

U.S. Environmental Protection Agency, Office of Research and Development

SAFE AND SUSTAINABLE WATER RESOURCES RESEARCH PROGRAM











The Impact of Algicide Exposure on

Cyanobacterial Responses to Oxidation

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Introduction

- Cyanobacteria are a successful phylum of microorganisms that have flourished for over 1 billion years
- Rising temperatures and nitrogen and phosphorous loadings can cause increased frequency of blooms
- Resource managers work to mitigate these blooms with techniques that include the application of coppercontaining algicides



Introduction

- Bloom material can contaminate drinking water treatment facilities
- Potentially toxic cyanobacteria cells are exposed to permanganate (MnO_4^-) oxidation
- Exposure to copper algicides or MnO₄ imposes pronounced effects on cyanobacterial cells
- What happens when a facility applies MnO₄⁻ to algicidetreated water pumped from a bloom-impacted reservoir?



Materials & methods

- Microcystin-producing Microcystis aeruginosa
- Cyanobacteria at 10⁶ cells/L in dechlorinated tap water
- pH = 6, 7, and 8
- $CuSO_4$ dose = 0, 0.1, 0.5, and 1.0 mg Cu/L
- $KMnO_4$ dose = 0 and 1.0 mg/L
- Analyze for:
 - extracellular microcystin and combined (extra + intracellular) microcystin by ELISA
 - extractive chlorophyll-a, in vivo chlorophyll-a, and in vivo phycocyanin
 - expression of toxin-producing genes



Materials & methods

Nine trials

рН	CuSO ₄	KMnO ₄
6	0.1	
	0.5	0 & 1
	1.0	
7	0.1	
	0.5	0 & 1
	1.0	
8	0.1	
	0.5	0 & 1
	1.0	



Materials & methods

Four jars per trial

Jar	1	2	3	4
CuSO ₄	-	X	-	X
KMnO ₄	-	-	X	X

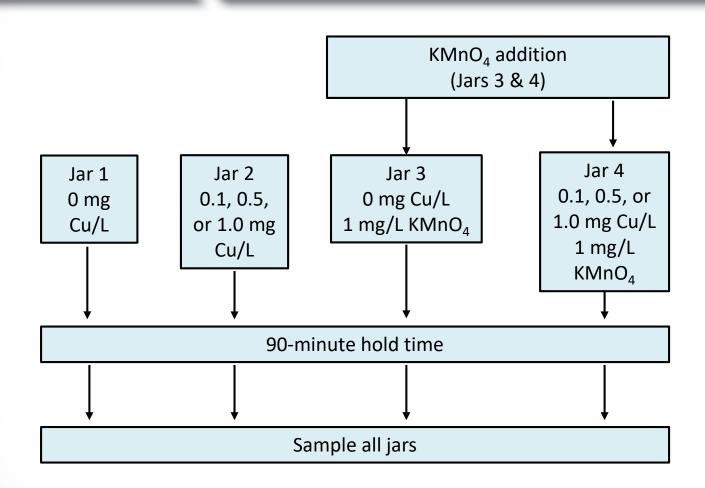


Materials & methods (0 – 24 hours)

pH adjustment and addition of cell suspension (all jars) Sample all jars CuSO₄ addition (Jars 2 & 4) Jar 2 Jar 3 Jar 1 Jar 4 0.1, 0.5 0 mg 0.1, 0.5 0 mg or 1.0 Cu/L Cu/L or 1.0 mg Cu/L mg Cu/L 24-hour hold time Sample all jars

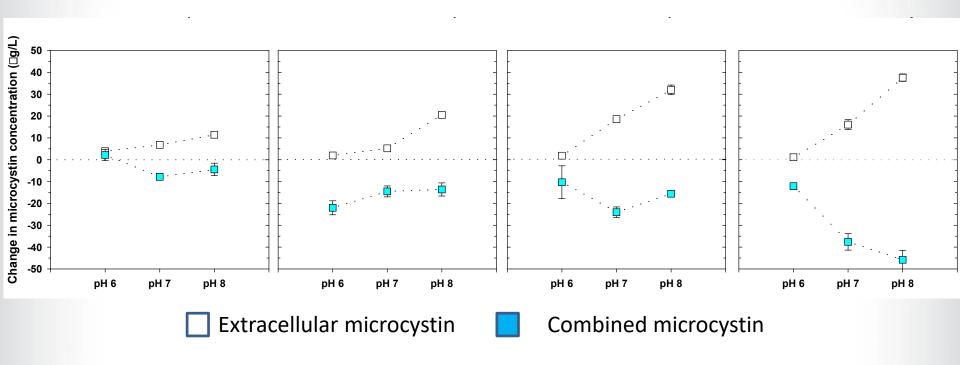


Materials & methods (24 – 25.5 hours)





Extracellular & combined microcystin (0 – 24 hours)

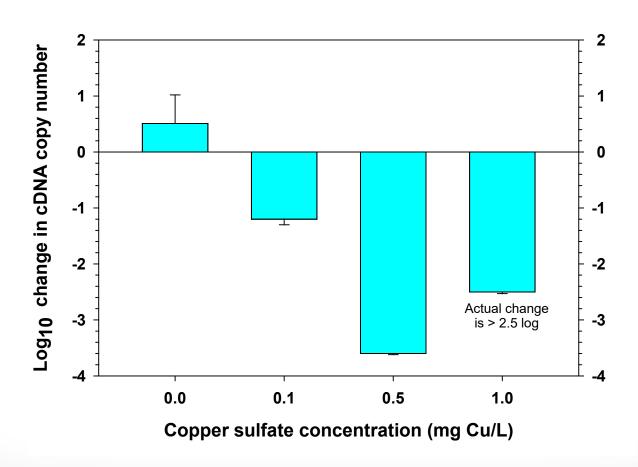


- Extracellular microcystins consistently increased
 - Greatest increases at higher pH
 - At pH 7 & 8, generally greater increases in the presence of copper sulfate
- Combined microcystins consistently decreased
 - Greater decreases in the presence of copper sulfate



Change in expression of gene associated with microcystin production (0 – 24 hours)

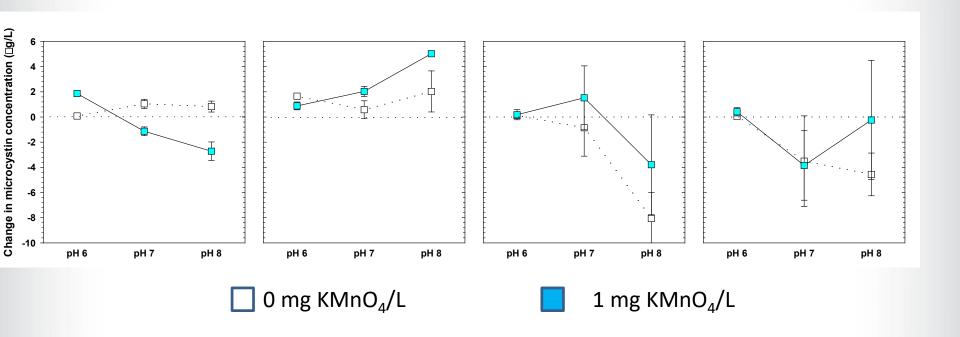
Change in expression of mcyA gene



Consistent order of magnitude decreases in microcystin gene expression in the presence of copper sulfate



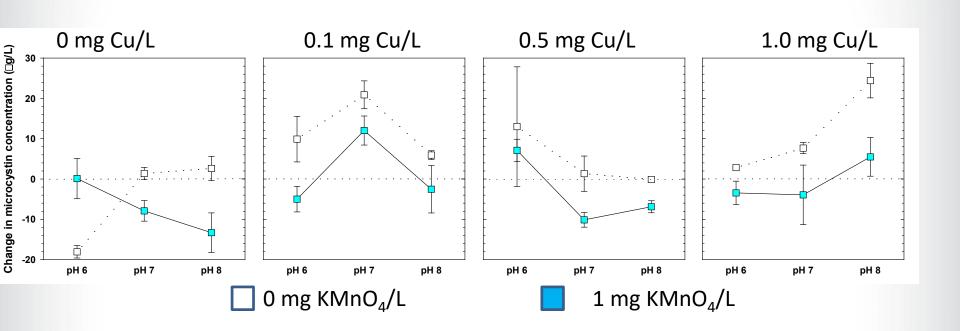
Extracellular microcystin (24 – 25.5 hours)



- In general, smallest changes in extracellular microcystins at pH 6
- At pH 7 & 8, in algicide-exposed suspensions, extracellular microcystins generally increased more or decreased less when treated with 1 mg/L KMnO₄, compared to 0 mg/l KMnO₄



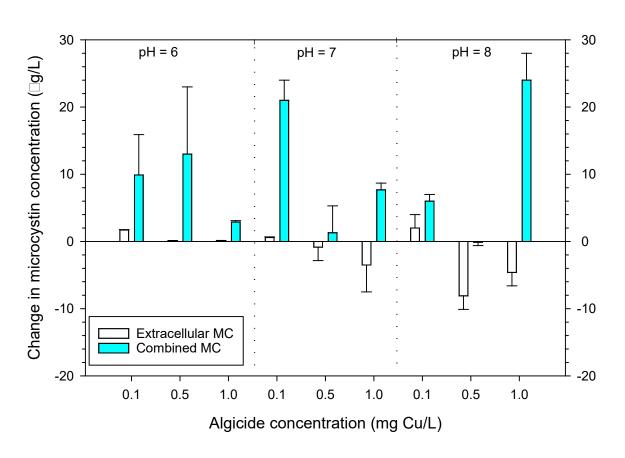
Combined microcystin (24 – 25.5 hours)



- Except at pH 6 & 0 mg Cu/L, combined microcystins in the absence of KMnO₄ remained steady or increased; while combined microcystins in KMnO₄ treated suspensions increased less or actually decreased
- Results imply the possibility that intracellular toxin concentrations can recover following algicide exposure and that the recovery can occur over representative time scales



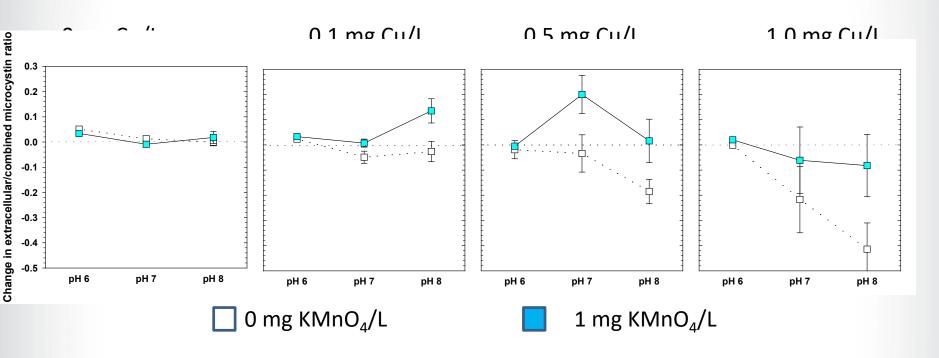
Combined microcystin (24 – 25.5 hours)



 Results imply the possibility that intracellular toxin concentrations can recover following algicide exposure and that the recovery can occur over time scales that are representative of drinking water treatment



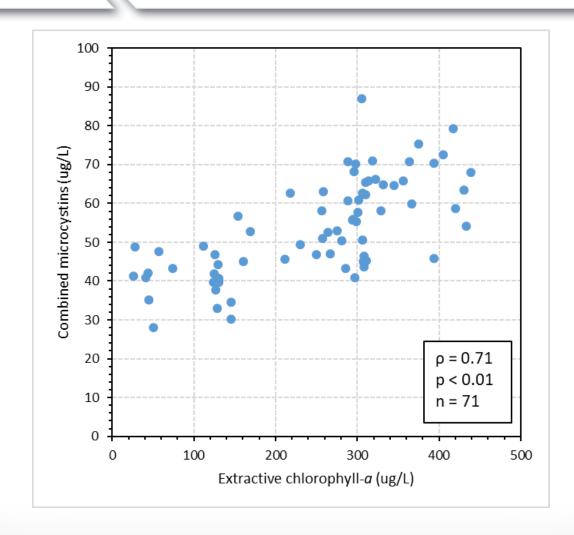
Extracellular/combined microcystin ratio (24 – 25.5 hours)



- For suspensions not exposed to KMnO₄, the extracellular/combined microcystin ratio remained stable or decreased, and the observed decreases were greater at higher pH levels and algicide exposures
- For suspensions exposed to KMnO₄, the extracellular/combined microcystin ratios remained stable, increased, or decreased less than in non-oxidized suspensions



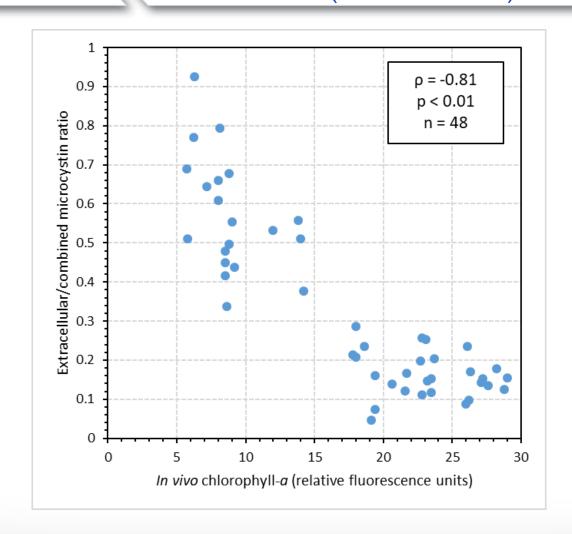
Extractive chlorophyll vs combined microcystins (0-24 hours)



The concentration of extractive chlorophyll, an indicator of intracellular pigments, moves in the same direction as the sum of extracellular and intracellular microcystins



In vivo chlorophyll vs extracellular/combined microcystin ratio (24 – 25.5 hours)



In vivo chlorophyll, an indicator of intracellular pigments, moves in the opposite direction of the extracellular/combined microcystin ratio \rightarrow potentially a rapid indicator of cellular stress



Correlation coefficients, pH 6, 7, & 8 (0 - 24 hours)

	Extracellular microcystin	Combined microcystin	Extracellular/combined microcystin ratio
Extractive chlorophyll-a	-0.55	0.71	-0.59
	(< 0.01)	(< 0.01)	(< 0.01)
	n = 72	n = 71	n = 71
<i>In vivo</i> chlorophyll-a	-0.55	0.30	-0.52
	(< 0.01)	(0.011)	(< 0.01)
	n = 72	n = 71	n = 71
<i>In vivo</i> phycocyanin	-0.14	0.50	-0.17
	(0.23)	(< 0.01)	(0.16)
	n = 72	n = 71	n = 71



Correlation coefficients, pH 7, & 8 (24 – 25.5 hours)

	Extracellular microcystin concentration	Combined microcystin concentration	Extracellular/combined microcystin ratio
Extractive chlorophyll- a	-0.52 (< 0.01) n = 48	0.56 (< 0.01) n = 48	-0.60 (< 0.01) n = 48
<i>In vivo</i> chlorophyll-a	-0.78 (< 0.01) n = 48	0.42 (< 0.01) n = 48	-0.81 (< 0.01) n = 48
<i>In vivo</i> phycocyanin	-0.50 (< 0.01) n = 48	0.60 (< 0.01) n = 48	-0.62 (< 0.01) n = 48



Conclusions

- Over the first 24 hours, algicide exposure was associated with increases in extracellular microcystins, decreases in combined microcystins, and decreases in the expression of a gene associated with microcystin production
- Over the final 1.5 hours, non-oxidized suspensions exhibited stable to decreasing extracellular microcystins and stable to increasing combined microcystins → smaller extracellular/combined microcystin ratio → greater relative fraction of toxins inside the cell
- Over the final 1.5 hours, oxidized suspensions tended to exhibit smaller decreases in the extracellular/combined microcystin ratio



Conclusions

- Difference versus non-oxidized suspensions increased at higher algicide doses and pH
- From a drinking water treatment perspective, a greater fraction of toxins inside cells is a favorable situation → higher probability that plant influent toxin burden can be removed through coagulation/flocculation/sedimentation
- Results indicated that extractive and in vivo indicators pigment concentrations are strongly correlated with changes in toxin concentrations



Disclaimer

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