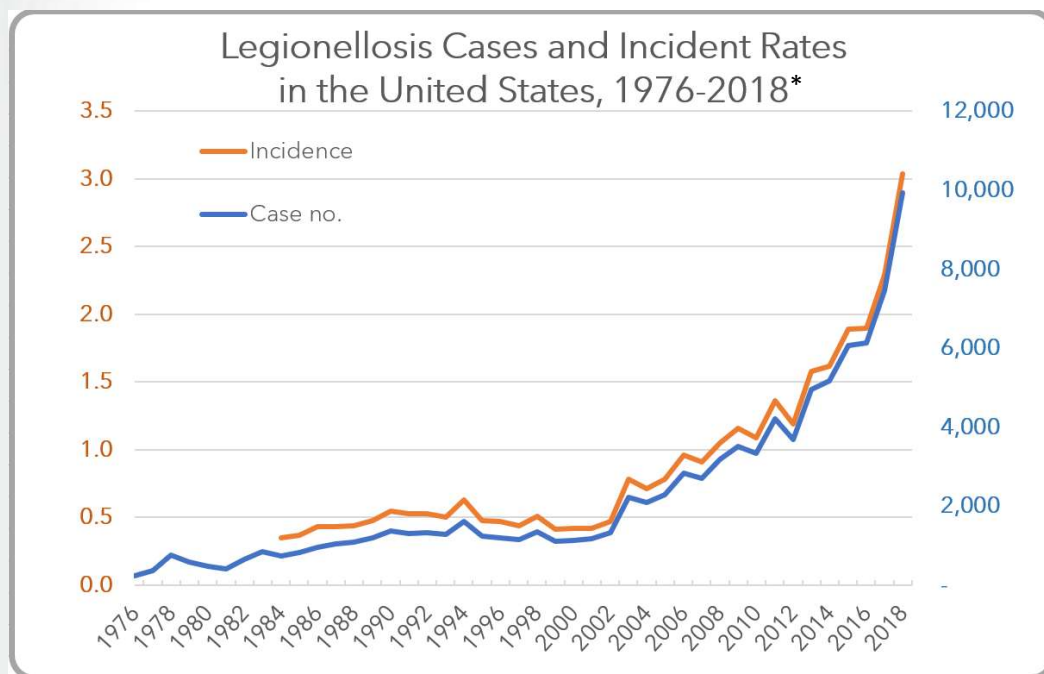


Acknowledgements

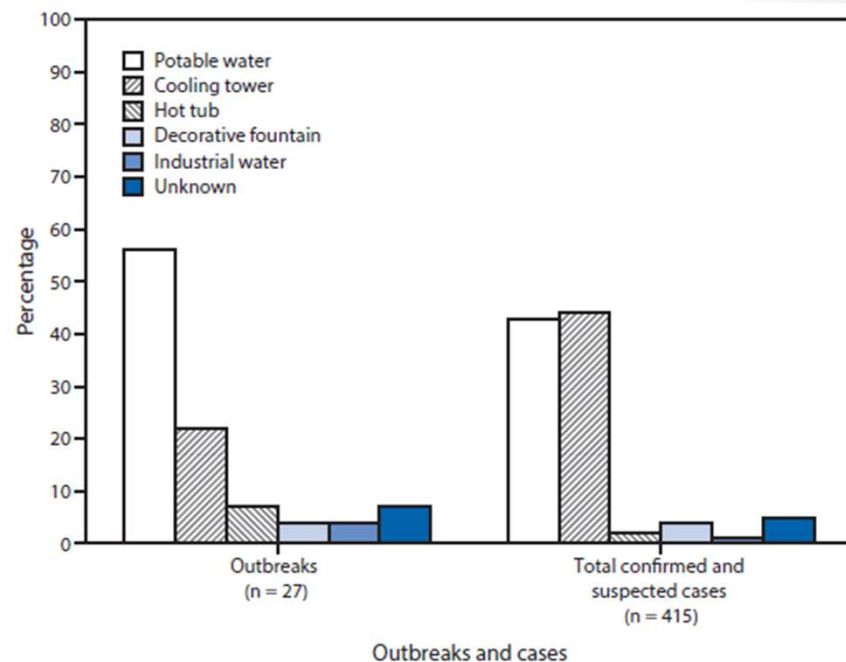




Legionellosis – A Public Health and Economic Burden



*National Notifiable Diseases Surveillance System



- Incidence rate up from 0.4 to 3.0 cases per 100,000 people; exposure to potable water responsible for the majority of outbreak cases
- In 2014, LD cases had an estimated healthcare cost of \$402 million (emergency department visits and hospitalizations)



Legionella spp.

60+ species (serogroups)

<i>L. adalaidensis</i>	<i>L. feeleeii</i> (2)	<i>L. monrovica</i>	<i>L. septentrionalis</i>
<i>L. anisa</i>	<i>L. geestiana</i>	<i>L. moravica</i>	<i>L. shakespearei</i>
<i>L. beliardensis</i>	<i>L. gormanii</i>	<i>L. nagasakiensis</i>	<i>L. spiritensis</i>
<i>L. birminghamensis</i>	<i>L. gratiana</i>	<i>L. nautarum</i>	<i>L. steelei</i>
<i>L. bozemanæ</i> (2)	<i>L. gresilensis</i>	<i>L. norrlandica</i>	<i>L. steigerwaltii</i>
<i>L. brunensis</i>	<i>L. hackeliae</i> (2)	<i>L. oakridgensis</i>	<i>L. saoudiensis</i>
<i>L. busanensis</i>	<i>L. impletisoli</i>	<i>L. parisiensis</i>	<i>L. taurinensis</i>
<i>L. cardiaca</i>	<i>L. israelensis</i>	<i>L. pittsburghensis</i>	<i>L. thermalis</i>
<i>L. cherrii</i>	<i>L. jamestowniensis</i>	<i>L. pneumophila</i> (15)	<i>L. tucsonensis</i>
<i>L. cincinnatiensis</i>	<i>L. jordanis</i>	<i>L. qingyii</i>	<i>L. tunisiensis</i>
<i>L. drancourtii</i>	<i>L. lansingensis</i>	<i>L. quateirensis</i>	<i>L. wadsworthii</i>
<i>L. dresdenensis</i>	<i>L. londiniensis</i>	<i>L. quinlivanii</i> (2)	<i>L. waltersii</i>
<i>L. drozanskii</i>	<i>L. longbeachae</i> (2)	<i>L. rowbothamii</i>	<i>L. worsleiensis</i>
<i>L. dumoffii</i>	<i>L. lytica</i>	<i>L. rubrilucens</i>	<i>L. yabuuchiae</i>
<i>L. erythra</i> (2)	<i>L. maceachernii</i>	<i>L. sainthelensi</i> (2)	
<i>L. fairfieldensis</i>	<i>L. massiliensis</i>	<i>L. santicrucis</i>	
<i>L. fallonii</i>	<i>L. micdadei</i>	<i>L. saoudiensis</i>	



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Dresden panel
of mAbs (17),
groups Lp into
sg 1 to 15

Lück, C. et al. 2013.
Typing Methods for
Legionella, p. 119-
148. In C.
Buchrieser and H.
Hilbi (ed.),
Legionella.

sg1, >80% clinical
isolates

About 1/3 of these species associated with human disease

Lipopolysaccharide and Serogroup Relationship

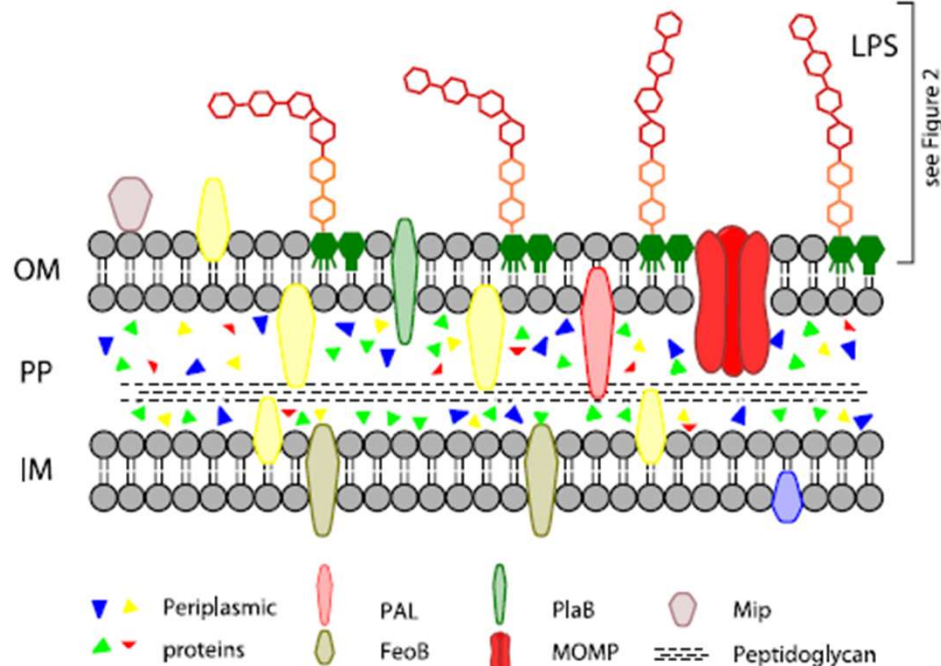


FIGURE 1 | Overview of the *L. pneumophila* cell envelope.

- LPS: 3 structural domains
 - Lipid A
 - inner and outer core
 - O-antigen

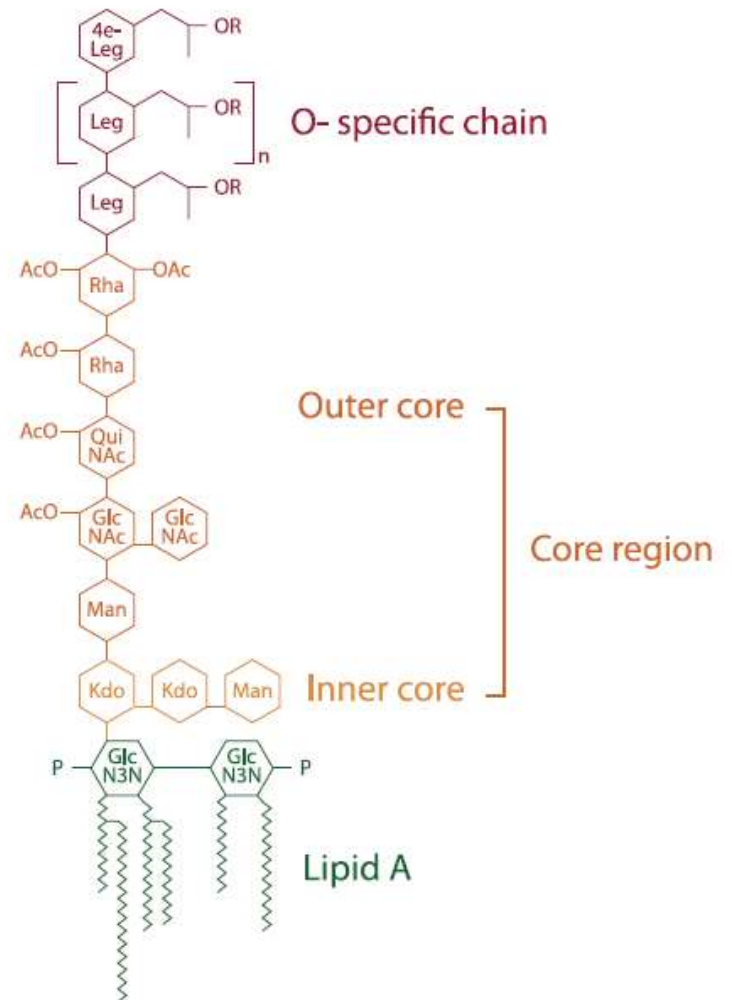


FIGURE 2 | Chemical structure of *L. pneumophila* LPS

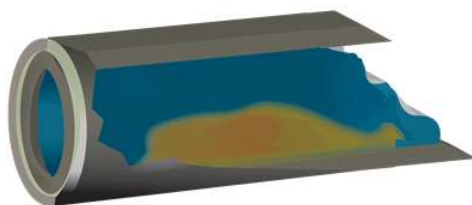
Adapted from Shevchuk et al. 2011 Front Microbiol



Legionella's Niche Within Premise Plumbing



generation of
respirable,
DW-derived aerosols



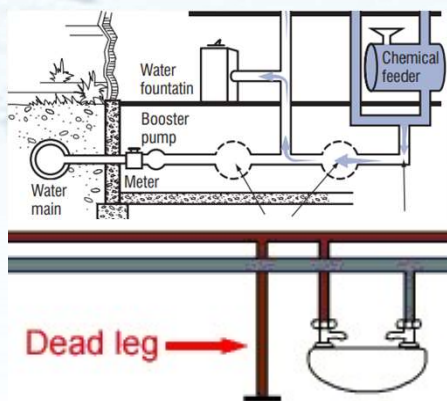
biofilms and sediments



amenable growth
temperatures and
gradients

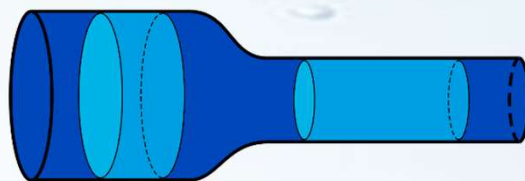


low to absent
residual



cross-connections

nutrients (metals, minerals, microbes, etc.)



high surface area to
volume ratios



eukaryotic hosts



water stagnation



Summary of Previous *Legionella* UV Studies

- UV low pressure (LP) and medium pressure (MP) studies:
 - LP: *L. gormanii*; *L. longbeachae*; *L. pneumophila* sg1 (clinical, drinking water, other environmental isolates), sg7, sg8
 - MP: *L. pneumophila* sg1
 - 2-log reduction: 0.9-5 mJ/cm²
 - 4-log reduction: 2.8-9.3 mJ/cm²

- UV light emitting diodes (LED) studies:
 - *L. pneumophila* sg1 (clinical and drinking water isolates); *L. rubrilucens*
 - 2-log reduction: 1.1 mJ/cm² (UV-C); 25 J/cm² (405nm violet LED); 4 mJ/cm² (LP)

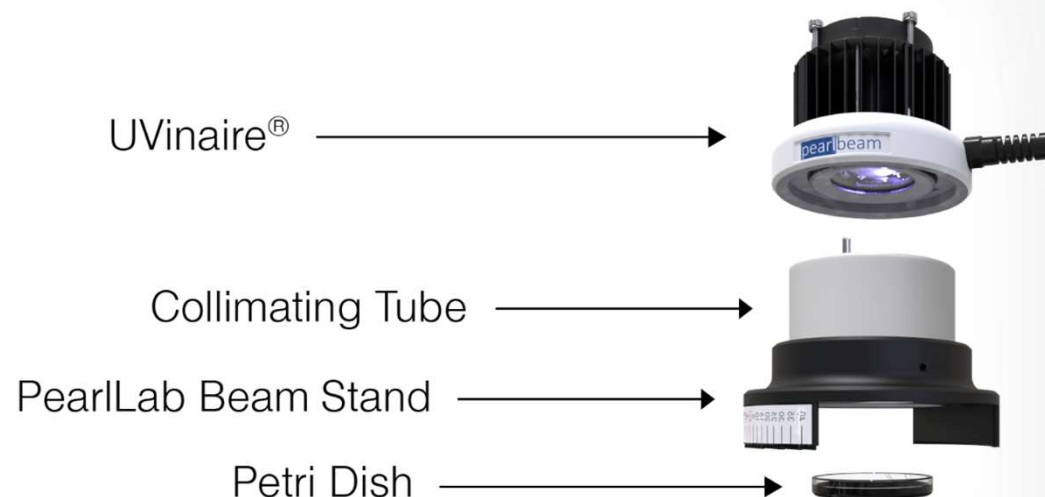
References for LP/MP studies:

Antopol & Ellner 1979 [Appl Environ Microbiol](#)
Cervero-Aragó et al. 2014 [Water Res](#)
Gilpin 1983 [2nd Intl Leg Symp](#)
Malayeri et al. 2016 [IUVA News](#)
Miyamoto et al. 2000 [Microbios](#)
Muraca et al. 1987 [Appl Environ Microbiol](#)
Oguma et al. 2004 [Water Res](#)
Wilson et al. 1992 [AWWA WQTC](#)

References for LED studies:

Carlson et al. *unpublished*
Hessling et al. 2018 [Hosp Pract Res](#)
Rattanakul & Oguma 2018 [Water Res](#)
Schmid et al. 2017 [GMS Hyg Infect Control](#)

Collimated Beam Method



fluence (mJ cm ⁻²)	255nm (UVT 79.7%)		265nm (UVT 86.5%)		280nm (UVT 91.7%)	
	min	sec	min	sec	min	sec
0.5	0	17	0	4	0	2
1	0	34	0	9	0	3
2	1	7	0	18	0	6
5	2	48	0	44	0	15
10	5	36	1	29	0	31
16					0	49
34					1	44

4 *L. pneumophila* strains:

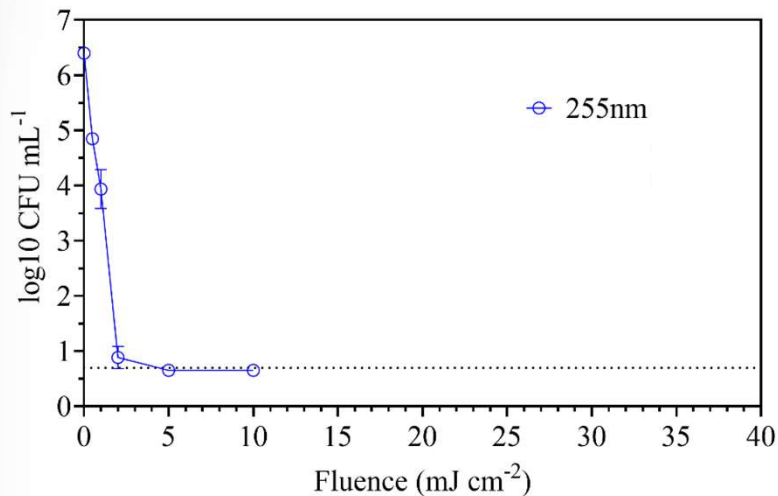
- sg1 clinical
- sg1 drinking water
- sg4 clinical
- sg6 clinical

Three replicates

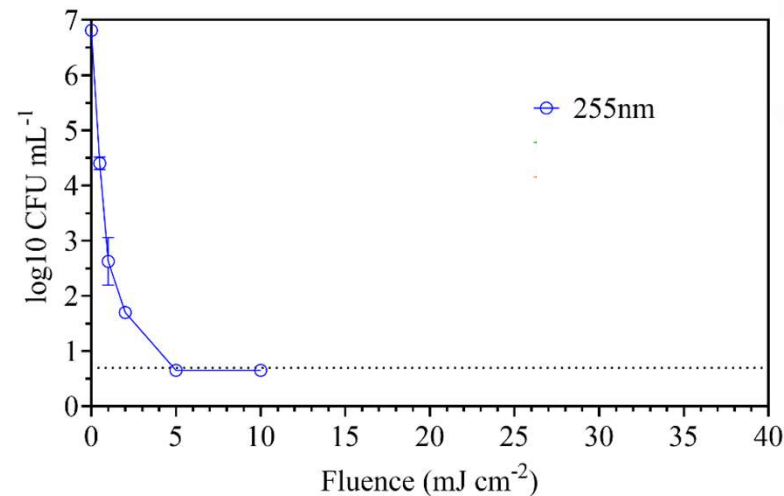


Collimated Beam Results – 255nm

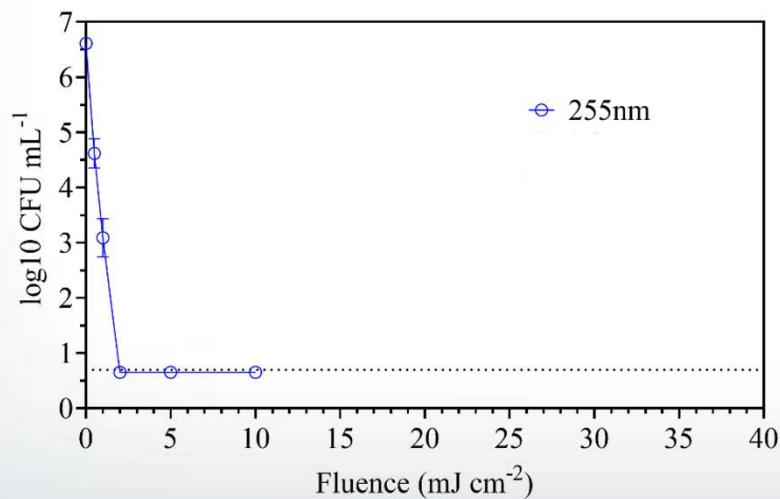
sg1



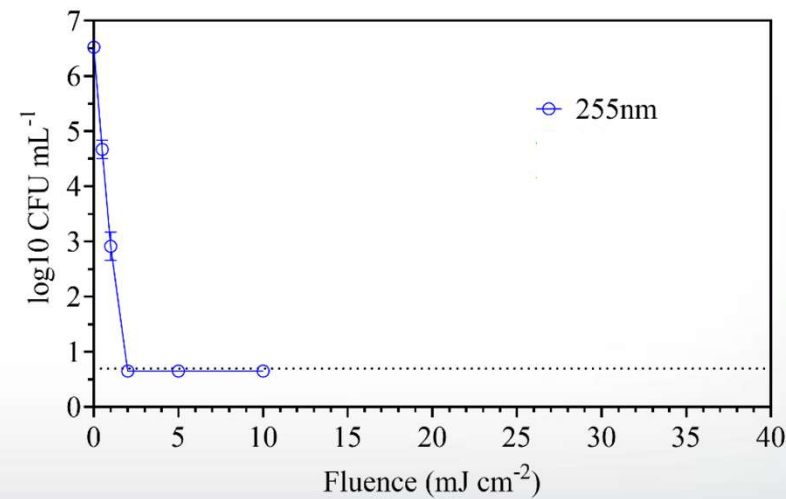
sg1 DW



sg4



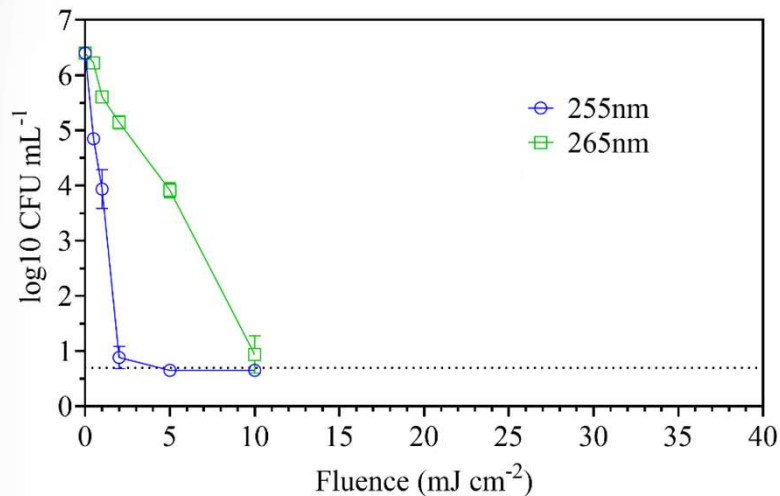
sg6



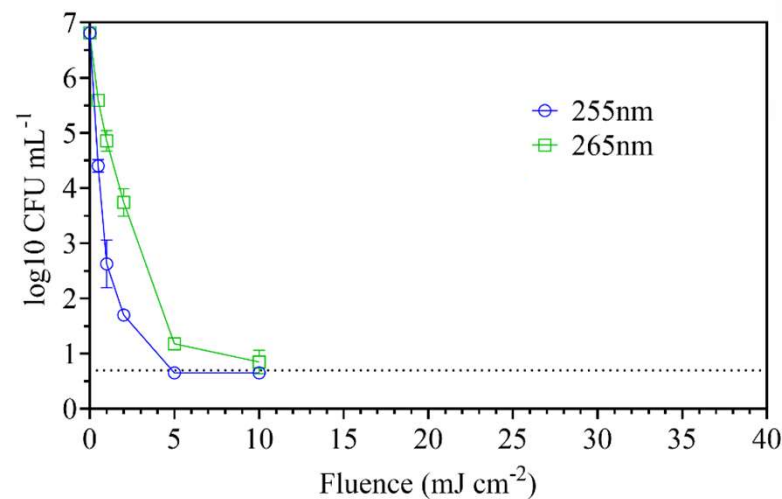


Collimated Beam Results – 265nm

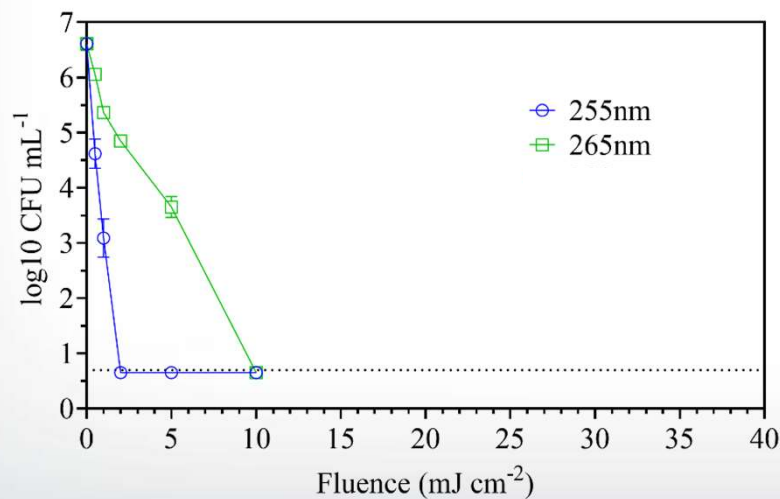
sg1



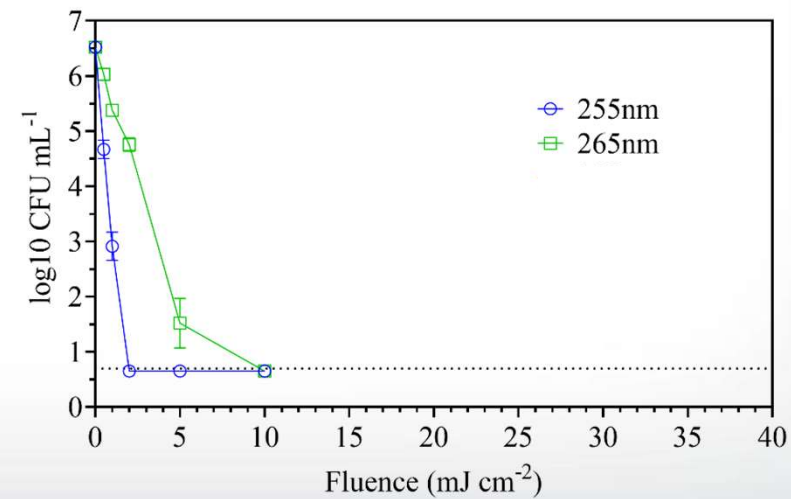
sg1 DW



sg4



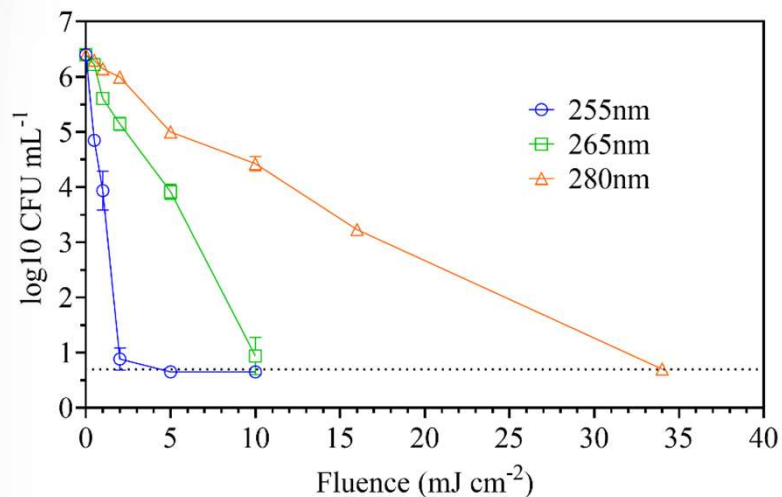
sg6



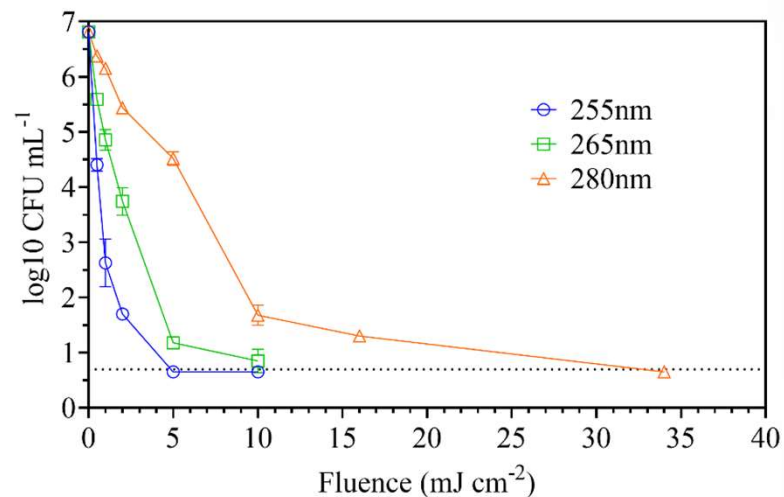


Collimated Beam Results – 280nm

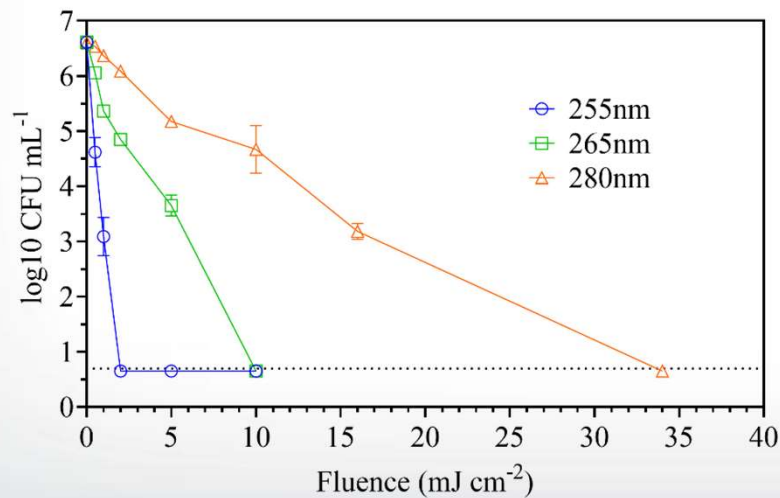
sg1



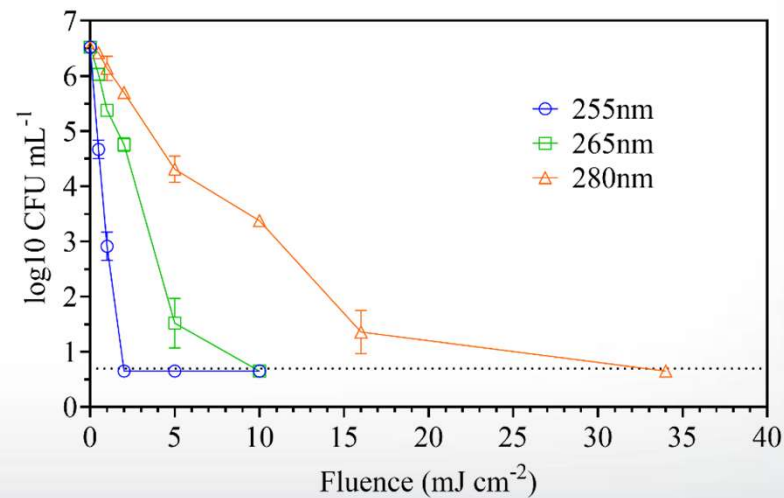
sg1 DW



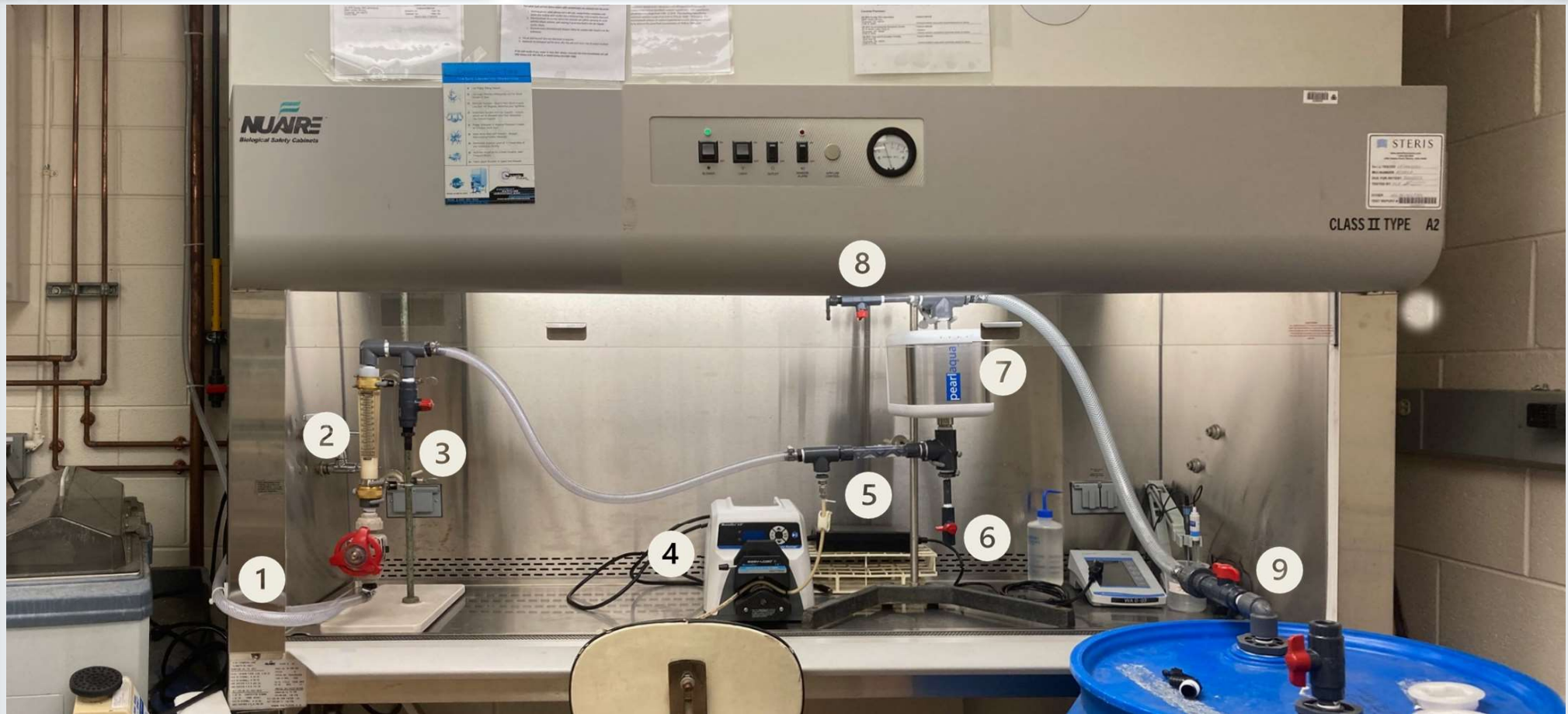
sg4



sg6



Experimental Setup of the UV-C LED POE Device



- | | | |
|---------------------|-----------------------------------|-------------------------------|
| 1. Tap water source | 4. Pump/injection port | 7. Water disinfection system |
| 2. Flow meter | 5. Pipe section with static mixer | 8. Post-treatment sample port |
| 3. Sample port | 6. Pre-treatment sample port | 9. Liquid waste collector |



Experimental Method



4 *L. pneumophila* strains:

- sg1 clinical
- sg1 drinking water
- sg4 clinical
- sg6 clinical

5 runs

- in sequence, turned on tap, Deca unit, pump, release *Legionella* inoculum tube clamp
- wait ~5s, collect 1L at the pre-treatment port (~30s)
- collect 1L at the post-treatment port (~30s)
- engage *Legionella* tube clamp and turn off pump
- flush influent then effluent port (~200mL ea)
- turn off Deca unit, then tap water





POE Device - Results

Table 1-1: Water quality specifications for optimal disinfection performance

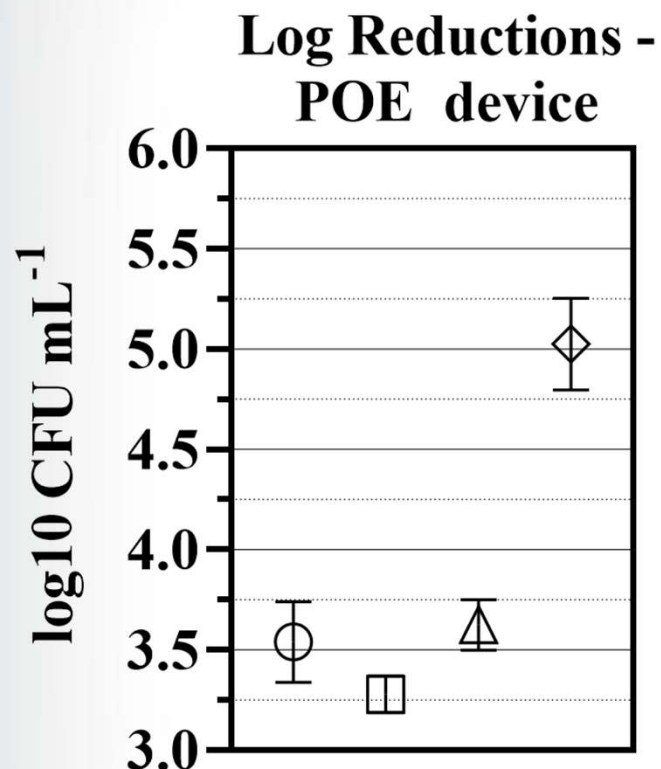
Parameter	Description	Limits	Unit
UV-Transmittance* (UV-T)	Measure of how well UV light can travel through a fluid. Defined as the ratio of UV light intensity after passing through a liquid sample to the UV light intensity at the light source.	≥ 90	%
Particulate Size**	Dirt, dust, rust, sediment, and other solid particles	≤ 10	micron (μm)
Hardness***	Lime scale	7/120	gpg/ppm (mg/L)
Iron***	Rust stains	0.3	ppm (mg/L)

Table. Influent water quality parameters for each experimental run

Parameter (units)	<i>Legionella pneumophila</i> strain used			
	sg1	sg1 DW	sg4	sg6
pH	8.60 ± 0.05	8.70 ± 0.04	9.0 ± 0.09	9.1 ± 0.01
temperature ($^{\circ}\text{C}$)	9.9 ± 0.1	15.1 ± 0.1	19.9 ± 0.1	17.1 ± 0.1
Hardness (mg/L CaCO_3)	130 ± 14	130 ± 14	120 ± 0	140 ± 0
Turbidity	0.25 ± 0.10	0.33 ± 0.00	0.64 ± 0.02	0.22 ± 0.00
% UVT 280nm	112.0 ± 0.1	93.4 ± 0.1	93.9 ± 1.3	111.3 ± 0.8
Free Chlorine (mg/L)	0.93 ± 0.00	0.92 ± 0.00	0.73 ± 0.00	0.92 ± 0.01
Total Chlorine (mg/L)	1.04 ± 0.00	1.04 ± 0.01	0.85 ± 0.01	1.05 ± 0.03
Ferrous Iron	0.00 ± 0.00	0.01 ± 0.00	0.00 ± 0.00	0.00 ± 0.00
Total Iron	0.05 ± 0.00	0.04 ± 0.01	0.00 ± 0.00	0.01 ± 0.00



POE Device - Results



- sg1
- sg1 DW
- △ sg4
- ◇ sg6

Influent	Effluent	Log Reduction
5.9 ± 0.05	2.4 ± 0.47	3.5 ±0.46
6.3 ± 0.05	3.0 ± 0.17	3.3 ±0.21
5.5 ± 0.84	1.9 ± 0.81	3.6 ±0.13
6.1 ± 0.19	1.5 ± 0.95	4.6 ±0.91

Data presented as log₁₀ ± SD CFU mL⁻¹

- sg6 isolate significantly more susceptible to inactivation
- Predicted UV dose at 2-2.5 gpm: ~80-100 mJ/cm²
- Based on collimated beam studies, ~3-4 log reductions occurred ~10-15 mJ/cm²

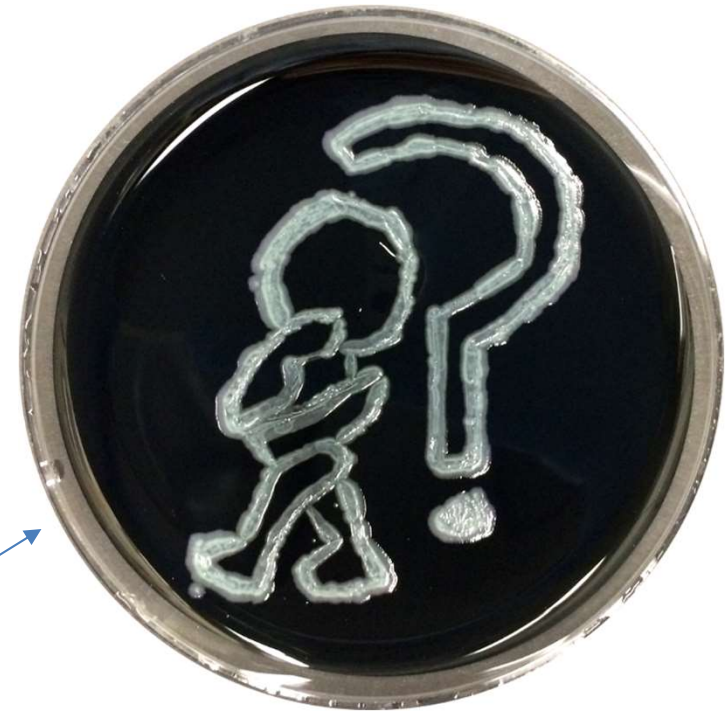
- Serogroup variations to UV inactivation – implications for DW treatment where known environmental strains are linked to clinical cases
- Simulating high contaminant loads, the POE AquiSense Technologies Deca device demonstrated at least 3-log reduction of planktonic *Legionella*
- Further studies needed to:
 - determine synergistic effects of chlorine, monochloramine, etc. and UV inactivation (also considering pH, temperature, other WQ parameters)
 - evaluate inactivation of shed biofilm-associated pathogens (e.g. microbial clumps, other suspended particles)

THANK YOU



Helen Y. Buse
buse.helen@epa.gov

L. pneumophila sgl
drinking water isolate
grown on BCYE agar plates



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