



## **Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA (anionic and acid forms) IRIS Assessments**

CASRN 335-76-2 (PFDA)  
CASRN 375-95-1 (PFNA)  
CASRN 307-24-4 (PFHxA)  
CASRN 355-46-4 (PFHxS)  
CASRN 375-22-4 (PFBA)

### **Supplemental Information—Appendix A**

*October 2019*  
*Updated February 2020 (in response to public comments)*

This document was posted for public comment on November 8, 2019 ([link to more information](#)), and subsequently updated in response to those comments (updates are outlined in Section 12). It does not represent and should not be construed to represent any Agency determination or policy. This document will serve as Appendix A of the Supplemental Materials for all five IRIS PFAS assessment.

Integrated Risk Information System  
Center for Public Health and Environmental Assessment  
Office of Research and Development  
U.S. Environmental Protection Agency  
Washington, DC

***Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments***

**DISCLAIMER**

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## ABBREVIATIONS

ADME	absorption, distribution, metabolism, and excretion	HED	human equivalent dose
AFFF	aqueous film-forming foam	HERO	Health and Environmental Research Online
AK DEC	Alaska Department of Environmental Conservation	HFPO	hexafluoropropylene oxide
ALT	alanine aminotransferase	hPPAR $\alpha$	humanized peroxisome proliferator-activated receptor alpha
AOP	adverse outcome pathway	HRL	health risk limit
AST	aspartate aminotransferase	i.p.	intraperitoneal
ATSDR	Agency for Toxic Substances and Disease Registry	IARC	International Agency for Research on Cancer
BMDL	benchmark dose lower confidence limit	IPCS	International Programme on Chemical Safety
BMI	body mass index	IRIS	Integrated Risk Information System
BMR	benchmark response	IUR	inhalation unit risk
BW <sup>3/4</sup>	body-weight scaling to the 3/4 power	K	potassium
CAR	constitutive androstane receptor	LD <sub>50</sub>	median lethal dose
CAS	Chemical Abstracts Service	LOAEL	lowest-observed-adverse-effect level
CASRN	Chemical Abstracts Service registry number	LOD	limit of detection
CBI	confidential business information	MAC	maximum acceptable concentration
CERCLA	Comprehensive Environmental Response, Compensation, and Liability Act	MCL	maximum contaminant level
CL <sub>A</sub>	clearance in animals	MDH	Minnesota Department of Health
CL <sub>H</sub>	clearance in humans	MF	modifying factor
CPAD	Chemical and Pollutant Assessment Division	MLR	mixed leukocyte reaction
CPHEA	Center for Public Health and Environmental Assessment	MOA	mode of action
CPN	chronic progressive nephropathy	MPPD	multiple path particle dosimetry
CRD	chemical reporting data	MRL	minimum reporting level
CT DPH	Connecticut Department of Health	Na	sodium
CTL	cytotoxic T lymphocyte	NAFLD	nonalcoholic fatty liver disease
CWA	Clean Water Act	ND	no data
DNA	deoxyribonucleic acid	NF- $\kappa$ B	nuclear factor kappa B pathway
DTH	delayed-type hypersensitivity	NH <sub>4</sub> <sup>+</sup>	ammonium
DWEL	drinking water equivalent level	NHANES	National Health and Nutrition Examination Survey
ECHA	European Chemicals Agency	NH DES	New Hampshire Department of Environmental Services
EFSA	European Food Safety Authority	NJ DEP	New Jersey Department of Environmental Protection
EPA	Environmental Protection Agency	NMD	normalized mean difference
FDA	Food and Drug Agency	NOAEL	no-observed-adverse-effect level
FIFRA	Federal Insecticide, Fungicide, and Rodenticide Act	NPDWR	National Primary Drinking Water Regulation
FOB	functional operational battery	NPL	National Priorities List
FXR	farnesoid X receptor	NR	nuclear receptor
GLP	good laboratory practice	NTP	National Toxicology Program
GRADE	Grading of Recommendations Assessment, Development, and Evaluation	OCSPP	Office of Chemical Safety and Pollution Prevention
HA	health advisory	OECD	Organisation for Economic Co-operation and Development
HAWC	Health Assessment Workspace Collaborative	OLEM	Office of Land and Emergency Management
		OR	odds ratio

*This document is a draft for review purposes only and does not constitute Agency policy.*

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ORD	Office of Research and Development	RfC	inhalation reference concentration
OSF	oral slope factor	RfD	oral reference dose
OW	Office of Water	ROBINS-I	Risk of Bias in Nonrandomized Studies of Interventions
PAC	protective action criteria	ROS	reactive oxygen species
PBPK	physiologically based pharmacokinetic	RXR	retinoid X receptor
PBTK	physiologically based toxicokinetic	SD	standard deviation
PCL	protective concentration level	SDWA	Safe Drinking Water Act
PECO	populations, exposures, comparators, and outcomes	$t_{1/2A}$	elimination half-life in animals
PFAS	per- and polyfluoroalkyl substances	$t_{1/2H}$	elimination half-life in humans
PFBA	perfluorobutanoic acid	TCEQ	Texas Commission on Environmental Quality
PFBS	perfluorobutane sulfonate	TD	toxicodynamic
PFCA	perfluoroalkyl carboxylic acid	TDI	tolerable daily intake
PFDA	perfluorodecanoic acid	TEEL	temporary emergency exposure limit
PFHxA	perfluorohexanoic acid	TNF $\alpha$	tumor necrosis factor alpha
PFHxS	perfluorohexanesulfonate	TRI	Toxics Release Inventory
PFNA	perfluorononanoic acid	TSCA	Toxic Substances Control Act
PFOA	perfluorooctanoic acid	TSCATS	Toxic Substances Control Act Test Submissions
PFOS	perfluorooctane sulfonate	UCMR	Unregulated Contaminant Monitoring Rule
PFSA	perfluoroalkane sulfonic acid	UF	uncertainty factor
PI3K-Akt	phosphatidylinositol-3-kinase-serine/threonine kinase Akt	1 UF <sub>A</sub>	animal-to-human uncertainty factor
PK	pharmacokinetic	2 UF <sub>C</sub>	composite uncertainty factor
POD	point of departure	3 UF <sub>D</sub>	database deficiencies uncertainty factor
PPAR $\alpha$	peroxisome proliferator-activated receptor alpha	4 UF <sub>H</sub>	human variation uncertainty factor
PPRTV	Provisional Peer-Reviewed Toxicity Value	5 UF <sub>L</sub>	LOAEL-to-NOAEL uncertainty factor
PR	preliminary review	6 UF <sub>S</sub>	subchronic-to-chronic uncertainty factor
pt.	point	7	
PVDF	polyvinylidene fluoride	$V_d$	volume of distribution
PWS	public water system	WHO	World Health Organization
PXR	pregnane X receptor	wt.	weight
RCRA	Resource Conservation and Recovery Act	XME	xenobiotic metabolizing enzymes

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## *Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments*

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<sup>1</sup>NCEA was reorganized (largely into CPHEA) during the 2019 Office of Research and Development reorganization.

# 1. INTRODUCTION

1 Per- and polyfluoroalkyl substances (PFAS) are a large class of synthetic (man-made)  
2 chemicals widely used in consumer products and industrial processes. The basic structure of PFAS  
3 consists of a carbon chain surrounded by fluorine atoms, with different chemicals possessing  
4 different end groups (see examples in Section 2.1.1); thousands of distinct PFAS exist in commerce.  
5 To help address this complex issue, the Environmental Protection Agency (EPA) is taking a  
6 proactive approach. Specifically, the development of human health toxicity assessments for  
7 exposure to individual PFAS represents only one component of the broader PFAS action plan  
8 underway at the EPA (<https://www.epa.gov/pfas/epas-pfas-action-plan>). The five toxicity  
9 assessments being developed according to the scope and methods outlined in this protocol build  
10 upon several other PFAS assessments that have already been developed (see Section 2.1.7).

11 This protocol document presents the methods for conducting the systematic reviews and  
12 dose-response analyses for assessments of perfluorodecanoic acid (PFDA), perfluorononanoic acid  
13 (PFNA), perfluorohexanoic acid (PFHxA), perfluorohexanesulfonate (PFHxS), and  
14 perfluorobutanoic acid (PFBA), and their related salts (see Figure 2-1). This includes a summary of  
15 why these specific PFAS were prioritized for evaluation, description of the objectives and specific  
16 aims of the assessments, draft populations, exposures, comparators, and outcomes (PECO) criteria,  
17 and identification of key areas of scientific complexity. This assessment protocol will be posted on  
18 the Integrated Risk Information System (IRIS) website (<https://cfpub.epa.gov/ncea/iris2/atoz.cfm>)  
19 for a 45-day comment period. The protocol will also be published in the Zenodo data repository  
20 (<https://zenodo.org/>). Public input received on the protocol is considered during preparation of  
21 the draft assessments, and any adjustments made to the protocol will be reflected in an updated  
22 version released in conjunction with the draft assessments. The literature search results for these  
23 five PFAS will also be posted to the Health and Environmental Research Online (HERO) database<sup>2</sup>  
24 upon public release of the protocol (the literature search results will be regularly updated during  
25 draft development and the subsequent stages of assessment review).

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<sup>2</sup>PFBA: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2632](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2632)  
PFHxA: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2628](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2628)  
PFHxS: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2630](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2630)  
PFNA: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2633](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2633)  
PFDA: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2614](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2614).

## 2. SCOPING AND PROBLEM FORMULATION SUMMARY

### 2.1. SUMMARY OF BACKGROUND INFORMATION

1 Section 2.1 provides a summary of background information for contextual purposes only.  
2 These brief overviews emphasize reviews and other summary information (e.g., in public  
3 databases) and are not intended to be comprehensive descriptions of the available information. In  
4 addition, the information in this section (developed in 2019) is not updated and thus may not  
5 represent the current state of the science at the time of review. The reader is encouraged to refer to  
6 the source materials and other updated information for current PFAS-specific details. The  
7 information in this section is not recommended for use in decision making.

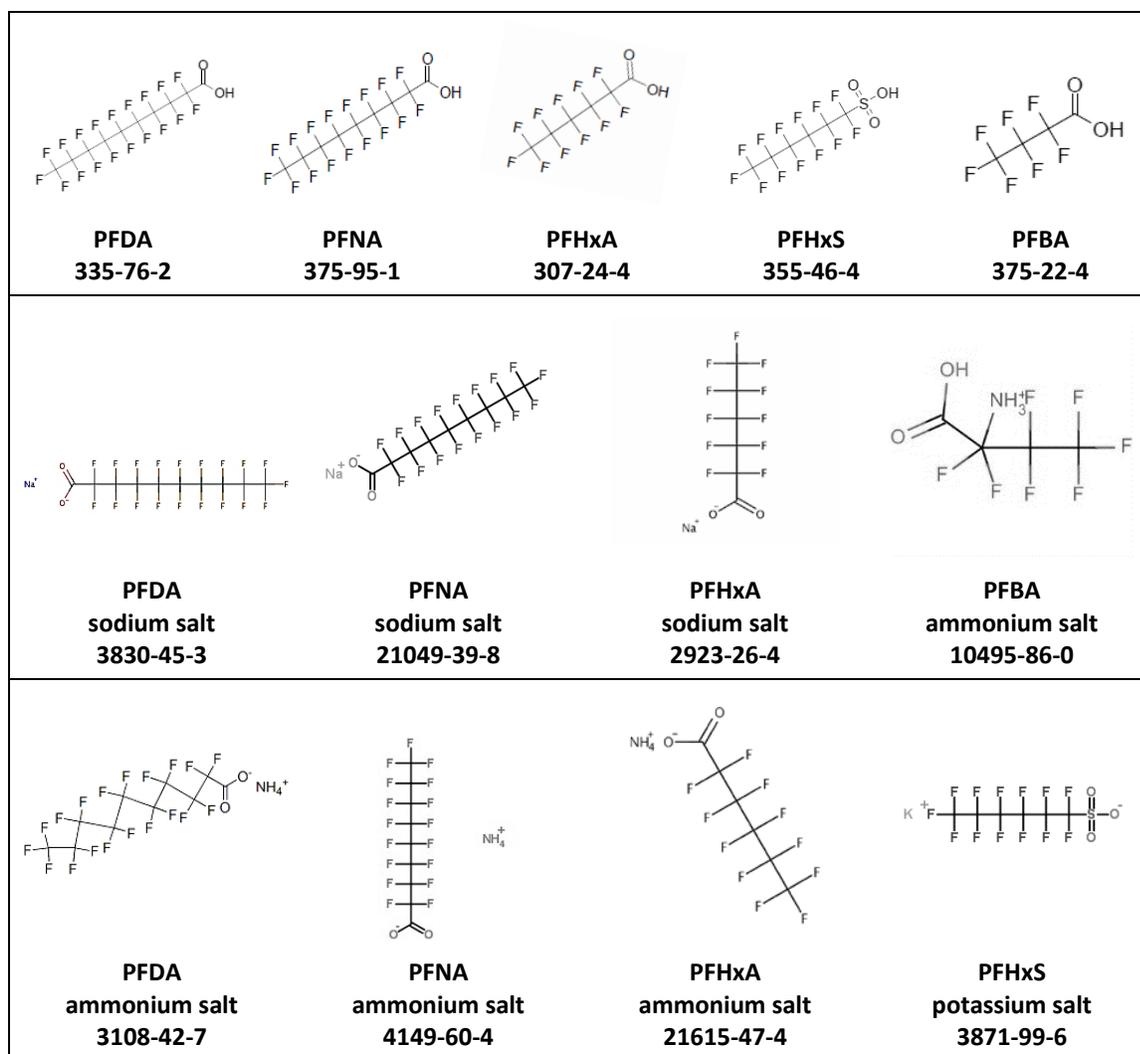
#### 2.1.1. Chemical and Physical Properties

8 Perfluorodecanoic acid (PFDA; CASRN 335-76-2), perfluorononanoic acid (PFNA;  
9 CASRN 375-95-1), perfluorohexanoic acid (PFHxA, CASRN 307-24-4), perfluorohexanesulfonic acid  
10 (PFHxS, CASRN 355-46-4), and perfluorobutanoic acid (PFBA, CASRN 375-22-4), and their related  
11 salts, are all PFAS. Section 2.2 (“Scoping Summary”) outlines the rationale for why these PFAS were  
12 prioritized for assessment. No single, consensus definition of PFAS exists. [Buck et al. \(2011\)](#)  
13 defined PFAS as fluorinated substances that “contain 1 or more C atoms on which all the H  
14 substituents (present in the nonfluorinated analogues from which they are notionally derived) have  
15 been replaced by F atoms, in such a manner that they contain the perfluoroalkyl moiety (C<sub>n</sub>F<sub>2n+1</sub>).”  
16 The definition in the EPA Chemistry Dashboard, which (as of late 2019) yields over 6,600 PFAS  
17 structures ([https://comptox.epa.gov/dashboard/chemical\\_lists/PFASTRUCT](https://comptox.epa.gov/dashboard/chemical_lists/PFASTRUCT)), includes all  
18 substances for which “the structure contains the substructure RCF<sub>2</sub>CFR’R” (R cannot be H); the  
19 Dashboard defines this substructure as “general enough to encompass the largest set of structures  
20 having sufficient levels of fluorination to potentially impart PFAS-type properties.” Regardless of  
21 the definition used, the PFAS being assessed in association with this protocol are members of a  
22 subset of PFAS called perfluoroalkyl acids (PFAAs; PFOA and PFOS are also members), which  
23 consist of a carbon backbone (typically 4–14 C atoms) that is fully fluorinated and bonded to a  
24 charged functional group [e.g., carboxylic acid, sulfonic acid, or phosphonic acid; [Lau et al. \(2007\)](#)].  
25 More specifically, PFDA, PFNA, PFHxA, and PFBA are classified as perfluoroalkyl carboxylic acids  
26 (PFCAs), and PFHxS is a perfluoroalkane sulfonic acid [PFSA; [OECD \(2015\)](#)]. PFCAs containing  
27 seven or more perfluorinated carbon groups and PFSAs containing six or more perfluorinated  
28 carbon units are considered long-chain PFAS ([ATSDR, 2018](#); [OECD, 2015](#); [Buck et al., 2011](#)). Thus,  
29 PFDA, PFNA, and PFHxS are considered long-chain, and PFHxA and PFBA are short-chain. To

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1 simplify the terminology used throughout this protocol and the subsequent assessments, PFBA,  
2 PFHxA, PFHxS, PFNA, and PFDA (and their salts) are referred to using the broad and more  
3 recognizable term, PFAS, rather than using the more specific terms PFAAs, PFSAs, or PFCAs. The  
4 chemical structures of PFDA, PFNA, PFHxA, PFHxS, and PFBA, and their related salts are shown in  
5 Figure 2-1 (along with their CASRNs), and estimated or experimental values for their  
6 physiochemical properties are provided in Table 2-1. Importantly, these values are intended for  
7 general context and may no longer be accurate or current at the time of review and should not be  
8 used for any purpose other than conveying generalities around physicochemical properties. For  
9 example, even though the logP may be difficult to predict, the possibility that PFAS exist in the  
10 ionized and nonionized ([Beesoon et al., 2012](#)) form cannot be ignored and understanding the PFAS  
11 dissociation and partitioning constants are important for understanding how PFAS interact with  
12 the environment.

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**Figure 2-1. Chemical structures of the per- and polyfluoroalkyl substances (PFAS) being assessed.**

**Table 2-1. Predicted or experimental physiochemical property values for the per- and polyfluoroalkyl substances (PFAS) being assessed (see <https://comptox.epa.gov/dashboard/>)**

Property (unit)	PFDA + salts			PFNA + salts			PFHxA	PFHxS + salts		PFBA + salts	
	PFDA <sup>a</sup>	NH <sub>4</sub> <sup>+</sup> salt <sup>b</sup>	Na salt <sup>c</sup>	PFNA <sup>d</sup>	NH <sub>4</sub> <sup>+</sup> salt <sup>e</sup>	Na salt <sup>f</sup>	PFHxA <sup>g</sup>	PFHxS <sup>h</sup>	K salt <sup>i</sup>	PFBA <sup>j</sup>	NH <sub>4</sub> <sup>+</sup> salt <sup>k</sup>
Molecular wt. (g/mol)	514	531	536	464	481	486	314	400	438	214	ND
Melting pt. (°C)	79.5	165*	71.4*	66.5	165*	ND	14.0	190	273	-17.5	ND
Boiling pt. (°C)	218	205*	205*	218	190*	196	157	345	236*	121	ND

*This document is a draft for review purposes only and does not constitute Agency policy.*

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Property (unit)	PFDA + salts			PFNA + salts			PFHxA	PFHxS + salts			PFBA + salts	
	PFDA <sup>a</sup>	NH <sub>4</sub> <sup>+</sup> salt <sup>b</sup>	Na salt <sup>c</sup>	PFNA <sup>d</sup>	NH <sub>4</sub> <sup>+</sup> salt <sup>e</sup>	Na salt <sup>f</sup>	PFHxA <sup>g</sup>	PFHxS <sup>h</sup>	K salt <sup>i</sup>	PFBA <sup>j</sup>	NH <sub>4</sub> <sup>+</sup> salt <sup>k</sup>	
Density (g/cm <sup>3</sup> )	1.79*	ND	ND	1.78*	ND	ND	1.69*	1.84*	1.84*	1.65	ND	
Vapor pressure (mm Hg)	3.89 × 10 <sup>-2*</sup>	3.51 × 10 <sup>-2*</sup>	3.51 × 10 <sup>-2*</sup>	7.06 × 10 <sup>-2*</sup>	7.38 × 10 <sup>-2*</sup>	1.71 × 10 <sup>-1</sup>	9.08 × 10 <sup>-1</sup>	9.95 × 10 <sup>-3*</sup>	9.95 × 10 <sup>-3*</sup>	6.37	ND	
Henry's law constant (atm·m <sup>3</sup> /mole)	3.55 × 10 <sup>-10*</sup>	3.55 × 10 <sup>-10*</sup>	3.55 × 10 <sup>-10*</sup>	1.64 × 10 <sup>-10*</sup>	1.64 × 10 <sup>-10*</sup>	ND	2.51 × 10 <sup>-10*</sup>	1.96 × 10 <sup>-10*</sup>	1.96 × 10 <sup>-10*</sup>	4.99 × 10 <sup>-5*</sup>	ND	
Water solubility (mol/L)	8.3 × 10 <sup>-6*</sup>	8.41 × 10 <sup>-1*</sup>	8.41 × 10 <sup>-1*</sup>	1.38 × 10 <sup>-5*</sup>	8.45 × 10 <sup>-1*</sup>	ND	9.39 × 10 <sup>-5</sup>	6.08 × 10 <sup>-4</sup>	2.96 × 10 <sup>-4*</sup>	2.09 × 10 <sup>-3</sup>	ND	
pKa	-0.17*	ND	ND	-0.17*	ND	ND	-0.16	0.14*	ND	0.08*	ND	
LogP	7.37*	4.36*	4.36*	6.59*	3.12*	ND	2.51	2.20	2.97*	1.43	ND	
Soil adsorption coefficient (L/kg)	397*	397*	397*	2,830*	2,830*	ND	1,070*	2,300*	2,300*	47.9*	ND	
Bioconcentration factor	789*	29.8*	29.8*	752*	4.95*	ND	41*	118*	5.94*	7.61*	ND	

K = potassium; ND = no data; NH<sub>4</sub><sup>+</sup> = ammonium; Na = sodium; pt. = point; wt. = weight.

<sup>a</sup>CASRN 335-76-2. [U.S. EPA \(2018a\)](https://comptox.epa.gov/dashboard/) Chemistry Dashboard (<https://comptox.epa.gov/dashboard/>; search = PFDA) for all values except pKa ([ATSDR, 2018](#)). Values are median experimental values (when available), or median or average predicted values.

<sup>b</sup>CASRN 3108-42-7. Predicted average values from the [U.S. EPA \(2018a\)](#) Chemistry Dashboard (search = 3108-42-7)

<sup>c</sup>CASRN 3830-45-3. Predicted average values from the [U.S. EPA \(2018a\)](#) Chemistry Dashboard (search = 3830-45-3)

<sup>d</sup>CASRN 375-95-1. [U.S. EPA \(2018a\)](#) Chemistry Dashboard (search = PFNA) for all values except pKa ([NLM, 2013](#)). Values are median experimental values (when available), or median or average predicted values.

<sup>e</sup>CASRN 4149-60-4. Predicted average values from the [U.S. EPA \(2018a\)](#) Chemistry Dashboard (search = 4149-60-4)

<sup>f</sup>CASRN 21049-39-8. ChemNet website:

<http://www.chemnet.com/cas/en/21049-39-8/sodium-heptadecafluorononanoate.html> for all values.

<sup>g</sup>CASRN: 307-24-4. [U.S. EPA \(2018a\)](#) Chemistry Dashboard (search = 307-24-4) for all values except pKa ([NLM, 2016](#)). Values are median or average experimental values (when available) or predicted values.

<sup>h</sup>CASRN 355-46-4. [U.S. EPA \(2018a\)](#) Chemistry Dashboard (search = 355-46-4) for all values except pKa ([NLM, 2017](#)). Values are median or average experimental values (when available) or predicted values.

<sup>i</sup>CASRN 3871-99-6. [U.S. EPA \(2018a\)](#) Chemistry Dashboard (search = 3871-99-6) for all values. Values are median or average experimental values (when available) or predicted values.

<sup>j</sup>CASRN 375-22-4. [U.S. EPA \(2018a\)](#) Chemistry Dashboard (search = 375-22-4)

<https://comptox.epa.gov/dashboard/dsstoxdb/results?utf8=%E2%9C%93&search=375-22-4> for all values except pKa ([ATSDR, 2018](#)). Values are median or average experimental values (when available) or predicted values.

<sup>k</sup>CASRN 10495-86-0.

\*Predicted value.

### 2.1.2. Sources, Production, and Use

- 1 PFAS are synthetic (man-made) compounds that have been used since the 1940s in
- 2 consumer products and industrial applications because of their resistance to heat, oil, stains,
- 3 grease, and water. They have been used in stain-resistant fabrics for clothing, carpets, and

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1 furniture; nonstick cookware; food packaging (e.g., popcorn bags, and fast-food containers); and  
2 personal care products [e.g., dental floss, cosmetics, and sunscreen; [ATSDR \(2018\)](#)]. Some PFAS  
3 have also been used in firefighting foam and as industrial surfactants, emulsifiers, wetting agents,  
4 additives, and coatings, and in the aerospace, automotive, building, and construction industries to  
5 help reduce friction ([ATSDR, 2018](#)). Because of the widespread use of PFAS and their persistence in  
6 the environment, most people in the United States have been exposed to them (see  
7 <https://www.epa.gov/pfas/epas-pfas-action-plan> for additional details). Although not exhaustive,  
8 the bulleted list below provides some examples of how the five PFAS of interest have been used:

- 9 • **PFDA** has been used in stain and grease-proof coatings on food packaging, furniture,  
10 upholstery, and carpet ([Harbison et al., 2015](#)), and as a lubricant, wetting agent, plasticizer,  
11 and corrosion inhibitor ([Kemi, 2015](#)).
- 12 • **PFNA** has been used as a processing aid in the production of fluoropolymers, primarily  
13 polyvinylidene fluoride (PVDF), which is a plastic designed to be temperature resistant and  
14 chemically nonreactive ([NJDWQI, 2017](#); [Prevedouros et al., 2006](#)). It has also been used in  
15 aqueous film-forming foam (AFFF) for fire suppression ([Laitinen et al., 2014](#)).
- 16 • **PFHxA** is not currently a commercial product; it is a breakdown product of “stain- and  
17 grease-proof coatings on food packaging and household products” ([NTP, 2018b](#)).
- 18 • **PFHxS** has been used as a surfactant to make fluoropolymers, and in water- and  
19 stain-protective coatings for carpets, paper, and textiles ([NTP, 2018a](#)). It may also be  
20 present in certain industrial and consumer products, such as “food-contact papers,  
21 water-proofing agents, cleaning and polishing products either for intentional uses (as  
22 surfactants or surface protection agents) or as unintentional impurities from industrial  
23 production processes” ([Norwegian Environment Agency, 2018](#)). It has also been used in  
24 AFFF for fire suppression ([Laitinen et al., 2014](#)).
- 25 • **PFBA** is a breakdown product of other PFAS that are used in stain-resistant fabrics, paper  
26 food packaging, and carpets; it was also used for manufacturing photographic film ([MDH,  
27 2009](#)).

28 The U.S. Environmental Protection Agency (EPA) has been working with companies in the  
29 fluorochemical industry since the early 2000s to phase out the production and use of long-chain  
30 PFAS ([https://www.epa.gov/assessing-and-managing-  
31 chemicals-under-tsca/risk-management-and-polyfluoroalkyl-substances-PFAS](https://www.epa.gov/assessing-and-managing-chemicals-under-tsca/risk-management-and-polyfluoroalkyl-substances-PFAS)). Although  
32 production of long-chain PFAS in Western Europe and Japan has declined ([OECD, 2015](#)), their  
33 production in emerging economies in Asia (China and India) has increased ([OECD, 2015](#)). Given the  
34 past production and use of these PFAS in some regions, and the increased production and use in  
35 others, PFAS have been and are being released to the environment through various waste streams  
36 ([NLM, 2016, 2013](#)). Also, because precursor products (e.g., fluorotelomer alcohols) or products  
37 containing PFAS are still in use, they continue to be a source of environmental PFAS contamination

1 through their disposal and subsequent breakdown into PFAS in the environment ([Kim and Kannan, 2007](#)).

3 Chemical reporting data (CRD) on production volumes are not available in EPA's ChemView  
4 ([U.S. EPA, 2019](#)) for PFDA, PFNA, PFHxA, PFHxS, PFBA, or their salts. Also, because there are no  
5 requirements to report releases to the environment from facilities manufacturing, processing, or  
6 otherwise using PFAS, quantitative information is not available in EPA's Toxics Release Inventory  
7 [TRI; [ATSDR \(2018\)](#); [U.S. EPA \(2019\)](#)].

### 2.1.3. Environmental Fate and Transport

8 PFAS are very stable and persistent in the environment ([ATSDR, 2018](#)), and many are found  
9 worldwide in the air, water, and soil and in the tissues of plants, animals, and humans  
10 ([https://www.epa.gov/assessing-and-](https://www.epa.gov/assessing-and-managing-chemicals-under-tsca/risk-management-and-polyfluoroalkyl-substances-pfas)  
11 [managing-chemicals-under-tsca/risk-management-and-polyfluoroalkyl-substances-pfas](https://www.epa.gov/assessing-and-managing-chemicals-under-tsca/risk-management-and-polyfluoroalkyl-substances-pfas)). They  
12 have been detected at a variety of sites, including private and federal facilities, and have been  
13 associated with various sources, including AFFF, chrome-plating facilities, PFAS manufacturers, and  
14 industries that use PFAS [e.g., textiles; [ATSDR \(2018\)](#)]. The environmental fate and transport of  
15 PFAS potentially includes releases to air to soil and surficial water bodies which can then lead to  
16 migration to subsurface soils and ground water contamination [[Guelfo et al. \(2018\)](#);  
17 <https://www.atsdr.cdc.gov/pfas/index.html>].

18 Some PFAS (PFNA, PFHxA, PFHxS) released to air are expected to exist solely in the vapor  
19 phase given their vapor pressures ([NLM, 2017, 2016, 2013](#); [Kim and Kannan, 2007](#)), although  
20 particle-bound concentrations have also been measured for PFNA and PFDA ([Kim and Kannan,](#)  
21 [2007](#)). Although vapor-phase PFAS are not susceptible to direct photolysis by sunlight ([NLM, 2017,](#)  
22 [2016, 2013](#)) and are generally resistant to photo-oxidation ([ATSDR, 2018](#)), they can be degraded by  
23 reaction with photochemically produced hydroxyl radicals ([NLM, 2017, 2016, 2013](#)). The  
24 atmospheric half-life for these reactions is estimated to be 31 days for PFNA and PFHxA, and  
25 115 days for PFHxS ([NLM, 2017, 2016, 2013](#)). Long-range atmospheric transport of PFAS is  
26 possible, as indicated by the detection of PFHxS in remote arctic and marine air samples ([NLM,](#)  
27 [2017](#)). Wet and dry deposition are potential removal processes for particle-bound PFAS in air  
28 [e.g., to surface water or soil; [ATSDR \(2018\)](#)]. Standardized analytical methods for measuring these  
29 five PFAS in ambient air is an area of ongoing research.

30 In soil, the mobility of PFAS will vary depending on their soil adsorption coefficients (see  
31 Table 2-1), with PFNA predicted to be the least mobile and PFBA the most mobile of the five PFAS  
32 addressed here. Volatilization of PFNA, PFHxA, and PFHxS from moist soil is not expected to be an  
33 important transport process ([NLM, 2017, 2016, 2013](#)). Uptake of soil PFAS to plants can occur  
34 ([ATSDR, 2018](#)). [Yoo et al. \(2011\)](#) estimated grass-soil accumulation factors (grass concentration  
35 divided by soil concentration) of 3.4, 0.12, and 0.10 for PFHxA, PFNA, and PFDA, respectively, based  
36 on samples collected from a site with bio-solids-amended soil. [Zhao et al. \(2016\)](#) observed that

1 shorter chain PFAS like PFBA were transported more readily from the roots to the shoots of wheat  
2 plants than longer chain PFAS.

3 PFNA, PFHxA, and PFHxS are expected to adsorb to suspended solids and sediments in  
4 water (NLM, 2017, 2016, 2013). The potential for PFAS to bioaccumulate in aquatic organisms can  
5 be generally assessed using the predicted bioconcentration factors, with the predicted potential for  
6 PFDA and PFNA to bioaccumulate being high compared with PFHxA, PFHxS, and PFBA (see  
7 Table 2-1). Note, however, that these predicted values may vary over a wide range depending on  
8 the variables (i.e., species, habitat, etc.). As described in Section 2.2, standardized analytical  
9 methods for measuring these five PFAS in drinking water exist (for four of the five PFAS to be  
10 assessed) or are under development (i.e., for PFBA). Standardized nondrinking water methods are  
11 currently under development.

#### 2.1.4. Environmental Concentrations

12 PFDA, PFNA, PFHxA, PFHxS, and PFBA have not been evaluated under the National Air  
13 Toxics Assessment program (<https://www.epa.gov/national-air-toxics-assessment>). However,  
14 PFDA, PFNA, and PFHxS were measured at concentrations ranging from below the limit of detection  
15 (LOD) to 1.56 pg/m<sup>3</sup> in the vapor and particle phases of air samples collected from an urban area of  
16 Albany, NY in 2006 (Kim and Kannan, 2007). PFAS have also been measured in indoor air and dust,  
17 and they may be associated with the indoor use of consumer products such as PFAS-treated carpets  
18 or other textiles (ATSDR, 2018). For example, Kato et al. (2009) analyzed dust samples collected  
19 from 39 homes in the United States, United Kingdom, Germany, and Australia for PFAS, including  
20 PFDA, PFNA, PFHxA, and PFHxS. These PFAS were detected in 38.5, 25.6, 46.2, and 79.5% of the  
21 samples, respectively. Likewise, Strynar and Lindstrom (2008) analyzed dust samples from  
22 110 homes and 10 day care centers in North Carolina and Ohio, and detected PFDA, PFNA, and  
23 PFHxA in 30.4, 42.9, and 92.9% of the samples, respectively. Indoor air samples ( $n = 4$ ) from a town  
24 in Norway had mean concentrations of 3.4 pg/m<sup>3</sup> for PFDA, 2.7 pg/m<sup>3</sup> for PFNA, and <4.1 pg/m<sup>3</sup> for  
25 PFHxS (Barber et al., 2007).

26 The levels of PFAS in soil and sediment surrounding perfluorochemical industrial facilities  
27 have been measured at concentrations ranging from less than the LOD to 124 ng/g for PFBA and  
28 less than the LOD to 3,470 ng/g for PFHxS (ATSDR, 2018). PFDA, PFNA, PFHxA, PFHxS, and PFBA  
29 were also detected at an Australian training ground where AFFFs had been used (Badel et al.,  
30 2015). PFDA, PFNA, PFHxA, PFHxS, and PFBA were detected at 10 U.S. military sites in 67.0, 71.4,  
31 70.3, 76.9, and 38.5% of the surface soil samples, respectively, and 48.5, 12.1, 63.6, 72.7, and 24.2%  
32 of the sediment samples, respectively (ATSDR, 2018). Table 2-2 shows the concentrations of these  
33 PFAS in soil and sediment at these military sites.

34 EPA conducted monitoring for several PFAS in drinking water as part of the third  
35 Unregulated Contaminant Monitoring Rule [UCMR; U.S. EPA (2016e)]. Under the UCMR, all public  
36 water systems (PWSs) serving more than 10,000 people and a representative sample of 800 PWSs  
37 serving 10,000 or fewer people were monitored for 30 unregulated contaminants between

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1 January 2013 and December 2015. PFNA and PFHxS were among the 30 contaminants monitored.  
2 PFNA was detected above the minimum reporting level (MRL) of 0.02 µg/L in 14 of the 4,920 PWSs  
3 tested and in 19 of the 36,972 samples collected. PFNA was also detected above the MRL  
4 (0.096 µg/L) in groundwater near an industrial site in New Jersey ([Post et al., 2013](#)). PFHxS was  
5 detected above the MRL of 0.03 µg/L in 55 of the 4,920 PWSs tested and in 207 of the  
6 36,971 samples collected. UCMR data were not available for PFDA, PFHxA, or PFBA. However,  
7 samples from seven municipal wells in Oakdale, MN were analyzed for PFHxA and PFBA. The  
8 concentrations ranged from <0.025 to 0.235 µg/L and 0.0855 to 2.04 µg/L, respectively ([U.S. EPA,](#)  
9 [2017b](#)). [Kim and Kannan \(2007\)](#) analyzed lake water, rainwater, snow, and surface water from  
10 Albany, NY, and reported concentrations of PFDA, PFNA, and PFHxS ranging from less than the LOD  
11 to 0.0135 µg/L. PFAS were detected at higher concentrations in groundwater samples from an  
12 industrial site (3M Cottage Grove) in Minnesota. PFHxS and PFBA were detected in all seven wells  
13 that were sampled at concentrations ranging from 6.47–40 µg/L and 23.3–318 µg/L, respectively  
14 [[WS \(2007\)](#) as cited in [ATSDR \(2018\)](#)]. The concentrations of these five PFAS measured at National  
15 Priorities List (NPL) sites are shown in Table 2-3 as reported in [ATSDR \(2018\)](#), and the  
16 concentrations of PFAS measured in surface water and groundwater at 10 military installations are  
17 given in Table 2-2 as reported in [ATSDR \(2018\)](#).

**Table 2-2. Levels of the per- and polyfluoroalkyl substances (PFAS) being assessed in environmental media at 10 military installations**

Media	Measure	PFDA	PFNA	PFHxA	PFHxS	PFBA
Surface soil	Frequency of detection (%)	67.03	71.43	70.33	76.92	38.46
	Reporting limit (µg/kg)	0.28	0.23	0.16	0.29	0.12
	Median (µg/kg)	0.980	1.30	1.75	5.70	1.00
	Maximum (µg/kg)	15.0	23.0	51.0	1,300	31.0
Subsurface soil	Frequency of detection (%)	12.50	14.42	65.38	59.62	29.81
	Reporting limit (µg/kg)	0.30	0.24	0.16	0.31	0.13
	Median (µg/kg)	1.40	1.50	1.04	4.40	0.960
	Maximum (µg/kg)	9.40	6.49	140	520	14.0
Sediment	Frequency of detection (%)	48.48	12.12	63.64	72.73	24.24
	Reporting limit (µg/kg)	0.46	0.38	0.26	0.48	0.21
	Median (µg/kg)	1.90	1.10	1.70	9.10	1.70
	Maximum (µg/kg)	59.0	59.0	710	2,700	140
Surface water	Frequency of detection (%)	52.00	36.00	96.00	88.00	84.00
	Reporting limit (µg/L)	0.008	0.017	0.003	0.007	0.010
	Median (µg/L)	0.067	0.096	0.320	0.710	0.076
	Maximum (µg/L)	3.20	10.0	292	815	110
Groundwater	Frequency of detection (%)	34.78	46.38	94.20	94.93	85.51
	Reporting limit (µg/L)	0.008	0.018	0.003	0.007	0.010
	Median (µg/L)	0.023	0.105	0.820	0.870	0.180
	Maximum (µg/L)	1.80	3.00	120	290	64.0

Source: [ATSDR \(2018\)](#).

**Table 2-3. Levels of the per- and polyfluoroalkyl substances (PFAS) being assessed in water, soil, and air at National Priorities List sites**

Media	Measure	PFDA	PFNA	PFHxA	PFHxS	PFBA
Water	Median (ppb)	ND	ND	0.25	0.26	2.15
	Geometric mean (ppb)	ND	ND	0.10	1.12	1.03
Soil	Median (ppb)	ND	27.2	1,175	5,585	1,600
	Geometric mean (ppb)	ND	27.2	1,175	5,585	1,600
Air	Median (ppbv)	ND	ND	ND	ND	ND
	Geometric mean (ppbv)	ND	ND	ND	ND	ND

ND = no data.

Source: [ATSDR \(2018\)](#).

- 1 [Schechter et al. \(2012\)](#) collected 10 samples of 31 food items from five grocery stores in
- 2 Texas and analyzed them for persistent organic pollutants, including PFDA, PFNA, PFHxA, and
- 3 PFHxS. PFDA, PFNA, and PFHxA were not detected in any of the foods targeted, and PFHxS was
- 4 detected in cod fish at a concentration of 0.07 ng/g wet weight. PFAS have been detected in fish
- 5 from U.S. lakes and rivers with concentrations ranging from less than the limit of quantification to

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1 15.0 ng/g for PFDA, and <1 to 0.47 ng/g for PFHxS ([ATSDR, 2018](#)). [Stahl et al. \(2014\)](#) characterized  
 2 PFAS in freshwater fish from 164 U.S. urban river sites and 157 Great Lakes sites. PFDA, PFNA,  
 3 PFHxA, PFHxS, and PFBA were detected in 92, 69, 15, 45, and 16% of the samples, at maximum  
 4 concentrations of 13.0, 9.7, 0.8, 3.5, and 1.3 ng/g, respectively. Apart from fish, overall dietary data  
 5 for the United States are limited; however, [Schneider et al. \(2017\)](#) detected PFAS in food packaging  
 6 collected from U.S. fast food restaurants. Data from other countries (e.g., South Korea, Brazil, Saudi  
 7 Arabia) suggest that these PFAS can sometimes be detected in samples of food products, including  
 8 shellfish, dairy products, meats, vegetables, food packaging materials, and water [both tap and  
 9 bottled; [Heo et al. \(2014\)](#); [Chen et al. \(2018\)](#); [Pérez et al. \(2014\)](#); [Moreta and Tena \(2014\)](#); [Surma et  
 10 al. \(2017\)](#)]. The relevance of these detects (and the associated PFAS levels) to U.S. products is  
 11 unknown. Information on detection limits is available in the referenced studies.

### **2.1.5. Potential for Human Exposure**

12 The general population may be exposed to PFAS through multiple routes, including  
 13 ingestion of drinking water and food, ingestion of dust, hand-to-mouth and dermal transfer in  
 14 products and materials containing these chemicals, and inhalation via indoor and outdoor air  
 15 ([ATSDR, 2018](#); [NLM, 2017, 2013](#)). The oral route of exposure has been considered the most  
 16 important exposure route for PFAS in the general population ([Klaunig et al., 2015](#)).

17 The presence of PFAS in human blood provides evidence of exposure among the general  
 18 population. PFAS have been monitored in the human population as part of the National Health and  
 19 Nutrition Examination Survey (NHANES). PFDA, PFNA, and PFHxS were measured in serum  
 20 samples collected in 2013–2014 from more than 2,000 survey participants. The results of these  
 21 analyses are presented in Table 2-4. PFDA and PFNA have also been observed in cord blood and  
 22 human milk ([ATSDR, 2018](#)). [Pinney et al. \(2014\)](#) and [Papadopoulou et al. \(2016\)](#) observed  
 23 associations between breastfeeding and elevated levels of PFHxS in the blood of children.

**Table 2-4. Serum concentrations of the per- and polyfluoroalkyl substances (PFAS) being assessed based on National Health and Nutrition Examination Survey (NHANES) 2013–2014 data (µg/L)**

Population group	Measure	PFDA	PFNA	PFHxA	PFHxS	PFBA
Total population (n = 2,168)	Geometric mean	0.185	0.675	ND	1.35	ND
	50th percentile	0.200	0.700	ND	1.40	ND
	95th percentile	0.700	2.00	ND	5.60	ND
3 to 5 yr (n = 181)	Geometric mean	<sup>a</sup>	0.764	ND	0.715	ND
	50th percentile	0.100	0.620	ND	0.740	ND
	95th percentile	0.370	3.49	ND	1.62	ND
6 to 11 yr (n = 458)	Geometric mean	<sup>a</sup>	0.809	ND	0.913	ND
	50th percentile	<LOD	0.750	ND	0.850	ND
	95th percentile	0.350	3.19	ND	4.14	ND

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12 to 19 yr (n = 402)	Geometric mean	0.136	0.599	ND	1.27	ND
	50th percentile	0.100	0.500	ND	1.10	ND
	95th percentile	0.400	2.00	ND	6.30	ND
20 yr and older (n = 1,766)	Geometric mean	0.193	0.685	ND	1.36	ND
	50th percentile	0.200	0.700	ND	1.40	ND
	95th percentile	0.800	2.00	ND	5.50	ND

LOD = limit of detection; 0.1 (µg/L); ND = no data.

<sup>a</sup>Not calculated because the proportion of results below the LOD was considered too high to provide a valid result.

Source: [CDC \(2018\)](#). Fourth National Report on Human Exposure to Environmental Chemicals.

### 2.1.6. Populations and Lifestages with Potentially Greater Exposures

1 In addition to exposure scenarios that are expected to apply to the general population (see  
2 Section 2.1.5), certain populations and lifestages may have greater exposures than the general  
3 population. These groups include individuals in occupations that require frequent contact with  
4 PFAS-containing products, such as firefighters or individuals who install and treat carpets ([ATSDR,  
5 2018](#)), infants and young children (due to placental transfer, breastfeeding, or their increased hand-  
6 to-mouth behaviors), and populations consuming contaminated drinking water. [Rotander et al.  
7 \(2015\)](#) analyzed serum samples from 149 Australian firefighters at an AFFF training facility. Mean  
8 and median PFHxS concentrations were 10 to 15 times higher than those in the general population  
9 of Australia and Canada. Populations living near fluorochemical facilities where environmental  
10 contamination has occurred may also be more highly exposed ([ATSDR, 2018](#)). Also, because these  
11 chemicals can be found in ski wax, individuals who engage in professional ski waxing may be more  
12 highly exposed because PFAS such as PFHxA, PFNA, and PFDA in dust or fumes may be inhaled  
13 during this process ([Harbison et al., 2015](#); [Nilsson et al., 2010a](#); [Nilsson et al., 2010b](#)). Populations  
14 living near military or airport fire training areas or industrial sites that use or manufacture PFAS  
15 may be more likely to have high-level PFAS exposure through consumption of contaminated  
16 drinking water ([Hu et al., 2016](#)). Further, due to the high water solubility and mobility of PFAS in  
17 groundwater (and lack of current remediation technology at many water treatment facilities) it is  
18 possible for populations consuming drinking water from a contaminated watershed to receive  
19 disproportionate PFAS exposure ([Sun et al., 2016](#)).

20 Populations that rely primarily on seafood for most of their diet, possibly including some  
21 native American tribes ([Byrne et al., 2017](#)), may also be disproportionately exposed. [Christensen et  
22 al. \(2017\)](#) and [Haug et al. \(2010\)](#) used data on serum PFAS levels and 30-day, self-reported fish and  
23 shellfish ingestion rates from NHANES 2007–2014 to explore potential relationships between PFAS  
24 exposures and fish consumption. PFDA, PFNA, and PFHxS were among the PFAS detected in the  
25 serum of at least 30% of the NHANES participants, and after adjusting for demographic  
26 characteristics, total fish consumption was associated with elevated serum PFDE and PFNA.  
27 Shellfish consumption was associated with elevated levels of all the PFAS examined.

1 PFAS exposures to fetuses and infants are also important to consider as studies show the  
2 potential for elevated exposures during these sensitive developmental periods. Animal testing  
3 ([Beesoon et al., 2012](#); [Hinderliter et al., 2005](#)), and human studies [e.g., [Fei et al. \(2007\)](#), [Gao et al.](#)  
4 [\(2016\)](#), [Mamsen et al. \(2019\)](#), [Mondal et al. \(2014\)](#), [Zhang et al. \(2013a\)](#)] suggest that PFAS cross  
5 the blood-placental barrier with transfer efficiencies in humans that may depend on PFAS chain  
6 length and binding affinity to serum and breastmilk-protein complexes. Studies also show that  
7 breastmilk appears to be an important route of exposure to long-chain PFAS in breastfed infants,  
8 although the extent of lactational transfer of the current long-chain PFAS—PFNA, PFDA, and  
9 PFHxS—is less clear [e.g., [Fromme et al. \(2010\)](#), [Haug et al. \(2011\)](#), [Mondal et al. \(2014\)](#), [Mogensen](#)  
10 [et al. \(2015\)](#), [Kärrman et al. \(2007\)](#)].

#### **2.1.7. Other Environmental Protection Agency (EPA) Assessments of Per- and Polyfluoroalkyl Substances (PFAS)**

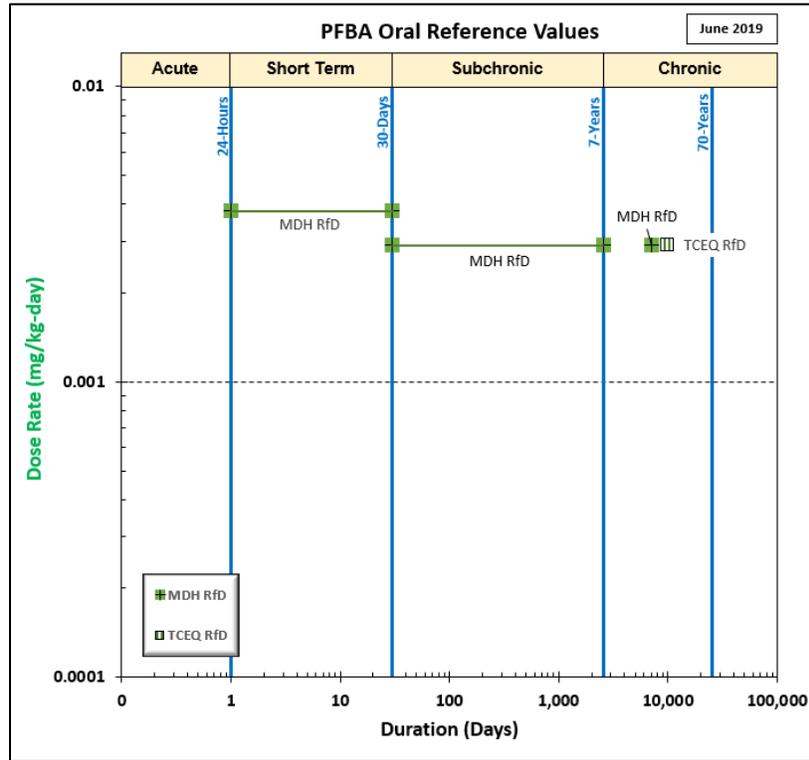
11 EPA released two PFAS assessments for peer review in 2018. Specifically, the draft  
12 assessments of (1) 2,3,3,3-tetrafluoro-2-(1,1,2,2,3,3,3-heptafluoropropoxy)propanoic acid (also  
13 called hexafluoropropylene oxide [HFPO] dimer acid) (CASRN 13252-13-6) and its ammonium salt  
14 2,3,3,3-tetrafluoro-2-(1,1,2,2,3,3,3-heptafluoropropoxy)propanoate (also called HFPO dimer acid)  
15 (CASRN 62037-80-3), referred to as GenX chemicals, and (2) perfluorobutane sulfonic acid  
16 (CASRN 375-73-5) and its salt potassium perfluorobutane sulfonate (CASRN 29420-49-3) referred  
17 to as PFBS. These assessments summarized the available data on the potential human health  
18 effects of lifetime exposure to these PFAS and included oral reference doses (RfDs), which estimate  
19 (with uncertainty spanning perhaps an order of magnitude) a level of daily oral exposure to the  
20 human population (including sensitive subgroups) that is likely to be without an appreciable risk of  
21 deleterious noncancer health effects during a lifetime, and qualitative descriptions of the  
22 carcinogenic potential of the chemicals. The PFBS assessment updates a Provisional Peer-Reviewed  
23 Toxicity Value (PPRTV) assessment that was developed in support of the Superfund Program and  
24 published in 2014 ([PFBS PPRTV 2014](#)). In addition, EPA released Drinking Water Health Advisories  
25 for PFOA and PFOS in 2016, along with health effect support documents ([Drinking Water Health](#)  
26 [Advisories for PFOA and PFOS](#)). Health advisories are nonenforceable and nonregulatory  
27 summaries of technical information on contaminants that can cause human health effects and are  
28 known or anticipated to occur in drinking water.

#### **2.1.8. Assessments and Toxicity Values from Other Sources**

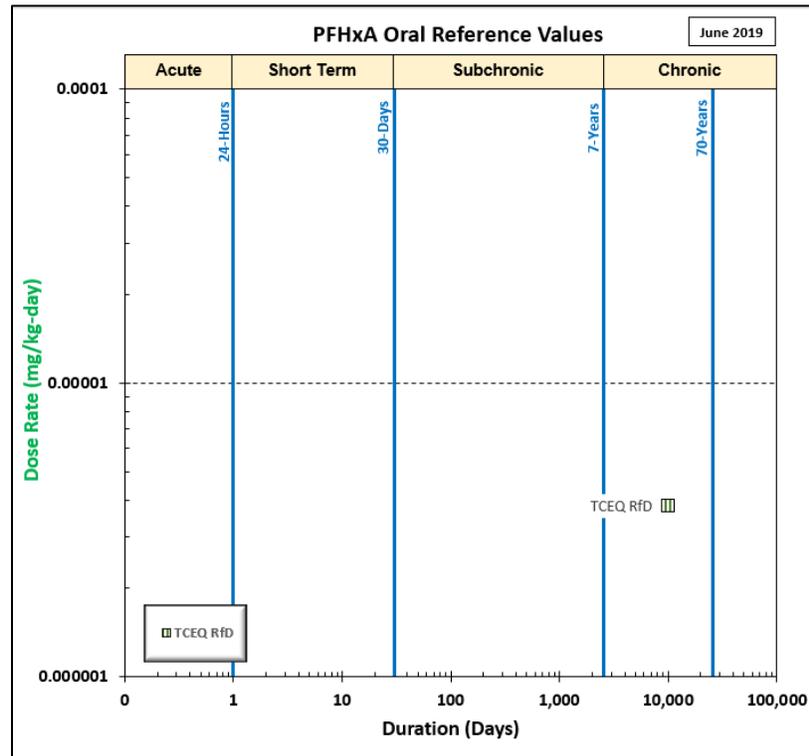
29 For the five PFAS addressed in this protocol, a summary of existing human health reference  
30 values from national, international, and state agencies (current as of March 2019), is provided in  
31 Figure 2-2 (see Addendum A for a tabular summary, including derivation details of the displayed  
32 values). The majority of current reference values are noncancer toxicity values based on oral  
33 exposure studies in rodents, although a few inhalation toxicity values exist (see Table A-1 in  
34 Addendum A for more details).

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(a)



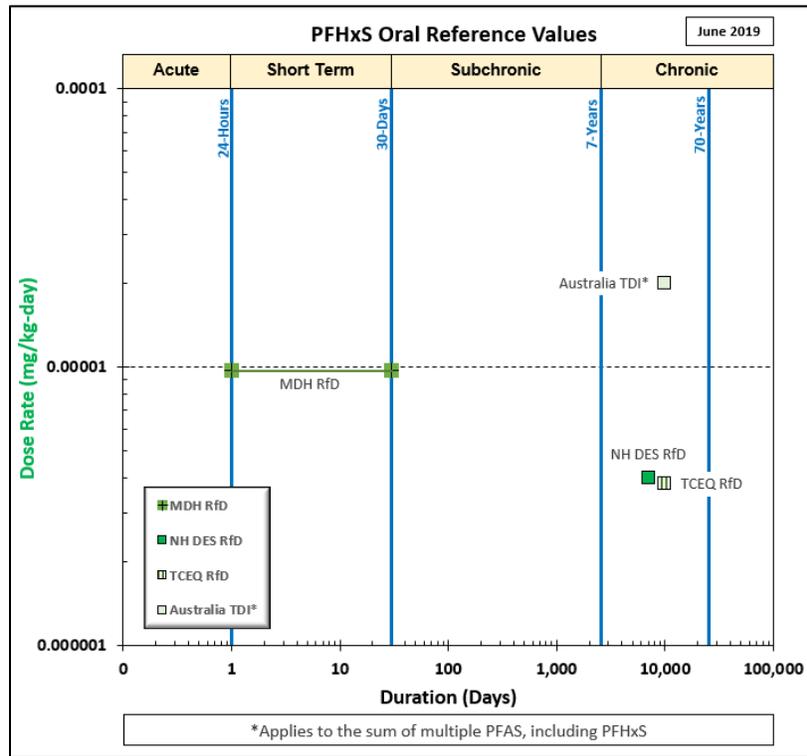
(b)



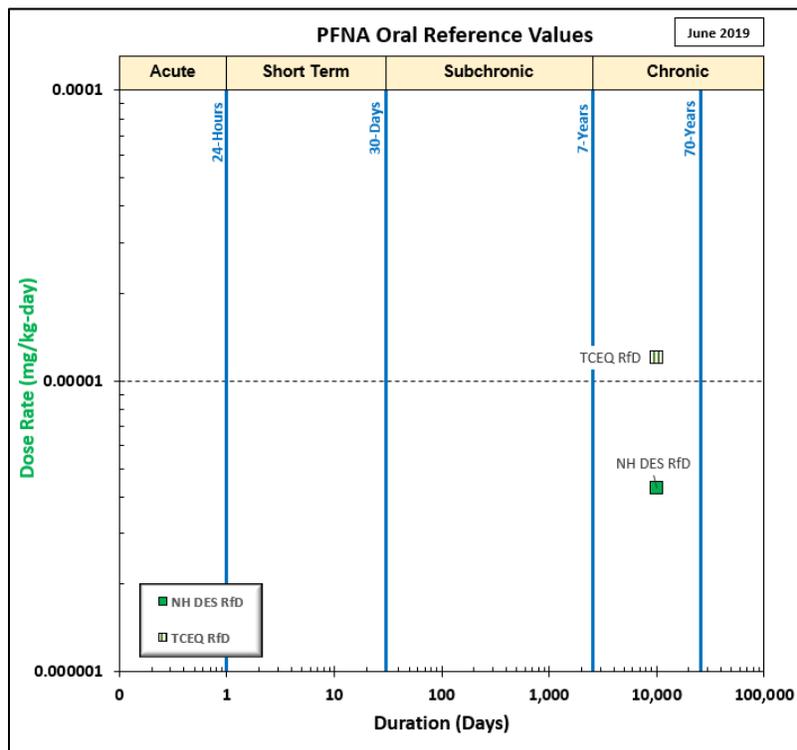
*This document is a draft for review purposes only and does not constitute Agency policy.*

**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

(c)



(d)



*This document is a draft for review purposes only and does not constitute Agency policy.*

(e)

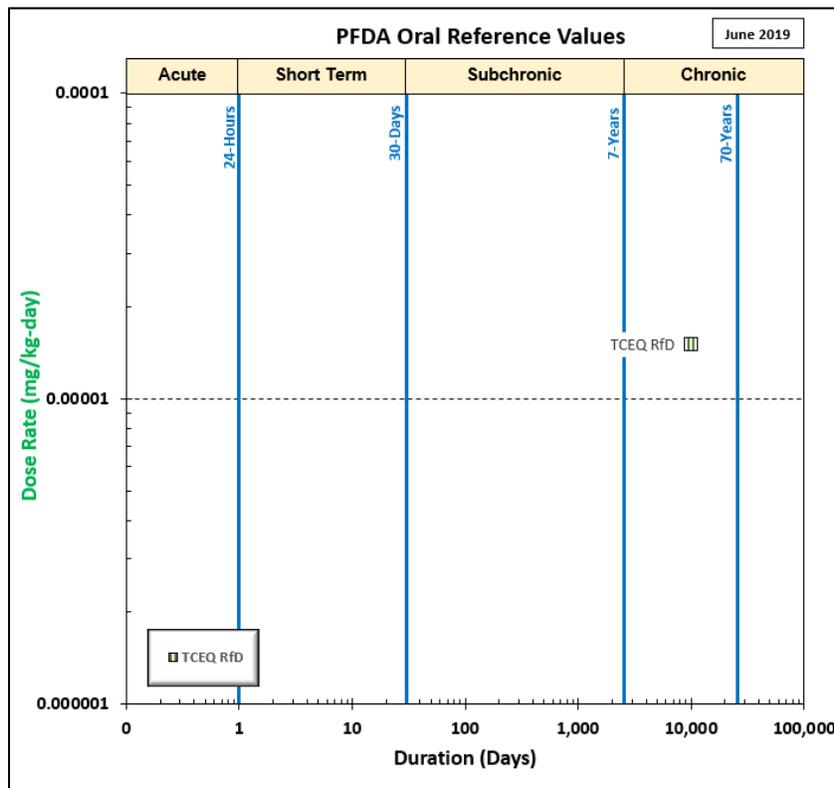


Figure 2-2. Existing oral reference values for (a) perfluorobutanoic acid (PFBA), (b) perfluorohexanoic acid (PFHxA), (c) perfluorohexanesulfonate (PFHxS), (d) perfluorononanoic acid (PFNA), and (e) perfluorodecanoic acid (PFDA). Abbreviations and additional details on the derivation of the values can be found in Addendum A.

## 2.2. SCOPING SUMMARY

1            Given the numerous PFAS of potential interest to the Agency, an extensive scoping effort  
 2            was undertaken to prioritize PFAS for review. This effort was coordinated across EPA program and  
 3            regional offices, where staff discussed specific assessment needs as well as the timeliness of those  
 4            needs. While additional factors were considered during this scoping effort, Table 2-5 summarizes  
 5            the primary considerations for selecting the five PFAS described in this protocol, as well as two  
 6            other PFAS that were recently assessed by EPA: PFBS and GenX chemicals  
 7            (<https://www.epa.gov/pfas/genx-and-pfbs-draft-toxicity-assessments>). In short, these PFAS:

- 8            • were identified as a priority to inform decision making for EPA’s Office of Water (OW),  
 9            Office of Land and Emergency Management (OLEM), Office of Chemical Safety and Pollution

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- 1 Prevention (OCSPP), Office of Children’s Health Protection (OCHP), EPA’s regional offices,  
 2 tribes, or state departments of environmental protection. Most of these PFAS were a  
 3 priority for multiple patrons;
- 4 • had been evaluated in in vivo studies of animals and thus might be used to derive toxicity  
 5 values; and
- 6 • had existing (or under development) standardized analytical methods to monitor  
 7 environmental levels to allow for site-specific application of toxicity values to regulatory  
 8 decision making.

**Table 2-5. Environmental Protection Agency (EPA) considerations for the selection of per- and polyfluoroalkyl substances (PFAS) for evaluation**

PFAS	EPA interest	Animal dose-response data available <sup>a</sup>	Analytical detection methods available <sup>b</sup>	
			Standards	Methods
PFDA	<ul style="list-style-type: none"> <li>OLEM priority<sup>c</sup></li> </ul>	Yes	Yes	Yes
PFNA	<ul style="list-style-type: none"> <li>OLEM priority<sup>c</sup></li> <li>OW (UCMR) priority</li> <li>Found in industrial effluent and AFFF</li> </ul>	Yes	Yes	Yes
PFHxA	<ul style="list-style-type: none"> <li>OCSPP priority<sup>d</sup></li> <li>OLEM priority<sup>c</sup></li> <li>Region 4 (Coosa and Tennessee Rivers)</li> <li>Found in AFFF</li> </ul>	Yes	Yes	Yes
PFHxS	<ul style="list-style-type: none"> <li>OCSPP priority</li> <li>OLEM priority<sup>c</sup></li> <li>OW (UCMR) priority</li> <li>Region 4 (Tennessee River)</li> <li>Found in AFFF</li> </ul>	Yes	Yes	Yes
PFBA	<ul style="list-style-type: none"> <li>OLEM priority<sup>c</sup></li> <li>OCSPP priority<sup>d</sup></li> <li>Found in AFFF</li> </ul>	Yes	Yes	Under development

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PFAS	EPA interest	Animal dose-response data available <sup>a</sup>	Analytical detection methods available <sup>b</sup>	
			Standards	Methods
PFBS	<ul style="list-style-type: none"> <li>OLEM priority<sup>c</sup></li> <li>OCSPP priority<sup>d</sup></li> <li>OW (UCMR) priority</li> <li>Found in AFFF</li> </ul>	Yes	Yes	Yes
GenX chemicals	<ul style="list-style-type: none"> <li>OCSPP priority<sup>e</sup></li> <li>Region 3 priority</li> <li>Region 4 priority</li> </ul>	Yes	Yes	Yes

GenX = perfluoro(2-methyl-3-oxahexanoic) acid (CASRN 13252-13-6); Unknown = status of validated standards and methods was unknown at scoping.

<sup>a</sup>A survey of publicly available literature on PFAS other than PFOA and PFOS (i.e., a broad PubMed search and review of recent assessments, including [ATSDR \(2018\)](#) was performed to identify in vivo animal studies that tested multiple PFAS exposure levels and evaluated health endpoints. The quality of the studies was not evaluated, and while multiple PFAS are evaluated in human studies, this was not a focus of the survey.

<sup>b</sup>As of March 2019. The methods noted are for drinking water; nondrinking water methods are being developed.

<sup>c</sup>Found at sites, including private and federal facilities and from various sources, including AFFF, chrome-plating facilities, PFAS manufacturers, and industries that use PFAS (e.g., textiles and electronics). These PFAS have also been detected in environmental media (e.g., surface water; biota).

<sup>d</sup>A significant number of new chemicals submitted to EPA are based on C6 and C4 chemistry. OCSPP often evaluates risk for these compounds based on PFHxA and PFBS, which are the terminal degradation products of certain C6 and C4 compounds.

<sup>e</sup>Replacement for PFOA (e.g., for emulsifiers) and perfluoroethers. GenX chemicals are of concern based on occurrence in NC and because EPA has received requests to review similar types of compounds (e.g., longer chain ethers that might break down to GenX chemicals) as new chemicals.

- 1 As described in Section 2.1.5, exposure to these five PFAS can occur via the oral, inhalation,
- 2 and dermal routes, with oral (e.g., through diet and drinking water) being the predominant one
- 3 ([Klaunig et al., 2015](#)). Given the potential regulatory applications of these PFAS assessments (see
- 4 Table 2-6), these assessments will consider PFAS exposures from all exposure routes. The
- 5 assessments will consider all potential health effects of exposure, both cancer and noncancer.

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**Table 2-6. Potential Environmental Protection Agency (EPA) needs and applications for five per- and polyfluoroalkyl substances (PFAS)**

EPA program or regional office	PFAS <sup>a</sup>	Oral	Inhalation	Dermal	Potential regulatory application and explanation (at the time scoping was conducted)
OLEM (in coordination with EPA Regions 1–10)	PFDA PFNA PFHxA PFHxS PFBA	✓	✓	✓	Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA). CERCLA authorizes EPA to conduct short- or long-term cleanups at Superfund sites and later recover cleanup costs from potentially responsible parties under Section 107. PFAS toxicological information may be used to make risk determinations for response actions (e.g., short-term removals, long-term remedial response actions). An evaluation of potential actions at Superfund sites considers all routes of exposure. Resource Conservation and Recovery Act (RCRA). RCRA can be drawn upon to help address waste management and cleanup needs, including accidental releases from potentially hazardous waste management facilities.
OW	PFNA PFHxS	✓			Safe Drinking Water Act (SDWA) and Clean Water Act (CWA). The SDWA requires EPA to periodically review the National Primary Drinking Water Regulation (NPDWR) for each contaminant and revise the regulation, if appropriate. These potential applications focus on oral exposure.
OCSPP	PFHxA PFHxS PFBA	✓	✓		New chemical submissions to the Office of Pollution Prevention and Toxics within OCSPP.
Region 4	PFHxA PFHxS	✓			Resource Conservation and Recovery Act (RCRA). RCRA can be drawn upon to help address waste management and cleanup needs, including accidental releases from potentially hazardous waste management facilities. For PFAS, the primary concern is potential oral exposure from rivers in Region 4.
OCHP	PFDA PFNA PFHxA PFHxS PFBA	✓	✓	✓	Executive Order 13045—Protection of Children from Environmental Health Risks and Safety Risks: Policy on Evaluating Health Risks to Children. In accordance with EPA’s 1995 policy and EO 13045, EPA instituted and reaffirmed an Agency-wide commitment to “consider the risks to infants and children consistently and explicitly as part of risk assessments generated during its decision-making process.”

<sup>a</sup>PFAS to which this protocol applies (i.e., excluding PFBS and GenX chemicals).

## **2.3. PROBLEM FORMULATION**

### **2.3.1. Preliminary Literature Inventory for the Five Per- and Polyfluoroalkyl Substances (PFAS) Being Assessed**

1           As described in Section 2.1.1, several of these five PFAS have associated salts of potential  
2 interest for human health assessment. Thus, the assessments will address each PFAS as follows:

- 3           • **PFBA:** PFBA (CASRN 375-22-4); PFBA ammonium salt (CASRN 10495-86-0)
- 4           • **PFHxA:** PFHxA (CASRN 307-24-4); PFHxA ammonium salt (CASRN 21615-47-4); PFHxA  
5 sodium salt (CASRN 2923-26-4)
- 6           • **PFHxS:** PFHxS (CASRN 355-46-4); PFHxS potassium salt (CASRN 3871-99-6)
- 7           • **PFNA:** PFNA (CASRN 375-95-1); PFNA ammonium salt (CASRN 4149-60-4); PFNA sodium  
8 salt (CASRN 21049-39-8)
- 9           • **PFDA:** PFDA (CASRN 335-76-2); PFDA ammonia salt (CASRN 3108-42-7); PFDA sodium salt  
10 (CASRN 3830-45-3)

11           The results of a preliminary literature inventory of health effect-related studies on these  
12 five PFAS and their associated salts are presented in Figure 2-3. The studies summarized in this  
13 preliminary literature inventory reflect searches conducted in mid-2019 (and will be updated in the  
14 context of the PFAS-specific assessments, but not this protocol) and are described on the project  
15 pages for these five PFAS assessments in HERO (<https://hero.epa.gov>; see Section 1 for links to the  
16 specific Health and Environmental Research Online [HERO] pages).

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**LEGEND:** +++ (~10+ studies)    ++ (~5 studies)    + (~1-2 studies)    - (Not Studied)

	PFDA and salts					PFNA and salts					PFHxA and salts					PFHxS and salts					PFBA and salts				
	Oral: Long <sup>1</sup>	Oral: Short <sup>1</sup>	Inhal.	Dermal	Human	Oral: Long <sup>1</sup>	Oral: Short <sup>1</sup>	Inhal.	Dermal	Human	Oral: Long <sup>1</sup>	Oral: Short <sup>1</sup>	Inhal.	Dermal	Human	Oral: Long <sup>1</sup>	Oral: Short <sup>1</sup>	Inhal.	Dermal	Human	Oral: Long <sup>1</sup>	Oral: Short <sup>1</sup>	Inhal.	Dermal	Human
Cardiovascular	-	+	-	-	++	-	+	-	-	+++	-	+	-	-	+	+	+	-	-	+++	-	+	-	-	+
Developmental	-	+	-	-	+++	-	++	-	-	+++	-	-	-	-	-	-	+	-	-	+++	-	+	-	-	+
Endocrine (Thyroid)	-	+	-	-	+++	-	+	-	-	+++	-	+	-	-	++	+	+	-	-	+++	-	+	-	-	+
Gastro-intestinal	-	+	-	-	-	-	-	-	-	-	-	+	-	-	-	-	+	-	-	-	-	-	-	-	-
Hematologic	-	+	-	-	+	-	+	-	-	+	+	++	-	-	+	+	+	-	-	+	-	+	-	-	-
Hepatic	-	+++	-	-	+++	-	+++	+	-	+++	+	+	-	-	++	+	++	-	-	+++	+	++	-	-	+
Immune	-	++	-	-	+++	-	++	-	-	+++	-	+	-	-	+	+	+	-	-	+++	-	-	-	-	-
Musculo-skeletal	-	+	-	-	-	-	-	-	-	+	-	-	-	-	-	-	+	-	-	+	-	-	-	-	-
Nervous	-	+	-	-	++	-	-	-	-	+++	+	+	-	-	-	+	+	-	-	+++	-	+	-	-	-
Ocular	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Reproductive	-	+	-	-	+++	-	++	-	-	+++	-	+	-	-	+	+	+	-	-	+++	-	+	-	-	+
Respiratory	-	+	-	-	-	-	-	-	-	-	-	+	-	-	-	-	+	-	-	-	-	-	-	-	-
Urinary	-	+	-	-	+	-	+	-	-	+++	+	+	-	-	+	+	+	-	-	+++	-	-	-	-	-
General Toxicity/ Other	-	+++	-	-	+	-	+++	+	-	+	+	++	-	-	+	+	++	-	-	+	+	++	-	-	-
Cancer	-	-	-	-	+	-	-	-	-	++	+	-	-	-	-	-	-	-	-	++	-	-	-	-	-

**Figure 2-3. Results of a preliminary literature inventory of five per- and polyfluoroalkyl substances (PFAS).** Data are approximated based on a cursory review of the literature search results for studies published through 2018 (see Section 4 for details; this includes at-the-time-unpublished reports from NTP, see Section 4.1). Health effects are based on groupings from the EPA’s Integrated Risk Information System (IRIS) website (<https://cfpub.epa.gov/ncea/iris/search/index.cfm>).<sup>a</sup> For this summary, metabolic effects are captured under “other” and “hepatic” includes lipid and lipoprotein measures.

<sup>a</sup>“Oral: long” indicates subchronic or chronic oral exposure duration studies in animals and “Oral: short” reflects short-term and acute oral exposure studies in animals, as well as reproductive and developmental studies.

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1           Based on the results from the preliminary literature inventory in Figure 2-3, the following  
2 health effects appear to be well studied for most PFAS of interest:

- 3           • Developmental effects
- 4           • Endocrine (primarily thyroid hormone) effects
- 5           • Hepatic effects, including lipid and lipoprotein measures
- 6           • Immune effects
- 7           • Reproductive effects in males or females
- 8           • Urinary effects
- 9           • General toxicity

10           As also shown in Figure 2-3, no studies of dermal exposure were identified. In addition,  
11 data are sparse for assessing the potential health effects from chronic or subchronic oral exposure  
12 or for inhalation exposure of any duration. Few studies have examined whether exposure to these  
13 PFAS may result in carcinogenicity.

14           Given the potential future utility of comparing evidence across PFAS assessments (including  
15 their respective data gaps), the five PFAS assessments will specifically address each of the potential  
16 health effects enumerated above as “well studied.” In addition, the potential for carcinogenicity will  
17 be explicitly addressed in each assessment. Data on several other, variably studied endpoints  
18 (i.e., cardiovascular effects, hematological effects, metabolic effects including diabetes, and nervous  
19 system effects) will also be summarized when available. These summaries may be developed in  
20 association with one of the health effects noted above, as a separate formal evaluation of hazard, or  
21 as part of a qualitative summary on “other effects,” depending on the assessment-specific data.  
22 Information on other health effects, such as gastrointestinal effects; musculoskeletal effects; ocular  
23 effects; and respiratory effects may be briefly summarized but will not be formally evaluated in any  
24 of these assessments because of the paucity of available studies and the absence of exceptional  
25 evidence in those that do exist. New literature relating to these outcomes will be monitored during  
26 literature search updates for potential inclusion.

---

### **2.4. KEY SCIENCE ISSUES**

27           This section describes critical areas of scientific complexity that were identified based on  
28 the preliminary literature inventory results summarized in the previous section. These scientific  
29 issues are essential to consider during development of these assessments, and the specific methods  
30 for doing so within these PFAS assessments are described in subsequent sections.

#### **2.4.1. Toxicokinetic Differences across Species and Sexes**

1           The PFAS being evaluated are not metabolized, and reported half-lives in humans range  
2 from several days (PFBA, PFBS) to multiple years (e.g., PFHxS). They are typically not stored in  
3 body fat (see Section 2.1 for PFAS-specific chemical properties, including predicted LogP), but  
4 accumulate in locations such as the blood, liver, and kidneys [and can be transferred to offspring  
5 through placental transfer and breast milk; [Post et al. \(2012\)](#); [ATSDR \(2018\)](#); [U.S. EPA \(2016d\)](#); [U.S.  
6 EPA \(2016c\)](#)]. However, as illustrated in Table 2-8, previous summaries of the existing literature  
7 suggest there are pronounced half-life differences across species, sex, and type of PFAS. In general,  
8 PFAS with longer chain lengths are reported to have a longer serum half-life. For the PFAS with  
9 data available, serum half-life variation across species generally exhibits the following pattern:  
10 rats<mice<monkeys<humans. The extent of this cross-species difference appears to be greater  
11 than would be predicted by standard allometric (body-weight scaling to the 3/4 power [BW<sup>3/4</sup>])  
12 scaling, and for mice versus rats is in the opposite direction (i.e., allometric scaling would predict  
13 mice<rats<monkeys<humans). Finally, some of the PFAS being assessed show a shorter serum  
14 half-life in females than in males (e.g., PFHxA in monkeys, PFBA in mice and rats), sometimes  
15 markedly so (e.g., PFNA in rats). Therefore, differences between males and females will generally  
16 be considered real, though a single half-life will be estimated if the difference appears negligible.  
17 The approach to validating and possibly refining these values (e.g., based on new data) in these  
18 PFAS assessments is outlined in Section 9.2.1. Notably, there is expected to be insufficient data to  
19 examine lifestage-specific differences in absorption, distribution, metabolism, and excretion  
20 (ADME).

21           While PK parameters can vary between animal strains, there are other potential sources of  
22 variability and uncertainty in reported values (differences in study design, analytic method, etc.)  
23 that would confound attempts to differentiate between strains. Different strains could also respond  
24 differently to the same dose because of differences in pharmacodynamic sensitivity. No PK studies  
25 that directly evaluate between-strain differences for a PFAS being addressed here  
26 (i.e., measurement of PK in more than one strain in a single study, hence eliminating other factors of  
27 study-to-study variability) have been identified. If, for example, several studies show that terminal  
28 phase clearance of a PFAS in one strain is in a range clearly different from that observed by another  
29 study in a different strain, then strain-specific half-life or clearance values will be determined. But  
30 considering the wide study-to-study variability found for experiments conducted with the same  
31 strain of animals, this outcome is unlikely.

32           Some toxicity studies report plasma PFAS levels measured at the end of the study. While  
33 these data can be a better measure of internal dose than the exposure dose rate and can potentially  
34 be converted directly to human equivalent doses (HEDs), this is less true for short-chain PFAS with  
35 half-lives on the order of hours. Calculation of HEDs requires a measure of clearance (CL) in  
36 humans, which depends on both the half-life ( $t_{1/2}$ ) and the volume of distribution ( $V_d$ ). For most of  
37 the PFAS only the  $t_{1/2}$  has been measured, in which case one would have to assume the  $V_d$  for

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1 humans is the same as for animals, making the approach equally reliant on animal PK data. Also,  
2  $t_{1/2}$  measurements in animals are most reliable when  $t_{1/2}$  is in the range of hours to a few days. On  
3 the other hand, measuring  $t_{1/2}$  is much more difficult when the value is very large, because it  
4 requires long-term observation during which one must account for animal growth. But when  $t_{1/2}$  is  
5 long, plasma levels will not vary rapidly from hour to hour, hence a measurement at the end of a  
6 toxicity study should be a reliable measure of average internal dose. PFNA and PFDA have the  
7 longest half-life in rats, averaging 1–2 months, but PFHxS also has  $t_{1/2}$  values reported up to 30 days  
8 in male rats and in mice. Conveniently, matched blood and urine data were obtained for these three  
9 in humans by [Zhang et al. \(2013b\)](#), allowing for a direct measure of CL in humans.

10 The apparent toxicokinetic differences may significantly affect the interpretation of toxic  
11 effects across species or sexes. More directly, substantive toxicokinetic differences would be  
12 expected to affect quantitative extrapolations of dose-response data from experimental animals to  
13 humans. Thus, the half-life estimates for these five PFAS are likely to impact multiple assessment  
14 decisions, and a critical review of the available ADME data for each PFAS will be important (see  
15 discussion in Sections 5 and 9.2).

16 Although not identified during the preliminary literature inventory shown above,  
17 physiologically based pharmacokinetic (PBPK) models for PFHxS ([Kim et al., 2018](#)) and PFDA and  
18 PFNA ([Kim et al., 2019](#)) parameterized for adult male and female rats and humans have recently  
19 been described. [Fàbrega et al. \(2015\)](#) also described a PBPK model for multiple PFAS in humans,  
20 including PFBS, PFHxS, PFHxA, PFNA, and PFDA. In addition, [Verner et al. \(2016\)](#) and [Goeden et al.  
21 \(2019\)](#) described models for evaluating gestational and lactational transfer of PFAS from mothers  
22 to their children, including PFHxS and PFOA. These models could prove useful for addressing  
23 toxicokinetic questions in these assessments (see discussion in Sections 6.4 and 11.2).

**Table 2-7. Preliminary serum half-life estimates of five per- and polyfluoroalkyl substances (PFAS) across species and sexes**

	PFBA (C4)		PFHxA (C6)		PFHxS (C6)		PFNA (C9)		PFDA (C10)	
	Female	Male	Female	Male	Female	Male	Female	Male	Female	Male
Rat	1.0–1.8 h	6–9 h	0.4–0.6 h	1.0–1.6 h	1.8 d	6.8 d	1.4 d	30.6 d	58.6 d	39.9 d
Mouse	3 h	12 h	~1.2 h	~1.6 h	24–27 d	28–30 d	26–68 d	34–69 d	ND	
Monkey	1.7 d		2.4 h	5.3 h	87 d	141 d	ND		ND	
Human	3 d		32 d		8.5 yr		4.3 yr		12 yr	

“C” = carbon chain length; ND = no data.

Data are summarized in [Lau \(2015\)](#). Note that these values do not necessarily represent those that would be used in qualitative or quantitative analyses for these PFAS assessments because the underlying data will be reviewed and possibly supplemented with additional (e.g., newer) studies. Darker shading indicates longer half-life (i.e., from hours to days to years).

#### 2.4.2. Human Relevance of Effects in Animals that Involve Peroxisome Proliferator-Activated Receptor Alpha (PPAR $\alpha$ )

1           Activation of the peroxisome proliferator-activated receptor alpha (PPAR $\alpha$ ) by PFAS has  
 2           been reported, with in vitro evidence that the potency of human and mouse PPAR $\alpha$  activation is  
 3           positively correlated with increasing PFCA chain length up to C9 (no human receptor activation was  
 4           noted for PFDA, although activation of the mouse receptor was only slightly less potent than PFNA)  
 5           and greater for carboxylates than sulfonates ([Wolf et al., 2014](#); [Wolf et al., 2008](#); [Takacs and Abbott,](#)  
 6           [2007](#); [Shipley et al., 2004](#); [Maloney and Waxman, 1999](#)). It is not known whether PFAS distribute to  
 7           the nucleus and bind directly to PPAR $\alpha$  in vivo, or whether these substances activate the receptor  
 8           indirectly. PPAR $\alpha$  ligand binding causes a conformational change in the protein, release of  
 9           corepressors, heterodimerization with the retinoid X receptor (RXR), and binding to cognate  
 10          peroxisome proliferator response elements in the promoters of target genes (perhaps most notably,  
 11          those related to fatty acid  $\beta$ -oxidation and energy homeostasis) to modulate gene transcription.

12          PPAR $\alpha$  is a ligand-activated nuclear receptor expressed in many tissues and has been at the  
 13          forefront of a longstanding debate as to whether chemical-induced PPAR $\alpha$  modulation in rodents,  
 14          particularly in the liver, is relevant to humans ([Corton et al., 2018](#); [Filgo et al., 2015](#); [Guyton et al.,](#)  
 15          [2009](#)). PPAR $\alpha$  is active in humans and responsive to the hypolipidemic effect of fibrate drugs that  
 16          lower serum lipid levels, but the human receptor is generally considered less sensitive than PPAR $\alpha$   
 17          in rodents ([Corton et al., 2014](#); [Wolf et al., 2014](#); [Wolf et al., 2008](#); [Maloney and Waxman, 1999](#)).  
 18          However, there are known human PPAR $\alpha$  and other hepatic nuclear receptor polymorphisms  
 19          associated with increased susceptibility to liver disease ([Li et al., 2016](#); [Li et al., 2012](#)). PPAR $\alpha$   
 20          activation has been extensively shown to induce peroxisome proliferation and result in  
 21          hepatocellular carcinoma. [reviewed in [Liss and Finck \(2017\)](#) and [Corton et al. \(2014\)](#)]. This

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1 phenomenon has not been observed in human models and is specific to rodents ([Corton et al., 2014](#);  
2 [Gonzalez and Shah, 2008](#); [Holden and Tugwood, 1999](#)). These effects are not observed in human  
3 models ([Corton et al., 2014](#)). Given the critical role PPARs' play as master regulators of lipid and  
4 glucose metabolism in multiple cell types as described above, the role this family of nuclear  
5 receptors plays in human metabolic diseases such as nonalcoholic liver disease (NAFLD) is an  
6 active area of research ([Liss and Finck, 2017](#)).

7 It continues to be difficult to evaluate the relative sensitivity of humans and animal models  
8 to PFAS-related PPAR $\alpha$  inductions and to determine the extent to which differences relate to  
9 differing toxicokinetics and/or intrinsic variations in biological sensitivities. For example, in some  
10 contrast to observations in rodent models, longer duration administration of PFOA to nonhuman  
11 primates (cynomolgus monkey) resulted in increasing absolute and relative liver-weight trends,  
12 with statistically significant increases in relative liver weight at the higher dose, but in the absence  
13 of histopathological changes ([Butenhoff et al., 2002](#)). These effects were concomitant with  
14 significantly increased enzymatic markers of mitochondrial proliferation (not dose dependent) and  
15 peroxisome proliferation at higher doses that further complicate interpretation. Evaluating the  
16 human relevance of animal PPAR $\alpha$  evidence is also complicated by a lack of comparable in vitro  
17 model systems, including widely used primary cell lines that rapidly lose the capability to express  
18 nuclear receptors such as PPAR $\alpha$  ([Soldatow et al., 2013](#)), and potential species-specific differences  
19 in transcriptional coactivators and other pathway components. Finally, while toxicity studies  
20 conducted with other more data-rich PFAS, such as PFOA and PFOS, may be informative to  
21 characterizing data gaps and uncertainties, caution needs to be exercised when extrapolating  
22 across PFAS, given that PFAS chain length, branching, and functional groups appear to be important  
23 drivers influencing toxicokinetic and toxicological properties.

24 PPAR $\alpha$  is also known to be important to other physiological processes in both rodents and  
25 humans, including energy homeostasis, inflammation, reproduction, musculoskeletal function, and  
26 development ([NJDWQI, 2017](#); [Corton et al., 2014](#); [Burri et al., 2010](#); [Abbott, 2009](#); [Peraza et al.,](#)  
27 [2006](#); [Corton et al., 2000](#)). Thus, although not extensively studied for PFAS, the modulation of  
28 PPAR $\alpha$  may be important to consider for developmental, metabolic, reproductive, and  
29 immunological effects, as well as for hepatic effects.

30 There are additional complexities when considering the dependence on, and human  
31 relevance of, PPAR $\alpha$  activation by PFAS for certain health effects. The extent of PPAR $\alpha$  activation is  
32 likely to differ by PFAS type, making it harder to apply read-across (specifically, drawing  
33 conclusions for one PFAS based on findings for another PFAS) or related approaches. In addition,  
34 based on conclusions from other PFAS assessments and review articles, there is evidence to  
35 indicate that many PFAS-mediated effects appear to include both PPAR $\alpha$ -dependent and  
36 PPAR $\alpha$ -independent mechanisms, the latter of which include activation of PPAR $\gamma$ ,  
37 phosphatidylinositol-3-kinase-serine/threonine kinase Akt (PI3K-Akt), constitutive androstane  
38 receptor (CAR), mitochondrial damage, nuclear factor kappa B pathway (NF- $\kappa$ B), farnesoid X

1 receptor, liver X receptor, and estrogen receptor  $\alpha$  ([Li et al., 2017](#); [NJDWQL, 2017](#); [Rosen et al.](#)  
2 [2017](#); [FSANZ, 2016](#); [U.S. EPA, 2014a, b](#); [Foreman et al., 2009](#)).

3 Despite the complexities involved, it is important to evaluate the human relevance of some  
4 PFAS exposure-mediated effects in animals (see discussion in Section 9.2.2).

### **2.4.3. Potential Confounding by Other Per- and Polyfluoroalkyl Substances (PFAS) Exposures in Epidemiology Studies**

5 Because different PFAS may be used in similar applications or result from similar sources,  
6 potential confounding of associations by PFAS coexposures is an important area of uncertainty for  
7 epidemiology studies. When associations are found for two or more moderately correlated PFAS in  
8 a study, including those not the focus of these assessments (e.g., PFOS and PFOA), confounding is a  
9 possible explanation. Based on a cursory review of studies identified during the preliminary  
10 literature inventory, a complicating factor is that correlations between PFAS pairs vary  
11 considerably across studies (see Section 6.2.1). When a study does not report the correlations in its  
12 population, the interpretation of the risk of bias from confounding is particularly challenging. Even  
13 when correlations are reported, there is no perfect method for eliminating confounding. Given this  
14 variability, assessing the likelihood and impact of this source of potential confounding based on  
15 reporting within individual studies is expected to be difficult (see discussion in Section 6.2).

### **2.4.4. Toxicological Relevance of Changes in Certain Urinary and Hepatic Endpoints in Rodents**

16 The scientific community has identified difficulties in interpreting the toxicological  
17 relevance of changes in certain urinary and hepatic endpoints available in rodent studies (based on  
18 the preliminary literature inventory) for some of the five PFAS assessments. The specific rodent  
19 endpoints in question are chronic progressive nephropathy and related urinary histopathological  
20 changes (including alpha 2u-globulin-mediated changes) and hepatic effects that may be  
21 considered adaptive (e.g., increased liver weight; cellular hypertrophy; single cell  
22 necrosis/apoptosis). For the former, some of these changes are not considered relevant to humans,  
23 and methods exist for evaluating the dependency of observed changes on this rodent-specific  
24 mechanism. For the latter, neither a clear scientific consensus nor specific EPA-wide guidance  
25 defines exactly what level of change or constellation of effects is necessary to establish a cause for  
26 concern. Thus, interpretations of the toxicological relevance of changes in these specific endpoints  
27 are expected to require additional consideration (see discussion in Section 9.2).

### **2.4.5. Characterizing Uncertainty Due to Missing Chemical-Specific Information**

28 Two PFAS, PFOA and PFOS (C8), have been studied more extensively than other PFAS.  
29 Thus, this existing knowledge base may be useful in helping to characterize existing data gaps and  
30 uncertainties in the current five PFAS assessments. Two recently developed EPA assessments  
31 (PFBS and GenX chemicals) could also provide information during the development of these

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1 current assessments. For example, given knowledge regarding the health effects of PFOA and PFOS,  
2 the potential lack of studies on immune effects for PFBA and developmental effects for PFHxA  
3 (based on the preliminary literature inventory; see Section 2.3.2) appear to represent important  
4 database uncertainties. In addition, given the potential for lifetime human exposure to PFAS by  
5 multiple routes of exposure (see Section 2.1.5), the apparent scarcity of data on most of these five  
6 PFAS other than short-term oral exposure studies in animals is expected to affect assessment  
7 decisions and characterization of uncertainties (see discussion in Sections 10.2 and 11.2.3).

### **3. OVERALL OBJECTIVES, SPECIFIC AIMS, AND POPULATIONS, EXPOSURES, COMPARATORS, AND OUTCOMES (PECO) CRITERIA**

1           The overall objective of these five assessments is to identify adverse human health effects  
2 and characterize exposure-response relationships for the effects of perfluorobutanoic acid (PFBA),  
3 perfluorohexanoic acid (PFHxA), perfluorohexanesulfonate (PFHxS), perfluorononanoic acid  
4 (PFNA), and perfluorodecanoic acid (PFDA) to support development of toxicity values. These  
5 assessments will use systematic review methods to evaluate the epidemiological and toxicological  
6 literature, including consideration of relevant mechanistic evidence (e.g., to inform key science  
7 issues; see Section 2.4). The evaluations conducted in these assessments will be consistent with  
8 relevant Environmental Protection Agency (EPA) guidance.<sup>3</sup>

9           The specific approach taken for these assessments of the potential health effects of PFBA,  
10 PFHxA, PFHxS, PFNA, and PFDA (and their associated salts) was based on input received during  
11 scoping, as well as a preliminary literature inventory of the health effects studied for these PFAS.  
12 As outlined in Section 2.3.2, these assessments will evaluate the potential for PFAS exposure via the  
13 oral or inhalation route to cause health effects in humans, specifically focusing on developmental  
14 effects; endocrine (primarily thyroid hormone) effects; hepatic effects, including lipid and  
15 lipoprotein measures; immune effects; reproductive effects in males or females; urinary effects;  
16 general toxicity; and carcinogenicity (see Section 5 for preliminary decisions for grouping outcomes  
17 and endpoints within each of these predetermined health effect categories). Data on cardiovascular  
18 effects, hematological effects, metabolic effects including diabetes, and nervous system effects will  
19 also be summarized when available. These summaries may be developed in association with one of  
20 the health effects noted above either as a separate formal evaluation of hazard or as part of a  
21 qualitative summary on “other effects,” depending on the assessment-specific data. Given the  
22 paucity of available studies and in the absence of exceptional evidence in any available studies,  
23 information on other health effects (i.e., gastrointestinal effects; musculoskeletal effects; ocular  
24 effects; and respiratory effects) will not be formally evaluated (these effects may be briefly  
25 summarized) in any of these assessments; although new literature relating to these outcomes will  
26 be monitored during literature search updates for potential inclusion. As outlined in the EPA PFAS  
27 action plan,<sup>4</sup> the characterization of the potential human health hazards from exposure to these

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<sup>3</sup>EPA guidance documents: <http://www.epa.gov/iris/basic-information-about-integrated-risk-information-system#guidance/>.

<sup>4</sup>EPA PFAS action plan: <https://www.epa.gov/pfas/epas-pfas-action-plan>.

1 individual PFAS will be coupled with data generated from new advances in computational and  
2 high-throughput toxicology to inform evaluations of other PFAS.

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### 3.1. SPECIFIC AIMS

3 The aims of these assessments are to:

- 4 • Identify epidemiological (i.e., human) and toxicological (i.e., experimental animal) literature  
5 reporting effects of exposure to PFBA, PFHxA, PFHxS, PFNA, and PFDA (and their associated  
6 salts), as outlined in the PECO. These five systematic reviews will focus on identifying  
7 studies following oral or inhalation exposure to PFAS.
  
- 8 • Evaluate mechanistic information (including toxicokinetic understanding) associated with  
9 exposure to PFBA, PFHxA, PFHxS, PFNA, and PFDA, to inform the interpretation of findings  
10 related to potential health effects in studies of humans and animals. The scope of these  
11 analyses of mechanistic information will be determined by the complexity and confidence in  
12 the phenotypic evidence in humans and animals, the likelihood of the analyses  
13 (e.g., considering the mechanistic studies available based on the literature inventory; see  
14 Section 4.2.2) to affect evidence synthesis conclusions for human health, and the directness  
15 or relevance of the available model systems for understanding potential human health  
16 hazards (see Section 9.2). The mechanistic evaluations will focus primarily on the key  
17 science issues identified in Section 2.4.
  
- 18 • Conduct study evaluations for individual epidemiological and toxicological studies  
19 (evaluating reporting quality, risk of bias, and sensitivity) and PBPK (scientific and technical  
20 review). The evaluation of epidemiology studies will specifically consider, to the extent  
21 possible, the likelihood and impact of potential confounding by other PFAS (see  
22 Section 6.2.1).
  
- 23 • Extract data on relevant health outcomes from epidemiological and toxicological studies of  
24 *high*, *medium*, and *low* confidence based on the study evaluations (full data extraction of low  
25 confidence studies may not be performed for poorly studied health effects or for health  
26 effects on which extensive *medium* and *high* confidence studies exist in the evidence base).
  
- 27 • Synthesize the evidence across studies, assessing similar health outcomes using a narrative  
28 approach. To inform future comparisons across a range of PFAS structures and properties  
29 (e.g., using high throughput screening, computational toxicology approaches, and chemical  
30 informatics to fill in data gaps; see [EPA PFAS action plan](#)), each of the five PFAS assessments  
31 will synthesize the available evidence (or lack thereof) for developmental effects; endocrine  
32 (primarily thyroid hormone) effects; hepatic effects, including lipid and lipoprotein  
33 measures (the latter of which are also applicable to interpreting the potential for  
34 cardiovascular toxicity); immune effects; reproductive effects in males or females; urinary  
35 effects; general toxicity; and carcinogenicity. Some assessments may include additional  
36 evidence syntheses for other health effects. The toxicological relevance of changes in some  
37 urinary and hepatic outcomes will be a point of focus in the evidence syntheses (see  
38 Section 9.2.3).

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- 1       • For each health outcome (or grouping of outcomes), evaluate the strength of evidence  
2       across studies (or subsets of studies) separately for studies of exposed humans and for  
3       animal studies. Based on the focused mechanistic analyses specific to each PFAS  
4       assessment (see Section 9.2), the mechanistic evidence will be used to inform evaluations of  
5       the available health effects evidence (or lack thereof).
  
- 6       • For each health outcome (or grouping of outcomes), develop an integrated expert judgment  
7       across evidence streams as to whether the evidence is sufficient (or insufficient) to indicate  
8       that exposure to the PFAS has the potential to be hazardous to humans (in rare instances,  
9       the evidence may be judged as sufficient to indicate that a hazard is unlikely). The judgment  
10      will be directly informed by the evidence syntheses and based on structured review of an  
11      adapted set of considerations for causality first introduced by Austin Bradford Hill [[Hill](#)  
12      (1965); see Sections 9 and 10], including consideration (e.g., based on available mechanistic  
13      information) and discussion of biological understanding. As part of the evidence  
14      integration narrative, characterize the strength of evidence for the available database of  
15      studies and its uncertainties, and identify and discuss issues concerning potentially  
16      susceptible populations and lifestages.
  
- 17      • Derive toxicity values (e.g., oral reference doses [RfDs], inhalation reference concentrations  
18      [RfCs], cancer risk estimates) as supported by the available data (see Section 10.2). Apply  
19      toxicokinetic and dosimetry modeling (possibly including PBPK modeling) to account for  
20      interspecies differences, as appropriate. Given the apparent species and sex differences in  
21      the toxicokinetic profile of the different PFAS (see Section 2.4), methods to address these  
22      potential differences will be a key consideration (see Section 9.2.1).
  
- 23      • Characterize uncertainties and identify key data gaps and research needs across each PFAS  
24      database, such as limitations of the available evidence, limitations of the systematic review,  
25      and consideration of dose relevance and toxicokinetic differences when extrapolating  
26      findings from higher dose animal studies to lower levels of human exposure.

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### **3.2. POPULATIONS, EXPOSURES, COMPARATORS, AND OUTCOMES (PECO) CRITERIA**

27           The PECO criteria are used to identify the evidence that addresses the specific aims of the  
28      assessment and to focus the literature screening, including study inclusion/exclusion, in a  
29      systematic review (see details on literature screening in Section 4.2). Given the expected lack of  
30      studies on carcinogenicity for these PFAS based on the preliminary literature inventory,  
31      genotoxicity studies were included in the PECO criteria (see Table 3-1).

32           In addition to those studies meeting the PECO criteria, studies containing supplemental  
33      material that are potentially relevant to the specific aims of the assessment were tracked during the  
34      literature screening process. Although these studies did not meet PECO criteria, they were not  
35      excluded from further consideration. The categories used to track studies as “potentially relevant  
36      supplemental material” are also described in Section 4.2.

**Table 3-1. Populations, exposures, comparators, and outcomes (PECO) criteria**

<b>PECO element</b>	<b>Evidence</b>
<u>Populations</u>	<p>Human: Any population and lifestage (occupational or general population, including children and other sensitive populations). The following study designs will be included: controlled exposure, cohort, case-control, and cross-sectional. (Note: Case reports and case series will be tracked as potential supplemental material.)</p> <p>Animal: Nonhuman mammalian animal species (whole organism) of any lifestage (including preconception, in utero, lactation, peripubertal, and adult stages).</p> <p>Other: In vitro, in silico, or nonmammalian models of genotoxicity. (Note: Other in vitro, in silico, or nonmammalian models will be tracked as potential supplemental material.)</p>
<u>Exposures</u>	<p>Human: Studies providing quantitative estimates of PFAS exposure based on administered dose or concentration, biomonitoring data (e.g., urine, blood, or other specimens), environmental or occupational-setting measures (e.g., water levels or air concentrations, residential location and/or duration, job title, or work title). (Note: Studies that provide qualitative, but not quantitative, estimates of exposure will be tracked as supplemental material.)</p> <p>Animal: Oral or Inhalation studies including quantified exposure to a PFAS of interest based on administered dose, dietary level, or concentration. (Note: Nonoral and noninhalation studies will be tracked as potential supplemental material.) PFAS mixture studies are included if they employ an experimental arm that involves exposure to a single PFAS of interest. (Note: Other PFAS mixture studies are tracked as potential supplemental material.)</p> <p>Studies must address exposure to one or more of the following: PFDA (CASRN 335-76-2), PFDA ammonia salt (CASRN 3108-42-7), PFDA sodium salt (CASRN 3830-45-3), PFNA (CASRN 375-95-1), PFNA ammonium salt (CASRN 4149-60-4), PFNA sodium salt (CASRN 21049-39-8), PFHxA (CASRN 307-24-4), PFHxA sodium salt (CASRN 2923-26-4), PFHxA ammonium salt (CASRN 21615-47-4), PFHxS (CASRN 355-46-4), PFHxS potassium salt (CASRN 3871-99-6), PFBA (CASRN 375-22-4), or PFBA ammonium salt (CASRN 10495-86-0). [Note: although while these PFAS are not metabolized or transformed in the body, there are precursor compounds known to be biotransformed to a PFAS of interest; for example, 6:2 fluorotelomer alcohol is metabolized to PFHxA and PFBA (<a href="#">Russell et al., 2015</a>). Thus, studies of precursor PFAS that identify and quantify a PFAS of interest will be tracked as potential supplemental material (e.g., for ADME analyses or interpretations).]</p>
<u>Comparators</u>	<p>Human: A comparison or reference population exposed to lower levels (or no exposure/exposure below detection levels) or for shorter periods of time.</p> <p>Animal: Includes comparisons to historical controls or a concurrent control group that is unexposed, exposed to vehicle-only or air-only exposures. (Note: Experiments including exposure to PFAS across different durations or exposure levels without including one of these control groups will be tracked as potential supplemental material [e.g., for evaluating key science issues; Section 2.4].)</p>
<u>Outcomes</u>	<p>All cancer and noncancer health outcomes. (Note: Other than genotoxicity studies, studies including only molecular endpoints [e.g., gene or protein changes; receptor binding or activation] or other nonphenotypic endpoints addressing the potential biological or chemical</p>

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<b>PECO element</b>	<b>Evidence</b>
	progression of events contributing towards toxic effects will be tracked as potential supplemental material [e.g., for evaluating key science issues; Section 2.4].)
PBPK models	Studies describing physiologically based pharmacokinetic (PBPK) and other PK models for PFDA (CASRN 335-76-2), PFDA ammonia salt (CASRN 3108-42-7), PFDA sodium salt (CASRN 3830-45-3), PFNA (CASRN 375-95-1), PFNA ammonium salt (CASRN 4149-60-4), PFNA sodium salt (CASRN 21049-39-8), PFHxA (CASRN 307-24-4), PFHxS (CASRN 355-46-4), PFHxS potassium salt (CASRN 3871-99-6), PFBA (CASRN 375-22-4), or PFBA ammonium salt (CASRN 10495-86-0).

ADME = absorption, distribution, metabolism, and excretion; PK = pharmacokinetic.

## 4. LITERATURE SEARCH AND SCREENING STRATEGIES

1           The initial literature search was completed in July 2017 as part of a cross-Environmental  
2 Protection Agency (EPA) workgroup that focused on a large set of PFAS, including the five PFAS  
3 addressed in this protocol. Subsequent literature searches were refined and are being updated  
4 regularly. In an effort to ensure that all pertinent studies were captured, the studies identified as  
5 relevant to PFBA, PFHxA, PFHxS, PFNA, and PFDA are being shared simultaneously with release of  
6 this protocol.<sup>5</sup> These search efforts reflect studies published through February 2018, several  
7 unpublished reports, and a few recent studies not yet identified through the formal literature  
8 updating process; the literature is currently being updated and will be updated regularly in the  
9 context of the five PFAS assessments (in their respective U.S. EPA Health and Environmental  
10 Research Online [HERO] pages) until several months before public release of the draft  
11 assessments.<sup>6</sup> Accordingly, the methods for literature search and screening (as well as some of the  
12 approaches to refining the evaluation plan based on the identified literature; see Section 5) are  
13 described in the protocol using the past tense, whereas approaches for the other assessment  
14 methods are outlined using the future tense.

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### 4.1. LITERATURE SEARCH STRATEGIES

15           The initial literature search strategy performed in July 2017 was designed to identify a  
16 broad range of topics relevant to PFAS, including studies on physicochemical properties,  
17 environmental fate and occurrences, human exposures, and biological effects representative of all  
18 types of evidence (i.e., human, animal, in vitro, in silico) and health outcomes. PFAS search terms  
19 included PFAS names (including salt, cationic, and anionic forms), all known synonyms, and CAS  
20 registry numbers. The literature search itself encompassed a non-date-limited query of the  
21 following databases:

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<sup>5</sup>PFBA: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2632](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2632)  
PFHxA: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2628](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2628)  
PFHxS: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2630](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2630)  
PFNA: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2633](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2633)  
PFDA: [https://hero.epa.gov/hero/index.cfm/project/page/project\\_id/2614](https://hero.epa.gov/hero/index.cfm/project/page/project_id/2614).

<sup>6</sup>Although not identified as part of the formal literature searches through early 2019, several more recent PBPK studies found through regular monitoring of new studies are included in this protocol (see Section 6.4) so that the process for evaluating those data can be outlined.

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- 1 • PubMed ([National Library of Medicine](#))
- 2 • Web of Science ([Thomson Reuters](#))
- 3 • Toxline ([National Library of Medicine](#))
- 4 • TSCATS ([Toxic Substances Control Act Test Submissions](#))

5 All literature identified in the initial search was loaded into the HERO database. In  
6 February 2018, the literature search was updated for the PFAS in this assessment (i.e., PFBA,  
7 PFHxA, PFHxS, PFNA, and PFDA). The updated literature query included all PFAS nomenclature  
8 from the initial search as well as a broader non-date-limited search of several new PFAS synonyms  
9 that were identified after the original search. This updated search was conducted by EPA's HERO  
10 tool to search the same databases that were included in the initial literature query.

11 Because each database has its own search architecture, the resulting search strategy was  
12 tailored to account for each database's unique search functionality. Full details of the July 2017 and  
13 February 2018 search strategies are presented in Addendum B. No literature was restricted by  
14 language.

15 Additional relevant literature not found through database searching was identified by:

- 16 • Review of studies cited in state, national (EPA, Food and Drug Administration [FDA], etc.),  
17 and international (International Agency for Research on Cancer [IARC], World Health  
18 Organization [WHO], European Chemicals Agency [ECHA], etc.) assessments on these five  
19 PFAS, including parallel assessment efforts in progress (e.g., the draft Agency for Toxic  
20 Substances and Disease Registry [ATSDR] assessment released publicly in 2018).
- 21 • Review of studies submitted to federal regulatory agencies and brought to the attention of  
22 EPA. For example, studies submitted to EPA by the manufacturers of these five PFAS in  
23 support of requirements under the Toxic Substances Control Act (TSCA). Such studies (or  
24 data summaries) will only be tracked in the literature flow diagrams released with each of  
25 the five assessments (and considered for inclusion in the assessment) when they can be  
26 made publicly available. To facilitate the timely completion of these assessments, if  
27 attempts to acquire a publicly accessible version of an identified study are unsuccessful  
28 after 3 months of the initial request, these studies will be considered unobtainable and will  
29 not be considered for inclusion in the assessment(s).
- 30 • Identification of studies during screening for other PFAS. For example, epidemiology  
31 studies relevant to more than one of these five PFAS were sometimes identified by searches  
32 focused on one PFAS, but not the others.
- 33 • Other gray literature (i.e., primary studies not indexed in typical databases, such as  
34 technical reports from government agencies or scientific research groups; unpublished  
35 laboratory studies conducted by industry; or working reports/white papers from research  
36 groups or committees) brought to the attention of EPA during problem formulation,

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1 engagement with technical PFAS experts, and during future solicitation of Agency,  
2 interagency, and public comment during the Integrated Risk Information System (IRIS)  
3 assessment development and review process. For example, one such study was brought to  
4 the attention of EPA on March 29, 2018 by the National Toxicology Program (NTP) when  
5 NTP published study tables and individual animal data from a 28-day toxicity study of  
6 multiple PFAS ([NTP, 2011](#)). A peer-reviewed NTP Technical Report was not yet available at  
7 the time this protocol was developed,<sup>7</sup> but these data have undergone standard NTP quality  
8 assurance/control processing and are publicly available, and a protocol outlining the NTP  
9 study methods is available in HERO  
10 ([https://hero.epa.gov/hero/index.cfm/reference/details/reference\\_id/4309741](https://hero.epa.gov/hero/index.cfm/reference/details/reference_id/4309741)).

11 The number of studies on PFBA, PFHxA, PFHxS, PFNA, and PFDA returned from the  
12 literature searches through February 2018 is documented in the literature flow diagrams in  
13 Figure 4-1, which also reflect the literature screening decisions (see Section 4.2). Notably, the  
14 identification and review of records submitted to EPA, which may include confidential business  
15 information (CBI), is ongoing. This includes exploring the possibility of making the data within any  
16 identified records publicly available. Any records included in the assessments will be reflected in  
17 updates to the protocol and in the draft assessments. In addition, any identified companion  
18 documents for the included studies, such as retractions, corrections and supplemental materials,  
19 will also be included, and the assessments will incorporate the most recent publication materials  
20 (note: these are tracked as separate, “included” records in the literature flow diagrams [see  
21 Section 4.2.2] and HERO; companion documents in other screening categories such as “excluded,”  
22 which are not relevant to the target PECO, are similarly tagged as separate records within that  
23 screening category).

24 The literature searches will be updated throughout the assessments’ development and  
25 review process to identify newly published literature. The last full literature search update will be  
26 conducted prior to (several months) the planned release of the draft document for public comment.  
27 The literature flow diagrams (see Section 4.2.2) presented in the assessment will be revised to  
28 reflect these updates. Although uncommon, it is possible that additional literature searches may be  
29 performed during assessment development and review (e.g., to supplement an analysis of a specific  
30 mechanism or biological linkage). Any such ancillary searches will be documented in updates to the  
31 protocol.

32 The IRIS Program takes extra steps to ensure identification of pertinent studies by  
33 encouraging the scientific community and the public to identify additional studies and ongoing  
34 research; by searching for publicly available data submitted under the TSCA and the Federal  
35 Insecticide, Fungicide, and Rodenticide Act (FIFRA); and by considering late-breaking studies that  
36 would affect the credibility of the conclusions, even during the review process. Studies identified

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<sup>7</sup>The 28-day oral studies of PFDA, PFNA, PFHxA, and PFHxS in male and female rats are now final (updated after public comments received). The published research reports will be cited in the assessments and are available on HERO; however, the protocol discusses these reports in relation to their unpublished version.

1 after peer review begins will only be considered for inclusion if they meet the PECO criteria and are  
2 expected to fundamentally alter the assessment’s conclusions. Release of the PECO-screened  
3 literature in parallel with release of the protocol for public comment provides an opportunity for  
4 stakeholders to identify any missing studies, which if identified, will be screened as outlined above  
5 for adherence to the PECO criteria.

#### **4.1.1. Non-Peer-Reviewed Data**

6 IRIS assessments rely mainly on publicly accessible, peer-reviewed studies. However, it is  
7 possible that gray literature (i.e., studies that are not reported in the peer-reviewed literature)  
8 directly relevant to the PECO may be identified during assessment development (e.g., good  
9 laboratory practice [GLP] studies submitted to EPA, dissertations, etc.). In this case, if the data  
10 make a substantial impact on assessment decisions or conclusions (i.e., have potential to affect the  
11 PECO statement, hazard conclusions, or dose-response analysis), EPA can obtain external peer  
12 review if the owners of the data are willing to have the study details and results made publicly  
13 accessible. This independent, contractor-driven peer review would include an evaluation of the  
14 study, as is done for peer review of a journal publication. The contractor would identify and select  
15 two to three scientists knowledgeable in scientific disciplines relevant to the topic as potential peer  
16 reviewers. Persons invited to serve as peer reviewers would be screened for conflict of interest  
17 before confirming their service. In most instances, the peer review would be conducted by letter  
18 review. The study authors would be informed of the outcome of the peer review and given an  
19 opportunity to clarify issues or provide missing details. The study and its related information, if  
20 used in the IRIS assessment, would become publicly available. In the assessment, EPA would  
21 acknowledge that the document underwent external peer review managed by EPA, and the names  
22 of the peer reviewers would be identified. In certain cases, IRIS will conduct an assessment for  
23 utility and data analysis based on having access to a description of study methods and raw data that  
24 have undergone rigorous quality assurance/quality control review (e.g., ToxCast/Tox21 data;  
25 results of National Toxicology Program [NTP] studies) but that have not yet undergone external  
26 peer-review.

27 Unpublished (e.g., raw) data from personal author communication can supplement a  
28 peer-reviewed study, if that information is made publicly available. If such ancillary information is  
29 acquired, it will be documented in either the Health Assessment Workspace Collaborative (HAWC)  
30 or HERO project page for the PFAS being assessed (depending on the nature of the information  
31 received).

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## **4.2. SCREENING PROCESS**

32 As described below, PECO criteria or predefined inclusion and exclusion criteria (i.e., the  
33 latter were used for the initial search) were used by two independent reviewers to screen and  
34 inventory studies at the title and abstract level. For those studies considered relevant at the title

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1 and abstract level, these criteria were then used to determine inclusion or exclusion of a reference  
2 based on the full text. In addition to the PECO criteria, the following exclusion criteria were  
3 applied:

- 4 • Review, commentary, other agency assessment, letter, or other record that does not contain  
5 original data (note that these records were tracked for potential use in identifying  
6 study-specific, original data relevant to specific scientific questions during assessment  
7 development, including scanning of reference lists for unidentified studies; any such studies  
8 incorporated into the assessment will be tracked under “other” as the reference source in  
9 updates to the protocol)
- 10 • Study available only as an abstract (e.g., conference abstract)
- 11 • Full text of the study is not available, and screening decisions could not be made at the  
12 title/abstract level

13 In addition to including studies that meet PECO criteria, other studies containing material  
14 that is potentially relevant to the assessments’ objectives and specific aims were tracked during the  
15 screening process as “potentially relevant supplemental material.” Importantly, these studies were  
16 not excluded, but they may not be incorporated into the assessments unless they are deemed to be  
17 relevant to addressing the key science issues, specific aims (see Sections 2.4 and 3.1), or key  
18 scientific uncertainties identified at later stages of assessment development (see Section 9). Studies  
19 categorized as “potentially relevant supplemental material” include the following:

- 20 • In vivo mechanistic or mode-of-action studies, including non-PECO routes of exposure and  
21 populations (e.g., nonmammalian models—generally, these are interpreted to be less  
22 directly relevant to evaluating the potential for human disease, although exceptions do exist  
23 for some endpoints)
- 24 • In vitro and in silico models
- 25 • ADME and toxicokinetic studies (excluding models)<sup>8</sup>
- 26 • Exposure assessment or characterization (no health outcome) studies
- 27 • PFAS mixture studies (no individual PFAS comparisons)
- 28 • Human case reports or case-series studies

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<sup>8</sup>Given the known importance of ADME data, this supplemental tagging was used as the starting point for a separate screening and review of toxicokinetics data (see Section 9.2.1 for details on the PECO and screening process for this separate literature identification effort).

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- 1 • Ecotoxicity studies
- 2 • Studies on PFAS manufacture/use
- 3 • Treatment/remediation studies
- 4 • Studies of PFAS analysis or other laboratory methods
- 5 • Environmental fate and transport studies
- 6 • Studies of other PFAS

7 Several of these categories of studies were further screened for consideration in addressing  
8 the key science issues (described in Section 9.2).

9 *Title and abstract screening.* Following a pilot phase to calibrate screening guidance, two  
10 screeners independently performed a title and abstract screen using a structured form in  
11 DistillerSR (Evidence Partners; [https://distillercer.com/products/distillersr-systematic-review-](https://distillercer.com/products/distillersr-systematic-review-software/)  
12 [software/](https://distillercer.com/products/distillersr-systematic-review-software/)). For citations with no abstract, the article was excluded if screening decisions could not  
13 be made based on the title and other citation information (e.g., page length) and if additional  
14 attempts to acquire the abstract or full text were unsuccessful. Screening conflicts were resolved  
15 by discussion between the primary screeners, with consultation by a third reviewer or technical  
16 advisor (if needed) to resolve any remaining disagreements. Eligibility status of non-English  
17 studies was assessed using the same approach with online translation tools used as needed to  
18 evaluate portions of the study text and assess eligibility at the title and abstract level.

19 Studies not meeting title/abstract criteria but identified as “potentially relevant  
20 supplemental material” were categorized (i.e., tagged) during the title and abstract screening  
21 process (further described in Section 4.3). Conflict resolution was not required during the  
22 screening process to identify supplemental information (i.e., tagging by a single screener was  
23 considered adequate to identify the study as potentially relevant supplemental material for possible  
24 inclusion during draft development).

25 Before beginning the Integrated Risk Information System (IRIS) PFAS assessments project,  
26 the EPA contractor that conducted the July 2017 literature search as part of an EPA-wide  
27 workgroup had performed a title and abstract screen to bin studies into different categories  
28 (e.g., human, in vivo animal, excluded). At the initiation of these PFAS assessments within IRIS, a  
29 formalized effort was deployed with a new title and abstract screen of all studies identified in the  
30 initial July 2017 search based on the PECO criteria in Table 3-1. For this initial literature screening,  
31 specific inclusion/exclusion criteria were applied in the formalized title and abstract screen (see  
32 Addendum B, Table B-6). Title and abstract screening of studies identified during literature search  
33 updates will be conducted using the PECO criteria in Table 3-1 in DistillerSR using forms that  
34 facilitate simultaneous initial tagging during screening (e.g., category of supplemental data;

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1 contains data on other PFAS of interest). An example of the questions and answers populating the  
2 DistillerSR form for title/abstract and full-text (below) screening during literature search updates  
3 is provided in Addendum B, Table B-7.

4 *Full-text screening.* Records that were not excluded based on the title and abstract were  
5 advanced to full-text review. Full-text copies of these potentially relevant records were retrieved,  
6 stored in the HERO database, and independently assessed by two screeners using a structured form  
7 in DistillerSR to confirm eligibility. Screening conflicts were resolved by discussion among the  
8 primary screeners with consultation by a third reviewer or technical advisor (as needed to resolve  
9 any remaining disagreements). As with the title and abstract screening, some studies were also  
10 identified as “potentially relevant supplemental material” based on full-text screening. Approaches  
11 for language translation included engagement of a native speaker from within EPA or use of  
12 fee-based translation services.

13 In addition to identifying studies as included, excluded, or potential supplemental material,  
14 the reviewers used the DistillerSR screening forms to confirm the specific PFAS (or multiple PFAS)  
15 evaluated and to document several important experimental features of the studies (see Section 4.3).

16 The results of this screening process are documented in the HERO database  
17 (<https://hero.epa.gov>; see Section 4.1 for links to the specific HERO pages) and literature flow  
18 diagrams (see Figure 4-1), with individual studies “tagged” in HERO according to their appropriate  
19 category descriptors (e.g., reference source; screening decisions regarding inclusion, exclusion, or  
20 identification as supplemental; type of study).

### **4.2.1. Multiple Publications of the Same Data**

21 When there are multiple publications using the same or overlapping data, all publications  
22 on the research were included, with one selected for use as the primary study; the others were  
23 considered as secondary publications with annotation in HAWC indicating their relationship to the  
24 primary record during data extraction. For epidemiology studies, the primary publication was  
25 generally the one with the longest follow-up, the largest number of cases, or the most recent  
26 publication date. For animal studies, the primary publication was typically the one with the most  
27 recent publication date, longest duration of exposure, or the one that assessed the outcome(s) most  
28 informative to the PECO. For both epidemiology and animal studies, the assessments will include  
29 relevant data from all publications of the study, although if the same data are reported in more than  
30 one study, the data will only be extracted once (see Section 8). For corrections, retractions, and  
31 other companion documents to the included publications, a similar approach to annotation was  
32 taken (see Section 4.1), and the most recently published data will be incorporated in the  
33 assessments.

#### **4.2.2. Literature Flow Diagrams**

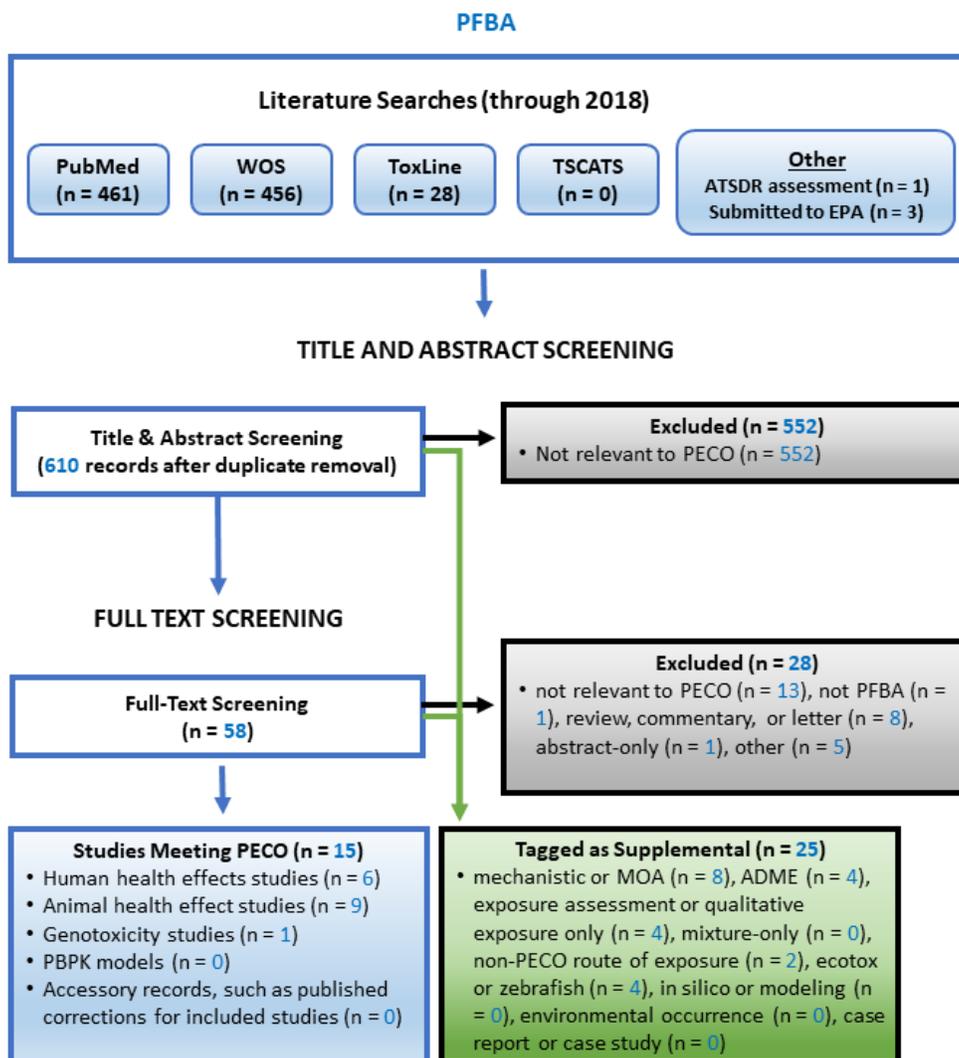
1            Figure 4-1 presents the literature flow diagrams for PFBA (a), PFHxA (b), PFHxS (c), PFNA  
2 (d), and PFDA (e).<sup>9</sup> These figures reflect literature searches through 2018. A literature search  
3 update has been conducted and the results will be reflected in the draft assessments (and the most  
4 current results can be viewed at any time in the HERO project pages provided in Section 4.1). Note  
5 that the potential for updates or revisions to these figures related to CBI data and other reference  
6 decisions is discussed in the previous sections.

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<sup>9</sup>Note that the literature searches included the associated salts for each of the five PFAS, as presented in Figure 2-1 (see Section 2.1.1). In addition, although not identified (yet) as part of the formal literature searches and not included in these diagrams, several recent PBPK studies found through regular monitoring of new studies are included in this protocol (see Section 6.4) so that the process for evaluating those studies can be outlined.

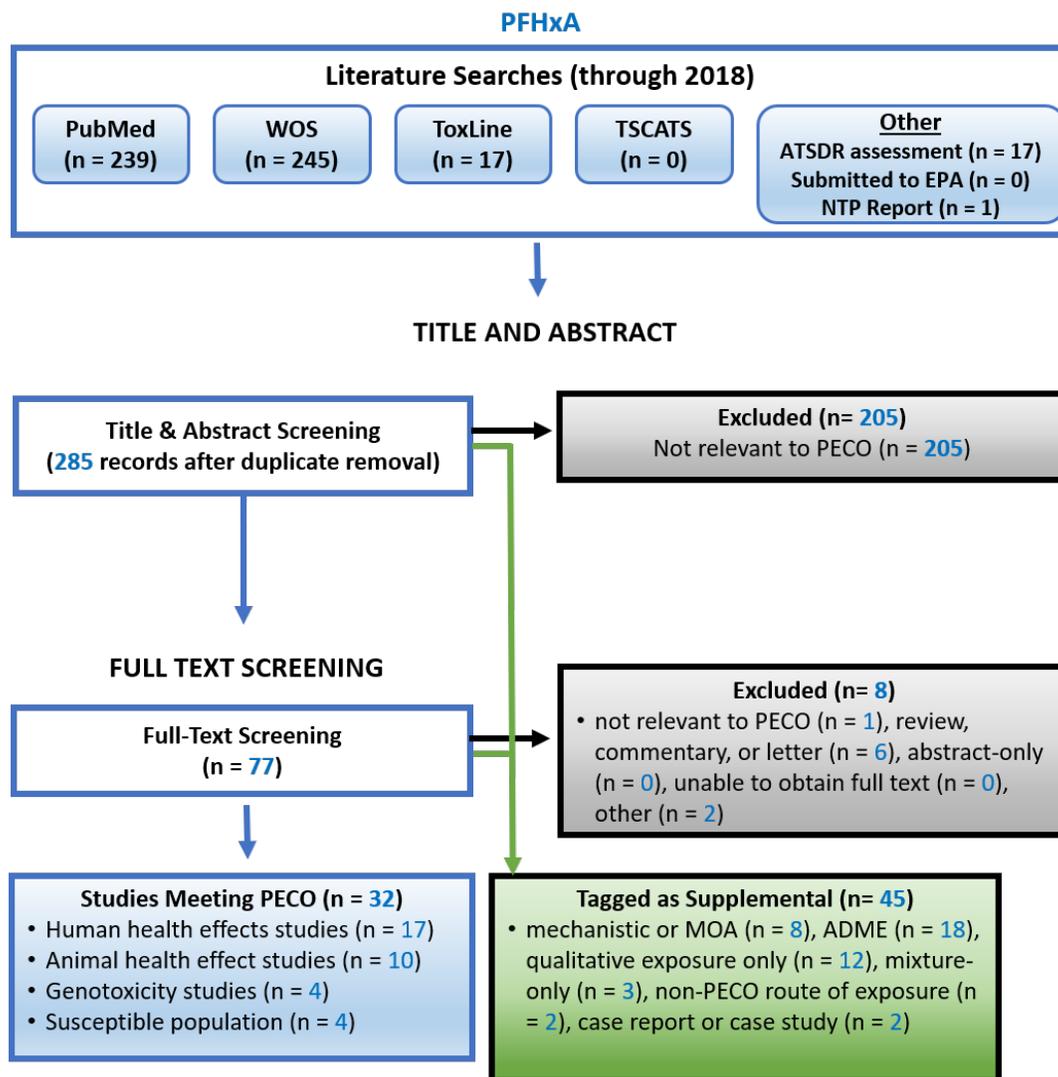
*Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments*

(a)



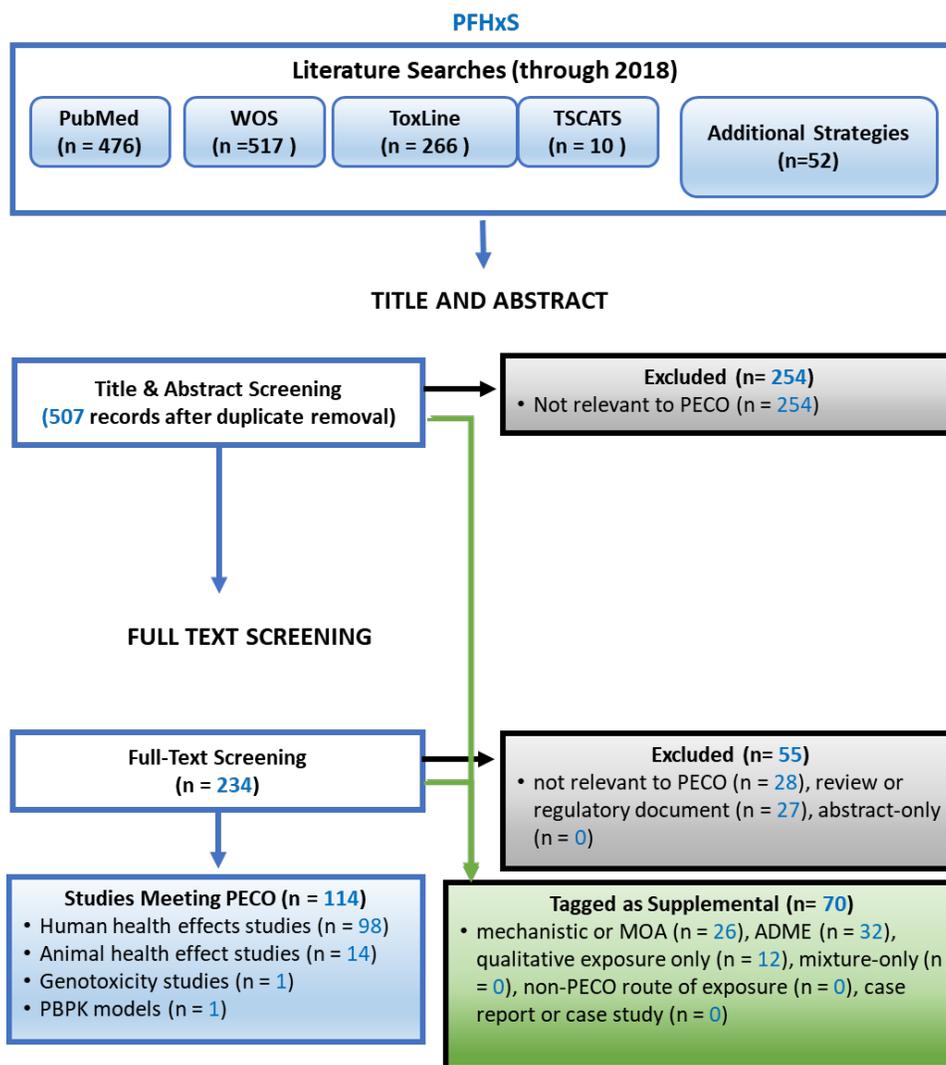
*Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments*

(b)



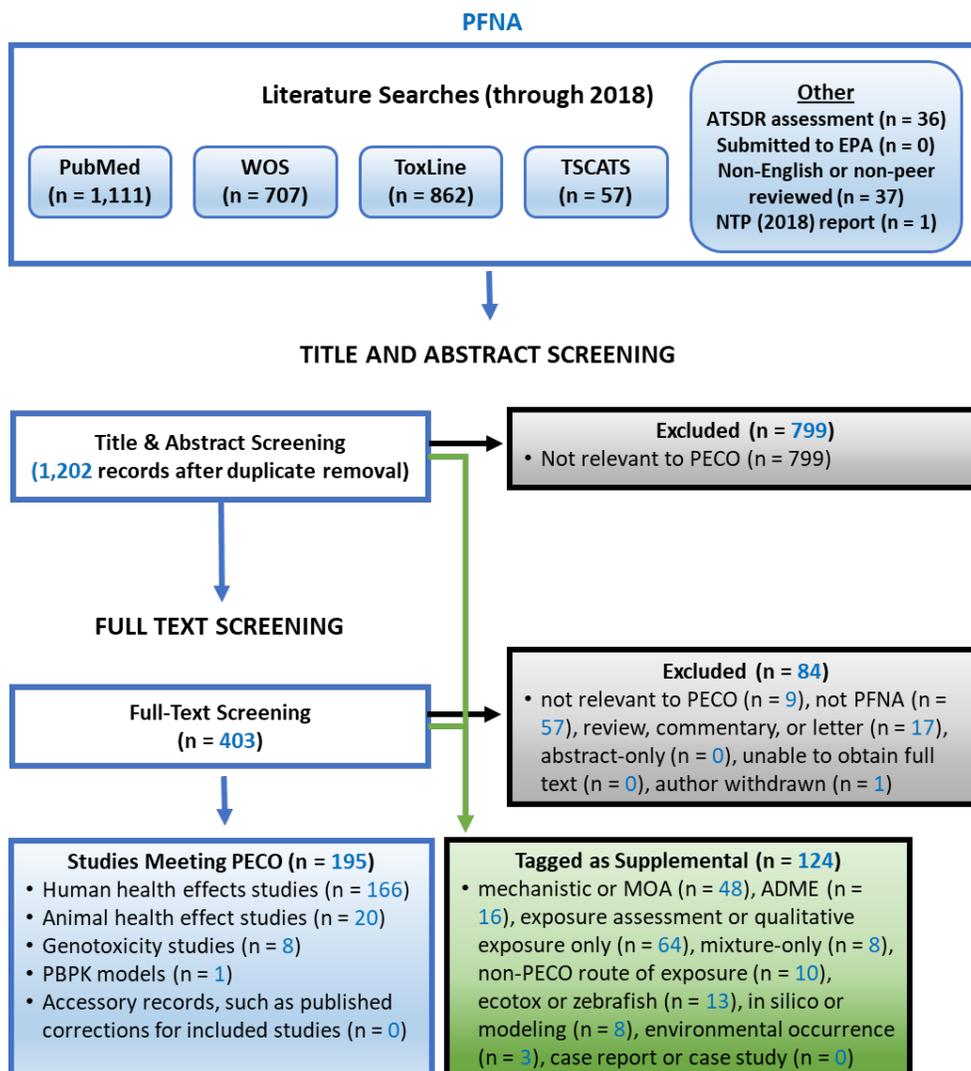
*Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments*

(c)



**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

(d)



(e)

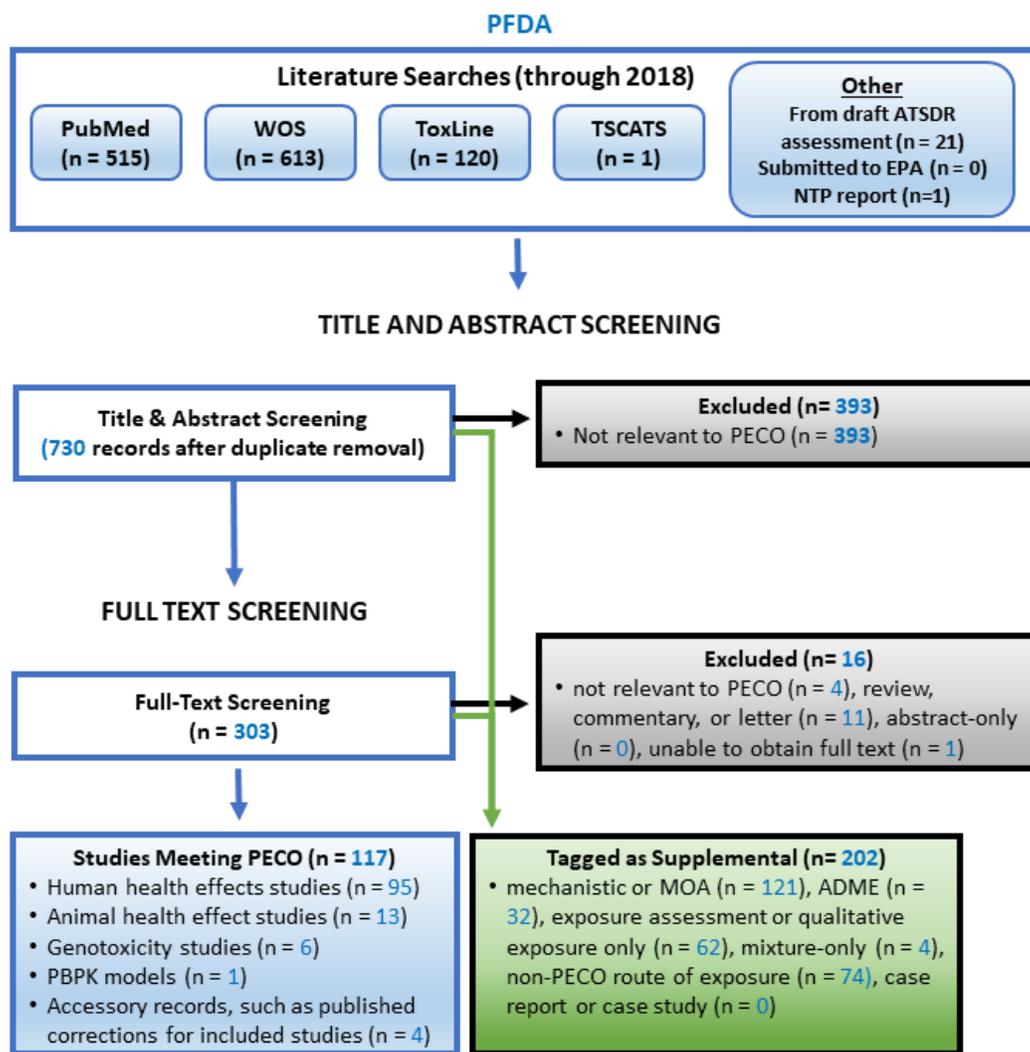


Figure 4-1. Literature flow diagrams for PFBA and its ammonium salt (a), PFHxA and its ammonium and sodium salts (b), PFHxS and its potassium salt (c), PFNA and its ammonium and sodium salts (d), and PFDA and its ammonium and sodium salts (e).

### 4.3. SUMMARY-LEVEL LITERATURE INVENTORIES

1 As noted in Section 4.2, during title/abstract or full-text level screening, studies tagged  
 2 based on PECO eligibility were further categorized based on features such as evidence type (human,  
 3 animal, mechanistic, PBPK, etc.), health outcome(s), and/or endpoint measure(s) included in the  
 4 study, and the specific PFAS (or multiple PFAS) addressed (see Addendum B, Table B-7 for  
 5 examples). Literature inventories for PECO-relevant studies were created to develop

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1 summary-level, sortable lists that include some basic study design information (e.g., study  
2 population, exposure information such as doses administered or biomarkers analyzed,  
3 age/lifestage<sup>10</sup> of exposure, endpoints examined, etc.). These working literature inventories are for  
4 internal use and facilitate subsequent review of individual studies or sets of studies by  
5 topic-specific experts.

6 Inventories were also created for studies that were tagged as “potentially relevant  
7 supplemental material” during screening, including in vitro or in silico models not addressing  
8 genotoxicity, ADME studies, and studies on endpoints or routes of exposure that did not meet the  
9 specific PECO criteria, but which may still be relevant to the research question(s). Here, the  
10 objective was to create an inventory of studies that can be tracked and further summarized as  
11 needed—for example, by model system, key characteristic [e.g., of carcinogens, [Smith et al. \(2016\)](#)],  
12 mechanistic endpoint, or key event—to support analyses of potentially critical mechanistic  
13 questions that arise at various stages of the systematic review (see Section 9.2 for a description of  
14 the process for determining the specific questions and pertinent mechanistic studies to be  
15 analyzed). For example, ADME data and related information are important to the next steps of  
16 evaluating the evidence from individual PECO-specific studies, and these data will be reviewed by  
17 subject matter experts early in the assessment process. Thus, the comprehensive identification of  
18 studies relevant to interpreting the ADME or toxicokinetic characteristics of these PFAS was  
19 prioritized (see additional discussion in Section 5, and the specifics of the approach in Section 9.2).

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<sup>10</sup>Age/lifestage was considered according to EPA’s [Guidance on Selecting Age Groups for Monitoring and Assessing Childhood Exposures to Environmental Contaminants](#) and EPA’s [A Framework for Assessing Health Risk of Environmental Exposures to Children](#).

## 5. REFINED EVALUATION PLAN

1           The primary purpose of this step is to outline any potential or expected refinements to the  
2 set of populations, exposures, comparators, and outcomes (PECO)-relevant studies that would  
3 narrow the scope of studies considered for use in evidence synthesis and beyond. This optional  
4 step is typically applied to focus an assessment with a very large number of PECO-relevant studies  
5 on review of the most informative evidence (e.g., when many studies examine the same health  
6 outcome, focusing on toxicity studies including exposures below a specified range, those studies  
7 examining more specific or objective measures of toxicity, or those that address lifestage- or  
8 exposure duration-specific knowledge on how the health outcome develops). Given the relatively  
9 small databases of animal toxicological studies for these five PFAS (see Section 2.3.2), this  
10 narrowing is not considered applicable to these data. Thus, for these five PFAS assessments, all  
11 relevant health outcomes in the animal toxicological studies meeting PECO criteria will be  
12 considered.

13           In contrast to the animal studies, there are many epidemiology studies. To make the  
14 systematic review of the epidemiology literature more pragmatic and efficient and focus the set of  
15 studies undergoing study evaluation, one epidemiologist per outcome performed an initial review  
16 of the available evidence examining at a high level the direction and consistency of observed  
17 associations. Based on this initial review, outcomes were classified into one of three tiers:  
18 (1) formal systematic review, (2) rapid review (reduced rigor; study evaluation with a single  
19 reviewer), or (3) no further review (no study evaluation or synthesis of the evidence, although the  
20 available database might be mentioned in the assessments to inform data gaps). Most outcomes  
21 were classified into the first tier (formal systematic review). Outcomes with an a priori serious  
22 concern for reverse causality (e.g., clear link to elimination of PFAS from the body, such as  
23 outcomes related to menstruation or renal function) were classified into the second tier (rapid  
24 review) because of the large amount of uncertainty in interpreting these results. These outcomes  
25 included renal function (e.g., glomerular filtration rate), menstrual cycle characteristics,  
26 endometriosis, polycystic ovary syndrome, and albumin. The third tier (no further review) was  
27 used primarily for outcomes where the results for available studies were null and the study  
28 sensitivity was poor, due to, for example, PFAS exposure levels being below or near the LOD. This  
29 included penile width, sex ratio, hematologic effects, and mortality. For the second and third tiers,  
30 it is possible that new data could change their classification. The outcomes included in the  
31 assessment are summarized in Table 5-1, which also indicates those undergoing only rapid review.

32           This approach of tiered reviews is consistent with recommendations from the National  
33 Academies of Science encouraging the U.S. Environmental Protection Agency (EPA) to explore ways  
34 to make systematic review more feasible, including conducting a “rapid review in which

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- 1 components of the systematic review process are simplified or omitted (e.g., the need for two
- 2 independent reviewers)” ([NASEM, 2017](#)).

**Table 5-1. Epidemiology outcome grouping categories**

Relevant human health effect category <sup>a,b</sup>	Examples of epidemiology outcomes included
<b>Hepatic</b> (toxicity)	<ul style="list-style-type: none"> <li>• Serum liver enzymes (e.g., ALT, AST, total bilirubin from clinical chemistry)</li> <li>• Liver disease</li> </ul>
<b>Cardiovascular</b> (toxicity)	<ul style="list-style-type: none"> <li>• Serum lipids (note: also, informative to hepatic)</li> <li>• Blood pressure</li> <li>• Atherosclerosis</li> <li>• Cardiovascular disease</li> <li>• Ventricular geometry</li> </ul>
<b>Immune</b> (effects)	<ul style="list-style-type: none"> <li>• Antibody response</li> <li>• Hypersensitivity (asthma, allergy, atopic dermatitis)</li> <li>• Infections</li> </ul>
<b>Urinary</b> (toxicity)	<ul style="list-style-type: none"> <li>• Renal function tests (e.g., glomerular filtration rate) (RR)</li> </ul>
<b>Endocrine</b> (effects)	<ul style="list-style-type: none"> <li>• Thyroid hormones</li> <li>• Thyroid disease</li> </ul>
<b>Metabolic</b> (effects)	<ul style="list-style-type: none"> <li>• Diabetes</li> <li>• Gestational diabetes</li> <li>• Insulin resistance</li> <li>• Serum glucose</li> <li>• Adiposity (e.g., BMI, weight gain)</li> <li>• Metabolic syndrome</li> </ul>
<b>Reproductive</b> (toxicity) Note: Evidence synthesis and evidence integration conclusions in assessments are developed separately for male and female reproductive effects (toxicity)	<ul style="list-style-type: none"> <li>• Reproductive hormones</li> <li>• Fecundity</li> <li>• Semen parameters</li> <li>• Anogenital distance</li> </ul>

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Relevant human health effect category <sup>a,b</sup>	Examples of epidemiology outcomes included
	<ul style="list-style-type: none"> <li>• Female reproductive conditions (endometriosis, polycystic ovary syndrome) (RR)</li> <li>• Ovarian reserve</li> <li>• Menstrual cycle characteristics (RR)</li> <li>• Pubertal development</li> </ul>
<p><b>Developmental</b> (effects)            Note: Evidence synthesis of these endpoints in the assessments is termed “offspring growth and early development,” but evidence integration conclusions will be drawn on the broader category of “developmental effects” (which also considers organ/system-specific effects after exposure during development)</p>	<ul style="list-style-type: none"> <li>• Birth size/fetal growth restriction</li> <li>• Preterm birth/gestational duration</li> <li>• Postnatal growth</li> <li>• Spontaneous abortion</li> </ul>

RR = rapid review; ALT = alanine aminotransferase; AST = aspartate aminotransferase; BMI = body mass index.

<sup>a</sup>The primary focus of these assessments will be on developmental effects; endocrine effects; hepatic effects, including lipid and lipoprotein measures; immune effects; reproductive effects in males or females; urinary effects; general toxicity; and carcinogenicity. Data on cardiovascular effects, hematological effects, metabolic effects including diabetes, and nervous system effects will be summarized when available. These summaries may be developed in association with one of the health effects noted above, as a separate formal evaluation of hazard, or as part of a qualitative summary on “other effects,” depending on the assessment-specific data.

<sup>b</sup>Some outcomes are relevant to multiple health effects. These outcomes may be categorized under only a single health effect in Table 5-2 for clarity. However, in the assessments, such outcome data would be discussed in the first relevant health effect synthesis (syntheses will generally follow the pattern of most to least available evidence) and then this synthesis will be cited in the syntheses of other relevant health effects. The evidence (for or against an effect) will contribute to evidence integration decisions for all relevant health effects.

1           To promote consistency in evaluation and presentation across assessments, preliminary  
 2 decisions were made regarding the grouping of related endpoints for outcome-specific study  
 3 evaluations and discussion in the evidence synthesis. This helps implement the study evaluation  
 4 criteria (see Section 6) because those evaluations are outcome and analysis specific. Preliminary  
 5 decisions for grouping of endpoints from animal toxicological studies for discussion within each  
 6 assessed human health effect category are described in Table 5-2. Parallel groupings for outcomes  
 7 assessed in the available epidemiology studies are captured in Table 5-1. These groupings are  
 8 meant to serve as a starting place for consistency in presentation and analysis across studies and  
 9 assessments, although assessment-specific deviations are possible (e.g., depending on the  
 10 assessment-specific database of endpoints in the available studies or PFAS-specific understanding  
 11 of mechanistic relationships across outcomes).

**Table 5-2. Animal endpoint grouping categories**

Relevant human health effect category <sup>a</sup>	Examples of animal endpoints included	Notes
<b>General toxicity</b>	<ul style="list-style-type: none"> <li>• Body weight (not maternal or pup weights, or weights after developmental-only exposure)</li> <li>• Mortality, survival, or LD<sub>50</sub>S</li> <li>• Growth curve</li> <li>• Clinical observations (nonbehavioral)</li> </ul>	<ul style="list-style-type: none"> <li>• Clinical chemistry endpoints are under Hepatic or Hematologic</li> <li>• Maternal or pup body-weight endpoints are under Developmental</li> <li>• Pathology (including gross lesions) is organ specific</li> </ul>
<b>Hepatic (toxicity)<sup>b</sup></b>	<ul style="list-style-type: none"> <li>• Liver weight and histopathology</li> <li>• Serum or tissue liver enzymes (e.g., ALT and AST from clinical chemistry)</li> <li>• Other liver tissue enzyme activity (e.g., catalase) or protein/DNA content</li> <li>• Other liver tissue biochemical markers (e.g., albumin; glycogen; glucose)</li> <li>• Liver-specific serum biochemistry (e.g., albumin; albumin/globulin)</li> <li>• Liver tissue lipids: triglycerides, cholesterol</li> <li>• Serum lipids (Note: also, informative to cardiovascular<sup>c</sup>)</li> </ul>	<ul style="list-style-type: none"> <li>• Biochemical markers such as albumin or glucose are under Hematological</li> <li>• Liver tissue cytokines are under Immune</li> <li>• Serum glucose is under Metabolic</li> </ul>
<b>Cardiovascular (toxicity)<sup>b,d</sup></b>	<ul style="list-style-type: none"> <li>• Heart weight and histopathology</li> <li>• Serum lipids (note: also, informative to Hepatic)</li> <li>• Blood pressure</li> <li>• Blood measures of cardiovascular risk (e.g., C-reactive protein)</li> </ul>	<ul style="list-style-type: none"> <li>• Other blood measures are under Hepatic, Immune, or Hematologic</li> </ul>
<b>Hematologic (effects)<sup>b,d</sup></b>	<ul style="list-style-type: none"> <li>• Red blood cells</li> <li>• Blood hematocrit or hemoglobin</li> <li>• Corpuscular volume</li> <li>• Blood platelets or reticulocytes</li> <li>• Blood biochemical measures (e.g., sodium, calcium, phosphorus)</li> </ul>	<ul style="list-style-type: none"> <li>• White blood cell count and globulin are under Immune</li> <li>• Serum lipids are under Cardiovascular</li> <li>• Serum liver markers are under Hepatic</li> </ul>

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Relevant human health effect category <sup>a</sup>	Examples of animal endpoints included	Notes
<b>Immune</b> (effects) <sup>b</sup>	<ul style="list-style-type: none"> <li>• Host resistance</li> <li>• Allergic, autoimmune or infectious disease</li> <li>• Hypersensitivity</li> <li>• General immune assays (e.g., white blood cell counts, immunological factors or cytokines in blood, lymphocyte phenotyping or proliferation)</li> <li>• Any measure in lymphoid tissues (weight; histopathology; cell counts; etc.)</li> <li>• Immune cell counts or immune-specific cytokines in nonlymphoid tissues</li> <li>• Other immune functional assays (e.g., antibody production, natural killer cell function, DTH, MLR, CTL, phagocytosis or bacterial killing by monocytes)</li> <li>• Immune responses in the respiratory system</li> <li>• Stress-related factors in blood (e.g., glucocorticoids or other adrenal markers)</li> </ul>	<ul style="list-style-type: none"> <li>• Red blood cells are under Hematological</li> <li>• Nonimmune measures of pulmonary function are under Respiratory</li> </ul>
<b>Urinary</b> (toxicity) <sup>b</sup>	<ul style="list-style-type: none"> <li>• Kidney weight and histopathology</li> <li>• Urinary measures (e.g., protein, volume, pH, specific gravity)</li> </ul>	
<b>Nervous system</b> (effects) <sup>b,d</sup>	<ul style="list-style-type: none"> <li>• Brain weight</li> <li>• Behavioral measures (including FOB and cage-side observations)</li> <li>• Nervous system histopathology</li> </ul>	
<b>Endocrine</b> (effects) <sup>b</sup>	<ul style="list-style-type: none"> <li>• Thyroid weight and histopathology</li> <li>• Hormonal measures in any tissue or blood (nonreproductive)</li> </ul>	<ul style="list-style-type: none"> <li>• Reproductive hormones are under Reproductive</li> </ul>

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Relevant human health effect category <sup>a</sup>	Examples of animal endpoints included	Notes
<b>Metabolic (effects)<sup>b,d</sup></b>	<ul style="list-style-type: none"> <li>• Free fatty acids</li> <li>• Serum glucose or insulin, or other measures related to diabetes</li> <li>• Pancreatic effects relevant to diabetes</li> <li>• Induced-obesity or BMI</li> <li>• Any of the above endpoints after developmental exposure will be primarily discussed in this health effect category, and then referenced under developmental effects</li> </ul>	
<b>Reproductive (toxicity)<sup>b</sup></b> Note: Evidence synthesis and evidence integration conclusions in assessments are developed separately for male and female reproductive effects (toxicity)	<ul style="list-style-type: none"> <li>• Reproductive organ weight and histopathology</li> <li>• Markers of sexual differentiation or maturation (e.g., preputial separation in males; vaginal opening or estrous cycling in females)</li> <li>• Mating parameters (e.g., success, mount latency)</li> <li>• Reproductive hormones</li> </ul>	<ul style="list-style-type: none"> <li>• Birth parameters (e.g., litter size; resorptions, implantations, viability) are under Developmental</li> <li>• If data indicate altered birth parameters are likely attributable to female fertility, these data may be discussed under Female Reproductive</li> </ul>
<b>Developmental (effects)<sup>b</sup></b> Note: Evidence synthesis of these endpoints in the assessments is termed “offspring growth and early development,” but evidence integration conclusions will be drawn on the broader category of “developmental effects” (which also considers organ/system-specific effects after exposure during development)	<ul style="list-style-type: none"> <li>• Dam health (e.g., weight gain, food consumption)</li> <li>• Pup viability/survival or other birth parameters (e.g., number of pups per litter)</li> <li>• Pup weight or growth (includes measures into adulthood after developmental-only exposure)</li> <li>• Developmental landmarks (eye opening, etc., but not including markers for other organ/system-specific toxicities)</li> </ul>	<ul style="list-style-type: none"> <li>• Histopathology and markers of development specific to other systems are organ/system-specific (e.g., vaginal opening is under Female Reproductive; tests of sensory maturation are under Nervous System)</li> </ul>
<b>Carcinogenicity<sup>b</sup></b>	<ul style="list-style-type: none"> <li>• Tumors</li> <li>• Precancerous lesions (e.g., dysplasia)</li> </ul>	

ALT = alanine aminotransferase; AST = aspartate aminotransferase; BMI = body mass index; CTL = cytotoxic T lymphocyte; DNA = deoxyribonucleic acid; DTH = delayed-type hypersensitivity; FOB = functional operational battery; LD<sub>50</sub> = median lethal dose; MLR = mixed leukocyte reaction.

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<sup>a</sup>Given the paucity of available studies and the absence of exceptional new evidence, information on gastrointestinal effects, musculoskeletal effects, ocular effects, and respiratory effects will not be formally evaluated in these assessments, although short summaries of the evidence may be included for context, and new literature relating to these outcomes will be monitored during literature search updates for potential inclusion.

<sup>b</sup>Any of the health effect-relevant endpoints observed after developmental exposure will be discussed primarily in the health effect category indicated, and then referenced under developmental effects.

<sup>c</sup>Some outcomes are relevant to multiple health effects. These outcomes may be categorized under only a single health effect in Table 5-2 for clarity. However, in the assessments, such outcome data would be discussed in the first relevant health effect synthesis (syntheses will generally follow the pattern of most-to-least available evidence) and then this synthesis will be cited in the syntheses of other relevant health effects. The evidence (for or against an effect) will contribute to evidence integration decisions for all relevant health effects.

<sup>d</sup>The primary focus of these assessments will be on developmental effects; endocrine effects; hepatic effects, including lipid and lipoprotein measures; immune effects; reproductive effects in males or females; urinary effects; general toxicity; and carcinogenicity. Data on cardiovascular effects, hematological effects, metabolic effects including diabetes, and nervous system effects will be summarized when available. These summaries may be developed in association with one of the health effects noted above, either as a separate formal evaluation of hazard or as part of a qualitative summary on “other effects,” depending on the assessment-specific data.

1           Assessment-specific refinements to the evaluation plan described in later sections of this  
2 protocol may be justified after review of the key areas of scientific complexity outlined in  
3 Section 2.4. Although not expected based on the relatively small database of studies for these PFAS,  
4 one such refinement includes the potential prioritization of studies testing specific (lower)  
5 exposure levels, exposure lifestages, or routes of exposure, as identified based on conclusions made  
6 regarding the ADME properties of these PFAS. As noted in Section 2.4, consideration of the  
7 available ADME data for these five PFAS will be prioritized (see additional discussion in  
8 Section 9.2). This will serve multiple purposes, including updating the data in Table 2-7 on serum  
9 half-lives across species and sexes. Notably, it is not expected that there will be enough data to  
10 examine lifestage-specific differences in ADME (including metabolic pathways for toxification or  
11 detoxification) that might inform evidence evaluation and synthesis decisions. (This is distinct  
12 from lifestage-specific differences in *exposure*, for example, due to the higher intake of food per kg  
13 body weight [BW] of young children, ingestion of dust, or maternal transfer via breastmilk.)  
14 However, a few anticipatory refinements will be applied to study evaluations based on the  
15 preliminary data presented in Table 2-7. Specifically, given the apparent sex-specific differences in  
16 PFAS half-life in rats and mice (note: toxicological studies in nonhuman primates were not  
17 identified for these PFAS), examining and reporting data for both sexes will be reviewed as a  
18 potential source of study insensitivity during study evaluation (see Section 6.3), particularly for  
19 PFAS that seem to vary largely for this parameter (e.g., PFHxS; PFNA). These half-life data will also  
20 be considered when evaluating the experiment-specific sensitivity of the frequency of exposures  
21 and the timing of endpoint testing after exposure in experimental animals (see Section 6.3), as well  
22 as the potential for using exposure biomarkers in exposed animals and humans (see Sections 6.2  
23 and 6.3). The apparent ADME differences across species will be a critical consideration in these  
24 assessments. This consideration will be applied to evidence synthesis and integration decisions  
25 (e.g., exploring ADME differences between rats and mice as a potential explanation if there are

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1 differences in sensitivity in outcome-specific responses; see Sections 9 and 10), as well as in  
2 extrapolating dosimetry (i.e., exposure levels and duration) from experiments in animal models to  
3 quantitative estimates relevant to humans, possibly including application within existing  
4 pharmacokinetic (PK) and PBPK models (see additional discussion in Sections 6.4 and 11.2).

5       Lastly, based on the key areas of scientific complexity outlined in Section 2.4, some of the  
6 analyses performed in support of these assessments may need to consider a broader array of  
7 studies than those available for these five PFAS. One example includes the need to consider the  
8 human relevance of certain outcomes observed in animals, including the role of receptors such as  
9 PPAR $\alpha$  (see additional discussion in Section 9.2). In addition, it is possible that additional literature  
10 identification and evaluation strategies will be developed to address the other key areas of  
11 scientific complexity outlined in Section 2.4, or other assessment-specific issues that arise during  
12 review. Any such approaches will be documented in updates to the PFAS-specific assessment  
13 protocol(s) (i.e., as assessment-specific updates to this document included as Addendum materials  
14 for each of the five PFAS assessments).

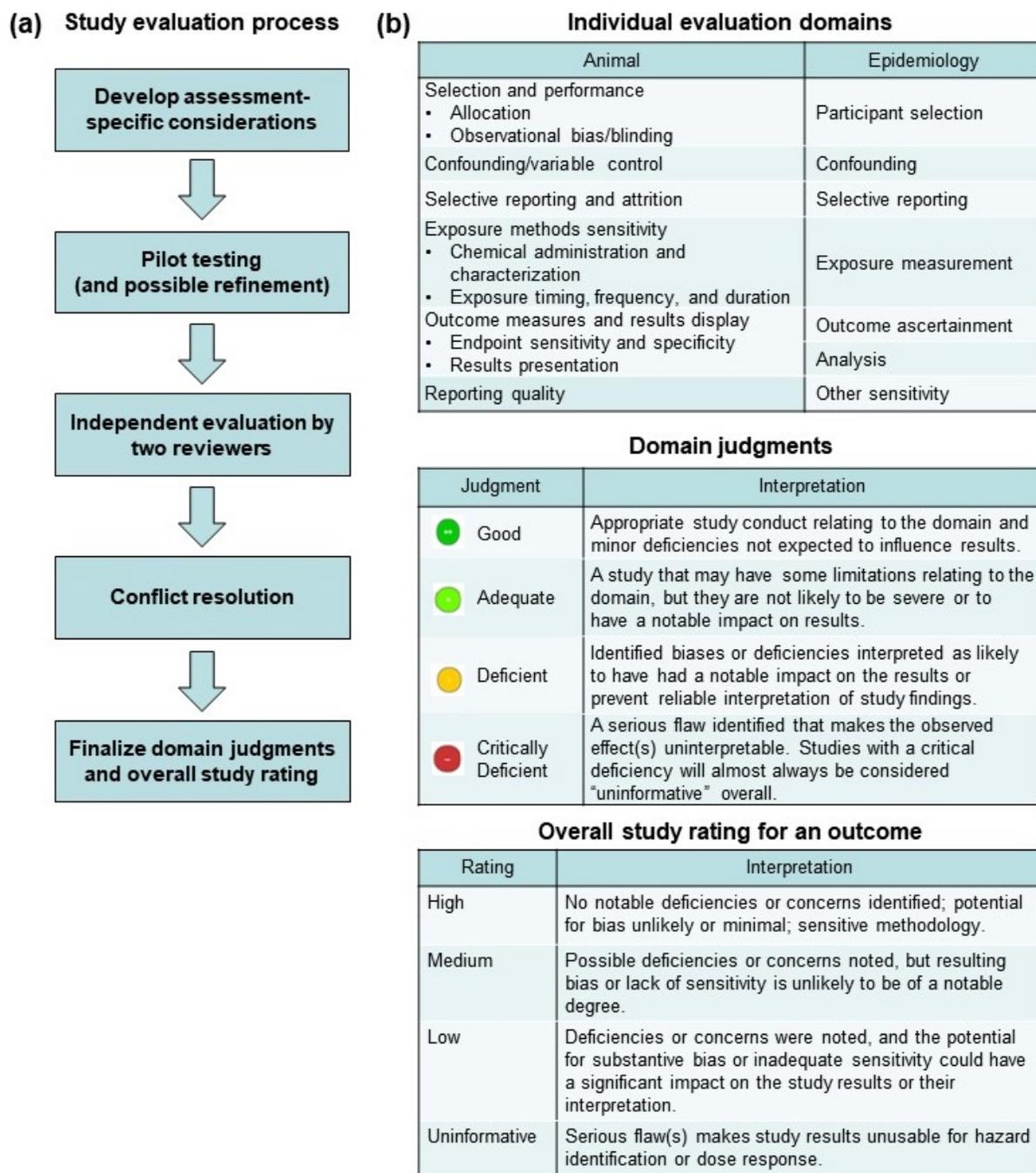
## **6. STUDY EVALUATION (REPORTING, RISK OF BIAS, AND SENSITIVITY) STRATEGY**

1           The general approach for evaluating PECO-relevant primary health effect studies is  
2 described in Section 6.1 and is the same for epidemiology studies and animal toxicology  
3 experiments, but the specifics of applying the approach differ; thus, they are described separately  
4 for epidemiology and animal toxicological studies in Sections 6.2 and 6.3, respectively. No  
5 controlled human exposure studies for these PFAS were identified (see Section 4). PBPK modeling  
6 studies were recently identified for PFHxS ([Kim et al., 2018](#)) and for PFDA and PFNA ([Kim et al.,](#)  
7 [2019](#)), although they were not formally identified by the systematic literature searches completed  
8 prior to posting of this protocol. In addition, a two-compartment PK model for gestational and  
9 lactational transfer of PFHxS in humans has been described by [Verner et al. \(2016\)](#). The specific  
10 approach for reviewing these studies is described in Section 6.4. Different approaches are used to  
11 evaluate mechanistic studies (see Sections 6.5 and 9.2).

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### **6.1. STUDY EVALUATION OVERVIEW FOR HEALTH EFFECT STUDIES**

12           Key concerns for the review of epidemiology and animal toxicological studies are potential  
13 bias (factors that affect the magnitude or direction of an effect in either direction) and insensitivity  
14 (factors that limit the ability of a study to detect a true effect; low sensitivity is a bias towards the  
15 null when an effect exists). The study evaluations are aimed at discerning the expected magnitude  
16 of any identified limitations (focusing on limitations that could substantively change a result),  
17 considering also the expected direction of the bias. The study evaluation approach is designed to  
18 address a range of study designs, health effects, and chemicals. The general approach for reaching  
19 an overall judgment for the study (or a specific analysis within a study) regarding confidence in the  
20 reliability of the results is illustrated in Figure 6-1.



**Figure 6-1. Overview of Integrated Risk Information System (IRIS) study evaluation process.** (a) An overview of the general evaluation process (note: see Section 5 for deviations from independent evaluation by two reviewers for some health outcomes in epidemiology studies). (b) The evaluation domains and definitions for ratings (i.e., domain and overall judgments, performed on an outcome-specific basis).

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1           With the exceptions noted in the refined evaluation plan for select outcomes reported in  
2 epidemiology studies (see Section 5), at least two reviewers will independently evaluate health  
3 effect studies to identify characteristics that bear on the informativeness (i.e., validity and  
4 sensitivity) of the results. The independent reviewers will use the structured platform for study  
5 evaluation housed within the Environmental Protection Agency’s (EPA’s) version of the Health  
6 Assessment Workplace Collaboration (HAWC)<sup>11</sup> to record separate judgements for each domain  
7 and the overall study for each outcome, to reach consensus between reviewers, and when  
8 necessary, resolve differences by discussion between the reviewers or consultation with additional  
9 independent reviewers. For some domains, additional chemical- or outcome-specific knowledge  
10 will be applied to evaluating the experimental design and methodology, as described below.

11           In general, considerations for reviewing a study with regard to its conduct for specific  
12 health outcomes is based on the use of existing guidance documents when available, including EPA  
13 guidance for carcinogenicity, neurotoxicity, reproductive toxicity, and developmental toxicity ([U.S.  
14 EPA, 2005a, 1998, 1996b, 1991a](#)). For some aspects of the study evaluations (e.g., review of  
15 exposure assessment in epidemiology studies), additional considerations are developed in  
16 consultation with topic-specific technical experts. To calibrate the assessment-specific  
17 considerations, the study evaluations will include a pilot phase to assess and refine the evaluation  
18 process. Additionally, as reviewers examine a group of studies, additional chemical-specific  
19 knowledge or methodologic concerns may emerge and a second pass of all pertinent studies may  
20 become necessary. Refinements to the study evaluation process made during the pilot phase and  
21 subsequent implementation across all relevant studies will be acknowledged as updates to the  
22 protocol.

23           Authors may be queried to obtain critical information, particularly that involving missing  
24 reporting quality information or other data (e.g., information on variability) or additional analyses  
25 that could address potential study limitations. The decision on whether to seek missing  
26 information includes consideration of what additional information would be useful, specifically  
27 with respect to any information that could result in a reevaluation of the overall study confidence  
28 for an outcome. Outreach to study authors will be documented and considered unsuccessful if  
29 researchers do not respond to an email or phone request within one month of the attempt to  
30 contact. Only information or data that can be made publicly available (e.g., within HAWC or Health  
31 and Environmental Research Online [HERO]) will be considered.

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<sup>11</sup>HAWC is a free and open source software application that provides a modular, web-based interface to help develop human health assessments of chemicals: <https://hawcproject.org/portal/>. Standard operating procedures provided to the reviewers to facilitate consistent and relevant documentation of their judgments using the HAWC software can be found as attachments embedded within the online tool (<https://hawcprd.epa.gov/assessment/10000039/>).

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1           When evaluating studies<sup>12</sup> that examine more than one outcome, the evaluation process will  
2 be performed separately for each outcome, because the utility of a study can vary for different  
3 outcomes. If a study examines multiple endpoints for the same outcome,<sup>13</sup> evaluations may be  
4 performed at a more granular level if appropriate, but these measures may still be grouped for  
5 evidence synthesis.

6           During review, the reviewers will reach a consensus judgment of *good*, *adequate*, *deficient*,  
7 *not reported*, or *critically deficient* for each evaluation domain. If a consensus is not reached, a third  
8 reviewer will perform conflict resolution. It is important to stress that these evaluations are  
9 performed in the context of the study’s utility for identifying individual hazards. While limitations  
10 specific to the usability of the study for dose-response analysis are useful to note for informing  
11 those later decisions, they do not contribute to the study confidence classifications.

12           These four categories are applied to each evaluation domain for each study as follows:

- 13           • *Good* represents a judgment that the study was conducted appropriately in relation to the  
14 evaluation domain and that any minor deficiencies noted would not be expected to  
15 influence the study results.
- 16           • *Adequate* indicates a judgment that there may be methodological limitations relating to the  
17 evaluation domain, but they are not likely to be severe or to have a notable impact on the  
18 results.
- 19           • *Deficient* denotes identified biases or deficiencies that are interpreted as likely to have had a  
20 notable impact on the results or that prevent interpretation of the study findings.
- 21           • *Not reported* indicates that the information necessary to evaluate the domain question was  
22 not available in the study. Generally, this term carries the same functional interpretation as  
23 *deficient* for the purposes of the study confidence classification (described below).  
24 Depending on the number of unreported items and severity of other limitations identified in  
25 the study, it may or may not be worth reaching out to the study authors for this information  
26 (see discussion above).
- 27           • *Critically deficient* reflects a judgment that the study conduct relating to the evaluation  
28 domain question introduced a serious flaw that is interpreted to be the primary driver of  
29 any observed effect(s) or makes the study uninterpretable. Studies with a determination of  
30 *critically deficient* in an evaluation domain will not be used for hazard identification or  
31 dose-response analysis, but they may be used to highlight possible research gaps. Given

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<sup>12</sup>“study” is used instead of a more accurate term (e.g., “experiment”) throughout these sections owing to an established familiarity within the field for discussing a study’s risk of bias or sensitivity, etc. However, all evaluations discussed herein are explicitly conducted at the level of an individual outcome within a population or cohort of humans or animals exposed in a similar manner (e.g., unexposed or exposed at comparable lifestages and for the same duration of exposure), or to a sample of the study population within a study.

<sup>13</sup>Note: “outcome” will be used throughout these methods; the same methods also apply to an endpoint assessed separately within a larger outcome. “Endpoint” refers to a more granular measurement (e.g., for the outcome of liver histopathology, different endpoints might include necrosis and cellular hypertrophy).

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1           this potential for exclusion, this classification is used infrequently and with extreme care;  
2           methodological limitations warranting this classification are defined a priori on an  
3           exposure- and outcome-specific basis and are inherently severe enough to warrant  
4           exclusion based on a single critical deficiency. Serious flaws that do not warrant study  
5           exclusion will be classified as *deficient*.

6           Once the evaluation domains have been rated, the identified strengths and limitations will  
7           be considered as a whole to reach a study confidence classification of *high, medium, or low*  
8           confidence, or *uninformative* for a specific health outcome. This classification is based on the  
9           reviewer judgments across the evaluation domains and considers the likely impact that inadequate  
10          reporting or the noted deficiencies in bias and sensitivity have on the outcome-specific results.  
11          There are no predefined weights for the domains, and the reviewers are responsible for applying  
12          expert judgment to make this determination. The classifications, which reflect a consensus  
13          judgment between reviewers, are defined as follows:

- 14          • *High* confidence: No notable deficiencies or concerns were identified; the potential for bias  
15          is unlikely or minimal, and the study used sensitive methodology. *High* confidence studies  
16          generally reflect judgments of *good* across all or most evaluation domains.
- 17          • *Medium* confidence: Possible deficiencies or concerns were noted, but the limitations are  
18          unlikely to be of a notable degree. Generally, *medium* confidence studies include *adequate*  
19          or *good* judgments across most domains, with the impact of any identified limitation not  
20          being judged as severe.
- 21          • *Low* confidence: Deficiencies or concerns are noted, and the potential for bias or inadequate  
22          sensitivity could have a significant impact on the study results or their interpretation.  
23          Typically, *low* confidence studies have a *deficient* evaluation for one or more domains,  
24          although some *medium* confidence studies may have a *deficient* rating in domain(s)  
25          considered to have less influence on the magnitude or direction of the outcome-specific  
26          results). *Low* confidence results are given less weight than *high* or *medium* confidence  
27          results during evidence synthesis and integration (see Section 10.1, Table 10-3 and  
28          Table 11-1), and are generally not used for hazard identification or dose-response analyses  
29          unless they are the only studies available or they inform data gaps unexamined in the *high*  
30          or *medium* confidence studies. Studies rated as *medium* or *low* confidence only because of  
31          sensitivity concerns about bias towards the null will be asterisked or otherwise noted  
32          because they may require additional consideration during evidence synthesis. Effects  
33          observed in studies biased toward the null may increase confidence in the results, assuming  
34          the study is otherwise well conducted (see Section 9).
- 35          • *Uninformative*: Serious flaw(s) make the study results unusable for hazard identification.  
36          Studies with *critically deficient* judgements in any evaluation domain are almost always  
37          classified as *uninformative* (see explanation above). Studies with multiple *deficient*  
38          judgments across domains may also be considered *uninformative*. *Uninformative* studies  
39          will not be considered further in the synthesis and integration of evidence, except perhaps  
40          to highlight possible research gaps.

1           As previously noted, study evaluation determinations reached by each reviewer and the  
2 consensus judgment between reviewers will be recorded in HAWC. Final study evaluations housed  
3 in HAWC, including for each domain and overall study confidence, will be made available when the  
4 draft is publicly released. These classifications and their rationales will be carried forward and  
5 considered as part of evidence synthesis (see Section 9) to help interpret the results across studies.

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## **6.2. EPIDEMIOLOGY STUDY EVALUATION**

6           Evaluation of epidemiology studies of health effects to assess risk of bias and study  
7 sensitivity will be conducted for the following domains: exposure measurement, outcome  
8 ascertainment, participant selection, potential confounding, analysis, study sensitivity, and selective  
9 reporting. Bias can result in false positives and negatives (i.e., Types I and II errors), while study  
10 sensitivity is typically concerned with identifying the latter.

11           The principles and framework used for evaluating epidemiology studies are based on the  
12 Cochrane Risk of Bias in Nonrandomized Studies of Interventions [ROBINS-I; [Sterne et al. \(2016\)](#)]  
13 but modified to address environmental and occupational exposures. The underlying philosophy of  
14 ROBINS-I is to describe attributes of an “ideal” study with respect to each of the evaluation domains  
15 (e.g., exposure measurement, outcome classification, etc.). Core and prompting questions are used  
16 to collect information to guide evaluation of each domain.

17           Core and prompting questions for each domain are presented in Table 6-1. Core questions  
18 represent key concepts, while the prompting questions help the reviewer focus on relevant details  
19 under each key domain. Table 6-1 also includes criteria that apply to all exposures and outcomes.  
20 PFAS-specific criteria are described in Section 6.2.1. As mentioned in Section 6.1, any additions to  
21 or refinements of the criteria will be documented as updates to the protocol.

**Table 6-1. Questions and criteria for evaluating each domain in epidemiology studies**

Domain and core question	Prompting questions	Follow-up questions	Criteria that apply to most exposures and outcomes
<p><u>Exposure measurement</u> Does the exposure measure reliably distinguish between levels of exposure in a time window considered most relevant for a causal effect with respect to the development of the outcome?</p>	<p>For all:</p> <ul style="list-style-type: none"> <li>Does the exposure measure capture the variability in exposure among the participants, considering intensity, frequency, and duration of exposure?</li> <li>Does the exposure measure reflect a relevant time window? If not, can the relationship between measures in this time and the relevant time window be estimated reliably?</li> <li>Was the exposure measurement likely to be affected by a knowledge of the outcome?</li> <li>Was the exposure measurement likely to be affected by the presence of the outcome (i.e., reverse causality)?</li> </ul> <p>For case-control studies of occupational exposures:</p> <ul style="list-style-type: none"> <li>Is exposure based on a comprehensive job history describing tasks, setting, time-period, and use of specific materials?</li> </ul>	<p>Is the degree of exposure misclassification likely to vary by exposure level?</p> <p>If the correlation between exposure measurements is moderate, is there an adequate statistical approach to ameliorate variability in measurements?</p> <p>If there is a concern about the potential for bias, what is the predicted direction or distortion of the bias on the effect estimate (if there is enough information)?</p>	<p><b>Good</b></p> <ul style="list-style-type: none"> <li>Valid exposure assessment methods used, which represent the etiologically relevant time-period of interest.</li> <li>Exposure misclassification is expected to be minimal.</li> </ul> <p><b>Adequate</b></p> <ul style="list-style-type: none"> <li>Valid exposure assessment methods used, which represent the etiologically relevant time-period of interest.</li> <li>Exposure misclassification may exist but is not expected to greatly change the effect estimate.</li> </ul> <p><b>Deficient</b></p> <ul style="list-style-type: none"> <li>Valid exposure assessment methods used, which represent the etiologically relevant time-period of interest. Specific knowledge about the exposure and outcome raise concerns about reverse causality, but there is uncertainty whether it is influencing the effect estimate.</li> <li>Exposed groups are expected to contain a notable proportion of unexposed or minimally exposed individuals, the method did not capture important temporal or spatial variation, or there is other evidence of exposure misclassification that would be expected to notably change the effect estimate.</li> </ul>

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<b>Domain and core question</b>	<b>Prompting questions</b>	<b>Follow-up questions</b>	<b>Criteria that apply to most exposures and outcomes</b>
	<p>For biomarkers of exposure, general population:</p> <ul style="list-style-type: none"> <li>• Is a standard assay used? What are the intra and interassay coefficients of variation? Is the assay likely to be affected by contamination? Are values less than the limit of detection dealt with adequately?</li> <li>• What exposure time-period is reflected by the biomarker? If the half-life is short, what is the correlation between serial measurements of exposure?</li> </ul>		<p><b>Critically deficient</b></p> <ul style="list-style-type: none"> <li>• Exposure measurement does not characterize the etiologically relevant time-period of exposure or is not valid.</li> <li>• There is evidence that reverse causality is very likely to account for the observed association.</li> <li>• Exposure measurement was not independent of outcome status.</li> </ul>
<p><u>Outcome ascertainment</u> Does the outcome measure reliably distinguish the presence or absence (or degree of severity) of the outcome?</p>	<p>For all:</p> <ul style="list-style-type: none"> <li>• Is outcome ascertainment likely to be affected by knowledge of, or presence of, exposure (e.g., consider access to health care, if based on self-reported history of diagnosis)?</li> </ul> <p>For case-control studies:</p> <ul style="list-style-type: none"> <li>• Is the comparison group without the outcome (e.g., controls in a case-control study) based on objective criteria with little or no likelihood of inclusion of people with the disease?</li> </ul>	<p>Is there a concern that any outcome misclassification is nondifferential, differential, or both?</p> <p>What is the predicted direction or distortion of the bias on the effect estimate (if there is enough information)?</p>	<p><b>Good</b></p> <ul style="list-style-type: none"> <li>• High certainty in the outcome definition (i.e., specificity and sensitivity), minimal concerns with respect to misclassification.</li> <li>• Assessment instrument was validated in a population comparable to the one from which the study group was selected.</li> </ul> <p><b>Adequate</b></p> <ul style="list-style-type: none"> <li>• Moderate confidence that outcome definition was specific and sensitive, some uncertainty with respect to misclassification but not expected to greatly change the effect estimate.</li> <li>• Assessment instrument was validated but not necessarily in a population comparable to the study group.</li> </ul>

**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

Domain and core question	Prompting questions	Follow-up questions	Criteria that apply to most exposures and outcomes
	<p>For mortality measures:</p> <ul style="list-style-type: none"> <li>How well does cause-of-death data reflect occurrence of the disease in an individual? How well do mortality data reflect incidence of the disease?</li> </ul> <p>For diagnosis of disease measures:</p> <ul style="list-style-type: none"> <li>Is the diagnosis based on standard clinical criteria? If it is based on self-report of the diagnosis, what is the validity of this measure?</li> </ul> <p>For laboratory-based measures (e.g., hormone levels):</p> <ul style="list-style-type: none"> <li>Is a standard assay used? Does the assay have an acceptable level of interassay variability? Is the sensitivity of the assay appropriate for the outcome measure in this study population?</li> </ul>		<p><b>Deficient</b></p> <ul style="list-style-type: none"> <li>Outcome definition was not specific or sensitive.</li> <li>Uncertainty regarding validity of assessment instrument.</li> </ul> <p><b>Critically deficient</b></p> <ul style="list-style-type: none"> <li>Invalid/insensitive marker of outcome.</li> <li>Outcome ascertainment is very likely to be affected by knowledge of, or presence of, exposure.</li> </ul> <p>Note: Lack of blinding should not be automatically construed to be <i>critically deficient</i>.</p>
<p><u>Participant selection</u> Is there evidence that selection into or out of the study (or analysis sample) was jointly related to exposure and to outcome?</p>	<p>For longitudinal cohort:</p> <ul style="list-style-type: none"> <li>Did participants volunteer for the cohort based on knowledge of exposure and/or preclinical disease symptoms? Was entry into the cohort or continuation in the cohort related to exposure and outcome?</li> </ul>	<p>Were differences in participant enrollment and follow-up evaluated to assess bias?</p> <p>If there is a concern about the potential for bias, what is the predicted direction or distortion of the bias on the effect</p>	<p><b>Good</b></p> <ul style="list-style-type: none"> <li>Minimal concern for selection bias based on description of recruitment process (e.g., selection of comparison population, population-based random sample selection, recruitment from sampling frame including current and previous employees).</li> <li>Exclusion and inclusion criteria specified and would not induce bias.</li> </ul>

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<b>Domain and core question</b>	<b>Prompting questions</b>	<b>Follow-up questions</b>	<b>Criteria that apply to most exposures and outcomes</b>
	<p>For occupational cohort:</p> <ul style="list-style-type: none"> <li>• Did entry into the cohort begin with the start of the exposure?</li> <li>• Was follow-up or outcome assessment incomplete, and if so, was follow-up related to both exposure and outcome status?</li> <li>• Could exposure produce symptoms that would result in a change in work assignment/work status (“healthy worker survivor effect”)?</li> </ul> <p>For case-control study:</p> <ul style="list-style-type: none"> <li>• Were controls representative of population and time periods from which cases were drawn?</li> <li>• Are hospital controls selected from a group whose reason for admission is independent of exposure?</li> <li>• Could recruitment strategies, eligibility criteria, or participation rates result in differential participation relating to both disease and exposure?</li> </ul> <p>For population-based survey:</p> <ul style="list-style-type: none"> <li>• Was recruitment based on advertisement to people with knowledge of exposure, outcome, and hypothesis?</li> </ul>	<p>estimate (if there is enough information)?</p> <p>Were appropriate analyses performed to address changing exposures over time in relation to symptoms?</p> <p>Is there a comparison of participants and nonparticipants to address whether differential selection is likely?</p>	<ul style="list-style-type: none"> <li>• Participation rate is reported at all steps of study (e.g., initial enrollment, follow-up, selection into analysis sample). If rate is not high, there is appropriate rationale for why it is unlikely to be related to exposure (e.g., comparison between participants and nonparticipants or other available information indicates differential selection is not likely).</li> </ul> <p><b>Adequate</b></p> <ul style="list-style-type: none"> <li>• Enough of a description of the recruitment process to be comfortable that there is no serious risk of bias.</li> <li>• Inclusion and exclusion criteria specified and would not induce bias.</li> <li>• Participation rate is incompletely reported but available information indicates participation is unlikely to be related to exposure.</li> </ul> <p><b>Deficient</b></p> <ul style="list-style-type: none"> <li>• Little information on recruitment process, selection strategy, sampling framework, and/or participation. Or aspects of these processes raise the potential for bias (e.g., healthy worker effect, survivor bias).</li> </ul> <p><b>Critically deficient</b></p> <ul style="list-style-type: none"> <li>• Aspects of the processes for recruitment, selection strategy, sampling framework, or participation result in concern that selection bias is likely to have had a large impact on effect estimates (e.g., convenience sample with no information about recruitment and selection, cases and controls are recruited from different sources with different likelihood of exposure, recruitment materials stated outcome of interest and potential participants are aware of or are concerned about specific exposures).</li> </ul>

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**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

<b>Domain and core question</b>	<b>Prompting questions</b>	<b>Follow-up questions</b>	<b>Criteria that apply to most exposures and outcomes</b>
<p><u>Confounding</u> Is confounding of the effect of the exposure likely?</p>	<p>Is confounding adequately addressed by considerations in:</p> <ul style="list-style-type: none"> <li>• Participant selection (matching or restriction)?</li> <li>• Accurate information on potential confounders and statistical adjustment procedures?</li> <li>• Lack of association between confounder and outcome, or confounder and exposure in the study?</li> <li>• Information from other sources?</li> </ul> <p>Is the assessment of confounders based on a thoughtful review of published literature, potential relationships (e.g., as can be gained through directed acyclic graphing), and minimizing potential overcontrol (e.g., inclusion of a variable on the pathway between exposure and outcome)?</p>	<p>If there is a concern about the potential for bias, what is the predicted direction or distortion of the bias on the effect estimate (if there is enough information)?</p>	<p><b>Good</b></p> <ul style="list-style-type: none"> <li>• Conveys strategy for identifying key confounders. This may include a priori biological considerations, published literature, causal diagrams, or statistical analyses, with the recognition that not all “risk factors” are confounders.</li> <li>• Inclusion of potential confounders in statistical models not based solely on statistical significance criteria (e.g., <math>p &lt; 0.05</math> from stepwise regression).</li> <li>• Does not include variables in the models that are likely to be influential colliders or intermediates on the causal pathway.</li> <li>• Key confounders are evaluated appropriately and considered to be unlikely sources of substantial confounding. This often will include:             <ul style="list-style-type: none"> <li>○ Presenting the distribution of potential confounders by levels of the exposure of interest and/or the outcomes of interest (with amount of missing data noted);</li> <li>○ Consideration that potential confounders were rare among the study population, or were expected to be poorly correlated with exposure of interest;</li> <li>○ Consideration of the most relevant functional forms of potential confounders;</li> <li>○ Examination of the potential impact of measurement error or missing data on confounder adjustment; or</li> <li>○ Presenting a progression of model results with adjustments for different potential confounders, if warranted.</li> </ul> </li> </ul>

**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

Domain and core question	Prompting questions	Follow-up questions	Criteria that apply to most exposures and outcomes
			<p><b>Adequate</b></p> <ul style="list-style-type: none"> <li>• Similar to <i>good</i> but may not have included all key confounders, or less detail may be available on the evaluation of confounders (e.g., sub-bullets in <i>good</i>). It is possible that residual confounding could explain part of the observed effect, but concern is minimal.</li> </ul> <p><b>Deficient</b></p> <ul style="list-style-type: none"> <li>• Does not include variables in the models that have been shown to be influential colliders or intermediates on the causal pathway.</li> <li>• And any of the following:               <ul style="list-style-type: none"> <li>○ The potential for bias to explain some of the results is high based on an inability to rule out residual confounding, such as a lack of demonstration that key confounders of the exposure-outcome relationships were considered;</li> <li>○ Descriptive information on key confounders (e.g., their relationship relative to the outcomes and exposure levels) are not presented; or</li> <li>○ Strategy of evaluating confounding is unclear or is not recommended (e.g., only based on statistical significance criteria or stepwise regression [forward or backward elimination]).</li> </ul> </li> </ul> <p><b>Critically deficient</b></p> <ul style="list-style-type: none"> <li>• Includes variables in the models that are colliders and/or intermediates in the causal pathway, indicating that substantial bias is likely from this adjustment; or</li> <li>• Confounding is likely present and not accounted for, indicating that all the results were most likely due to bias.</li> </ul>

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**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

Domain and core question	Prompting questions	Follow-up questions	Criteria that apply to most exposures and outcomes
<p><u>Analysis</u> Does the analysis strategy and presentation convey the necessary familiarity with the data and assumptions?</p>	<ul style="list-style-type: none"> <li>• Are missing outcome, exposure, and covariate data recognized, and if necessary, accounted for in the analysis?</li> <li>• Does the analysis appropriately consider variable distributions and modeling assumptions?</li> <li>• Does the analysis appropriately consider subgroups or lifestages of interest (e.g., based on variability in exposure level or duration or susceptibility)?</li> <li>• Is an appropriate analysis used for the study design?</li> <li>• Is effect modification considered, based on considerations developed a priori?</li> <li>• Does the study include additional analyses addressing potential biases or limitations (i.e., sensitivity analyses)?</li> </ul>	<p>If there is a concern about the potential for bias, what is the predicted direction or distortion of the bias on the effect estimate (if there is enough information)?</p>	<p><b>Good</b></p> <ul style="list-style-type: none"> <li>• Use of an optimal characterization of the outcome variable, including presentation of subgroup- or lifestage-specific comparisons (as appropriate for the outcome).</li> <li>• Quantitative results presented (effect estimates and confidence limits or variability in estimates) (i.e., not presented only as a <i>p</i>-value or “significant”/“not significant”).</li> <li>• Descriptive information about outcome and exposure provided (where applicable).</li> <li>• Amount of missing data noted and addressed appropriately (discussion of selection issues—missing at random vs. differential).</li> <li>• Where applicable, for exposure, includes LOD (and percentage below the LOD), and decision to use log transformation.</li> <li>• Includes analyses that address robustness of findings, for example, examination of exposure-response (explicit consideration of nonlinear possibilities, quadratic, spline, or threshold/ceiling effects included, when feasible); relevant sensitivity analyses; effect modification examined based only on a priori rationale with sufficient numbers.</li> <li>• No deficiencies in analysis evident. Discussion of some details may be absent (e.g., examination of outliers).</li> </ul> <p><b>Adequate</b></p> <ul style="list-style-type: none"> <li>• Same as <i>good</i>, except:</li> <li>• Descriptive information about exposure provided (where applicable) but may be incomplete; might not have discussed missing data, cutpoints, or shape of distribution(s).</li> </ul>

**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

Domain and core question	Prompting questions	Follow-up questions	Criteria that apply to most exposures and outcomes
			<ul style="list-style-type: none"> <li>• Includes analyses that address robustness of findings (examples in <i>good</i>), but some important analyses are not performed.</li> </ul> <p><b>Deficient</b></p> <ul style="list-style-type: none"> <li>• Does not conduct analysis using optimal characterization of the outcome variable.</li> <li>• Descriptive information about exposure levels not provided (where applicable).</li> <li>• Effect estimate and <i>p</i>-value presented, without standard error or confidence interval.</li> <li>• Results presented as statistically “significant”/“not significant.”</li> </ul> <p><b>Critically deficient</b></p> <ul style="list-style-type: none"> <li>• Results of analyses of effect modification examined without clear a priori rationale and without providing main/principal effects (e.g., presentation only of statistically significant interactions that were not hypothesis driven).</li> <li>• Analysis methods are not appropriate for design or data of the study.</li> </ul>
<p><u>Selective reporting</u> Is there reason to be concerned about selective reporting?</p>	<ul style="list-style-type: none"> <li>• Were results provided for all the primary analyses described in the methods section?</li> <li>• Is there appropriate justification for restricting the amount and type of results that are shown?</li> <li>• Are only statistically significant results presented?</li> </ul>	<p>If there is a concern about the potential for bias, what is the predicted direction or distortion of the bias on the effect estimate (if there is enough information)?</p>	<p><b>Good</b></p> <ul style="list-style-type: none"> <li>• The results reported by study authors are consistent with the primary and secondary analyses described in a registered protocol or methods paper.</li> </ul> <p><b>Adequate</b></p> <ul style="list-style-type: none"> <li>• The authors described their primary (and secondary) analyses in the methods section, and results were reported for all primary analyses.</li> </ul>

**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

Domain and core question	Prompting questions	Follow-up questions	Criteria that apply to most exposures and outcomes
			<p><b>Deficient</b></p> <ul style="list-style-type: none"> <li>• Concerns were raised based on previous publications, a methods paper, or a registered protocol indicating that analyses were planned or conducted that were not reported, or that hypotheses originally considered to be secondary were represented as primary in the reviewed paper.</li> <li>• Only subgroup analyses were reported, suggesting that results for the entire group were omitted.</li> <li>• Only statistically significant results were reported.</li> </ul>
<p><u>Sensitivity</u> Is there a concern that sensitivity of the study is not adequate to detect an effect?</p>	<ul style="list-style-type: none"> <li>• Is the exposure range adequate to detect associations and exposure-response relationships?</li> <li>• Was the appropriate population or lifestage included?</li> <li>• Was the length of follow-up adequate? Is the time/age of outcome ascertainment optimal given the interval of exposure and the health outcome?</li> <li>• Are there other aspects related to risk of bias or otherwise that raise concerns about sensitivity?</li> </ul>		<p><b>Adequate</b></p> <ul style="list-style-type: none"> <li>• The range of exposure levels provides adequate variability to evaluate the associations of relevance.</li> <li>• The population was exposed to levels expected to have an impact on response.</li> <li>• The study population was sensitive to the development of the outcomes of interest (e.g., ages, lifestage, sex).</li> <li>• The timing of outcome ascertainment was appropriate given expected latency for outcome development (i.e., adequate follow-up interval).</li> <li>• The study was adequately powered to observe an effect.</li> <li>• No other concerns raised regarding study sensitivity.</li> </ul> <p><b>Deficient</b></p> <ul style="list-style-type: none"> <li>• Concerns were raised about the issues described for <i>adequate</i> that are expected to notably decrease the sensitivity of the study to detect associations for the outcome.</li> </ul>

**6.2.1. Epidemiology Study Evaluation Criteria Specific to These Five Per- and Polyfluoroalkyl Substances (PFAS)**

1 The exposure criteria described in Table 6-2 below are modified from the criteria  
2 developed by NTP OHAT<sup>14</sup> for their assessment of the association between PFOA and immune  
3 effects.

4 The estimated serum half-lives of PFAS in humans were presented in Table 2-7 (see  
5 Section 2.4.1). In considering temporality concerns, some PFAS (PFHxS, PFDA, and PFNA) are  
6 persistent compounds with longer (multiple year) half-lives in humans, so current exposure levels  
7 may be indicative of critical exposure windows that were narrow or past exposures that extended  
8 beyond the anticipated half-lives. In contrast, other PFAS appear to have half-lives of 1 month or  
9 less (PFBA, PFHxA), and current exposure levels may not be indicative of past exposures that  
10 extend beyond the anticipated half-lives. Some evidence suggests that the half-lives vary based on  
11 sex, parity, interval between pregnancy, reproductive hormones, and gynecological disorders ([Lau  
12 et al., 2007](#)); therefore, these factors will be considered depending on the population(s), critical  
13 windows, and outcomes being examined.

14 Standard analytical methods of individual PFAS in serum or whole-blood using quantitative  
15 techniques such as liquid chromatography-triple quadrupole mass spectrometry are considered to  
16 be well-established methods ([CDC, 2019a, b](#); [ATSDR, 2018](#); [CDC, 2015](#); [U.S. EPA, 2014a, b](#); [CDC,  
17 2009](#)).

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<sup>14</sup>National Toxicology Program (NTP) Report:

[https://ntp.niehs.nih.gov/ntp/ohat/pfoa\\_pfos/pfoa\\_pfosmonograph\\_508.pdf](https://ntp.niehs.nih.gov/ntp/ohat/pfoa_pfos/pfoa_pfosmonograph_508.pdf).

NTP protocol: [https://ntp.niehs.nih.gov/ntp/ohat/pfoa\\_pfos/protocol\\_201506\\_508.pdf](https://ntp.niehs.nih.gov/ntp/ohat/pfoa_pfos/protocol_201506_508.pdf).

**Table 6-2. Criteria for evaluating exposure measurement in epidemiology studies of per- and polyfluoroalkyl substances (PFAS) and health effects**

Rating	Criteria
<i>Good</i>	<ul style="list-style-type: none"> <li>• Evidence that exposure was consistently assessed using well-established analytical methods that directly measure exposure (e.g., measurement of PFAS in blood, serum, or plasma).</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Exposure was assessed using less established methods (e.g., measurement of PFAS in breast milk) or methods that indirectly measure exposure (e.g., drinking water concentrations and residential location/history, questionnaire or occupational exposure assessment by a certified industrial hygienist) that are validated against well-established direct methods (i.e., intermethods validation: one method vs. another) in the target population of interest.</li> </ul> <p><b>And all the following:</b></p> <ul style="list-style-type: none"> <li>• Exposure was assessed in a relevant time window (i.e., temporality is established, and sufficient latency occurred before disease onset) for development of the outcome based on current biological understanding.</li> <li>• There is evidence that sufficient exposure data measurements are above the limit of quantification for the assay.</li> <li>• The laboratory analysis included data on standard quality control measures with demonstrated precision and accuracy.</li> </ul>
<i>Adequate</i>	<ul style="list-style-type: none"> <li>• Exposure was assessed using less established methods or indirect measures that are validated but not in the target population of interest.</li> </ul> <p><b>OR</b></p> <ul style="list-style-type: none"> <li>• Evidence that exposure was consistently assessed using methods described in <i>good</i>, but there were some concerns about quality control measures or other potential for nondifferential misclassification.</li> </ul> <p><b>And all the following:</b></p> <ul style="list-style-type: none"> <li>• Exposure was assessed in a relevant time window for development of the outcome</li> <li>• There is evidence that sufficient exposure data measurements are above the limit of quantification for the assay.</li> <li>• The laboratory analysis included some data on standard quality control measures with demonstrated precision and accuracy.</li> </ul>
<i>Deficient</i>	<p><b>Any of the following:</b></p> <ul style="list-style-type: none"> <li>• Some concern, but no direct evidence, that the exposure was assessed using methods that have not been validated or empirically shown to be consistent with methods that directly measure exposure.</li> </ul>

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Rating	Criteria
	<ul style="list-style-type: none"> <li>Exposure was assessed in a relevant time window(s) for development of the outcome, but there could be some concern about the potential for bias due to reverse causality<sup>a</sup> between exposure and outcome, yet no direct evidence that it is present.</li> </ul>
<i>Critically deficient</i>	<p><b>Any of the following:</b></p> <ul style="list-style-type: none"> <li>Exposure was assessed in a time window that is unknown or not relevant for development of the outcome. This could be due to clear evidence of bias from reverse causality between exposure and outcome, or other concerns such as the lack of temporal ordering of exposure and disease onset, insufficient latency, or having exposure measurements that are not reliable measures of exposure during the etiologic window(s).</li> <li>Direct evidence that bias was likely because the exposure was assessed using methods with poor validity.</li> <li>Evidence of differential exposure misclassification (e.g., differential recall of self-reported exposure).</li> <li>There is evidence that an insufficient number of the exposure data measurements were above the limit of quantification for the assay.</li> </ul>

<sup>a</sup>Reverse causality refers to a situation in which an observed association between exposure and outcome is not due to causality from exposure to outcome, but rather due to the outcome of interest causing a change in the measured exposure.

1            In addition, there are PFAS-specific considerations for the evaluation of confounding. As  
2 discussed in Section 2.4.3, confounding across PFAS is an important area of uncertainty when  
3 interpreting the results of epidemiology studies for individual PFAS (i.e., quantifying the effect of  
4 an individual PFAS can potentially be confounded by other PFAS). Based on preliminary analyses,  
5 correlations differ across the PFAS (see Figure 6-2). While some pairs have correlation coefficients  
6 consistently above 0.6 (e.g., PFNA and PFDA), the correlations for most vary from 0.1 to 0.6  
7 depending on the study, and little data is available on correlations with less commonly occurring or  
8 detected PFAS like PFBA and PFHxA.

	PFBA	PFDA	PFHxA	PFHxS	PFNA	PFOA	PFOS
PFBA*	1.00	0.01	0.45	0.14	0.15	0.03	0.07
PFDA		1.00	-0.03	0.28	0.73	0.42	0.48
PFHxA*			1.00	0.08	-0.07	0.19	-0.04
PFHxS				1.00	0.35	0.43	0.50
PFNA					1.00	0.54	0.51

**Figure 6-2. Preliminary mean correlation coefficients across per- and polyfluoroalkyl substances (PFAS) among studies in the inventory, for all media types.**

\*PFBA and PFHxA correlations were based on three studies for PFOS, PFOA, and PFHxS, two studies for each other, and one study for PFNA and PFDA, so these estimates are less stable than the other PFAS, which were all based on >10 studies.

PFOS = perfluorooctane sulfonate.

1            Rather than rating each study with lower confidence because of this issue, potential  
2            confounding by other PFAS will be considered during the evidence synthesis phase, primarily when  
3            there is some support for an association with adverse health effects in the epidemiology evidence.  
4            This may include looking across studies in populations with different exposure profiles  
5            (e.g., observing an association in a population with much higher exposure to one PFAS due to  
6            proximity to an industrial plant would increase confidence for that PFAS). In situations where  
7            there is considerable uncertainty regarding the impact of residual confounding across PFAS, this  
8            will be captured as a factor that decreases evidence strength (see Section 10).

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### 6.3. EXPERIMENTAL ANIMAL STUDY EVALUATION

9            Using the principles described in Section 6.1, the evaluation of animal studies of health  
10            effects to assess risk of bias and sensitivity will be conducted for the following domains: reporting  
11            quality, risk of bias (selection or performance bias, confounding/variable control, and reporting or  
12            attrition bias), and study sensitivity (exposure methods sensitivity, and outcome measures and  
13            results display) (see Table 6-3). Several additional considerations specific to assessing these five  
14            PFAS are outlined in Section 6.3.1.

15            The rationale for judgments will be documented clearly and consistently at the outcome  
16            level. In addition, for domains other than reporting quality, the evaluation documentation in HAWC  
17            will include the identified limitations and consider their impact on the overall confidence level, a  
18            procedure similar to the evaluation of epidemiology studies. This will, to the extent possible, reflect  
19            an interpretation of the potential influence on the outcome-specific results (including the direction  
20            and/or magnitude of influence).

**Table 6-3. Considerations to evaluate domains from animal toxicological studies**

Evaluation concern	Domain—core question	Prompting questions	General considerations
Reporting quality	<p><b>Reporting quality</b> Does the study report information for evaluating the design and conduct of the study for the endpoints/outcomes of interest? <i>Note:</i> <i>This domain is limited to reporting. Other aspects of the exposure methods, experimental design, and endpoint evaluation methods are evaluated using the domains related to risk of bias and study sensitivity.</i></p>	<p>Does the study report the following?</p> <p><b>Critical information</b> necessary to perform study evaluation:</p> <ul style="list-style-type: none"> <li>• Species, test article name, levels and duration of exposure, route (e.g., oral; inhalation), qualitative or quantitative results for at least one endpoint of interest</li> </ul> <p><b>Important information</b> for evaluating the study methods:</p> <ul style="list-style-type: none"> <li>• Test animal: strain, sex, source, and general husbandry procedures</li> <li>• Exposure methods: source, purity, method of administration</li> <li>• Experimental design: frequency of exposure, animal age, and lifestage during exposure and at endpoint/outcome evaluation</li> <li>• Endpoint evaluation methods: assays or procedures used to measure the endpoints/outcomes of interest</li> </ul>	<p>A judgment and rationale for this domain will generally be given for the study. In the rationale, reviewers will also indicate when a study adhered to GLP, or to OECD (or similar) testing guidelines.</p> <ul style="list-style-type: none"> <li>• <b>Good:</b> All <b>critical and important information</b> is reported or inferable for the endpoints/outcomes of interest.</li> <li>• <b>Adequate:</b> All <b>critical information</b> is reported, but some <b>important information</b> is missing. However, the missing information is not expected to significantly impact the study evaluation.</li> <li>• <b>Deficient:</b> All <b>critical information</b> is reported, but <b>important information</b> is missing that is expected to significantly reduce the ability to evaluate the study.</li> <li>• <b>Critically deficient:</b> Study report is missing any pieces of <b>critical information</b>. Studies that are <i>critically deficient</i> for reporting are <i>uninformative</i> for the overall rating and not considered further.</li> </ul>

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Evaluation concern	Domain—core question	Prompting questions	General considerations
<p align="center"><b>Risk of bias</b></p>	<p><b>Allocation</b> Were animals assigned to experimental groups using a method that minimizes selection bias?</p>	<p>For each study:</p> <ul style="list-style-type: none"> <li>• Did each animal or litter have an equal chance of being assigned to any experimental group (i.e., random allocation<sup>a</sup>)?</li> <li>• Is the allocation method described?</li> <li>• Aside from randomization, were any steps taken to balance variables across experimental groups during allocation?</li> </ul>	<p>A judgment and rationale for this domain will be given for each cohort or experiment in the study.</p> <ul style="list-style-type: none"> <li>• <i>Good</i>: Experimental groups were randomized, and any specific randomization procedure was described or inferable (e.g., computer-generated scheme). (Note that normalization is not the same as randomization [see response for <i>adequate</i>].)</li> <li>• <i>Adequate</i>: Authors report that groups were randomized but do not describe the specific procedure used (e.g., “animals were randomized”). Alternatively, authors used a nonrandom method to control for important modifying factors (i.e., with respect to the outcome of interest) across experimental groups (e.g., body-weight normalization).</li> <li>• <i>Not reported</i> (interpreted as <i>deficient</i>): No indication of randomization of groups or other methods (e.g., normalization) to control for important modifying factors across experimental groups.</li> <li>• <i>Critically deficient</i>: Bias in the animal allocations was reported or inferable.</li> </ul>

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Evaluation concern	Domain—core question	Prompting questions	General considerations
<p align="center">Risk of bias (continued)</p> <p align="center">Selection and performance bias (continued)</p>	<p><b>Observational bias/blinding</b> Did the study implement measures to reduce observational bias?</p>	<p>For each endpoint/outcome or grouping of outcomes in a study:</p> <ul style="list-style-type: none"> <li>• Does the study report blinding or other methods/procedures for reducing observational bias, as appropriate for the assays of interest?</li> <li>• If not, did the study use a design or approach for which such procedures can be inferred?</li> <li>• What is the expected impact of failure to implement (or report implementation) of these methods/procedures on results?</li> </ul>	<p>A judgment and rationale for this domain will be given for each endpoint/outcome or group of outcomes investigated in the study.</p> <ul style="list-style-type: none"> <li>• <i>Good</i>: Measures to reduce observational bias were described (e.g., blinding to conceal treatment groups during endpoint evaluation; consensus-based evaluations of histopathology lesions<sup>a</sup>).</li> <li>• <i>Adequate</i>: Methods for reducing observational bias (e.g., blinding) can be inferred or were reported but described incompletely.</li> <li>• <i>Not reported</i>: Measures to reduce observational bias were not described.             <ul style="list-style-type: none"> <li>○ (Interpreted as <i>adequate</i>): The potential concern for bias was mitigated based on use of automated/computer-driven systems, standard laboratory kits, relatively simple objective measures (e.g., body or tissue weight), or screening-level evaluations of histopathology.</li> <li>○ (Interpreted as <i>deficient</i>): The potential impact on the results is large (e.g., outcome measures are highly subjective).</li> </ul> </li> <li>• <i>Critically deficient</i>: Strong evidence for observational bias that impacted the results.</li> </ul>

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Evaluation concern	Domain—core question	Prompting questions	General considerations	
Risk of bias (continued)	Confounding/variable control	<p><b>Confounding</b> Are variables with the potential to confound or modify results controlled for and consistent across all experimental groups?</p>	<p>For each study:</p> <ul style="list-style-type: none"> <li>• Are there differences across the treatment groups (e.g., coexposures, vehicle, diet, palatability, husbandry, health status, surgery) that could bias the results?</li> <li>• If differences are identified, to what extent are they expected to impact the results?</li> </ul>	<p>A judgment and rationale for this domain will be given for each cohort or experiment in the study, noting when the potential for confounding is restricted to specific endpoints/outcomes.</p> <ul style="list-style-type: none"> <li>• <i>Good</i>: Outside of the exposure of interest, variables that are likely to confound or modify results appear to be controlled for and consistent across experimental groups.</li> <li>• <i>Adequate</i>: Some concern that variables likely to confound or modify the results were uncontrolled or inconsistent across groups, but these are expected to have a minimal impact on the results.</li> <li>• <i>Deficient</i>: Notable concern that potentially confounding variables were uncontrolled or inconsistent across groups and that they are expected to substantially impact the results.</li> <li>• <i>Critically deficient</i>: Confounding variables were presumed to be uncontrolled or inconsistent across groups, and they are expected to be a primary driver of the results.</li> </ul>

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Evaluation concern	Domain—core question	Prompting questions	General considerations	
Risk of bias (continued)	Selective reporting and attrition bias	<p><b>Selective reporting and attrition</b> Did the study report results for all prespecified outcomes and tested animals?</p> <p><i>Note:</i> <i>This domain does not consider the appropriateness of the comparisons/results presentation. This aspect of study quality is evaluated in another domain.</i></p>	<p>For each study: Selective reporting bias:</p> <ul style="list-style-type: none"> <li>• Are all results presented for endpoints/outcomes described in the methods (see note)?</li> </ul> <p>Attrition bias:</p> <ul style="list-style-type: none"> <li>• Do the results account for all animals?</li> <li>• If there are discrepancies, do the authors provide an explanation (e.g., death or unscheduled sacrifice during the study)?</li> <li>• If unexplained results omissions and/or attrition are identified, what is the expected impact on the interpretation of the results?</li> </ul>	<p>A judgment and rationale for this domain will be given for each cohort or experiment in the study.</p> <ul style="list-style-type: none"> <li>• <i>Good:</i> Quantitative or qualitative results were reported for all prespecified outcomes (explicitly stated or inferred), exposure groups, and evaluation time points. Data not reported in the primary article are available from supplemental material. If results omissions or animal attrition are identified, the authors provide an appropriate explanation, and the omissions or attrition are not expected to impact the interpretation of the results.</li> <li>• <i>Adequate:</i> Quantitative or qualitative results are reported for most prespecified outcomes (explicitly stated or inferred), exposure groups, and evaluation time points. Omissions and/or attrition are not explained, but they are not expected to significantly impact the interpretation of the results.</li> <li>• <i>Deficient:</i> Quantitative or qualitative results are missing for many prespecified outcomes (explicitly stated or inferred), exposure groups and evaluation time points, or there is high animal attrition; omissions and/or attrition are not explained and are expected to significantly impact the interpretation of the results.</li> <li>• <i>Critically deficient:</i> Extensive results omission and/or animal attrition are identified and prevent comparisons of results across treatment groups.</li> </ul>

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Evaluation concern	Domain—core question	Prompting questions	General considerations
<p align="center"><b>Sensitivity</b></p> <p align="center"><b>Exposure methods sensitivity</b></p>	<p><b>Chemical administration and characterization</b>            Did the study adequately characterize exposure to the chemical of interest and the exposure administration methods?</p> <p><i>Note:            These considerations are limited to oral exposure, as only a single inhalation study focusing on acute toxicity (i.e., after PFNA exposure) was identified (see Section 2.3.2).</i></p>	<p>For each study:</p> <ul style="list-style-type: none"> <li>• Does the study report the source and purity and/or composition (e.g., identity and percent distribution of different isomers) of the chemical? If not, can the purity and/or composition be obtained from the supplier (e.g., as reported on the website)?</li> <li>• Was independent analytical verification of the test article purity and composition performed?</li> <li>• Are there concerns about the methods used to administer the chemical (e.g., gavage volume)?</li> <li>• If necessary, based on consideration of chemical-specific knowledge (e.g., instability in solution; volatility) and/or exposure design (e.g., the frequency and duration of exposure), were the chemical concentrations in the dosing solutions or diet analytically confirmed?</li> </ul>	<p>A judgment and rationale for this domain will be given for each cohort or experiment in the study.</p> <ul style="list-style-type: none"> <li>• <i>Good:</i> Chemical administration and characterization is complete (i.e., source, purity, and analytical verification of the test article are provided). There are no concerns about the composition, stability, or purity of the administered chemical, or the specific methods of administration.</li> <li>• <i>Adequate:</i> Some uncertainties in the chemical administration and characterization are identified, but these are expected to have minimal impact on interpreting the results (e.g., source and vendor-reported purity are presented, but not independently verified; purity of the test article is suboptimal but not concerning).</li> <li>• <i>Deficient:</i> Uncertainties in the exposure characterization are identified and expected to substantially impact the results (e.g., source of the test article is not reported; levels of impurities are substantial or concerning; deficient administration methods, such as use of a gavage volume considered too large for the species and/or lifestage at exposure).</li> <li>• <i>Critically deficient:</i> Uncertainties in the exposure characterization are identified, and there is reasonable certainty that the results are largely attributable to factors other than exposure to the chemical of interest (e.g., identified impurities are expected to be a primary driver of the results).</li> </ul>

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Evaluation concern		Domain—core question	Prompting questions	General considerations
Sensitivity (continued)	Exposure methods sensitivity (continued)	<p><b>Exposure timing, frequency, and duration</b> Was the timing, frequency, and duration of exposure sensitive for the endpoint(s)/outcome(s) of interest?</p>	<p>For each endpoint/outcome or grouping of outcomes in a study:</p> <ul style="list-style-type: none"> <li>• Does the exposure period include the full critical window of sensitivity, based on current biological understanding?</li> <li>• Was the duration and frequency of exposure sensitive for detecting the endpoint of interest?</li> </ul>	<p>A judgment and rationale for this domain will be given for each endpoint/outcome or group of outcomes investigated in the study.</p> <ul style="list-style-type: none"> <li>• <i>Good</i>: The duration and frequency of the exposure was sensitive, and the exposure included the critical window of sensitivity (if known).</li> <li>• <i>Adequate</i>: The duration and frequency of the exposure was sensitive, and the exposure covered most of the critical window of sensitivity (if known).</li> <li>• <i>Deficient</i>: The duration and/or frequency of the exposure is not sensitive and did not include most of the critical window of sensitivity (if known). These limitations are expected to bias the results towards the null.</li> <li>• <i>Critically deficient</i>: The exposure design was not sensitive and is expected to strongly bias the results towards the null. The rationale should indicate the specific concern(s).</li> </ul>

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Evaluation concern	Domain—core question	Prompting questions	General considerations
<p align="center">Sensitivity (continued)</p> <p align="center">Outcome measures and results display</p>	<p><b>Endpoint sensitivity and specificity</b>            Are the procedures sensitive and specific for evaluating the endpoint(s)/outcome(s) of interest?</p> <p><i>Note:</i></p> <ul style="list-style-type: none"> <li>• <i>Sample size alone is not a reason to conclude an individual study is critically deficient.</i></li> <li>• <i>Considerations related to adjustments/corrections to endpoint measurements (e.g., organ weight corrected for body weight) are addressed under results presentation.</i></li> </ul>	<p>For each endpoint/outcome or grouping of outcomes in a study:</p> <ul style="list-style-type: none"> <li>• Are there concerns regarding the sensitivity, specificity, and/or validity of the outcome measurement protocols?</li> <li>• Are there serious concerns regarding the sample size?</li> <li>• Are there concerns regarding the timing of the endpoint assessment?</li> </ul>	<p>A judgment and rationale for this domain will be given for each endpoint/outcome or group of outcomes investigated in the study. Examples of potential concerns include:</p> <ul style="list-style-type: none"> <li>• Selection of protocols that are insensitive or nonspecific for the endpoint of interest.</li> <li>• Evaluations did not include all treatment groups (e.g., only control and high dose).</li> <li>• Use of unreliable or invalid methods to assess the outcome.</li> <li>• Assessment of endpoints at inappropriate or insensitive ages, or without addressing known endpoint variation (e.g., due to circadian rhythms, estrous cyclicity).</li> <li>• Decreased specificity or sensitivity of the response due to the timing of endpoint evaluation, as compared with exposure (e.g., immediate endpoint assessment after exposure to chemicals with short-acting depressant or irritant effects; insensitivity due to prolonged period of nonexposure before testing).</li> </ul>

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Evaluation concern	Domain—core question	Prompting questions	General considerations
<p align="center"><b>Sensitivity (continued)</b></p> <p align="center"><b>Outcome measures and results display (continued)</b></p>	<p><b>Results presentation</b> Are the results presented in a way that makes the data usable and transparent?</p> <p><i>Note:</i> <i>Potential issues associated with statistical analyses will be flagged for review by EPA statisticians and possible reanalysis (if information is available to do so, any reanalysis will be transparently presented). Any remaining limitations will be discussed during evidence synthesis or dose-response analyses (depending on the identified issue).</i></p>	<p>For each endpoint/outcome or grouping of outcomes in a study:</p> <ul style="list-style-type: none"> <li>• Does the level of detail allow for an informed interpretation of the results?</li> <li>• Are the data analyzed, compared, or presented in a way that is inappropriate or misleading?</li> </ul>	<p>A judgment and rationale for this domain will be given for each endpoint/outcome or group of outcomes investigated in the study. Examples of potential concerns include:</p> <ul style="list-style-type: none"> <li>• Nonpreferred presentation (e.g., developmental toxicity data averaged across pups in a treatment group, when litter responses are more appropriate; presentation of absolute organ-weight data when relative weights are more appropriate).</li> <li>• Failing to present quantitative results either in tables or figures.</li> <li>• Pooling data when responses are known or expected to differ substantially (e.g., across sexes or lifestages).</li> <li>• Failing to report on or address overt toxicity when exposure levels are known or expected to be highly toxic.</li> <li>• Lack of full presentation of the data (e.g., presentation of mean without variance data; concurrent control data are not presented).</li> </ul>
<p align="center"><b>Overall confidence</b></p>	<p><b>Overall confidence</b> Considering the identified strengths and limitations, what is the overall confidence rating for the endpoint(s)/outcome(s) of interest?</p> <p><i>Note:</i> <i>Reviewers will mark studies that are rated lower than high confidence due only to low sensitivity (i.e., bias towards the null) for additional consideration during evidence synthesis. If the study is otherwise well conducted and an effect is observed, the confidence may be increased.</i></p>	<p>For each endpoint/outcome or grouping of outcomes in a study:</p> <ul style="list-style-type: none"> <li>• Were concerns (i.e., limitations or uncertainties) related to the reporting quality, risk of bias, or sensitivity identified?</li> <li>• If yes, what is their expected impact on the overall interpretation of the reliability and validity of the study results, including (when possible) interpretations of impacts on the magnitude or direction of the reported effects?</li> </ul>	<p>The overall confidence rating considers the likely impact of the noted concerns (i.e., limitations or uncertainties) in reporting, bias and sensitivity on the results. A confidence rating and rationale will be given for each endpoint/outcome or group of outcomes investigated in the study. Confidence rating definitions are described above (see Section 6.1).</p>

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<sup>a</sup>Several studies have characterized the relevance of randomization, allocation concealment, and blind outcome assessment in experimental studies ([Hirst et al., 2014](#); [Krauth et al., 2013](#); [Macleod, 2013](#); [Higgins and Green, 2011](#)).

GLP = good laboratory practice; OECD = Organisation for Economic Co-operation and Development.

### **6.3.1. Animal Toxicological Study Evaluation Considerations Specific to These Five Per- and Polyfluoroalkyl Substances (PFAS)**

1           A key uncertainty in these assessments involves the toxicokinetics of the five PFAS. The  
2           apparent differences in toxicokinetics across animal species will not be addressed at the individual  
3           study level but will be considered during evidence integration (see Section 10) and is expected to  
4           be most influential when developing toxicity values for potential human health hazards (see  
5           Section 11). However, based on Table 2-7 (see Section 2.4.1), the clearance of some of these PFAS,  
6           and the sex-specific differences in serum half-lives, represent important considerations for  
7           potential sources of insensitivity during study evaluation. Specifically, studies may be judged as  
8           insensitive if they fail to account for the short serum half-lives of PFBA in female rats and mice  
9           (half-lives of ~1–3 hours; half-lives in males are close to half a day and of less concern) and PFHxA  
10          in rats and mice of both sexes (half-lives of ~0.5–1.5 hours) by including, for example, multiple  
11          daily exposures. Half-lives in rodents for PFDA, PFNA, and PFHxS are on the order of days or  
12          longer, so insensitivity due to the short half-life in rodents does not represent a concern for these  
13          PFAS (note: no nonhuman primate health effect studies were identified). Similarly, given the  
14          profound apparent differences in clearance between male and female rats for PFNA (i.e., females  
15          appear to clear PFNA 25× faster), studies that examined both sexes are preferred, and any study  
16          that tested female rats only may be judged as insensitive. This consideration may also be applied,  
17          but to a lesser extent, for studies of PFHxS in rats and PFBA in mice or rats (females appear to clear  
18          these PFAS ~4–6× faster than males).

19          These five PFAS are considered stable and nonreactive, and the presence of potentially toxic  
20          impurities within these readily available chemicals has not been identified as an issue in the  
21          literature. Thus, failure to describe preparation and storage of dosing solutions will not be  
22          considered an issue of concern, and a lack of information on chemical purity will not be considered  
23          a significant limitation. This interpretation is consistent with quality control and solubilization  
24          information on these PFAS performed by the EPA as part of the ongoing ToxCast testing  
25          ([https://comptox.epa.gov/dashboard/chemical\\_lists/EPAPFASINV](https://comptox.epa.gov/dashboard/chemical_lists/EPAPFASINV)). None of these PFAS were  
26          flagged as problematic (e.g., based on volatility and solubility, or degradation-type issues), or raised  
27          concerns during analytical testing, although C10 (PFDA) can become less soluble in water at very  
28          high concentrations. Given the more relevant possibility of contamination of these five PFAS with  
29          other PFAS, a lack of analytical verification of the test article will be flagged as a limitation, although  
30          this alone will not significantly affect overall study confidence ratings.

31          A wide variety of outcomes have been assessed in the available animal studies for these five  
32          PFAS. Considerations specific to each outcome are not included in this protocol (outcome-specific

1 concerns will be available in HAWC when the assessments are released). As examples, a few  
2 specific considerations that will be applied include better domain ratings for studies that address  
3 potential differences in time of day for evaluations of hormone levels (due to fluctuations  
4 throughout the day), and for studies that address fasting status for metabolic-related  
5 measurements.

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#### **6.4. PHARMACOKINETIC MODEL EVALUATION**

6 A similar approach for evaluation will be applied to the full PBPK models for PFHxS ([Kim et](#)  
7 [al., 2018](#)) and for PFDA and PFNA ([Kim et al., 2019](#)), as well as to the two-compartment PK model  
8 for gestational and lactational transfer of PFHxS in humans described by [Verner et al. \(2016\)](#).  
9 Models will be preferred for use in these assessments when an applicable one exists and no equal  
10 or better alternative for dosimetric extrapolation is available. Given these preferences, sound  
11 justification will be provided for *not* using a PBPK (or classical PK) model when an applicable one  
12 exists and no equal or better alternative for dosimetric extrapolation is available. Note, however,  
13 that these preferences *only* apply to models that faithfully represent current scientific knowledge  
14 and accurately translate the science into computational code in a reproducible, transparent  
15 manner. In practice, many models have errors that affect their predictions to varying degrees;  
16 hence, an evaluation of a model is required before it can be used in an assessment. Thus, the  
17 currently available models and any other models identified at later stages of developing these  
18 assessments will be evaluated as described below.

19 Considerations for judging the suitability of a model are separated into two categories:  
20 scientific and technical. The scientific criteria focus on whether the biology, chemistry, and other  
21 information available for chemical mode(s) of action (MOA[s]) are appropriately represented by  
22 the model structure and equations. Scientific criteria are easier to evaluate in judging a model's  
23 suitability because they can be judged by reading the publication or report that describes the  
24 model, without requiring an evaluation of the computer code. Preliminary technical criteria include  
25 the availability of the computer code and apparent completeness of parameter listing and  
26 documentation. The in-depth technical and scientific criteria focus on the accurate implementation  
27 of the conceptual model in the computational code, use of correct or biologically consistent  
28 parameters in the model, and reproducibility of model results reported in journal publications and  
29 other documents. Specific details for this evaluation are provided in the Quality Assurance Project  
30 Plan for PBPK models ([U.S. EPA, 2018b](#)).

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#### **6.5. MECHANISTIC STUDY EVALUATION**

31 Sections 9 and 10 outline an approach for focused consideration of information from  
32 mechanistic studies (including *in vitro*, *in vivo*, *ex vivo*, and *in silico* studies) where the specific  
33 analytical approach is targeted to the assessment needs, depending in part on the extent and nature  
34 of the phenotypic human and animal evidence. In this way, the mechanistic synthesis for a given

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1 health effect might range from a high-level summary (or detailed analysis) of potential mechanisms  
2 of action to specific, focused questions needed to address important and impactful assessment  
3 uncertainties unaddressed by the available phenotypic studies (e.g., expected shape of the  
4 dose-response curve in the low-dose region, applicability of the animal evidence to humans,  
5 addressing susceptible populations). Individual study-level evaluation of mechanistic endpoints  
6 will not typically be pursued. However, it may be necessary to identify assay-specific  
7 considerations for study endpoint evaluations on a case-by-case basis to provide a more detailed  
8 summary and evaluation for the most relevant individual mechanistic studies addressing a key  
9 assessment uncertainty. This may be done, for example, when the scientific understanding of a  
10 critical mechanistic event or MOA lacks scientific consensus, when the reported findings on a  
11 critical mechanistic endpoint are conflicting, when the available mechanistic evidence addresses a  
12 complex and influential aspect of the assessment, or when in vitro or in silico data make up the bulk  
13 of the evidence base and there is little or no evidence from epidemiological studies or animal  
14 bioassays. As noted in Section 3 and Section 4, genotoxicity studies were identified as meeting  
15 PECO criteria; these data will be summarized in each PFAS assessment to describe evidence  
16 relevant to carcinogenicity even in the absence of more phenotypic data. Based on the  
17 considerations above, if these studies are interpreted as adequate to draw a hazard conclusion  
18 (i.e., other than “insufficient”), individual study-level evaluations of some or all the genotoxicity  
19 studies will be informative to this decision. If necessary, based on the assessment-specific issues  
20 identified during study evaluation and evidence synthesis (see Section 9.2), the specific approach to  
21 evaluating individual studies other than those addressed in Sections 6.2–6.4 will be outlined in the  
22 assessments and included as an update to the protocol.

## **7. ORGANIZING THE HAZARD REVIEW**

1           The organization and scope of the hazard evaluation is determined by the available  
2 evidence for each PFAS regarding routes of exposure, metabolism and distribution, outcomes  
3 evaluated, and number of studies pertaining to each outcome, as well as the results of the  
4 evaluation of sources of bias and sensitivity. The hazard evaluations will be organized around  
5 organ systems (e.g., nervous system) informed by one or multiple related outcomes, as described in  
6 Section 5, and a decision will be made as to what level (e.g., organ system or subsets of outcomes  
7 within an organ system) to organize the synthesis.

8           Table 7-1 lists some questions that may be asked of the evidence to assist with this decision.  
9 These questions extend from considerations and decisions made during development of the refined  
10 evaluation plan to include review of the concerns raised during individual study evaluations as well  
11 as the direction and magnitude of the study-specific results. Resolution of these questions will then  
12 inform critical decisions about the organization of the hazard evaluation and help determine what  
13 studies may be useful in dose-response analyses.

**Table 7-1. Querying the evidence to organize syntheses for human and animal evidence**

Evidence	Questions	Follow-up questions
ADME	Given the known ADME issues for these PFAS, do the data appear to differ by route of exposure studied, lifestage when exposure occurred, sex, species, or dosing regimens used?	Will separate analyses be needed by factors such as sex, route of exposure, or by methods of dosing within a route of exposure (e.g., are large differences expected between gavage and dietary exposures)? Are data available to inform which lifestages and what dosing regimens are more relevant to human exposure scenarios?
	Is there toxicity information for metabolites that also should be evaluated for hazard?	What exposures will be included in the evaluation?
Outcomes	What outcomes are reported in studies? Are the data reported in a comparable manner across studies (similar output metrics at similar levels of specificity, such as adenomas and carcinomas quantified separately)?	At what level (hazard, grouped outcomes, or individual outcomes) will the synthesis be conducted? What commonalities will the outcomes be grouped by:
	Are there interrelated outcomes? If so, consider whether some outcomes are more useful and/or of greater concern than others.	<ul style="list-style-type: none"> <li>• health effect,</li> <li>• exposure levels,</li> </ul>
	Does the evidence indicate greater sensitivity to effects (at lower exposure levels or severity) in certain subgroups (by age, sex, ethnicity, lifestage)? Should the hazard evaluation include a subgroup analysis?	<ul style="list-style-type: none"> <li>• functional or population-level consequences (e.g., endpoints all ultimately leading to decreased fertility or impaired cognitive function),</li> </ul>
	Does incidence or severity of an outcome increase with duration of exposure or a particular window of exposure. What exposure time frames are relevant to development or progression of the outcome?	<ul style="list-style-type: none"> <li>• involvement of related biological pathways</li> </ul>
	Is there mechanistic evidence that informs how outcomes might be grouped together?	How well do the assessed human and animal outcomes relate within a level of grouping?
	<p>How robust is the evidence for specific outcomes?</p> <ul style="list-style-type: none"> <li>• What outcomes are reported by both human and animal studies and by one or the other? Were different animal species and sexes (or other important population-level differences) tested?</li> <li>• In general, what are the study confidence conclusions of the studies (<i>high, medium, low, uninformative</i>) for the different outcomes? Is there enough evidence from <i>high</i> and <i>medium</i> confidence studies to draw conclusions about causality?</li> </ul>	<p>What outcomes should be highlighted? Should the others be synthesized at all? Would comparisons by specific limitations be informative?</p>

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<b>Evidence</b>	<b>Questions</b>	<b>Follow-up questions</b>
Dose-response	Did some outcomes include better coverage of exposure ranges that may be most relevant to human exposure than others?	What outcomes and studies are informative for developing toxicity values?
	For which outcomes are there sufficient data available to draw conclusions about dose-response? Are there outcomes with study results of sufficient similarity (e.g., an established linkage in a biological pathway) to allow examination or calculation of common measures of effect across studies? Do the mechanistic data identify surrogate or precursor outcomes that are adequate for dose-response analysis?	
	Are there subgroups that exhibit responses at lower exposure levels than others?	
	Are there findings from ADME studies that could inform data-derived extrapolation factors, or link toxicity observed via different routes of exposure, or link effects between humans and experimental animals?	What studies might be used to develop nondefault UFs? Is there a common internal dose metric that can be used to compare species or routes of exposure?

ADME = absorption, distribution, metabolism, and excretion; UF = uncertainty factor.

## 8. DATA EXTRACTION OF STUDY METHODS AND RESULTS

1 Data extraction and content management will be carried out using the Health Assessment  
2 Workplace Collaborative (HAWC; web links will be shared in the individual assessments). A  
3 consistent approach to data extraction will be applied across these PFAS assessments to facilitate  
4 their anticipated future use in addressing poorly studied PFAS (e.g., through coupling with  
5 computational toxicology data generated as described in the Environmental Protection Agency  
6 [EPA] PFAS action plan). Data extraction elements that may be collected from epidemiological,  
7 controlled human exposure, animal toxicological, and in vitro studies are described in HAWC  
8 (<https://hawcprd.epa.gov/about/>). Not all studies that meet the PECO criteria go through data  
9 extraction. Studies evaluated as being *Uninformative* are not considered further and therefore  
10 would not undergo data extraction. In addition, outcomes determined to be less relevant during  
11 PECO refinement (see Section 5) may not go through data extraction or may have only minimal data  
12 extraction. The same may be true for *low* confidence studies if enough *medium* and *high* confidence  
13 studies (e.g., on an outcome) are available. All findings are considered for extraction, regardless of  
14 statistical significance. The level of extraction for specific outcomes within a study may differ  
15 (i.e., ranging from a narrative to full extraction of dose-response effect size information). In part,  
16 this extraction level is determined based on the level of detail to be discussed in the evidence  
17 synthesis for that health effect (e.g., a detailed extraction will not be necessary for health effects  
18 with very few available studies; these will only be briefly summarized in a short narrative).  
19 Similarly, decisions about data extraction for *low* confidence studies are typically made while  
20 implementing the protocol and are based on the quality and extent of the available evidence. If  
21 necessary, the version of the protocol released with the draft assessment will outline how *low*  
22 confidence studies were treated for extraction and evidence synthesis.

23 The data extraction results for included studies will be presented in the assessment (and  
24 made available for download from EPA HAWC in Excel format) when the draft is publicly released.  
25 (Note: The following browsers are supported for accessing HAWC: Google Chrome [preferred],  
26 Mozilla Firefox, and Apple Safari. There are errors in functionality when viewed with Internet  
27 Explorer.) For quality control, data extraction will be performed by one member of the evaluation  
28 team and independently verified by at least one other member. Discrepancies in data extraction  
29 will be resolved by discussion or consultation with a third member of the evaluation team. Digital  
30 rulers, such as WebPlotDigitizer (<http://arohatgi.info/WebPlotDigitizer/>), will be used to extract  
31 numerical information from figures, and their use will be documented during extraction.

1 As previously described, routine attempts will be made to obtain missing information from  
2 human and animal health effect studies, if it is considered influential during study evaluations (see  
3 Section 6) or when it can provide information important for dose-response analysis or  
4 interpretations of significance (e.g., missing group size or variance descriptors such as standard  
5 deviation or confidence interval). Missing data from individual mechanistic (e.g., in vitro) studies  
6 generally will not be sought. Outreach to study authors or designated contact persons will be  
7 documented and considered unsuccessful if they do not respond to email or phone requests within  
8 1 month of initial attempt(s) to contact.

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## 8.1. STANDARDIZING REPORTING OF EFFECT SIZES

9 In addition to providing quantitative outcomes in their original units for all study groups,  
10 results from outcome measures will be transformed, when possible, to a common metric to help  
11 compare distinct but related outcomes that are measured with different scales. These standardized  
12 effect size estimates facilitate systematic evaluation and evidence integration for hazard  
13 identification (see Section 9.1). The following summary of effect size metrics by data type outlines  
14 issues in selecting the most appropriate common metric for a collection of related endpoints  
15 ([Vesterinen et al., 2014](#)).

16 Common metrics for continuous outcomes include:

- 17 • *Absolute difference in means.* This metric is the difference between the means in the control  
18 and treatment groups, expressed in the units in which the outcome is measured. When the  
19 outcome measure and its scale are the same across all studies, this approach is the simplest  
20 to implement.
- 21 • *Percent control response (or normalized mean difference [NMD]).* Percent control group  
22 calculations are based on means. Standard deviation (or standard error) values presented  
23 in the studies for these normalized effect sizes can also be estimated if sufficient  
24 information has been provided. Note that some outcomes reported as percentages, such as  
25 mean percentage of affected offspring per litter, can lead to distorted effect sizes when  
26 further characterized as percentage change from control. Such measures are better  
27 expressed as absolute difference in means, or rather preferably transformed to incidences  
28 using approaches for event or incidence data (see below).
- 29 • *Standardized mean difference.* The NMD approach above is relevant to ratio scales, but  
30 sometimes it is not possible to infer what a “normal” animal would score, such as when data  
31 for animals without lesions are not available. In these circumstances, standardized mean  
32 differences can be used. The difference in group means is divided by a measure of the  
33 pooled variance to convert all outcome measures to a standardized scale with units of  
34 standard deviations. This approach can also be applied to data for which different  
35 measurement scales are reported for the same outcome measure (e.g., different measures of  
36 lesion size such as infarct volume and infarct area).

1 Common metrics for event or incidence data include:

- 2 • Percent change from control. This metric is analogous to the NMD approach described for  
3 continuous data above.
- 4 • For binary outcomes such as the number of individuals that developed a disease or died,  
5 and with only one treatment evaluated, data can be represented in a  $2 \times 2$  table. Note that  
6 when the value in any cell is 0, 0.5 is added to each cell to avoid problems with the  
7 computation of the standard error. For each comparison, the odds ratio (OR) and its  
8 standard error can be calculated. Odds ratios are normally combined on a logarithmic scale.

9 An additional approach for epidemiology studies is to extract adjusted statistical estimates  
10 when possible rather than unadjusted or raw estimates.

11 It is important to consider the variability associated with effect size estimates, with better  
12 powered studies generally showing more precise estimates. Effect size estimation can be affected,  
13 however, by such factors as variances that differ substantially across treatment groups, or by lack of  
14 information to characterize variance, especially for animal studies in biomedical research  
15 ([Vesterinen et al., 2014](#)). The assessments will consider the nature of any variance issues and  
16 ensure that the associated uncertainties are clarified and accounted for during the evidence  
17 synthesis process (see Section 9).

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## **8.2. STANDARDIZING ADMINISTERED DOSE LEVELS/CONCENTRATIONS**

18 Exposures will be standardized to common units. Exposure levels in oral studies will be  
19 expressed in units of mg PFAS/kg-day. When the study authors provide exposure levels in  
20 concentrations in the diet or drinking water, dose conversions will be made using study-specific  
21 food or water consumption rates and body weights when available. Otherwise, EPA defaults will be  
22 used ([U.S. EPA, 1988](#)), addressing age and study duration as relevant for the species/strain and sex  
23 of the animal of interest. Exposure levels in inhalation studies will be expressed in units of mg/m<sup>3</sup>.  
24 Assumptions used in performing dose conversions will be documented in HAWC or the specific  
25 assessments.

## 9. SYNTHESIS OF EVIDENCE

1 For the purposes of this assessment, evidence synthesis and integration are considered  
2 distinct, but related processes. As described below, for each assessed health effect the evidence  
3 syntheses provide a summary discussion of each body of evidence considered in the review  
4 (i.e., human, animal, and mechanistic evidence). These separate summaries directly inform  
5 interpretations regarding the support for causation provided by each body of evidence and the  
6 evidence as a whole. In other words, the syntheses of separate bodies of evidence described in this  
7 section will directly inform the integration across evidence to draw an overall judgment for each of  
8 the assessed human health effects (as described in Section 10). The phrase “evidence integration”  
9 used here is analogous to the phrase “weight of evidence” used in some other assessment processes  
10 ([EFSA, 2017](#); [U.S. EPA, 2017a](#); [NRC, 2014](#); [U.S. EPA, 2005a](#)).<sup>15</sup>

11 For each potential human health effect (or smaller subset of related outcomes), the U.S.  
12 Environmental Protection Agency (EPA) will separately synthesize the available phenotypic human  
13 and animal evidence pertaining to that potential health effect. Mechanistic evidence will also be  
14 considered in targeted analyses conducted before, during, and after developing syntheses of the  
15 phenotypic human and animal evidence. The results of the analyses of mechanistic evidence will be  
16 used to help resolve key uncertainties; as a result, the scope of the mechanistic analyses will  
17 generally depend on the extent and nature of the human and animal evidence (see Sections 9.2 and  
18 10). Thus, apart from the predefined mechanistic analyses (see Sections 9.2.1–9.2.3), the human  
19 and animal evidence syntheses (or the lack of phenotypic data in humans and animals) help  
20 determine the approach to be taken in synthesizing the available mechanistic evidence (see  
21 Section 9.2.4). In this way, a mechanistic evidence synthesis might range from a high-level  
22 summary of potential toxicity mechanisms discussed in the published literature to a detailed  
23 analysis of multiple potential modes of action, or it might evaluate specific, focused questions that  
24 inform key uncertainties unaddressed by the phenotypic human and animal evidence (e.g., shape of  
25 the dose-response curve at low doses, applicability of the animal evidence to humans, addressing  
26 susceptible populations). Each synthesis will provide a summary discussion of the available  
27 evidence that addresses considerations regarding causation (see Table 9-1). These considerations

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<sup>15</sup>This revision has been adopted primarily based on the 2014 NAS review of IRIS ([NRC, 2014](#)): “The present committee found that the phrase *weight of evidence* has become far too vague as used in practice today and thus is of little scientific use. In some accounts, it is characterized as an oversimplified balance scale on which evidence supporting hazard is placed on one side and evidence refuting hazard on the other... The present committee found the phrase *evidence integration* to be more useful and more descriptive of what is done at this point in an IRIS assessment—that is, IRIS assessments must come to a judgment about whether a chemical is hazardous to human health and must do so by integrating a variety of evidence.”

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1 are adapted from considerations for causality introduced by Austin Bradford Hill ([Hill](#)  
2 [1965](#)): consistency, dose-response relationship, strength of the association, temporal relationship,  
3 biological plausibility, coherence, and “natural experiments” in humans [see additional discussion  
4 in [U.S. EPA \(2005a\)](#) and [U.S. EPA \(1994\)](#)]. Importantly, the evidence synthesis process explicitly  
5 considers and incorporates the conclusions from the individual study evaluations (see Section 6).

**Table 9-1. Information most relevant to describing primary considerations for assessing causality during evidence syntheses**

Consideration	Description of the consideration and its application in IRIS syntheses
Study confidence	<p><u>Description:</u> Incorporates decisions about study confidence within each of the considerations.</p> <p><u>Application:</u> In evaluating the evidence for each of the causality considerations described in the following rows, the syntheses will consider study confidence decisions. <i>High</i> confidence studies carry the most weight. The syntheses will consider the specific limitations and strengths identified during study evaluation and describe how these informed each consideration.</p>
Consistency	<p><u>Description:</u> Examines the similarity of results (e.g., direction; magnitude) across studies.</p> <p><u>Application:</u> Syntheses will evaluate the homogeneity of findings on a given outcome or endpoint across studies. When inconsistencies exist, the syntheses consider whether results were “conflicting” (i.e., unexplained positive and negative results in similarly exposed human populations or in similar animal models) or “differing” (i.e., mixed results explainable by, for example, differences between human populations, animal models, exposure conditions, or study methods) (<a href="#">U.S. EPA, 2005a</a>). These considerations are based on analyses of potentially important explanatory factors such as:</p> <ul style="list-style-type: none"> <li>• Confidence in studies’ results, including study sensitivity (e.g., some study results that appear to be inconsistent may be explained by potential biases or other attributes that affect sensitivity).</li> <li>• Exposure, including route (if applicable) and administration methods, levels, duration, timing with respect to outcome development (e.g., critical windows), and exposure assessment methods (i.e., in epidemiology studies), including analytical units and specific groups being compared.</li> <li>• Specificity and sensitivity of the endpoint for evaluating the health effect in question (e.g., functional measures can be more sensitive than organ weights).</li> <li>• Populations or species, including consideration of potential susceptible groups or differences across lifestyle at exposure or endpoint assessment.</li> <li>• Toxicokinetic information explaining observed differences in responses across route of exposure, other aspects of exposure, species, sexes, or lifestages.</li> </ul> <p>The interpretation of consistency will emphasize biological significance, to the extent that it is understood, over statistical significance. Statistical significance from suitably applied tests (this may involve consultation with an EPA statistician) adds weight when biological significance is not well understood. Consistency in the direction of results increases confidence in that association even in the absence of statistical significance. In some cases, it may be helpful to consider the potential for publication bias to provide context to interpretations of consistency.<sup>a</sup></p>

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Consideration	Description of the consideration and its application in IRIS syntheses
Strength (effect magnitude) and precision	<p><u>Description:</u> Examines the effect magnitude or relative risk, based on what is known about the assessed endpoint(s), and considers the precision of the reported results based on analyses of variability (e.g., confidence intervals; standard error). This may include consideration of the rarity or severity of the outcomes.</p> <p><u>Application:</u> Syntheses will analyze results both within and across studies and may consider the utility of combined analyses (e.g., meta-analysis). While larger effect magnitudes and precision (e.g., <math>p &lt; 0.05</math>) help reduce concerns about chance, bias, or other factors as explanatory, syntheses should also consider the biological or population-level significance of small effect sizes.</p>
Biological gradient/dose-response	<p><u>Description:</u> Examines whether the results (e.g., response magnitude; incidence; severity) change in a manner consistent with changes in exposure (e.g., level; duration), including consideration of changes in response after cessation of exposure.</p> <p><u>Application:</u> Syntheses will consider relationships both within and across studies, acknowledging that the dose-response relationship (e.g., shape) can vary depending on other aspects of the experiment, including the biology underlying the outcome and the toxicokinetics of the chemical. Thus, when dose-dependence is lacking or unclear, the synthesis will also consider the potential influence of such factors on the response pattern.</p>
Coherence	<p><u>Description:</u> Examines the extent to which findings are cohesive across different endpoints that are related to, or dependent on, one another (e.g., based on known biology of the organ system or disease, or mechanistic understanding such as toxicokinetic/dynamic understanding of the chemical or related chemicals). In some instances, additional analyses of mechanistic evidence from research on the chemical under review or related chemicals that evaluate linkages between endpoints or organ-specific effects may be needed to interpret the evidence. These analyses may require additional literature search strategies.</p> <p><u>Application:</u> Syntheses will consider potentially related findings, both within and across studies, particularly when relationships are observed within a cohort or within a narrowly defined category (e.g., occupation; strain or sex; lifestage of exposure). Syntheses will emphasize evidence indicative of a progression of effects, such as temporal- or dose-dependent increases in the severity of the type of endpoint observed. If an expected coherence between findings is not observed, possible explanations should be explored, including those related to the biology of the effects as well as the sensitivity and specificity of the measures used.</p>

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Consideration	Description of the consideration and its application in IRIS syntheses
<p>Mechanistic evidence related to biological plausibility</p>	<p><u>Description:</u> There are multiple uses for mechanistic information, and this consideration overlaps with “coherence.” This consideration examines the biological support (or lack thereof) for findings from the human and animal health effect studies and becomes more influential on the hazard conclusions when notable uncertainties in the strength of those sets of studies exist. These analyses can also improve understanding of dose- or duration-related development of the health effect. In the absence of human or animal evidence of apical health endpoints, the synthesis of mechanistic information may drive evidence integration conclusions (when such information is available).</p> <p><u>Application:</u> Syntheses can evaluate evidence on precursors, biomarkers, or other molecular or cellular changes related to the health effect(s) of interest to describe the likelihood that the observed effects result from exposure. This evaluation will entail an analysis of existing evidence, and not simply speculate whether a theoretical pathway can be postulated. This analysis may not be limited to evidence relevant to the PECO but may also include evaluations of biological pathways (e.g., for the health effect; established for other, possibly related, chemicals). Any such synthesis of mechanistic evidence will consider the sensitivity of the mechanistic changes and the potential contribution of alternative or previously unidentified mechanisms of toxicity.</p>
<p>Natural experiments</p>	<p><u>Description:</u> Specific to epidemiology studies and rarely available, this consideration examines effects in populations that have experienced well-described, pronounced changes in chemical exposure (e.g., lead exposures before and after banning lead in gasoline).</p> <p><u>Application:</u> Compared with other observational designs, natural experiments have the benefit of dividing people into exposed and unexposed groups without them influencing their own exposure status. During synthesis, associations in <i>medium</i> and <i>high</i> confidence natural experiments can substantially reduce concerns about residual confounding.</p>

<sup>a</sup>Publication bias involves the influence of the direction, magnitude, or statistical significance of the results on the likelihood of a paper being published; it can result from decisions made, consciously or unconsciously, by study authors, journal reviewers, and journal editors ([Dickersin, 1990](#)). When evidence of publication bias is present for a set of studies, less weight may be placed on the consistency of the findings for or against an effect during evidence synthesis and integration.

PECO = populations, exposures, comparators, and outcomes.

- 1 Data permitting, the syntheses will also discuss analyses relating to potential susceptible
- 2 populations.<sup>16</sup> These analyses will be based on knowledge about the health outcome or organ
- 3 system affected, demographics, genetic variability, lifestage, health status, behaviors or practices,
- 4 and social determinants (see Table 9-2). This information will be used to draw conclusions

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<sup>16</sup>Various terms have been used to characterize populations that may be at increased risk of developing health effects from exposure to environmental chemicals, including “susceptible,” “vulnerable,” and “sensitive.” Furthermore, these terms have been inconsistently defined across the scientific literature. The term susceptibility is used in this protocol to describe populations or lifestages at increased risk, focusing on intrinsic biological factors that can modify the effect of a specific exposure, but also considering social determinants or behaviors that may increase susceptibility. However, factors resulting in higher exposures to specific groups (e.g., proximity, housing, occupation) will typically not be analyzed to describe increased risk among specific populations or subgroups.

1 regarding potential susceptibility among specific populations or subgroups in a separate section  
 2 (see Section 10.3). This summary will describe concerns across the available evidence for all  
 3 potential human health effects and will be used for both hazard identification and dose-response  
 4 analyses.

**Table 9-2. Individual and social factors that may increase susceptibility to exposure-related health effects**

Factor	Examples
Demographic	Sex, age, race/ethnicity, education, income, occupation, geography
Genetic variability	Polymorphisms in genes regulating cell cycle, DNA repair, cell division, cell signaling, cell structure, gene expression, apoptosis, and metabolism
Lifestage	In utero, childhood, puberty, pregnancy, women of childbearing age, old age
Health status	Pre-existing conditions or disease such as psychosocial stress, elevated body mass index, frailty, nutritional status, chronic disease
Behaviors or practices	Diet, mouthing, smoking, alcohol consumption, pica, subsistence or recreational hunting and fishing
Social determinants	Income, socioeconomic status, neighborhood factors, health care access, and social, economic, and political inequality

EPA ExpoBox Exposure Assessment Tools, based on EPA's *Guidelines for Exposure Assessment* (U.S. EPA, 1992).  
 DNA = deoxyribonucleic acid.

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## 9.1. HUMAN AND ANIMAL HEALTH EFFECTS EVIDENCE

5 The syntheses of the human and animal health effects evidence will focus on describing  
 6 aspects of the evidence that best inform causal interpretations, including the exposure context  
 7 examined in the sets of studies. Each evidence synthesis will be based primarily on studies of *high*  
 8 and *medium* confidence. *Low* confidence studies may be used if few or no studies with higher  
 9 confidence are available to help evaluate consistency, or if the study designs of the *low* confidence  
 10 studies address notable uncertainties in the set of *high* or *medium* confidence studies on a given  
 11 health effect. If *low* confidence studies are used, then a careful examination of risk bias and  
 12 sensitivity with potential impacts on the evidence synthesis conclusions will be included in the  
 13 narrative.

14 As previously described, these syntheses will articulate the strengths and the weaknesses of  
 15 the available evidence organized around the considerations described in Table 9-1, as well as issues  
 16 that stem from the evaluation of individual studies (e.g., concerns about bias or sensitivity). If  
 17 possible, results across studies will be compared using graphs and charts or other data  
 18 visualization strategies. The analysis will typically include examination of results stratified by any

1 or all of the following: study confidence classification (or specific issues within confidence  
2 evaluation domains), population or species, exposures (e.g., level, patterns [intermittent or  
3 continuous], duration, intensity), sensitivity (e.g., low vs. high), and other factors that may have  
4 been identified during study evaluation or analyses of key science issues (see Section 2.4). The  
5 number of studies and the differences encompassed by the studies will determine the extent to  
6 which specific factors can be examined for use in stratifying study results. Additional analyses  
7 across studies (e.g., meta-analyses) may also be conducted for both the human and animal evidence  
8 syntheses, if supported by available data.

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## 9.2. MECHANISTIC INFORMATION

9 The synthesis of mechanistic information informs the integration of health effects evidence  
10 for both hazard identification (i.e., biological plausibility or coherence of the available human or  
11 animal evidence; inferences regarding human relevance, or the identification of susceptible  
12 populations and lifestages across the human and animal evidence) and dose-response evaluation.  
13 As introduced in previous sections, several key science issues that are essential to consider in these  
14 five assessments will involve a focused analysis and synthesis of mechanistic information (see  
15 Sections 9.2.1–9.2.3). Other potential assessment-specific uncertainties for which mechanistic  
16 analyses might be conducted, and the considerations for including those analyses in an assessment,  
17 are outlined in Section 9.2.4. Deviations from the approaches described in Sections 9.2.1–9.2.3, as  
18 well as the specific methods for any analyses conducted based on the considerations described in  
19 Section 9.2.4, will be tracked as updates to the protocol.

20 Mechanistic evidence includes any experimental measurement related to a health outcome  
21 that provides information about the biological or chemical events associated with phenotypic  
22 effects; these measurements can improve understanding of the mechanisms involved in the toxic  
23 effects following exposure to a chemical but are not generally considered adverse outcomes.  
24 Mechanistic data are reported in a diverse array of observational and experimental studies across  
25 species, model systems, and exposure paradigms, including in vitro, in vivo (by various routes of  
26 exposure), ex vivo, and in silico studies, and across a wide spectrum of diverse endpoints.

27 Evaluations of mechanistic information typically differ from evaluations of phenotypic  
28 evidence (e.g., from routine toxicological studies). This is primarily because mechanistic data  
29 evaluations consider the support for and involvement of specific events or sets of events within the  
30 context of a broader research question (e.g., support for a hypothesized mechanism; consistency  
31 with known biological processes), rather than evaluations of individual apical endpoints considered  
32 in relative isolation. Such analyses are complicated because a chemical may operate through  
33 multiple mechanistic pathways, even if one hypothesis dominates the literature ([U.S. EPA, 2005a](#)).  
34 Similarly, multiple mechanistic pathways might interact to cause an adverse effect. Thus, pragmatic  
35 and stepwise approaches to considering and reviewing this evidence for these PFAS assessments  
36 are outlined below. The format of these syntheses is expected to vary from a short narrative

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1 summary of existing knowledge to an in-depth analysis and weighing of the evidence underlying  
2 multiple mechanistic events, depending on data availability and the criticality of the  
3 assessment-specific uncertainties.

### **9.2.1. Toxicokinetic Information and Pharmacokinetic (PK)/Physiologically Based Pharmacokinetic (PBPK) Models**

4 One key mechanistic issue involves the toxicokinetics of these five PFAS, particularly their  
5 serum half-life values because these values are useful for extrapolating doses from exposed animals  
6 to humans. Toxicokinetic studies were extracted for consideration (from the broad PFAS literature  
7 searches) by subject matter experts using two different methods: (1) tagging of studies during  
8 literature screening (see Sections 4.2–4.3), noting that this tagging was not conducted by ADME  
9 subject matter experts, and (2) use of SWIFT Review software  
10 [<https://www.sciome.com/swift-review/>; Howard et al. (2016)] to categorize the literature via  
11 health outcome tags for ADME from the title and abstract. For identification of ADME-related  
12 studies to be reviewed using SWIFT Active Screener  
13 (<https://www.sciome.com/swift-activescreener/>), the results using the health outcome tags for  
14 ADME embedded within SWIFT Review were confirmed using a search string developed by experts  
15 in toxicokinetics within the IRIS Program.<sup>17</sup> This tagging resulted in 813 potentially relevant  
16 studies that were imported into SWIFT Active Screener for review by two independent reviewers  
17 with demonstrated expertise in ADME (conflicts were resolved through discussion). A basic set of  
18 PECO criteria were used for this review:

- 19 • Population: in vivo studies in humans, nonhuman primates, rats, or mice. (Note: in vitro  
20 studies in these species were tagged as potentially supportive; see explanation below.)
- 21 • Exposure: any route of administration of a single chemical compound that is expected to  
22 occur for human exposure for PFBA, PFHxS, PFHxA, PFDA, or PFNA. Exposure to metabolic  
23 precursors of these chemicals was also included. (Note: intraperitoneal [i.p.] injection  
24 studies and in vitro studies were tagged as potentially supportive; see explanation below.)

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<sup>17</sup>tiab: (adme OR admet OR bile OR biliary OR bioavail\* OR biodistribut\* OR biologic-avail\* OR biological-avail\* OR biologically-avail\* OR biotrans\* OR clearance OR detox\* OR distribut\* OR dosim\* OR eliminat\* OR endocytosis OR enterohepatic OR "entero hepatic" OR excret\* OR exhalation OR hepatobiliary OR inhalation OR metaboli\* OR "partition coefficient" OR permeability OR persistence OR phagocytosis OR pharmacokinetic\* OR physiologic-avail\* OR physiological-avail\* OR physiologically-avail\* OR pinocytosis OR protein-bind\* OR reabsorption OR retention OR secretion\* OR toxicokinetic\* OR transport OR uptake OR urination OR ( (absorb OR absorbs OR absorbed OR absorption\* OR deposition) NOT (atomic OR optical OR spectra\* OR spectros\* OR spectrum\* OR infrared)) OR title : ( "gas exchange" AND (alveolar OR lung OR lungs OR pulmonary OR respirat\*)) OR mesh\_mh: ("biological transport" OR "enterohepatic circulation" OR pharmacokinetics) OR mesh\_sh: (pharmacokinetics) OR mesh\_mh: (toxicokinetics)).

This string identified two fewer potentially relevant studies than the SWIFT review (including all studies identified using the non-SWIFT string). So, the studies identified by SWIFT Review were screened in SWIFT Active Screener.

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- 1 • Comparator: vehicle control or reference population.
- 2 • Outcome: data to quantify ADME processes, steady state analysis, empirical
- 3 pharmacokinetic (PK), full PBPK.

4 This screening (i.e., to 96% predicted completion based on the machine-learning software)  
5 resulted in the identification of 99 studies relevant to toxicokinetics across the five PFAS  
6 assessments. These data will be considered for use in the assessments as described below.

7 All PK and PBPK models will be formally evaluated for use in the assessments, as described  
8 in Section 6.4. The specific approaches for determining the most appropriate method for  
9 dosimetric extrapolation, if necessary for these assessments (note: this is likely to be necessary,  
10 based on the preliminary literature inventory), as well as other potential quantitative approaches  
11 for using the PK/PBPK models and ADME data, are outlined in Section 11.2.

12 To draw conclusions regarding the most appropriate serum half-life measures, the ADME  
13 studies identified by the screening methods described above will be considered as outlined in [U.S.](#)  
14 [EPA \(2018b\)](#). Briefly, the studies relevant to updating the data presented in Table 2-7 in  
15 Section 2.4.1, including the studies underlying the current data in the table, will be reviewed, and  
16 data that are highly unreliable will be excluded (e.g., data points below the limit of detection [LOD];  
17 values based on uncertain exposure estimates, or other unvalidated assumptions). Study  
18 characteristics that will be reviewed by subject matter experts to determine whether studies are  
19 informative to the PFAS-specific half-life values include appropriateness of the analytic method, the  
20 number of exposure levels tested, the human relevance of the exposure range, and the number of  
21 time points and tissues sampled. Although ADME data from in vitro studies and i.p. injection  
22 studies were tracked as potentially relevant during screening, additional considerations will apply  
23 to the potential incorporation of these data into the assessments, given their inherent uncertainties  
24 (e.g., difficulties interpreting the relevance of bioavailability or peak concentration data from i.p.  
25 injection studies). Specifically, regarding in vitro studies, it is expected that there may be no in vivo  
26 toxicokinetic data on the rate of conversion of precursor compounds to the PFAS of interest, in  
27 which case conversion rates measured in vitro can be extrapolated to in vivo as the next best means  
28 of predicting this mechanism of exposure. Even if such extrapolation is determined to be  
29 quantitatively uncertain, these data might still provide useful qualitative information.

30 While data and careful PBPK modeling of PFOA and PFOS have revealed nonlinear kinetics  
31 attributed to a mechanism of saturable renal resorption [e.g., [Loccisano et al. \(2011\)](#)], initial  
32 evaluation of PFAS data for the compounds addressed in this protocol do not show such a clear  
33 pattern; that is, studies evaluating PK parameters at high and low doses do not show a significant  
34 dose-dependence in clearance. Such dose-dependence is taken to be distinct from time-dependent  
35 biphasic distribution patterns, whereby an initial, relatively rapid distribution phase is followed by  
36 a slower (terminal) elimination phase. The distribution phase is more rapid because the decline in  
37 blood or plasma concentration reflects both elimination and distribution to peripheral tissues, but

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1 the corresponding half-lives may be independent of dose. EPA's analysis of the PK data will seek to  
2 identify a common elimination-phase half-life (or clearance) among all doses and studies for a given  
3 PFAS in each animal species and sex, or among humans, separated into men and women given  
4 sufficient data. Variation in the rate of absorption (for oral dosing) and distribution phases is  
5 expected to occur between and within studies from random factors that are not dose dependent,  
6 and between tissues within a study due to differing distribution characteristics. Various features of  
7 study design will be considered in evaluating apparent variation (e.g., if the duration of a study is  
8 too short or the sensitivity of the analytic method too low to observe the terminal elimination  
9 phase, such that the apparent clearance is likely to be due to distribution within the animal or  
10 subject). If this analysis reveals a clear dose dependence (nonlinearity) in the elimination phase,  
11 separate from these other sources of variation, the analysis will then focus on identifying and using  
12 the half-life at low doses, considered most relevant to animal-human extrapolation.

13 Because significant differences in the half-life between males and females of a given species  
14 have been observed for some PFAS, these sex differences will be assumed to be real in general  
15 across species. Specifically, when feasible, the data for males and females of each species, for each  
16 PFAS, will be analyzed separately, even if the difference is not statistically significant. If the values  
17 for the elimination-phase half-life differ significantly across studies for the same species/strain/sex,  
18 a more detailed review of the study methods indicated above will be conducted to determine  
19 whether one study is more likely to provide accurate information than another.

20 Given that PFAS inhalation exposure is expected to be via adsorption to particulates, if  
21 sufficient data are available for any of the assessed PFAS, inhalation exposure rates for  
22 PFAS-containing particles will be predicted using the multiple-path particle dosimetry (MPPD)  
23 model (<https://www.ara.com/products/multiple-path-particle-dosimetry-model-mppd-v-304>).  
24 This model predicts inhaled particle deposition in laboratory animals and humans as a function of  
25 particle size. Particle sizes used in controlled animal studies or measured in ambient  
26 environmental or workplace exposure studies in humans will be used as inputs. If PK data are  
27 identified that allow the bioavailability of inhaled particulate PFAS to be estimated, the mass  
28 deposition predicted by the MPPD model will be adjusted accordingly. Otherwise 100% absorption  
29 of PFAS from inhalation deposition will be assumed. Note that while some inhaled particles are  
30 later moved by the mucociliary apparatus to the larynx and swallowed, the PK bioavailability for  
31 oral ingestion can then be applied to that fraction. Any predictions will be considered for use in  
32 comparing findings across oral and inhalation routes of exposure during evidence integration (see  
33 Section 10). In addition, see Section 11.2 for the application of these predictions to developing  
34 quantitative estimates. If necessary, inhalation of PFAS in free ionic form can be estimated based  
35 on the inhalation uptake of other chemicals with high liquid:air partition coefficients (i.e., assuming  
36 nearly complete absorption of any free ions that contact the airway lining).

### **9.2.2. Peroxisome Proliferator-Activated Receptor Alpha (PPAR $\alpha$ ) Dependence for Health Effect(s) Observed in Animals**

1           A second area of focused mechanistic analysis is evaluating the human relevance of effects  
2 in animals that appear to involve (at least in part) a PPAR $\alpha$ -mediated MOA. The approach outlined  
3 below focuses on hepatic effects, which are expected to be the primary health effect area in these  
4 assessments for which this analysis is useful, and for which there are likely to be data for analysis.  
5 The specifics of applying this approach may vary across the five PFAS assessments, depending on  
6 the availability of data to address this question and the strength of the evidence indicating PPAR $\alpha$   
7 involvement. During assessment development, for other health effects with evidence that a  
8 PPAR $\alpha$ -mediated MOA might be operant, the mechanistic syntheses will include consideration of  
9 this issue. These analyses will depend on the amount of information available and the strength of  
10 the evidence indicating PPAR $\alpha$  involvement. Thus, the analyses might range from a short summary  
11 of the available evidence when data are sparse to an evaluation approximating the one described  
12 below when extensive data are available.

13           To identify the literature most relevant to addressing the question of the  
14 PPAR $\alpha$ -dependence of hepatic effects observed in experimental animals, a PFAS assessment with  
15 extensive evidence of liver effects and potential PPAR $\alpha$  involvement will screen<sup>18</sup> the “potentially  
16 relevant supplemental material” studies on a given PFAS at the full-text level as follows:

- 17           • Population: in vivo animal studies in mammalian models; in vitro and human experiments  
18           using primary or immortalized liver cell lines
- 19           • Exposure: PFAS of interest (parent compound only)
- 20           • Comparator: vehicle control
- 21           • Outcome: mechanistic outcomes relevant to the hepatobiliary system (e.g., in liver tissues or  
22           cells)

23           Any additional assessment-specific strategies for identifying other information of potential  
24 relevance on molecular mechanistic data for these five PFAS, or from the more extensive literature  
25 on perfluorooctanoic acid (PFOA) and PFAS (e.g., as points of comparison), will be described as  
26 updates to this protocol.

27           The pool of studies identified based on the strategies outlined above will be inventoried into  
28 a database to allow for the organization and evaluation of these data. Specifically, the following  
29 information will be extracted for each reference: a reference identifier; test compound; exposure

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<sup>18</sup>Although the specifics of this screening process may vary across PFAS, this protocol describes that screening will occur by at least two reviewers and use of DistillerSR to track decisions and resolve differences. Any deviations from this will be tracked on an assessment-specific basis as updates to the protocol.

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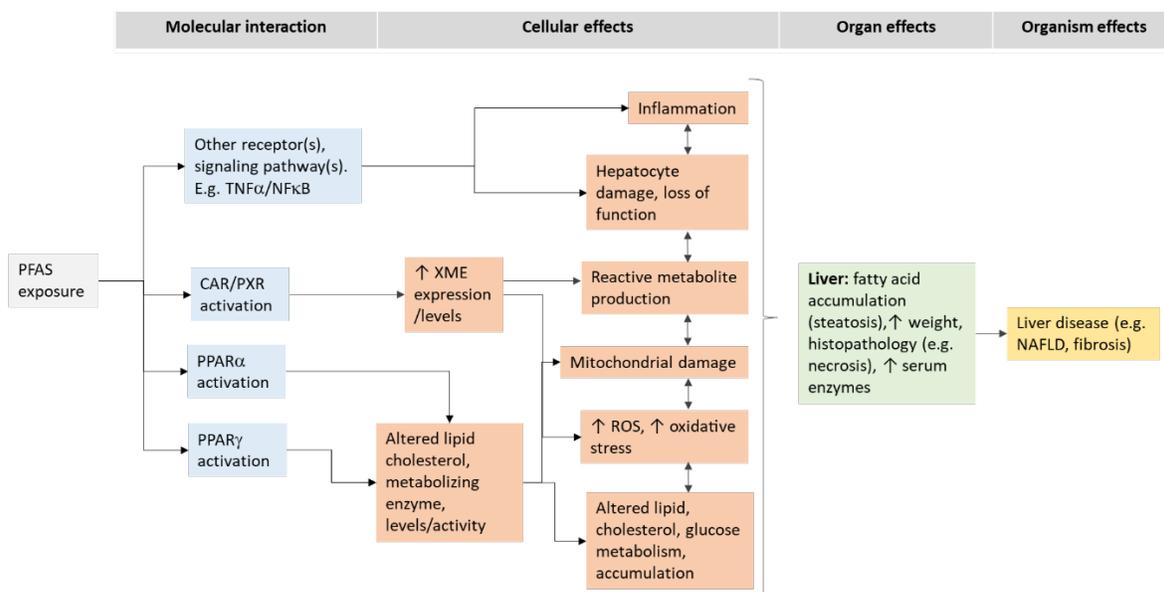
1 route and duration; the sex, species, and strain of the organism; age at exposure; and endpoint  
2 evaluation of the test organism or test system. Additionally, the inventory(ies) will capture a  
3 succinct description of the assessed endpoints and the potential mechanistic event(s) informed by  
4 those endpoints in each study. The mechanistic events in the proposed mechanisms pathway for  
5 which there are data will then be organized according to the following levels of biological  
6 organization: molecular target(s), cellular response(s), tissue/organ response(s), and organism  
7 response(s), in accordance with the levels of biological organization used to develop adverse  
8 outcome pathways (AOPs).<sup>19</sup>

9 Although refinements based on the assessment-specific evidence are anticipated, these  
10 assessments will first consider the use of the preliminary pathway outlined in Figure 9-1 as an  
11 organizing AOP for these data. The preliminary, proposed AOP displayed in Figure 9-1 is based on  
12 molecular initiating events, key events, and adverse outcomes identified in previous evaluations on  
13 PFOS and PFOA and proposed AOPs for chemical-induced noncancer liver toxicity [see [Li et al.,  
14 2017](#), [Mellor et al. \(2016\)](#), [Wang et al. \(2014\)](#), [U.S. EPA \(2016d\)](#), [U.S. EPA \(2016d\)](#), [ATSDR \(2018\)](#),  
15 and [NJDWQI \(2017\)](#)]. Prior evaluations of PFOS and PFOA have discussed studies using wild-type,  
16 PPAR $\alpha$  knockout and humanized PPAR $\alpha$  (hPPAR $\alpha$ ) mice showing that exposure leads to fatty acid  
17 and triglyceride accumulation in the liver and steatosis via both PPAR $\alpha$ -dependent  
18 and -independent pathways ([ATSDR, 2018](#); [Li et al., 2017](#); [Viberg and Eriksson, 2017](#)). In addition  
19 to PPAR $\alpha$ , these reviews have implicated other nuclear receptor (NR) and cell signaling pathways  
20 with PFOA- and PFOS-induced noncancer liver effects, including PPAR $\beta/\delta$ , PPAR $\gamma$ , constitutive  
21 androstane receptor (CAR) and pregnane X receptor (PXR), the farnesoid X receptor (FXR), the  
22 phosphatidylinositol 3-kinase-serine/threonine kinase Akt (PI3K-Akt) signal transduction pathway,  
23 and the nuclear factor kappa B pathway (NF- $\kappa$ B) ([Li et al., 2017](#); [Viberg and Eriksson, 2017](#)).  
24 Activation of these pathways can be associated with alterations in lipid and glucose metabolism,  
25 increased cellular stress, and inflammation ([Mackowiak et al., 2018](#); [Li et al., 2017](#); [Mellor et al.,  
26 2016](#); [Wang et al., 2014](#)). Thus, the potential involvement and contribution of these different  
27 signaling responses to hepatic effects after exposure to the five PFAS will also be considered.

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<sup>19</sup>Although the World Health Organization (WHO)-International Programme on Chemical Safety (IPCS)-MOA and the Organisation for Economic Co-operation and Development (OECD)-AOP frameworks are similar in the identification and analysis of key events following modified Bradford Hill criteria ([Meek et al., 2014](#)), AOPs are chemical agnostic whereas MOA analyses are intended to inform health assessments of individual (or groups of) chemical(s) ([Edwards et al., 2016](#)).

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**Figure 9-1. Preliminary proposed mechanistic pathway for per- and polyfluoroalkyl substances (PFAS)-induced noncancer liver effects.** Based on previous reviews of perfluorooctane sulfonate (PFOS)- and perfluorooctanoic acid (PFOA)-induced noncancer liver effects in animals ([ATSDR, 2018](#); [Li et al., 2017](#); [Viberg and Eriksson, 2017](#); [U.S. EPA, 2016c, d](#)), and proposed adverse outcome pathways for hepatic steatosis ([Mellor et al., 2016](#)).

NAFLD = nonalcoholic fatty liver disease; ROS = reactive oxygen species; TNF $\alpha$  = tumor necrosis factor alpha; XME = xenobiotic metabolizing enzymes.

1           The analysis of the involvement of PPAR $\alpha$  and these other signaling cascades in hepatic  
 2 toxicity after exposure to these five PFAS will focus on the concordance of changes in the specific  
 3 mechanistic events or separate pathways to effects (i.e., in Figure 9-1, and as otherwise identified  
 4 during assessment-specific evaluations) across species to ascertain the relevance of animal studies  
 5 to human health. The analyses of evidence for each mechanistic event and potential pathway will  
 6 be qualitatively analyzed for various aspects of the Hill considerations outlined in the EPA Cancer  
 7 Guidelines framework for MOA analysis ([U.S. EPA, 2005a](#)). Given the focus of these analyses, the  
 8 review will stress the aspects of consistency, coherence, and biological plausibility to ascertain the  
 9 level of support (or lack thereof), depending on the availability of data. To facilitate this analysis,  
 10 the following prompting questions and clarifying considerations will be used, depending on the  
 11 assessment-specific data:

- 12           • What is the level of evidentiary support (or lack thereof) for the mechanistic events or  
 13 signaling pathways, based on the assessment-specific PFAS data? In parallel, are  
 14 assessment-specific data available to inform the strength of the linkages between events in  
 15 the pathway or across pathways? In general, well-conducted, independent studies using  
 16 different experimental models and reporting consistent or coherent findings would provide

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1 strong supportive evidence for a mechanistic (potentially key) event or pathway (or  
2 linkages between events in a pathway), with a lesser degree of support provided by  
3 individual experimental observations or sets of studies reporting some consistent or  
4 coherent findings as well as some equivocal results or findings that vary from one model to  
5 another without explanation.

- 6 • Are sufficient assessment-specific data available to inform exposure duration- or  
7 level-dependencies for any of the evaluated mechanistic events or pathways?
- 8 • Is the assessment-specific evidence (on specific events or pathways in general) consistent  
9 with the general biology of the human liver or mechanisms known to be associated with  
10 noncancer liver effects in humans? To consider this question, assessments will compare the  
11 endpoint-level results across studies on a particular PFAS against the mechanistic  
12 understanding/underlying biology for similar effects in the human liver. (Note: this analysis  
13 might be informed by studies or reviews on the more robust PFOA/perfluorooctane  
14 sulfonate [PFOS] evidence bases.)
- 15 • Are responses across studies for these five PFAS assessments indicative of activation of  
16 specific mechanisms or signaling pathways conserved across experimental models and  
17 designs? To consider this question, assessments will include an evaluation of consistency  
18 and coherence across different species and strains of animals, human and animal cell  
19 culture models, and in vivo humanized animal models, depending on data availability.
- 20 • Does the assessment-specific mechanistic information indicate the likelihood of populations  
21 or lifestyles that may be more susceptible to PFAS-induced liver effects?

22 The assessment-specific conclusions (and attendant uncertainties) regarding these  
23 questions will be used to draw judgments regarding the human relevance of these animal effects,  
24 and the rationale for these judgments will be documented transparently within each assessment.  
25 As described in EPA guidance ([U.S. EPA, 2005a](#)), human relevance is the default, and mechanistic  
26 evidence will need to be compelling and strong to conclude otherwise (i.e., to conclude that findings  
27 in animals are not relevant to humans).

### **9.2.3. Toxicological Relevance of Select Outcomes Observed in Animals**

28 The preliminary literature inventory identified studies on several health outcomes relating  
29 to potential urinary and hepatic effects (see below) for which it is expected to be difficult to  
30 determine whether any observed changes (or a lack of changes) are toxicologically relevant. It is  
31 expected that in some instances, the synthesis will need to address this issue to inform whether the  
32 effects in animals are relevant to interpreting the potential for PFAS exposure to cause a human  
33 health effect, and in other instances addressing this issue may be necessary to identify a level of  
34 change for use in determining the potential for adversity or in dose-response analysis. It is possible  
35 that additional outcomes with similar questions of health relevance might be identified during the  
36 development of these assessments. If so, the specifics of the approach selected to address those

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1 outcomes will be documented in the assessment(s) and as an update to this protocol. For the  
2 aforementioned outcomes, different approaches will be taken, specifically:

- 3 1) *Kidney changes in rats, including chronic progressive nephropathy (CPN) and effects that*  
4 *appear to be mediated by an alpha 2u-globulin MOA.* Because the rodent (i.e., male  
5 rat)-specific alpha 2u-globulin MOA is not considered relevant to humans, assessments with  
6 evidence indicating its involvement will include an evaluation and judgment of the evidence  
7 supporting (or not supporting) dependence on this MOA. Specifically, these data will be  
8 evaluated against the predefined criteria established by the [U.S. EPA \(1991a\)](#) and/or more  
9 recently established criteria, such as those published by [Swenberg and Lehman-McKeeman](#)  
10 [\(1999\)](#). Relatedly (and possibly overlapping the evaluation of alpha 2u-globulin, because  
11 this MOA may exacerbate CPN), there is no human disease analog to the constellation of  
12 changes observed in rodent CPN. CPN represents a complex disease process in rats, and its  
13 etiology is unknown. Thus, these evaluations will include judgments as to whether all or a  
14 subset of the observed changes have adequate evidence to identify dependence on  
15 rodent-specific processes, including whether it can be concluded (i.e., based on biological  
16 understanding) that the observed kidney endpoints are associated with CPN and the  
17 potential for exacerbation of human-relevant disease processes can be ruled out. Data  
18 permitting, the assessments will consider whether these conclusions vary by exposure level.
- 19 2) *Hepatic changes.* Some individual liver endpoints (and even some constellations of  
20 endpoints) might be considered adaptive in nature, possibly leading to the interpretation  
21 that some statistically significant changes are not indicative of adverse effects. These  
22 endpoints include increased liver weight, cellular hypertrophy, and single cell  
23 necrosis/apoptosis. To draw inferences regarding the adversity of these types of liver  
24 effects, these assessments will consider the panel recommendations outlined by [Hall et al.](#)  
25 [\(2012\)](#) to draw assessment-specific judgments regarding adversity. Briefly, these include  
26 evaluation of the available histological data and results suggesting structural degeneration  
27 or cellular demise (e.g., apoptosis, oncosis, and/or necrosis), and clinical evidence of  
28 hepatocyte damage. As the recommendations were developed in the context liver tumor  
29 formation, consultation of additional relevant information will be considered to interpret  
30 the adversity of noncancer liver effects over a lifetime exposure, taking into account that  
31 effects perceived as adaptive can progress into more severe responses and lead to cell  
32 injury ([Hall et al., 2012](#)). These considerations include the EPA 2002 Guidance Document on  
33 Hepatocellular Hypertrophy ([U.S. EPA, 2002a](#)), reference materials on clinical and  
34 histopathology data ([Thoolen et al., 2010](#); [EMEA, 2008](#); [Boone et al., 2005](#)) and publications  
35 describing potential mechanisms of chemical-induced liver disease such as fatty liver  
36 disease/steatohepatitis ([Wahlang et al., 2019](#); [Joshi-Barve et al., 2015](#); [Wahlang et al., 2013](#)).  
37 Each assessment will include an explanatory rationale documenting the application of the  
38 [Hall et al. \(2012\)](#) recommendations (and any other considerations) to the available  
39 evidence.

#### **9.2.4. Other Focused Mechanistic Analyses**

1 Other analyses within the syntheses of mechanistic information will focus on the evidence  
2 most useful for informing key uncertainties in the human or animal health effect evidence, both  
3 qualitative and quantitative.

4 This means that, for example, if extensive and consistent *high* confidence human or animal  
5 evidence is available, the need to synthesize all relevant mechanistic evidence will likely be  
6 diminished. In these cases, the analyses will focus on reviewing and interpreting smaller sets of  
7 mechanistic studies that specifically address controversial or outstanding issues that are expected  
8 to have a substantial impact on the assessment conclusions. Generally, key uncertainties will be  
9 addressed in the mechanistic evidence syntheses by considering the biological understanding of  
10 how the effect(s) in question develop or are related. In this way, the analyses can provide  
11 information on, for example, (1) potential precursor events when the apical data are uncertain (or  
12 unusable for dose-response analyses), (2) animal results for which the human relevance is unclear  
13 or controversial and the human evidence is weak, (3) the shape of the dose-response curve at low  
14 exposure levels when this understanding is highly uncertain and data informing this uncertainty  
15 are known to exist, or (4) the identification of likely susceptible populations and lifestages. Thus,  
16 consideration of biological understanding represents an important component of the evidence  
17 analysis. However, mechanistic understanding is not a prerequisite for drawing a conclusion that a  
18 chemical causes a given health effect ([NTP, 2015](#); [NRC, 2014](#)).

19 To identify the focused set(s) of studies to use in analyzing critical mechanistic questions  
20 other than those outlined in Sections 9.2.1–9.2.3, a stepwise approach will be applied to  
21 progressively define the scope of the mechanistic information to be considered throughout  
22 assessment development. This stepwise scoping begins during the literature search and screening  
23 steps and depends primarily on the potential health hazard signals that arise from the individual  
24 human and/or animal health effect studies, or from mechanistic studies that signal potential health  
25 hazards not examined in studies of phenotypic, potentially adverse effects. Examples of the focused  
26 questions or scenarios triggering these mechanistic evaluations, as well as when during the  
27 systematic review they are likely to apply, are listed in Table 9-3. While the specific methods for  
28 evaluating the evidence most relevant to each question will vary, some general considerations for  
29 judging the evidence strength in these syntheses are provided below, and if necessary,  
30 assessment-specific refinements will be included as updates to the protocol.

**Table 9-3. Examples of questions and considerations that can trigger focused analysis and synthesis of mechanistic information**

Key assessment-specific uncertainties	Examples of questions and PFAS-specific considerations for identifying the uncertainties and key evidence to analyze
Addressing database completeness based on literature inventories of human, animal, and mechanistic information	<ul style="list-style-type: none"> <li>• Are there mechanistic studies on an organ system or potential health hazard that were not examined by human or animal studies meeting the PECO criteria?                             <ul style="list-style-type: none"> <li>○ Depending on the extent of the available data, consider the utility of developing a separate synthesis of evidence versus the utility of a concise, narrative summary (or evidence mapping) to describe these knowledge gaps. Consider whether the mechanistic evidence might be sufficient to substantiate a conclusion on its own (if so, a separate synthesis will be developed).</li> </ul> </li> </ul>
Addressing questions of inconsistency within the human and animal evidence	<ul style="list-style-type: none"> <li>• For the health effects of potential concern, is a mechanistic evaluation(s) warranted to inform questions regarding the consistency of the available human or animal studies? Typically, this consideration would focus on health effects that show some indication of an association in epidemiological studies or causality in experimental studies during evidence synthesis. Based on the literature inventory, consider whether mechanistic data are available to inform the specific, key uncertainties in question. Examples of specific scenarios for evaluation include:                             <ul style="list-style-type: none"> <li>○ If cancer has been observed and tumor types appear to differ across populations (e.g., species or sex), review the literature inventory for mechanistic data that might be relevant to interpreting such differences, and conduct analyses as warranted based on that review. Approaches outlined in the EPA Cancer Guidelines (<a href="#">U.S. EPA, 2005a</a>) that may be relevant to these analyses will be applied, as appropriate.</li> <li>○ If pronounced and unexplained differences in health effect(s)-specific responses are observed across lifestages or populations (e.g., animal strain; human demographic), first consider toxicokinetic differences for the specific PFAS, and then the mechanistic evidence relevant to assessing the potential for health effect-specific biological differences in response (toxicodynamics). Further, inconsistent evidence (i.e., heterogeneous results) across different animal species or human populations might be clarified by a review of the evidence relevant to whether different mechanisms may be operant in the different populations (e.g., evidence demonstrating that certain species are more or less sensitive to a certain biological perturbation; evidence that gene polymorphisms are related to variability in response).</li> </ul> </li> </ul>

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Key assessment-specific uncertainties	Examples of questions and PFAS-specific considerations for identifying the uncertainties and key evidence to analyze
Addressing questions of biological plausibility <sup>a</sup> and coherence within the human and animal evidence, and coherence across bodies of evidence	<ul style="list-style-type: none"> <li>• For the health effects of potential concern, would a mechanistic evaluation(s) of biological plausibility (usually for an individual outcome) or coherence (usually across outcomes) provide meaningful information for interpreting the evidence strength? Typically, this consideration would focus on effects for which the evidence strength for an individual outcome (either for or against an effect) is questionable (e.g., primarily studies of <i>low</i> confidence), when a substantial outstanding methodological concern(s) across the relevant studies exists, or when evidence exists for multiple, potentially related (e.g., biologically) outcomes. Based on the literature inventory, consider whether there are mechanistic data available to inform the specific, key uncertainties in question. Examples of specific scenarios for evaluation may include:               <ul style="list-style-type: none"> <li>○ If the evidence for a given outcome is weak or uncertain, or when unaddressed methodological concerns identify critical uncertainties in the human or animal findings for a health effect, identify data on mechanistic changes in exposed humans or animals that are likely to be linked to the development or occurrence of the health outcome in question. If enough suitable studies are available, analyze data on changes expected to be related to the phenotypic finding(s) of interest, which can either increase or decrease the evidence strength that the finding(s) is real. It is important to note that the absence of a mechanistic explanation for an association (e.g., the MOA is not understood) will not be used to reduce confidence in observations from human or animal studies. However, the plausibility of an association observed in human or animal studies may be diminished if expected mechanistic findings (e.g., based a known biological dependence) are tested and not apparent. The mechanistic evidence on possible precursors or effects that are known to co-occur with the health outcome of interest are particularly impactful when the changes are observed in the same exposed population presenting the outcome of interest. An understanding of mechanistic pathways (e.g., by identifying and analyzing mechanistic precursor events linked qualitatively or quantitatively to apical health effect[s]; see Section 9.2.2 for additional context) can inform the strength of the evidence integration judgments (see Section 10).</li> <li>○ If evidence on multiple health outcomes within an organ system, or possibly across organ systems (e.g., thyroid and nervous system), is available and the strength of the evidence for any single outcome is uncertain, identify biological data that can inform understanding of the relatedness of outcomes within and across systems. Biological understanding or strong mechanistic support (e.g., a shared mechanistic event) for linkages across outcomes can increase the strength of the evidence when changes are related. However, evidence strength may be diminished if an expected pattern among biologically linked outcomes is not observed. Interpretation of the pattern of changes across the outcomes will consider the underlying biological understanding (e.g., one outcome may be expected to precede the other, or be more sensitive). These same considerations inform analyses of the coherence of observed effects across evidence streams during evidence integration (see Section 10.2).</li> </ul> </li> </ul>

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Key assessment-specific uncertainties	Examples of questions and PFAS-specific considerations for identifying the uncertainties and key evidence to analyze
Addressing questions on the human relevance of findings in animals	<ul style="list-style-type: none"> <li>• For the health effects of potential concern, does the available evidence raise questions of human relevance? Typically, this consideration applies when human evidence is lacking or has results that differ from animal studies, given that responses can differ between humans and animals [e.g., for cancer, site concordance is not a requirement for determining the relevance of animal data for humans (<a href="#">U.S. EPA, 2005a</a>); for noncancer nervous system effects, behavioral changes can manifest differently between animals and humans]. The identification of potential differences will also consider ADME information across species, primarily relating to distribution (e.g., to the likely target tissue) and PFAS half-life. Examples of information to identify from the literature inventory, as well as specific scenarios and considerations for these analyses may include:               <ul style="list-style-type: none"> <li>○ If there is no evidence indicating that the animal results are irrelevant to humans, summarize existing knowledge on the development of the health effect in each species, including potential differences in PFAS toxicokinetics, and assess the relatedness across species. Note that in the absence of sufficient evidence to the contrary, effects in animal models are assumed to be relevant to humans (<a href="#">ATSDR, 2018</a>; <a href="#">NTP, 2015</a>; <a href="#">U.S. EPA, 2005a</a>).<sup>b</sup></li> <li>○ If there is evidence indicating that the mechanisms underlying the effects in animals may not operate in humans, or that the available animal model(s) may not be suitable for the human health outcome(s) of interest, present and analyze the strength of the evidence for and against the human relevance of the observed findings. In addition to considerations specific to the outcome of interest, the analysis will evaluate observations of mechanistic changes in exposed humans for similarities or biological coherence with mechanistic or toxicological changes in experimental animals interpreted to be associated with the health outcome under evaluation. It may also include an evaluation of findings across species known or presumed to be more or less relevant for interpreting potential human toxicity for the health effect(s) in question. In rare instances or for controversial decisions that are likely to drive key assessment conclusions, the analysis may extend to a detailed analysis of a plausible mechanistic pathway(s) or MOA(s) within which each key event and key event relationship is evaluated regarding the likelihood of similarities (e.g., in presence or function) across species. These analyses, regardless of their rigor, will lead to a definitive judgment about whether the animal response is relevant to humans during evidence integration (see Section 10).</li> </ul> </li> </ul>

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<p align="center"><b>Key assessment-specific uncertainties</b></p>	<p align="center"><b>Examples of questions and PFAS-specific considerations for identifying the uncertainties and key evidence to analyze</b></p>
<p><b>Addressing questions on potential susceptibility for hazard identification and dose-response analysis</b></p>	<ul style="list-style-type: none"> <li>• For the health effects of potential concern, do the results from the human and animal health effect studies appear to differ by categories that indicate the apparent presence of susceptible populations (e.g., across demographics, species, strains, sexes, or lifestages)? Separately, are there human or animal study data that could identify or clarify population differences in response (e.g., experiments testing sensitivity of responses across lifestages or across genetic variations; observed differences attributable to genetic polymorphisms)? Are there mechanistic data (i.e., based on the literature inventory) that address potential susceptibility factors?<sup>c</sup> If evidence exists for any of these scenarios, information on susceptibility will be reviewed and, if impactful to assessment conclusions, analyzed in detail. Examples of when these analyses are important include:             <ul style="list-style-type: none"> <li>○ If the analysis of evidence indicates the likely presence of a sensitive population or lifestage in humans, the groups likely to be at greatest risk will be captured in the evidence integration narrative (see Section 10). In addition, this narrative will discuss whether the appropriate analogous exposures and populations or lifestages were adequately represented or tested in the available human or animal studies, and if not, will identify studies on the most susceptible populations or lifestages as key research needs (see Section 10).</li> <li>○ If the analysis of evidence indicates the likely presence of a sensitive population or lifestage in humans, this information will be used to select studies for quantitative analysis (e.g., prioritizing those studies that include such populations [see Section 11]). If specific studies addressing these susceptibilities are unusable for quantitative analysis, susceptibility data may be used to support refined human variability uncertainty factors or probabilistic uncertainty analyses (see Section 11).</li> </ul> </li> </ul>

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Key assessment-specific uncertainties	Examples of questions and PFAS-specific considerations for identifying the uncertainties and key evidence to analyze
Addressing questions on biological understanding to optimize dose-response analysis	<ul style="list-style-type: none"> <li>• If the human and/or animal health effect data amenable to dose-response analysis are weak<sup>d</sup> or only at high exposure levels, or if the selection of critical parameters for modeling is uncertain, the following analyses will be considered:               <ul style="list-style-type: none"> <li>○ When the apical health effect data are highly uncertain or cannot be used with confidence for the purpose of deriving quantitative estimates, mechanistic precursor events linked qualitatively or quantitatively to the phenotypic effect can be evaluated for use as surrogate markers (e.g., based on the strength and completeness of the linkage between mechanistic and phenotypic effects) for deriving quantitative estimates.</li> <li>○ When understanding of the appropriate exposure metric, biomarker, or modeling parameter for developing quantitative estimates is notably lacking, then toxicokinetic and mechanistic understanding of the development of the health effect can inform the most biologically appropriate measure.</li> <li>○ When there are dose-response modeling decisions or uncertainties that would be substantially improved by biological or toxicokinetic understanding, mechanistic analyses can improve selection of particular models (e.g., a linear, nonlinear, or threshold model) and help evaluate the appropriateness of integrating/combining data across related outcomes (e.g., based on biological coherence or a conserved MOA). For cancer toxicity values, existing guidance will be consulted (<a href="#">U.S. EPA, 2005a</a>).</li> </ul> </li> </ul>

<sup>a</sup>As applied herein, biological plausibility describes mechanistic information that either strengthens or weakens an interpretation of the likelihood of an association between exposure and the health effect. The interpretation of biological plausibility considers the existing biological understanding of how the health effect develops and can involve analyses of information at different levels of biological complexity (e.g., molecular, cellular, tissue).

<sup>b</sup>As described in the EPA RfD/RfC Technical Report (2002), “one of the major default assumptions in EPA’s risk assessment guidelines is that animal data are relevant for humans [e.g., [U.S. EPA \(1998\)](#), [U.S. EPA \(1991a\)](#), and [U.S. EPA \(1996a\)](#)]. Such defaults are intended to be used in the absence of experimental data that can provide direct information on the relevance of animal data” ([U.S. EPA, 2002b](#)).

<sup>c</sup>Susceptibility factors include lifestage, demographics and social determinants, behavioral factors, health status, and genetic variability. Although not considered in these analyses, factors that can increase vulnerability, such as other pollutant exposures or differential proximity to exposure sources, are typically considered during a full risk assessment.

<sup>d</sup>Note that “weak” here refers to the study’s usability for dose-response analysis specifically. Such studies may be judged to be of *medium* or *high* confidence for the purposes of identifying potential hazards but possess limitations preventing their use for deriving reliable quantitative estimates.

PECO = populations, exposures, comparators, and outcomes; RfC = inhalation reference concentration; RfD = oral reference dose.

- 1 If focused areas for additional mechanistic evaluations are identified that can help address
- 2 key assessment-specific uncertainties (e.g., by applying Table 6-3), the assessments will identify the
- 3 most influential studies for evaluation. This could represent only a subset of the potentially
- 4 relevant studies, particularly if there are many mechanistic studies relevant to the specific

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1 question(s). Because the potential influence of the information provided by the available studies  
2 can vary depending on the question(s) or the associated mechanistic events or pathways, the rigor  
3 of the analyses will likewise vary from cursory insights drawn from sets of unanalyzed results to  
4 detailed evaluations of a subset of the most relevant, individual mechanistic studies. Although the  
5 specifics that might be applicable across potential mechanistic topic areas cannot be predefined, the  
6 analyses will first consider the studies based on their toxicological relevance to answering the  
7 specific question (e.g., model system; specificity of the assay for the effect of interest), potentially  
8 refining the focus to a subset of the most relevant studies. This will be particularly important when  
9 the set(s) of studies are inconsistent and potentially conflicting. If available, emphasis will  
10 generally be placed on more informative studies that challenge the necessity of proposed  
11 mechanistic relationships between exposure and an apical effect (e.g., altering a receptor-mediated  
12 pathway through chemical intervention or using knockout animals). The analysis may also  
13 consider whether particular study design aspects in some or all of the relevant studies are likely to  
14 have significant flaws or important uncertainties (e.g., for certain questions, a preliminary review of  
15 the exposure methods across the relevant mechanistic studies can flag serious deficiencies). In  
16 general, across these assessments, relevant mechanistic information from in vivo studies will be  
17 prioritized, with preference given to PFAS- and endpoint-relevant exposure routes and exposure  
18 designs. Analysis of ex vivo and in vitro studies will then be considered, prioritizing those most  
19 informative for evaluating the mechanistic events indicated by the in vivo data, including studies  
20 conducted under conditions most relevant to human exposures and in model systems best  
21 replicating in vivo human biology.

22 In some instances, additional literature searches may be warranted, targeting mechanistic  
23 events or biological pathways that are not specific to a particular PFAS or group of PFAS. When  
24 more rigorous mechanistic analyses are deemed necessary, the review will be aided by using  
25 pathway-based organizational methods and, if available, established evidence evaluation  
26 frameworks. These approaches provide transparency and objectivity to integrate and interpret the  
27 mechanistic events and pathways anchored to the specific questions that have been identified  
28 (e.g., anchored to a specific health effect) across diverse sets of relevant data (e.g., human, animal,  
29 and in vitro studies). The approaches may be facilitated by using organizational tools or  
30 frameworks, such as AOPs (see example in Section 9.2.2). As noted above, any additional  
31 assessment-specific literature searches and evaluation methods will be described in updates to the  
32 protocol.

33 Based on the analyses and considerations outlined in Sections 9.2.1–9.2.4, the results of the  
34 health effect- and assessment-specific mechanistic evidence syntheses will inform both evidence  
35 integration and dose-response analyses (see Sections 10 and 11).

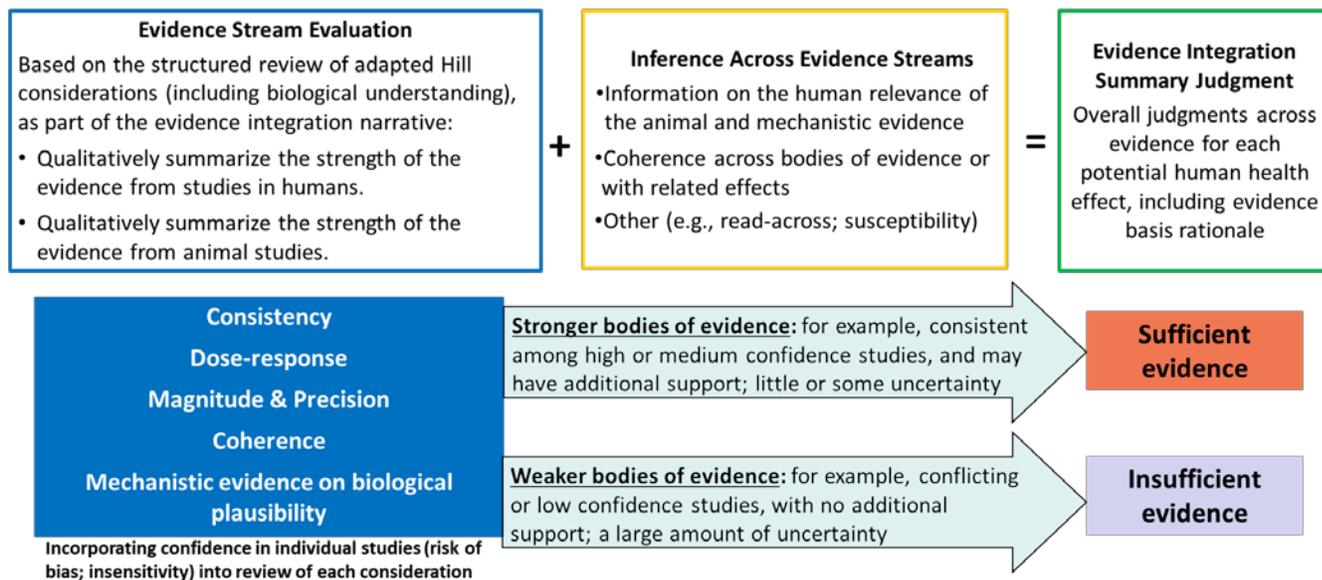
## 10. EVIDENCE INTEGRATION

1 For analyzing human health outcomes that might result from chemical exposure, these PFAS  
2 assessments will draw integrated judgments across human, animal, and mechanistic evidence for  
3 each assessed health effect. The evidence integration judgments include interpretations drawn  
4 regarding the support provided by the individual bodies of evidence (i.e., human, animal, and  
5 mechanistic evidence) based on the structured application of an adapted set of considerations first  
6 introduced by Austin Bradford Hill ([Hill, 1965](#)), which are directly informed by the summary  
7 discussions of each body of evidence during evidence synthesis (see Section 9). As previously  
8 discussed in Section 9.2, the approach to evaluating the mechanistic evidence relevant to each  
9 assessed health effect will follow a stepwise approach and is expected to vary depending on the  
10 nature and impact of the uncertainties identified within each evidence base, as well as the specific  
11 mechanistic information available to address those uncertainties. This includes evaluations of  
12 mechanistic evidence relevant to the identified key science issues (see Section 2.4) prior to or in  
13 parallel with evaluations of the phenotypic data in human and animal studies, as well as other  
14 focused mechanistic analyses identified during draft development to address key assessment  
15 uncertainties (see Section 9.2.4 for a discussion of these scenarios). During evidence integration, a  
16 structured and documented process will be used, as follows (and depicted in Figure 10-1):

- 17 • Building from the separate syntheses of the human and animal evidence (see Section 9.1),  
18 the strength of the evidence from the available human and animal health effect studies will  
19 be summarized in parallel, but separately, using a structured evaluation of an adapted set of  
20 considerations first introduced by Austin Bradford Hill ([Hill, 1965](#)). Table 10-2 describes  
21 these structured evaluations and the explicit consideration of study confidence within each  
22 evaluation domain. Based on the approaches and considerations described in Section 9.2,  
23 these summaries will incorporate the relevant mechanistic evidence (or MOA  
24 understanding) that informs the biological plausibility and coherence within the available  
25 human or animal health effect studies.
- 26 • The strength of the animal and human evidence will be considered together in light of  
27 inferences across evidence streams. Specifically, the inferences considered during this  
28 integration include the human relevance of the animal and mechanistic evidence, coherence  
29 across the separate bodies of evidence, and other important information (e.g., judgments  
30 regarding susceptibility). Note that without evidence to the contrary, the human relevance  
31 of animal findings is assumed.

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- 1 • A summary judgment is drawn as to whether the available evidence base for each potential  
 2 human health effect as a whole is sufficient (or insufficient) to indicate that PFAS exposure  
 3 has the potential to be hazardous to humans.<sup>20</sup>



**Figure 10-1. Process for evidence integration.** Note that “sufficient evidence” could indicate a judgment of “sufficient evidence for hazard” or “sufficient evidence to judge that a hazard is unlikely,” depending on the nature and extent of the available evidence (see Table 10-3).

4 The decision points within the structured evidence integration process will be summarized  
 5 in an evidence profile table for each health effect category (see Table 10-1 for a preliminary  
 6 template version) in support of the evidence integration narrative. The specific decision  
 7 frameworks for the structured evaluation of the strength of the human and animal evidence  
 8 streams and for drawing the overall evidence integration judgment are described in Section 10.1.  
 9 This process is similar to that used by the Grading of Recommendations Assessment, Development,  
 10 and Evaluation [GRADE; [Morgan et al. \(2016\)](#); [Guyatt et al. \(2011\)](#); [Schünemann et al. \(2011\)](#)],  
 11 which arrives at an overall integration conclusion based on consideration of each body of evidence.  
 12 As described in Section 9, the human, animal, and mechanistic evidence syntheses serve as inputs  
 13 providing a foundation for the evidence integration decisions; thus, the major conclusions from  
 14 these syntheses will be summarized in the evidence profile table (see Table 10-1 for a preliminary  
 15 template version) supporting the evidence integration narrative. The evidence profile tables on

<sup>20</sup>Due to the expected rarity of scenarios where there is “sufficient evidence to judge that a hazard is unlikely” (see description in Table 10-3 and Section 10.2) and to improve readability, this judgment is not specified in some instances.

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- 1 each potential human health effect evaluated will summarize the judgments and evidence basis for
- 2 each step of the structured evidence integration process. Separate sections are included for
- 3 summarizing the human and animal evidence, for the inference drawn across evidence streams, and
- 4 for the overall evidence integration judgment. The table presents the key information from the
- 5 different bodies of evidence that informs each decision.

Table 10-1. Evidence profile table template

Evidence integration summary judgment (for each health effect or outcome grouping)					
Describe judgment regarding whether there is <i>sufficient</i> (or <i>insufficient</i> ) evidence to identify a potential human health hazard, providing the primary interpretations from each evidence stream as well as a summary of the models and range of dose levels in the studies upon which the judgment is primarily reliant.					
Summary of human, animal, and mechanistic evidence					
Studies, outcomes, and confidence	Factors that increase strength or certainty	Factors that decrease strength or certainty	Key findings and interpretation	Evidence stream summary	Inferences across evidence streams
<b>Evidence from studies of exposed humans (may be separated by exposure route or other study design characteristic<sup>a</sup>)</b>					<ul style="list-style-type: none"> <li>• Human relevance of findings in animals</li> <li>• Cross-stream coherence</li> <li>• Summary of potential susceptible populations or lifestages</li> </ul>
<p><u>May be separate rows by outcome</u></p> <ul style="list-style-type: none"> <li>• References</li> <li>• Study confidence</li> <li>• Study design description (if informative)</li> </ul>	<ul style="list-style-type: none"> <li>• Consistency</li> <li>• Dose-response gradient</li> <li>• Coherence of observed effects</li> <li>• Effect size</li> <li>• Mechanistic evidence providing plausibility</li> <li>• <i>Medium or high</i> confidence studies<sup>b</sup></li> </ul>	<ul style="list-style-type: none"> <li>• Unexplained inconsistency</li> <li>• Imprecision</li> <li>• <i>Low</i> confidence studies<sup>b</sup></li> <li>• Evidence demonstrating implausibility</li> </ul>	<ul style="list-style-type: none"> <li>• Description of the primary results and interpretation</li> <li>• If sensitivity issues were identified, describe the impact on reliability of the reported findings</li> </ul>	<p>Summary of the strength of the evidence from human studies based on the factors at left, including the primary evidence basis and considering:</p> <ul style="list-style-type: none"> <li>• Results across human epidemiological and controlled exposure studies</li> <li>• Interpretations regarding any human mechanistic evidence informing biological plausibility (e.g., precursor events linked to adverse outcomes)</li> </ul>	
<b>Evidence from in vivo animal studies (may be separated by exposure route or other study design characteristic<sup>a</sup>)</b>					<ul style="list-style-type: none"> <li>• Other inferences:                             <ul style="list-style-type: none"> <li>○ MOA analysis inferences</li> <li>○ Relevant information from other sources</li> </ul> </li> </ul>
<p><u>May be separate rows by outcome</u></p> <ul style="list-style-type: none"> <li>• References</li> <li>• Study confidence</li> </ul>	<ul style="list-style-type: none"> <li>• Consistency and/or replication</li> <li>• Dose-response gradient</li> <li>• Coherence of observed effects</li> <li>• Effect size</li> </ul>	<ul style="list-style-type: none"> <li>• Unexplained inconsistency</li> <li>• Imprecision</li> <li>• <i>Low</i> confidence studies<sup>b</sup></li> </ul>	<ul style="list-style-type: none"> <li>• Description of the primary results and interpretation</li> <li>• If sensitivity issues were identified, describe the impact on</li> </ul>	<p>Summary of the strength of the evidence for an effect in animals based on the factors at left, including the primary evidence basis and considering:</p> <ul style="list-style-type: none"> <li>• Results across animal toxicological studies</li> </ul>	

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<ul style="list-style-type: none"> <li>• Study design description (if informative)</li> </ul>	<ul style="list-style-type: none"> <li>• Mechanistic evidence providing plausibility</li> <li>• <i>Medium or high</i> confidence studies<sup>b</sup></li> </ul>	<ul style="list-style-type: none"> <li>• Evidence demonstrating implausibility</li> </ul>	reliability of the reported findings	<ul style="list-style-type: none"> <li>• Interpretations regarding any animal mechanistic evidence informing biological plausibility (e.g., precursor events linked to adverse outcomes)</li> </ul>	(e.g., read-across)
<b>Biological events or pathways (or other information)</b>	<b>Species or model systems</b>	<b>Key findings, limitations, and interpretation (for each row below)</b>		<b>Evidence stream summary</b>	
<b>Mechanistic evidence or supplemental information (may be separated by, for example, exposure route<sup>a</sup> or focused topic of analysis)</b>					
<u>Separate rows by biological events or other feature of the approach used for analysis</u> <ul style="list-style-type: none"> <li>• Generally, studies are not listed, but will cite the synthesis section</li> <li>• Does not have to be chemical-specific (e.g., read-across)</li> </ul>	<u>May be multiple rows (e.g., by measurement method)</u> <p>Typically includes:</p> <ul style="list-style-type: none"> <li>• Evidence base(s) (e.g., number of studies; suite of HTS assays; new approach methods [NAMs])</li> <li>• Species (may include lifestage- or sex-specific description, if important to interpretation)</li> <li>• System (in vivo; in vitro; in silico)</li> <li>• Range of exposure levels and durations tested</li> </ul>	<p><i>Interpretation:</i> Summary of expert interpretation for the body of evidence and supporting rationale</p> <p><i>Key findings:</i> Summary of findings across the body of evidence (may focus on or emphasize highly informative designs or findings)</p> <p><i>Limitations:</i> summary of key sources of uncertainty or limitations of the study designs tested (e.g., regarding the biological event or pathway being examined)</p>	Overall summary of expert interpretation across the assessed set of biological events, potential mechanisms of toxicity, or other analysis approach (e.g., AOP).	<ul style="list-style-type: none"> <li>• Includes the primary evidence supporting the interpretation(s)</li> <li>• Describes and substantiates the extent to which the evidence influences inferences across evidence streams</li> <li>• Characterizes the limitations of the evaluation and highlights existing data gaps</li> <li>• May have overlap with factors summarized for other streams</li> </ul>	

<sup>a</sup>In addition to exposure route, the summaries of each evidence stream may include multiple rows (e.g., by study confidence, population, or species, if they informed the analysis of results heterogeneity or other features of the evidence). When data within an evidence stream are lacking or otherwise not informative to the evidence integration decisions, the summary subrows for that evidence stream may be abbreviated to more easily present this information.

<sup>b</sup>Study confidence, based on evaluation of risk of bias and study sensitivity (see Section 6), and information on susceptibility will be considered when evaluating each of the other factors that increase or decrease strength (e.g., consistency). Notably, lack of findings in studies deemed insensitive neither increases nor decreases strength.

## **10.1. EVALUATING THE STRENGTH OF THE HUMAN AND ANIMAL EVIDENCE STREAMS**

1           Before drawing overall evidence integration judgments about whether exposure to one of  
2 these five PFAS has the potential to cause certain health effect(s) in humans given relevant  
3 exposure circumstances, the strength of evidence for the available human and animal evidence will  
4 be evaluated and summarized. For each assessed health effect or health effect grouping (see  
5 Section 5 for examples of the endpoints that will be considered within each health effect category),  
6 the relevant mechanistic evidence in exposed humans and animals (or in their cells, relevant new  
7 approach methods [NAMs] or in silico models), which will be synthesized based on the approaches  
8 and considerations in Section 9.2, will be integrated with the evidence from the available studies of  
9 phenotypic effects in humans and animals. The considerations previously outlined in Table 9-1 (the  
10 different features of the evidence considered and summarized during evidence synthesis) will be  
11 evaluated by the specific PFAS assessment teams within the context of how they affect judgments of  
12 the strength of evidence (see Table 10-2), which will directly inform the overall evidence  
13 integration judgment (see Section 10.2). The evaluation of the strength of the human or animal  
14 health effects evidence will preferably occur at the most specific health outcome level possible  
15 (e.g., an analysis at the level of decreased pulmonary function is generally preferable to an analysis  
16 of respiratory system effects), if there is an adequate set of studies for analyses at this level and  
17 considering the interrelatedness of the available outcomes. If studies on a target system are sparse  
18 or varied, or if the evidence strength relies largely on the interpretation of coherence across related  
19 outcomes, then the analyses may need to be conducted at a broader health effect level. The factors  
20 judged to increase or decrease the strength of the evidence will be summarized in tabular format  
21 using the evidence profile table template in Table 10-1 to transparently convey expert judgments  
22 made throughout the evidence synthesis and integration processes. The evidence profile table  
23 allows for consistent documentation of the supporting rationale for each decision.

**Table 10-2. Considerations that inform evaluations of the strength of the human and animal evidence**

Consideration	Increased evidence strength (of the human or animal evidence)	Decreased evidence strength (of the human or animal evidence)
<p>The structured categories and criteria in Table 10-3 (see Section 10.2) will guide the application of strength-of-evidence judgments for an outcome or health effect. Evidence synthesis scenarios that do not warrant an increase or decrease in evidence strength for a given consideration will be considered “neutral” and are not described in this table (and, in general, will not be captured in the assessment-specific evidence profile tables).</p>		
<p>Risk of bias; sensitivity (across studies)</p>	<ul style="list-style-type: none"> <li>An evidence base of <i>high</i> or <i>medium</i> confidence studies increases strength.</li> </ul>	<ul style="list-style-type: none"> <li>An evidence base of mostly <i>low</i> confidence studies decreases strength. An exception to this is an evidence base of studies in which the primary issues resulting in <i>low</i> confidence are related to insensitivity. This may increase evidence strength in cases where an association is identified because the expected impact of study insensitivity is towards the null.</li> <li>Decisions to increase strength for other considerations in this table should generally not be made if there are serious concerns for risk of bias.</li> </ul>
<p>Consistency</p>	<ul style="list-style-type: none"> <li>Similarity of findings for a given outcome (e.g., of a similar magnitude, direction) across independent studies or experiments increases strength, particularly when consistency is observed across populations (e.g., geographical location) or exposure scenarios in human studies, and across laboratories, populations (e.g., species), or exposure scenarios (e.g., duration; route; timing) in animal studies.</li> </ul>	<ul style="list-style-type: none"> <li>Unexplained inconsistency [i.e., conflicting evidence; see <a href="#">U.S. EPA (2005a)</a>] decreases strength. Generally, strength should not be decreased if discrepant findings can be reasonably explained by study confidence conclusions; variation in population or species, sex, or lifestage; exposure patterns (e.g., intermittent or continuous); exposure levels (low or high); or exposure duration.</li> </ul>

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Consideration	Increased evidence strength (of the human or animal evidence)	Decreased evidence strength (of the human or animal evidence)
Strength (effect magnitude) and precision	<ul style="list-style-type: none"> <li>Evidence of a large magnitude effect (considered either within or across studies) can increase strength. Effects of a concerning rarity or severity can also increase strength, even if they are of a small magnitude.</li> <li>Precise results from individual studies or across the set of studies increases strength, noting that biological significance is prioritized over statistical significance.</li> </ul>	<ul style="list-style-type: none"> <li>Strength may be decreased if effect sizes that are small in magnitude are concluded not to be biologically significant, or if there are only a few studies with imprecise results.</li> </ul>
Biological gradient/dose-response	<ul style="list-style-type: none"> <li>Evidence of dose-response increases strength. Dose-response may be demonstrated across studies or within studies and it can be dose- or duration-dependent. It also may not be a monotonic dose-response (monotonicity should not necessarily be expected, e.g., different outcomes may be expected at low vs. high doses because of activation of different mechanistic pathways or induction of systemic toxicity at very high doses).</li> <li>Decreases in a response after cessation of exposure (e.g., symptoms of current asthma) also may increase strength by increasing certainty in a relationship between exposure and outcome (this is most applicable to epidemiology studies because of their observational nature).</li> </ul>	<ul style="list-style-type: none"> <li>A lack of dose-response when expected based on biological understanding and having a wide range of doses/exposures evaluated in the evidence base can decrease strength.</li> <li>In experimental studies, strength may be decreased when effects resolve under certain experimental conditions (e.g., rapid reversibility after removal of exposure). However, many reversible effects are of high concern. Deciding between these situations is informed by factors such as the toxicokinetics of the chemical and the conditions of exposure [see <a href="#">U.S. EPA (1998)</a>], endpoint severity, judgments regarding the potential for delayed or secondary effects, as well as the exposure context focus of the assessment (e.g., addressing intermittent or short-term exposures).</li> <li>In rare cases, and typically only in toxicological studies, the magnitude of effects at a given exposure level might decrease with longer exposures (e.g., due to tolerance or acclimation). Like the discussion of reversibility above, a decision about whether this decreases evidence strength depends on the exposure context focus of the assessment and other factors.</li> <li>If the data are not adequate to evaluate a dose-response pattern, then strength is neither increased nor decreased.</li> </ul>

*This document is a draft for review purposes only and does not constitute Agency policy.*

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<b>Consideration</b>	<b>Increased evidence strength (of the human or animal evidence)</b>	<b>Decreased evidence strength (of the human or animal evidence)</b>
Coherence	<ul style="list-style-type: none"> <li>Biologically related findings within an organ system, or across populations (e.g., sex) increase strength, particularly when a temporal- or dose-dependent progression of related effects is observed within or across studies, or when related findings of increasing severity are observed with increasing exposure.</li> </ul>	<ul style="list-style-type: none"> <li>An observed lack of expected coherent changes (e.g., well-established biological relationships) will typically decrease evidence strength. However, the biological relationships between the endpoints being compared and the sensitivity and specificity of the measures used need to be carefully examined. The decision to decrease evidence strength depends on the availability of evidence across multiple related endpoints for which changes would be anticipated, and it considers factors (e.g., dose and duration of exposure, strength of expected relationship) across the studies of related changes.</li> </ul>
Mechanistic evidence related to biological plausibility	<ul style="list-style-type: none"> <li>Mechanistic evidence of precursors or health effect biomarkers in well-conducted studies of exposed humans or animals, in appropriately exposed human or animal cells, or other relevant human, animal, or in silico models (including new approach methods [NAMs]) increases strength, particularly when this evidence is observed in the same cohort/population exhibiting the phenotypic health outcome.</li> <li>Evidence of changes in biological pathways or support for a proposed MOA in appropriate models also increases strength, particularly when support is provided for rate-limiting or key events or across multiple components of the pathway or MOA.</li> </ul>	<ul style="list-style-type: none"> <li>Mechanistic understanding is not a prerequisite for drawing a conclusion that a chemical causes a given health effect; thus, an absence of knowledge should not be used as a basis for decreasing strength (<a href="#">NTP, 2015</a>; <a href="#">NRC, 2014</a>).</li> <li>Mechanistic evidence in well-conducted studies (see examples of evidence types at left) that demonstrates that the health effect(s) are unlikely to occur, or only likely to occur under certain scenarios (e.g., above certain exposure levels), can decrease evidence strength. A decision to decrease strength depends on an evaluation of the strength of the mechanistic evidence for and against biological plausibility, as well as the strength of the health effect-specific findings (e.g., stronger health effect data require more certainty in mechanistic evidence opposing plausibility).</li> </ul>

1 For human and animal evidence, the analyses of each consideration in Table 10-2 will be  
2 used to qualitatively summarize the strength-of-evidence for the separate evidence streams in the  
3 evidence integration narrative. Table 10-3 provides the criteria that will guide how to draw the  
4 judgment for each health effect, and the terms that will be used to summarize those evidence  
5 integration judgments.

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## 10.2. OVERALL EVIDENCE INTEGRATION JUDGMENTS

6 Evidence integration combines decisions regarding the strength of the animal and human  
7 evidence with considerations regarding mechanistic information on the human relevance of the  
8 animal evidence, relevance of the mechanistic evidence to humans (especially in cases where  
9 animal evidence is lacking), coherence across bodies of evidence, and information on susceptible  
10 populations and lifestages, based on the considerations and analyses outlined in Section 9.2. This  
11 evidence integration decision process will culminate in an evidence integration narrative that  
12 summarizes the judgments regarding the evidence for each potential health effect (i.e., each  
13 noncancer health effect and specific type of cancer, or broader grouping of related outcomes). For  
14 each health effect, this narrative will include:

- 15 • A descriptive summary of the primary judgments about the evidence informing the  
16 potential for health effects in exposed humans, based on the following analyses:
  - 17 ○ evaluations of the strength of the available human and animal evidence (see  
18 Section 10.1);
  - 19 ○ consideration of the coherence of findings (i.e., the extent to which the evidence for  
20 health effects and relevant mechanistic changes are similar) across human and animal  
21 studies;
  - 22 ○ other information on the human relevance of findings in animals (see Section 9.2); and
  - 23 ○ conclusions drawn based on the predefined mechanistic analyses (see  
24 Sections 9.2.1–9.2.3), as well as those based on analyses identified during stepwise  
25 consideration of the health effect-specific evidence during draft development (see  
26 Section 9.2.4).
- 27 • A summary of key evidence supporting these judgments, highlighting the evidence that was  
28 the primary driver of these judgments and any notable issues (e.g., data quality; coherence  
29 of the results), and a narrative expression of confidence (a summary of strengths and  
30 remaining uncertainties) for these judgments.
- 31 • Information on the general conditions of expression of these health effects (e.g., exposure  
32 routes and levels in the studies that were the primary drivers of these judgments), noting  
33 that these conditions will be clarified during dose-response analysis (see Section 11).

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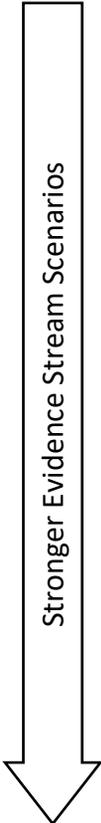
- 1 • Indications of potentially susceptible populations or lifestages (i.e., an integrated summary  
2 of the available evidence on potential susceptible populations and lifestages drawn across  
3 the syntheses of the human, animal, and mechanistic evidence).<sup>21</sup>
- 4 • A summary of key assumptions used in the analysis, which are generally based on EPA  
5 guidelines and which are largely captured in this protocol.
- 6 • Strengths and limitations of the evidence integration judgments, including key uncertainties  
7 and data gaps, as well as the limitations of the systematic review. As noted in Section 4.2.2,  
8 for one or more of these five PFAS assessments, characterization of the uncertainties in the  
9 animal evidence is expected to include a discussion of the reliance on short-term oral  
10 exposure studies in rats. Similarly, the characterization of uncertainty in the human  
11 evidence is expected to include a discussion of potential confounding by PFAS other than  
12 the PFAS of interest.

13 In short, the evidence integration narrative will present a qualitative summary of the  
14 strength of each evidence stream and an overall judgment across all relevant evidence, with  
15 exposure context provided. For each health effect or specific cancer type of potential concern, the  
16 first sentence of the evidence integration narrative will include the summary judgment [see  
17 description below for how these judgments help inform selection of a descriptor for carcinogenicity  
18 ([U.S. EPA, 2005a](#))]. Assessments will also include an evidence profile table (see Table 10-1) to  
19 support the evidence integration narrative by providing the major decisions and supporting  
20 rationale. Table 10-3 describes the categories of evidence integration judgments that will be used  
21 in these PFAS assessments and provides examples of database scenarios that fit each category of  
22 evidence. These summary judgments provide a succinct and clear representation of the decisions  
23 from the more detailed analyses of whether (or not) the evidence strength indicates that PFAS  
24 exposure has the potential to cause the human health effect(s) under the necessary conditions of  
25 exposure. Consistent with EPA noncancer and cancer guidelines, a judgment that the evidence  
26 supports an apparent lack of an effect of PFAS exposure on the health effect(s) will only be used  
27 when the available data are considered robust for deciding that there is no basis for human hazard  
28 concern; lesser levels of evidence suggesting a lack of an effect will be characterized as  
29 “insufficient.”

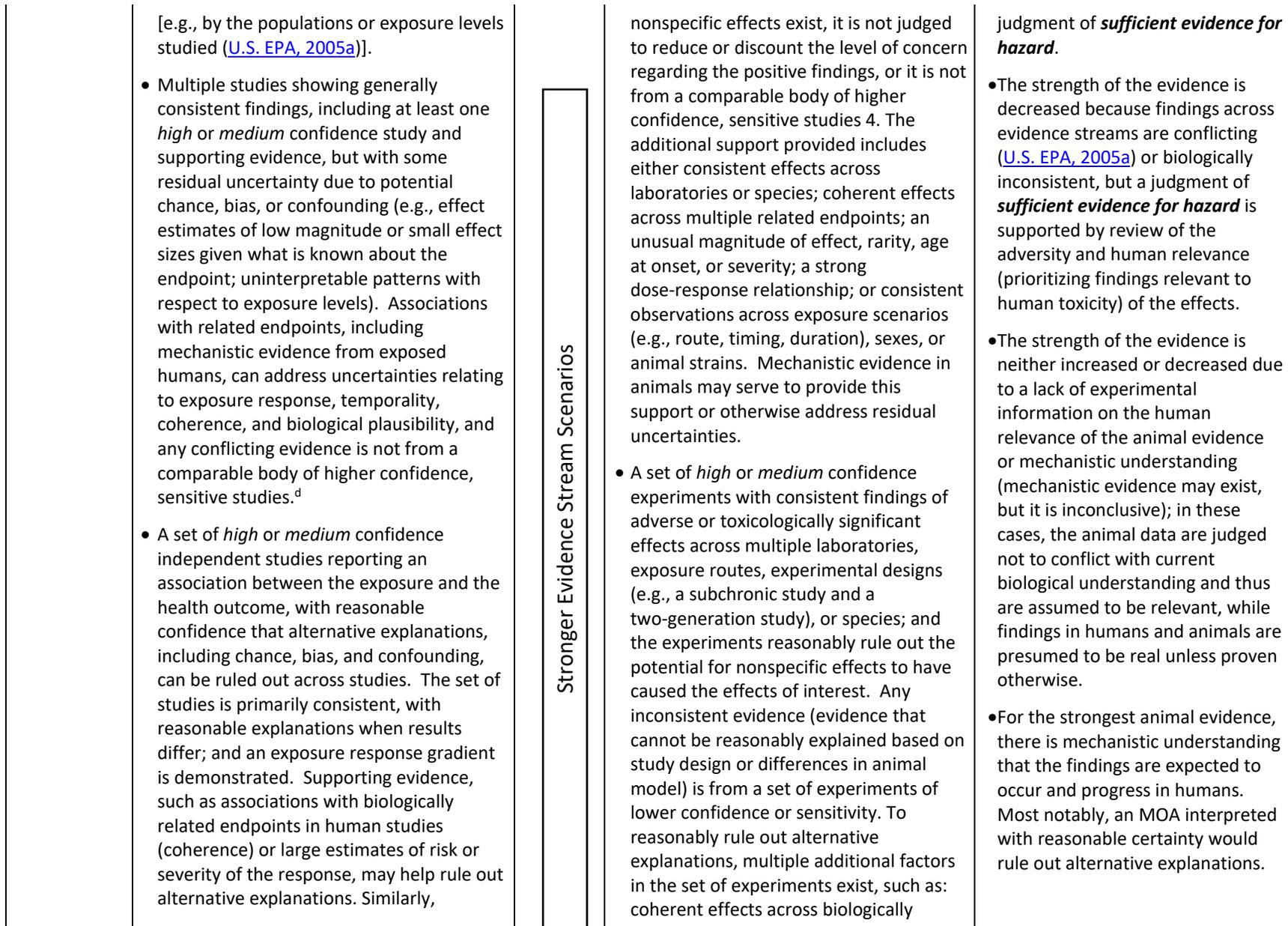
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<sup>21</sup>One or more of these five PFAS assessments may include consideration of information outside of their PFAS-specific database to address this aspect of the evidence integration narrative. These PFAS-specific data gaps and uncertainties appear to extend beyond poorly studied health effects, and the discussion of missing information on potential populations, sexes, or lifestages that are likely to be more susceptible to developing a specific health effect may consider information from reviews of other PFAS.

**Table 10-3. Evidence integration judgments for characterizing potential human health hazards in the evidence integration narrative**

Evidence integration judgment <sup>a</sup>	Evidence in studies of humans	Evidence in animal studies	Inferences across evidence streams
<p><b>Sufficient evidence for hazard</b></p>	<p>A judgment of <b>sufficient evidence for hazard</b> requires that a scenario below is met for <u>either</u> the evidence in studies of humans or evidence in animal studies, incorporating the considerations outlined under inferences across evidence streams. The scenarios justifying this judgment span a broad range of overall evidence strength and examples are provided below, starting with the weakest evidence.<sup>b</sup></p> <ul style="list-style-type: none"> <li>• Strong mechanistic evidence in well-conducted studies of exposed humans (<i>medium</i> or <i>high</i> confidence) or human cells (including NAMs), in the absence of other substantive data, in which an informed evaluation has determined that the data are reliable for assessing toxicity relevant to humans and the mechanistic events have been causally linked to the development of the health effect of interest.<sup>c</sup></li> <li>• A single <i>high</i> or <i>medium</i> confidence study demonstrating an effect with one or more factors that increase evidence strength, such as: a large magnitude or severity of the effect, a dose-response gradient, unique exposure or outcome scenarios (e.g., a natural experiment), or supporting coherent evidence, including mechanistic evidence from exposed humans. There are no comparable studies of similar confidence and sensitivity providing conflicting evidence, or if there are, the differences can be reasonably explained</li> </ul>	<p style="text-align: center;">Stronger Evidence Stream Scenarios</p>  <ul style="list-style-type: none"> <li>• Strong mechanistic evidence in well-conducted studies of animals or animal cells (including NAMs), in the absence of other substantive data, in which an informed evaluation has determined the assays are reliable for assessing toxicity relevant to humans and the mechanistic events have been causally linked to the development of the health effect.<sup>c</sup></li> <li>• A single <i>high</i> or <i>medium</i> confidence experiment in the absence of comparable experiment(s) of similar confidence and sensitivity providing conflicting evidence<sup>d</sup> [evidence that cannot be reasonably explained, e.g., by respective study designs or differences in animal model; (U.S. EPA, 2005a)].</li> <li>• At least one <i>high</i> or <i>medium</i> confidence study with supporting information increasing the strength of the evidence. Although the results are largely consistent, notable uncertainties remain. However, in scenarios when inconsistent evidence or evidence indicating</li> </ul>	<ul style="list-style-type: none"> <li>• Supplemental evidence (e.g., structure-activity data; chemical class information; other NAMs) is judged to increase the strength of limited or near-equivocal, chemical-specific human or animal evidence to <b>sufficient evidence for hazard</b>.</li> <li>• Coherent or biologically consistent findings across evidence streams increases the strength of limited or near-equivocal human or animal evidence (e.g., single or few <i>high</i> or <i>medium</i> confidence studies with some conflicting evidence) to <b>sufficient evidence for hazard</b>.</li> <li>• The strength of the evidence is decreased because mechanistic information (even if it does not provide MOA understanding) raises uncertainties regarding the human and/or animal evidence, but overall the evidence is still considered strong enough to result in a</li> </ul>

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	<p>mechanistic evidence from exposed humans may serve to address uncertainties related to exposure-response, temporality, coherence, and biological plausibility (i.e., providing evidence consistent with an explanation for how exposure could cause the health effect based on current biological knowledge).</p>		<p>related endpoints; an unusual magnitude of effect, rarity, age at onset, or severity; a strong dose-response relationship; or consistent observations across animal lifestages, sexes, or strains. Similarly, mechanistic evidence (e.g., precursor events linked to adverse outcomes) in animal models may exist to address uncertainties in the evidence base.</p>	<ul style="list-style-type: none"> <li>•For the strongest evidence, there is adequate testing of potentially susceptible lifestages and populations, based on the effect(s) of interest and chemical knowledge (e.g., toxicokinetics).</li> </ul>
<p><b>Insufficient evidence</b></p>	<p>A judgment of <b>insufficient evidence</b> requires that a scenario below is met for <u>both</u> the <i>evidence in studies of humans</i> and <i>evidence in animal studies</i>, incorporating the considerations outlined under <i>inferences across evidence streams</i>.</p>			
<ul style="list-style-type: none"> <li>• A body of evidence, including scenarios with one or more <i>high</i> or <i>medium</i> confidence studies reporting an association between exposure and the health outcome, where either (1) conflicting evidence exists in studies of similar confidence and sensitivity<sup>d,e</sup>, (2) a single study without a factor that increases evidence strength as described in <b>sufficient evidence for hazard</b>), or (3) considerable methodological uncertainties remain across the body of evidence (typically related to exposure or outcome ascertainment, including temporality), and there is no supporting coherent evidence that increases the overall evidence strength.</li> <li>• A set of only <i>low</i> confidence studies.</li> <li>• No studies of exposed humans or well-conducted studies of human cells.</li> </ul>	<ul style="list-style-type: none"> <li>• A body of evidence, including scenarios with one or more <i>high</i> or <i>medium</i> confidence experiments reporting effects but without supporting coherent evidence that increases the overall evidence strength, where conflicting evidence exists from a set of sensitive experiments of similar or higher confidence (can include mechanistic evidence).<sup>d,e</sup></li> <li>• A set of only <i>low</i> confidence experiments.</li> <li>• No animal studies or well-conducted studies of animal cells.</li> <li>• The available endpoints are not informative to the hazard question under evaluation.</li> <li>• A set of largely null studies that does not meet the criteria for <b>sufficient evidence to judge that a hazard is unlikely</b>.</li> </ul>	<ul style="list-style-type: none"> <li>•The evidence in animal studies meets a scenario for <b>sufficient evidence for hazard</b>, but strong experimental evidence (e.g., an MOA interpreted with reasonable certainty) indicates the findings in animals are unlikely to be relevant to humans.</li> <li>•The evidence meets a scenario for <b>sufficient evidence to judge that a hazard is unlikely</b>, but there is inadequate testing of susceptible populations and lifestages or of data conflict across evidence streams.</li> <li>•The evidence in animal studies meets a scenario for <b>sufficient evidence to judge that a hazard is unlikely</b>, but the database lacks experimental support that the models are relevant to humans for the effect of interest.</li> </ul>		

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	<ul style="list-style-type: none"> <li>• A set of largely null studies that does not meet a scenario for <b>sufficient evidence to judge that a hazard is unlikely</b>.</li> </ul>		
<p><b>Sufficient evidence to judge that a hazard is unlikely<sup>f</sup></b></p>	<p>A judgment of <b>sufficient evidence to judge that a hazard is unlikely</b> requires that a scenario below is met for <u>either</u> the <i>evidence in studies of humans</i> or <i>evidence in animal studies</i>, incorporating the considerations outlined under <i>inferences across evidence streams</i>.</p> <ul style="list-style-type: none"> <li>• Several <i>high</i> confidence studies showing null results (for example, an odds ratio of 1.0), ruling out alternative explanations including chance, bias, and confounding with reasonable confidence. Each of the studies should have used an optimal outcome and exposure assessment and adequate sample size (specifically for higher exposure groups and for susceptible populations). The overall set should include the full range of levels of exposures that human beings are known to encounter, and an evaluation of an exposure-response gradient.</li> </ul>	<ul style="list-style-type: none"> <li>• A set of <i>high</i> confidence experiments examining a reasonable spectrum of endpoints relevant to a type of toxicity that demonstrate a lack of biologically significant effects across multiple species, both sexes (if applicable), and a broad range of exposure levels. The data are compelling in that the experiments have examined the range of scenarios across which health effects in animals could be observed, and an alternative explanation (e.g., inadequately controlled features of the studies' experimental designs; inadequate sample sizes) for the observed lack of effects is not available. The experiments were designed to specifically test for the effects of interest, including suitable exposure timing and duration, postexposure latency, and endpoint evaluation procedures.</li> </ul>	<ul style="list-style-type: none"> <li>• There is adequate testing of susceptible populations and lifestages.</li> <li>• When the <i>evidence in animal studies</i> meets a scenario for this judgment, there is experimental support that the models are relevant to humans for the effect of interest and no conflicting human evidence exists.</li> <li>• When the <i>evidence in studies of humans</i> meets a scenario for this judgment and conflicting animal data exist, mechanistic information indicates that the animal data are unlikely to be relevant to humans.</li> <li>• When multiple <i>high</i> confidence animal experiments and studies in humans indicate lack of an effect, but the evidence does not meet a scenario for <b>sufficient evidence to judge that a hazard is unlikely</b>, strong mechanistic evidence in models relevant to humans supports lack of an effect such that the totality of evidence supports this judgment.</li> </ul>

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<sup>a</sup>These categories are based on those indicated for use in hazard characterization from the existing EPA guidelines for noncancer health effects [i.e., [U.S. EPA \(1988\)](#); [U.S. EPA \(1991b\)](#); [U.S. EPA \(1996b\)](#)]. As described in those guidance documents, the judgments depend heavily on expert judgment (note: as applied herein, the process of “evidence integration” is synonymous with “weight of evidence”). The evidence integration judgment for each assessed health effect will be included as part of an evidence integration narrative, the specific documentation of the various expert decisions and evidence-based (or default) rationales are summarized in an evidence profile table, and the judgement will be contextualized based on the primary supporting evidence (experimental model or observed population, and exposure levels tested or estimated). Importantly, as discussed in Section 10.1, these judgments may be based on analyses of grouped outcomes at different levels of granularity (e.g., motor activity vs. neurobehavioral effects vs. nervous system effects) depending on the specifics of the health effect evidence base. Health effects characterized as having ***sufficient evidence for hazard*** will be evaluated for use in dose-response assessment.

<sup>b</sup>Qualitative descriptions of differences in the strength of the evidence across different health effects judged as having ***sufficient evidence for hazard*** are useful for other assessment decisions, including prioritizing outcomes in quantitative analyses and characterizing assessment uncertainties. Thus, for all evidence scenarios, but particularly for those in the lower end of this range, it is important to characterize the uncertainties in the evidence base within the evidence integration narrative and convey the evidence strength to subsequent steps, including toxicity values developed based on those effects. Existing guidance defines the minimum evidence necessary to judge that a health hazard could exist as one adverse endpoint from one well-conducted study ([U.S. EPA, 1998](#)); this has been expanded in this table to better incorporate mechanistic evidence, including new approach methods (NAMs) data.

<sup>c</sup>Scientific understanding of toxicity mechanisms and of the human implications of new toxicity testing methods (e.g., from high-throughput screening, from short-term in vivo testing of alternative species, or from new in vitro and in silico testing and other NAMs) will continue to increase. Thus, the sufficiency of mechanistic evidence alone for identifying potential human health hazards is expected to increase as the science evolves. The decision to identify a potential human hazard based on these data is an expert judgment dependent on the state of the science at the time of review.

<sup>d</sup>Scenarios with unexplained heterogeneity across sets of studies with similar confidence and sensitivity can be considered either ***sufficient evidence for hazard*** or ***insufficient evidence***, depending on the expert judgment of the overall weight of evidence. Specifically, this judgment considers the level of support (or lack thereof) provided by evaluations of the magnitude or severity of the effects, coherence of related findings (including mechanistic evidence), dose-response, and biological plausibility, as well as the comparability of the supporting and conflicting evidence (e.g., the specific endpoints tested, or the methods used to test them; the specific sources of bias or insensitivity in the respective sets of studies). The evidence-specific factors supporting either evidence integration judgment will be clearly articulated in the evidence integration narrative.

<sup>e</sup>When the database includes at least one well-conducted study and a hazard characterization judgment of ***insufficient evidence*** is drawn, quantitative analyses may still be useful for some purposes (e.g., providing a sense of the magnitude and uncertainty of estimates for health effects of potential concern, ranking potential hazards, or setting research priorities), but not for others [see related discussions in [U.S. EPA \(2005a\)](#)]. It is critical to transparently convey the extreme uncertainty in any such estimates.

<sup>f</sup>The criteria for this category are intentionally more stringent than those justifying a conclusion of ***sufficient evidence for hazard***, consistent with the “difficulty of proving a negative” [as discussed in [U.S. EPA \(1988\)](#); [U.S. EPA \(1991b\)](#); [U.S. EPA \(1996b\)](#)].

1 Evaluations of carcinogenicity will be consistent with EPA’s Cancer Guidelines ([U.S. EPA,](#)  
2 [2005a](#)). One of EPA’s standardized cancer descriptors will be used as a shorthand characterization  
3 of the evidence integration narrative, describing the overall potential for human carcinogenicity  
4 across all potential cancer types. These are (1) *carcinogenic to humans*, (2) *likely to be carcinogenic*  
5 *to humans*, (3) *suggestive evidence of carcinogenic potential*, (4) *inadequate information to assess*  
6 *carcinogenic potential*, or (5) *not likely to be carcinogenic to humans*. More than one descriptor can  
7 be used when a chemical’s effects differ by exposure level or route ([U.S. EPA, 2005a](#)); if the  
8 database supports such an analysis, these decisions will be clarified based on a more thorough  
9 review of the mechanistic evidence or more detailed dose-response analysis (see Section 11). In  
10 some cases, mutagenicity will also be evaluated (e.g., when there is evidence of carcinogenicity),  
11 because it influences the approach to dose-response assessment and subsequent application of  
12 adjustment factors for exposures early in life ([U.S. EPA, 2005a, b](#)).

13 An appropriate cancer descriptor will be selected as described in EPA Cancer Guidelines  
14 ([U.S. EPA, 2005a](#)). For each cancer subtype, an evidence integration narrative and summary  
15 judgment will be provided, as described above. The cancer descriptor will consider the  
16 interrelatedness of cancer types potentially due to PFAS exposure, consistency across the human  
17 and animal evidence for any cancer type [noting that site concordance is not required ([U.S. EPA,](#)  
18 [2005a](#))], and the uncertainties associated with each assessment-specific conclusion. In general,  
19 however, if a systematic review of more than one cancer type was conducted, then the overall  
20 judgment and discussion of evidence strength in the evidence integration narrative for the cancer  
21 type(s) with the strongest evidence for hazard will be used to inform selection of the cancer  
22 descriptor, with each assessment providing a transparent description of the decision rationale. The  
23 cancer descriptor and evidence integration narrative for potential carcinogenicity, including  
24 application of the MOA framework, will consider the conditions of carcinogenicity, including  
25 exposure (e.g., route; level) and susceptibility (e.g., genetics; lifestage), as the data allow ([Farland,](#)  
26 [2005; U.S. EPA, 2005a, b](#)).

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### **10.3. HAZARD CONSIDERATIONS FOR DOSE-RESPONSE**

27 This section outlines how these assessments will consider and describe the transition from  
28 hazard identification to dose-response analysis, highlighting (1) information that will inform the  
29 selection of outcomes or broader health effect categories for which toxicity values will be derived,  
30 (2) whether toxicity values can be derived to protect specific populations or lifestages, (3) how  
31 dose-response modeling will be informed by toxicokinetic information, and (4) information aiding  
32 the identification of biologically based benchmark response (BMR) levels. The pool of outcomes  
33 and study-specific endpoints will be discussed to identify which categories of effects and study  
34 designs are considered the strongest and most appropriate for quantitative assessment of a given  
35 health effect. Health effects that were analyzed in human studies in relation to exposure levels  
36 within or closer to the range of exposures encountered in the environment will be considered

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1 particularly informative, as are animal studies testing a broad range of exposure levels and  
2 including levels in the lower dose region. When there are multiple endpoints for an organ/system,  
3 considerations for characterizing the overall impact on this organ/system will be discussed,  
4 including the severity and longevity of the effects. For example, if there are multiple  
5 histopathological alterations relevant to liver function changes, liver necrosis may be selected as  
6 the most representative endpoint to consider for dose-response analysis. This section may review  
7 or clarify which endpoints or combination of endpoints in each organ/system characterize the  
8 overall effect for dose-response analysis. For cancer types, consideration will be given to the  
9 overall risk of multiple types of tumors. Multiple tumor types (if applicable) will be discussed and a  
10 rationale given for any grouping.

11 Biological considerations that are important for dose-response analysis (e.g., that could help  
12 with selection of a BMR) will be discussed. The impact of route of exposure on toxicity to different  
13 organs/systems will be examined, if appropriate. The existence and validity of PBPK models or  
14 toxicokinetic information that may allow the estimation of internal dose for route-to-route  
15 extrapolation will be presented (see additional discussion and decision points in Section 11.2). In  
16 addition, mechanistic evidence analyses that will influence the dose-response analyses will be  
17 highlighted (see Section 9.2 for specific considerations), for example, evidence related to  
18 susceptibility or potential shape of the dose-response curve.

19 This section will also describe the evidence regarding populations and lifestages that  
20 appear to be susceptible to the health hazards identified and factors that are likely to increase the  
21 risk of developing (or exacerbating) these health effects, depending on the available evidence. This  
22 section will include this discussion even if there are no specific data on the effects of exposure to  
23 the PFAS of interest in the potentially susceptible population. Table 9-2 in Section 9 outlines some  
24 of the specific factors that will be considered for discussion and summaries of the evidence with  
25 respect to patterns across studies pertinent to consistency, coherence, and the magnitude and  
26 direction of effect measures. At a minimum, consideration will be given to discussion of  
27 information relevant to infants and children, pregnant women, and women of childbearing age.

28 The section will consider options for using susceptible population data in the dose-response  
29 analysis. In particular, an attempt will be made to highlight where it might be possible to develop  
30 separate risk estimates for a specific population or lifestage or to determine whether evidence is  
31 available to select a data-derived uncertainty factor.

## 11. DOSE-RESPONSE ASSESSMENT: SELECTING STUDIES AND QUANTITATIVE ANALYSIS

1           The previous sections of this protocol describe how systematic review principles will be  
2 applied to evaluate studies (for potential bias and sensitivity) and reach evidence integration  
3 conclusions on potential human health effects associated with exposure to the PFAS of interest.  
4 Selection of specific data sets for dose-response assessment and performance of the dose-response  
5 assessment will be conducted after hazard identification is complete and involves database- and  
6 chemical-specific biological judgments that build from decisions made at earlier stages of  
7 assessment development. Several Environmental Protection Agency (EPA) guidance and support  
8 documents describe data requirements and other considerations for dose-response modeling,  
9 especially EPA's *Benchmark Dose Technical Guidance* ([U.S. EPA, 2012](#)), EPA's *Review of the Reference*  
10 *Dose and Reference Concentration Processes* ([U.S. EPA, 2002b](#)), *Guidelines for Carcinogen Risk*  
11 *Assessment* ([U.S. EPA, 2005a](#)), and *Supplemental Guidance for Assessing Susceptibility from Early-Life*  
12 *Exposure to Carcinogens* ([U.S. EPA, 2005b](#)). This section of the protocol provides an overview of  
13 considerations for conducting the dose-response assessment, particularly statistical considerations  
14 specific to dose-response analysis that support quantitative risk assessment. Importantly, these  
15 considerations do not supersede existing EPA guidance.

16           Dose-response assessments will be performed for both noncancer and cancer health  
17 hazards, and for both oral and inhalation routes of exposure following exposure<sup>22</sup> to the chemical of  
18 interest, if supported by existing data. For noncancer hazards, an oral reference dose (RfD) and/or  
19 an inhalation reference concentration (RfC) will be derived when possible. An RfD or an RfC is an  
20 estimate, with uncertainty spanning perhaps an order of magnitude, of an exposure to the human  
21 population (including susceptible subgroups) that is likely to be without an appreciable risk of  
22 deleterious health effects over a lifetime ([U.S. EPA, 2002b](#)). In addition to an RfD and/or RfC, when  
23 feasible and if the available data are appropriate for doing so, the assessments will derive a less-  
24 than-lifetime toxicity value (a "subchronic" reference value) for noncancer hazards. Likewise, part  
25 of the process for deriving an oral or inhalation reference value will include developing separate  
26 values specific to each hazard ("organ- or system-specific" reference values). Both less-than-  
27 lifetime and hazard-specific values may be useful to EPA risk assessors within specific decision

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<sup>22</sup>For most health outcomes (e.g., this would not apply to outcomes related to developmental toxicity), dose-response assessments will be preferably based on studies of chronic exposure. However, analyses will also be conducted for shorter durations, particularly when the evidence base for a PFAS indicates potential risks associated with shorter exposures to the chemical ([U.S. EPA, 2002b](#)).

1 contexts. Reference values are not predictive risk values; that is, they provide no information about  
2 risks at higher or lower exposure levels.

3 Reference values may also be derived for cancer effects [e.g., in a case where a nonlinear  
4 MOA is concluded that indicates a key precursor event necessary for carcinogenicity does not occur  
5 below a specific exposure level ([U.S. EPA, 2005a](#)); see Section 11.2.3]. When low-dose linear  
6 extrapolation for cancer effects is supported, particularly for chemicals with direct mutagenic  
7 activity or those for which the data indicate a linear component below the point of departure (POD),  
8 an oral slope factor (OSF) and/or an inhalation unit risk (IUR) will be used to estimate human  
9 cancer risks. In general, this will also be the case when no data are available to inform the  
10 evaluation of linearity. An OSF is a plausible upper bound lifetime cancer risk from chronic  
11 ingestion of a chemical per unit of mass consumed per unit body weight per day (mg/kg-day). An  
12 IUR is a plausible upper bound lifetime cancer risk from chronic inhalation of a chemical per unit of  
13 air concentration (expressed as ppm or  $\mu\text{g}/\text{m}^3$ ). In contrast with reference values (RfVs), an OSF or  
14 IUR can be used in conjunction with exposure information to predict cancer risk at a given dose.

15 As discussed in Section 2 “Scoping and Problem Formulation Summary” for these PFAS  
16 assessments, the Integrated Risk Information System (IRIS) Program will conduct the assessments  
17 with a goal of developing any toxicity values that are reasonably supported by the available data,  
18 based on judgments of the evidence drawn during hazard identification and the suitability of  
19 studies for dose-response analysis.

20 The derivation of reference values and cancer risk estimates will depend on the nature of  
21 the health hazard conclusions drawn during evidence integration (see Section 10.2). Specifically,  
22 EPA generally conducts dose-response assessments and derives cancer values for chemicals that  
23 are classified as *carcinogenic* or *likely to be carcinogenic* to humans. When there is *suggestive*  
24 *evidence* of carcinogenicity to humans, EPA generally would not conduct a dose-response  
25 assessment or derive a cancer value except when the evidence includes a well-conducted study and  
26 quantitative analyses may be useful for some purposes, for example, providing a sense of the  
27 magnitude and uncertainty of potential risks, ranking potential hazards, or setting research  
28 priorities ([U.S. EPA, 2005a](#)). A parallel approach will be taken for potential noncancer health effects  
29 in these assessments. Specifically, for noncancer outcomes these assessments will generally  
30 include dose-response assessments when the evidence integration judgments indicate there is  
31 ***sufficient evidence for hazard***, with preference given to health effects with stronger evidence  
32 scenarios within that category (see Section 10.2), and quantitative analyses generally will not be  
33 attempted for ***insufficient evidence***.

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## 11.1. SELECTING STUDIES FOR DOSE-RESPONSE ASSESSMENT

34 The dose-response assessment will begin with a review of the important health effects  
35 highlighted during hazard identification, particularly among the studies of highest quality and that  
36 exemplify the study attributes summarized in Table 11-1. This review will also consider whether

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1 there are opportunities for quantitative evidence integration, although it is considered unlikely that  
2 the data available to do so will be available for these assessments based on the preliminary  
3 literature inventory. Examples of quantitative integration, from simplest to more complex, include  
4 (1) the combination of results for an outcome across sex (within a study); (2) characterizing overall  
5 toxicity, as in combining effects that constitute a syndrome, or occur on a continuum  
6 (e.g., precursors and overt toxicity, benign tumors that progress to malignant tumors); and  
7 (3) meta-analysis or metaregression of all studies addressing a category of important health effects.

8         Some studies that were used qualitatively for hazard identification may or may not be  
9 considered useful quantitatively for dose-response analysis in these five assessments because of  
10 factors like the lack of quantitative measures of exposure or of variability measures for response  
11 data. If the needed information cannot be located (e.g., by contacting study authors and making any  
12 information publicly available), a semiquantitative analysis (e.g., via no-observed-adverse-effect  
13 level [NOAEL]/lowest-observed-adverse-effect level [LOAEL]) will be considered. Studies of low  
14 sensitivity may be considered less useful if they failed to detect an effect or reported points of  
15 departure with wide confidence limits, but such studies will still be considered for inclusion in a  
16 meta-analysis.

17         Among the studies that support the evidence integration conclusions, those that are most  
18 useful for dose-response analysis will generally have at least one exposure level in the region of the  
19 dose-response curve near the benchmark response (the response level to be used for deriving  
20 toxicity values) to minimize low-dose extrapolation. Such studies will also have more exposure  
21 levels and larger sample sizes overall ([U.S. EPA, 2012](#)). These attributes support a more complete  
22 characterization of the shape of the exposure-response curve and decrease the uncertainty in the  
23 associated exposure-response metric (e.g., IUR or RfC) by reducing statistical uncertainty in the  
24 POD and minimizing the need for low-dose extrapolation. In addition to these more general  
25 considerations, specific issues that may be considered for their potential to affect the feasibility of  
26 dose-response modeling for individual data sets are described in more detail in the *Benchmark Dose*  
27 *Technical Guidance* ([U.S. EPA, 2012](#)).

**Table 11-1. Attributes used to evaluate studies for deriving toxicity values**

Study attributes		Considerations	
		Human studies	Animal studies
Rationale for choice of species		Human data are preferred over animal data to eliminate interspecies extrapolation uncertainties (e.g., in toxicodynamics, relevance of specific health outcomes to humans, and in toxicokinetics, especially given minimal human TK data).	Animal studies provide supporting evidence when adequate human studies are available and are considered principal studies when adequate human studies are not available. For some hazards, studies of animal species known to respond similarly to humans would be preferred over studies of other species.
Relevance of exposure paradigm	Exposure route	Studies involving human environmental exposures (oral, inhalation).	Studies by a route of administration relevant to human environmental exposure are preferred. A validated toxicokinetic model can also be used to extrapolate across exposure routes.
	Exposure durations	When developing a chronic toxicity value, chronic- or subchronic-duration studies are preferred over studies of acute exposure. Exceptions exist, such as when a susceptible population or lifestage is more sensitive in a certain time window (e.g., developmental exposure).	
	Exposure levels	Exposures near the range of typical environmental human exposures are preferred. Studies with a broad exposure range and multiple exposure levels are preferred to the extent that they can provide information about the shape of the exposure-response relationship [see the EPA <i>Benchmark Dose Technical Guidance</i> ( <a href="#">U.S. EPA, 2012</a> )] and facilitate extrapolation to more relevant (generally lower) exposures.	
Subject selection		Studies that provide risk estimates in the most susceptible groups are preferred.	
Controls for possible confounding <sup>a</sup>		Studies with a design (e.g., matching procedures, blocking) or analysis (e.g., covariates or other procedures for statistical adjustment) that adequately address the relevant sources of potential critical confounding for a given outcome are preferred.	
Measurement of exposure		Studies that can reliably distinguish between levels of exposure in a time window considered most relevant for a causal effect with respect to the development of the outcome are preferred. Exposure assessment methods that reduce measurement error and methods that provide measurement of exposure at the level of the individual are preferred. Measurements of exposure should not be influenced by knowledge of health outcome status.	Studies providing actual measurements of exposure (e.g., analytical inhalation concentrations vs. target concentrations) are preferred. Relevant internal dose measures may facilitate extrapolation to humans, as would availability of a suitable animal PBPK model in conjunction with an animal study reported in terms of administered exposure.

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Study attributes	Considerations	
	Human studies	Animal studies
Measurement of health outcome(s)	Studies that can reliably distinguish the presence or absence (or degree of severity) of the outcome are preferred. Outcome ascertainment methods using generally accepted, standardized approaches) are preferred.	
	Studies with individual data are preferred in general. Examples include characterizing experimental variability more realistically and characterizing overall incidence of individuals affected by related outcomes (e.g., phthalate syndrome).	
Study size and design	Preference is given to studies using designs reasonably expected to have power to detect responses of suitable magnitude. <sup>b</sup> This does not mean that studies with substantial responses but low power would be ignored, but that they should be interpreted in the context of a confidence interval or variance for the response. Studies that address changes in the number at risk (through decreased survival, loss to follow-up) are preferred.	

<sup>a</sup>An exposure or other variable that is associated with both exposure and outcome but is not an intermediary between the two.

<sup>b</sup>Power is an attribute of the design and population parameters, based on a concept of repeatedly sampling a population; it cannot be inferred post hoc using data from one experiment ([Hoening and Heisey, 2001](#)).

## **11.2. CONDUCTING DOSE-RESPONSE ASSESSMENTS**

1 Consistent with EPA practice, these PFAS assessments will apply a two-step approach for  
2 dose-response assessment that distinguishes analysis of the dose-response data in the range of  
3 observation from any inferences about responses at lower environmentally relevant exposure  
4 levels ([U.S. EPA, 2012, 2005a](#)):

- 5 1) Within the observed dose range, the preferred approach will be to use dose-response  
6 modeling to incorporate as much of the data set as possible into the analysis. This modeling  
7 to derive a POD should include an exposure level ideally near the lower end of the range of  
8 observation, without significant extrapolation to lower exposure levels (see Section 11.2.1  
9 for more details).
- 10 2) As derivation of cancer risk estimates and reference values nearly always involves  
11 extrapolation to exposures lower than the POD; the approaches to be applied in these  
12 assessments are described in more detail in Section 11.2.2 and Section 11.2.3, respectively.

13 When sufficient and appropriate human and laboratory animal data are available for the  
14 same outcome, human data will be generally preferred for the dose-response assessment because  
15 its use eliminates the need to perform interspecies extrapolations.

16 For reference values, these assessments will typically derive a candidate value from each  
17 suitable data set, whether in humans or animals (see Section 11.1). Evaluation of these candidate  
18 values grouped within a given organ/system will yield a single organ/system-specific value for each  
19 organ/system under consideration. Next, evaluation of these organ/system-specific values will  
20 result in the selection of a single overall reference value to cover all health outcomes across all  
21 organs/systems. While this overall reference value represents the focus of these dose-response  
22 assessments, the organ/system-specific values can be useful for subsequent cumulative risk  
23 assessments that consider the combined effect of multiple PFAS (or other agents) acting at a  
24 common organ/system.

25 For cancer, if there are multiple tumor sites that can be quantified individually, the final  
26 cancer risk estimate(s) will typically address overall cancer risk, to the extent the data allow.

27 For both cancer and noncancer toxicity values, uncertainties in these estimates will be  
28 transparently characterized and discussed.

### **11.2.1. Dose-Response Analysis in the Range of Observation**

29 Toxicodynamic (“biologically based”) modeling is generally preferred when there are  
30 sufficient, reliable data to ascertain the MOA and quantitatively support model parameters that  
31 represent rates and other quantities associated with the key precursor events of the MOA. Such  
32 data, however, do not appear to be available for these five PFAS.

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1           Because a toxicodynamic model will not be available for dose-response assessment,  
2 empirical modeling will be used to fit the data (on the apical outcome or a key precursor event) in  
3 the range of observation. For this purpose, EPA has developed a standard set of models  
4 (<http://www.epa.gov/ncea/bmds>) that can be applied to typical data sets, including those that are  
5 nonlinear. In situations where there are alternative models with significant biological support  
6 (e.g., when the available evidence provides strong support for a threshold MOA), the decision  
7 maker will be informed by the presentation of these alternatives in the assessment(s) along with  
8 the models' strengths and uncertainties. EPA has developed guidance on modeling dose-response  
9 data, assessing model fit, selecting suitable models, and reporting modeling results [see the EPA  
10 *Benchmark Dose Technical Guidance* ([U.S. EPA, 2012](#))]. Additional judgment or alternative analyses  
11 will be used if the procedure fails to yield reliable results; for example, if the fit is poor, modeling  
12 may be restricted to the lower doses, especially if there is competing toxicity at higher doses.

13           For each modeled response, a POD from the observed data will be estimated to mark the  
14 beginning of extrapolation to lower doses. The POD is an estimated dose (expressed in  
15 human-equivalent terms) near the lower end of the observed range without significant  
16 extrapolation to lower doses. The POD will be used as the starting point for subsequent  
17 extrapolations and analyses. For linear extrapolation of cancer risk, the POD will be used to  
18 calculate an OSF or IUR, and for nonlinear extrapolation, the POD will be used in the calculation of  
19 an RfD or RfC.

20           The response level at which the POD is calculated will be guided by the severity of the  
21 endpoint. If linear extrapolation is used, standard values near the low end of the observable range  
22 will generally be used (for example, 10% extra risk for cancer bioassay data, 1% for epidemiologic  
23 data, lower for rare cancers). For nonlinear approaches, both statistical and biological significance  
24 will be considered. For dichotomous data, a response level of 10% extra risk will generally be used  
25 for minimally adverse effects, 5% or lower for more severe effects. For continuous data, a response  
26 level ideally will be based on an established definition of biologic significance. In the absence of  
27 such definition, one control standard deviation from the control mean will generally be used for  
28 minimally adverse effects, and one-half standard deviation for more severe effects. The point of  
29 departure will be the 95% lower bound on the dose associated with the selected response level.

30           EPA has developed standard approaches to determine the relevant dose for use in  
31 dose-response modeling in the absence of appropriate toxicokinetic modeling. These standard  
32 approaches can also aide comparison across exposure patterns and species in the absence of a  
33 validated pharmacokinetic (PK) model (see below). The general approaches and considerations to  
34 be used to extrapolate PFAS dosimetry from (1) shorter to longer durations within studies, (2) from  
35 animals to humans, and (3) across routes of exposure are outlined below:

- 36           • Intermittent study exposures will be standardized to a daily average over the duration of  
37 exposure. For chronic effects, daily exposures will be averaged over the life span.

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- 1 Exposures during a critical period, however, will not be averaged over a longer duration  
2 ([U.S. EPA, 2005a, 1991a](#)). Note that this will typically be done after modeling because the  
3 conversion is linear.
- 4 • The preferred approach for dosimetry extrapolation from animals to humans will be  
5 through PBPK or PK modeling. This approach will be considered first for PFAS and  
6 lifestages with existing PBPK models or where an existing model structure can be readily  
7 adapted (see Section 6.4 on PBPK modeling).
  - 8 • Because there are PK data for the PFAS being evaluated in at least one relevant animal  
9 species (rats or monkeys) and in humans (see Section 2.4.1), a data-informed extrapolation  
10 approach will also be considered for any PFAS that either lacks a PK model or has a model  
11 determined to be of inadequate quality. Briefly, the ratio of the elimination half-life in  
12 animal to that in the human,  $t_{1/2A}: t_{1/2H}$ , or the ratio of the clearance in the human to the  
13 animal,  $CL_H:CL_A$ , will be considered for use in converting an oral dose-rate in animals  
14 (mg/kg-day) to a human equivalent dose rate (i.e., the human exposure that should yield  
15 the same blood concentration as the animal exposure from which it is being extrapolated).  
16 Note that clearance and half-life are inversely related. The assessments will consider these  
17 metrics as follows:
    - 18 ◦ Of these two metrics,  $t_{1/2}$  and CL, the half-life is a less complete measure of elimination  
19 but one that can be evaluated from more minimal PK data. A half-life can be estimated  
20 by observing the decline in an individual's blood concentration of a compound after an  
21 exposure has ended. In this way, the total exposure or body burden of the chemical  
22 does not have to be known. However, PFAS elimination may go through several phases  
23 during which distinct half-lives apply, and the blood concentration that occurs during  
24 ongoing exposure may effectively reflect an average among these. The specific  
25 approaches and considerations for estimating PFAS half-life are outlined in  
26 Section 9.2.1.
    - 27 ◦ The clearance, on the other hand, is a measure of average elimination but requires more  
28 data to estimate. One must also quantify a companion variable, the volume of  
29 distribution ( $V_d$ ), which in turn requires a measure of total exposure or dose in  
30 well-conducted studies. Although more rigorous assessment-specific evaluations will  
31 be performed, based on a preliminary review of studies in the literature inventory, the  
32 data necessary for the reliable quantification of  $V_d$  in humans are expected to be lacking.  
33 Specifically, accurate estimates of dose do not seem to be available in human exposure  
34 studies, and the identified animal studies demonstrate considerable interstudy  
35 variability in  $V_d$  estimates.
    - 36 ◦ Using an estimate of human CL based on  $V_d$  measured in rats, for example, yields the  
37 same rat-human conversion factor as using the ratio of half-lives, but in a less  
38 transparent way: that is, because the underlying assumption that human  $V_d$  equals rat  
39  $V_d$  is not clear, a reviewer might assume that use of CL indicates a more complete  
40 evaluation of human PK. However, it is expected that  $V_d$  in humans will be similar to  
41 that in non-human primates based on the more similar physiology and biochemical  
42 parameters, so it is reasonable to use a primate  $V_d$  to estimate human CL. Hence human  
43 CL values will only be used if they are based on an independent direct measurement of  
44 CL in humans (e.g., using subject-paired measurements of a PFAS in human serum and

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1 urine), direct estimation of Vd in humans using controlled exposures to PFAS, or using  
2 T<sub>0.5</sub> determined from human data together with a Vd from non-human primates.

3 ° As indicated in Section 9.2.1, if the PK data clearly indicate a dose-dependent half-life,  
4 the t<sub>1/2</sub> at lower doses, most relevant to human health extrapolation, will be used.

5 • Based on the selection of half-life as the preferred metric and a POD identified from a  
6 health-effects study in animals, the human equivalent dose (HED) will be calculated as:

$$\begin{aligned} 7 \quad \text{HED} &= (\text{CL}_{\text{H}[\text{s}]} / \text{CL}_{\text{A}[\text{s}]}) \times \text{POD} \text{ or} \\ 8 \quad \text{HED} &= (t_{1/2\text{A}[\text{s}]} / t_{1/2\text{H}[\text{s}]}) \times \text{POD} \end{aligned} \quad (11-1)$$

9 ° Here, the [s] in the subscript indicates that the value may be sex specific. When there  
10 are sex-specific values (significant differences between males and females) in both  
11 animals and humans, the CL or t<sub>1/2</sub> values for females would be used to extrapolate  
12 health effects in female animals to women, the CL or t<sub>1/2</sub> values for males used to  
13 extrapolate male animal health effects to men. If human data are available to estimate  
14 separate half-lives for women and men, the CL or t<sub>1/2</sub> for women will likewise be used to  
15 estimate HED values in women and the CL or t<sub>1/2</sub> in men used to estimate HEDs in men.  
16 If human data are not sufficient to provide distinct values for men and women, a  
17 common t<sub>1/2</sub> for humans will be used.

18 • In the absence of PK data/half-lives, oral doses will be scaled allometrically using BW<sup>3/4</sup> as  
19 the equivalent dose metric across species. Allometric scaling pertains to equivalence across  
20 species, not across lifestages, and will not be used to scale doses from adult humans or  
21 mature animals to infants or children ([U.S. EPA, 2011a](#), [2005a](#), [1994](#)). Using this approach,  
22 the HED will be calculated as:

$$23 \quad \text{HED} = (\text{BW}_{\text{H}} / \text{BW}_{\text{A}})^{0.25} \times \text{POD} \text{ (mg/kg-day)} \quad (11-2)$$

24 ° If half-life data are available in humans and rats but not mice, for example, then  
25 allometric scaling may be used to estimate the mouse half-life from the rat value  
26 (i.e., using two species closer in BW). This extrapolated mouse half-life can then be used  
27 with the measured human half-life to estimate an HED as described above, making the  
28 greatest possible use of available TK data.

29 • Inhalation exposures will be scaled using dosimetry models that apply species-specific  
30 physiologic and anatomic factors and consider whether the effect occurs at the site of first  
31 contact or after systemic circulation ([U.S. EPA, 2012](#), [1994](#)).

32 • It can be informative to convert doses across exposure routes. If this is done, the  
33 assessment will describe the underlying data, algorithms, and assumptions ([U.S. EPA](#),

1 [2005a](#)). Depending on the availability of sufficient data (see Section 9.2) and/or suitable  
2 models (see Section 6.4), route-to-route extrapolations in these assessments will be  
3 accomplished by using the inhalation exposure rates for PFAS-containing particles  
4 predicted using the MPPD model (see Section 9.2) as an ingestion rate in the PK analysis  
5 (PBPK/PK model or ADME adjustment), under the assumption that once absorbed into  
6 general circulation, the toxic effect is only a function of the body burden or blood  
7 concentration.

- 8 • In the absence of study-specific data on, for example, intake rates or body weight, the EPA  
9 has developed recommended values for use in dose-response analysis ([U.S. EPA, 1988](#)).

### **11.2.2. Extrapolation: Slope Factors and Unit Risk**

10 An OSF or IUR will be used to estimate human cancer risks when low-dose linear  
11 extrapolation for cancer effects is supported by the PFAS-specific evidence, particularly for PFAS  
12 with direct mutagenic activity or those for which the data indicate a linear component below the  
13 POD. Low-dose linear extrapolation will also be used as a default when the data are insufficient to  
14 establish the MOA ([U.S. EPA, 2005a](#)). If the PFAS-specific data are sufficient to ascertain that one or  
15 more modes of action are consistent with low-dose nonlinearity, or to support their biological  
16 plausibility, low-dose extrapolation will use the reference-value approach when suitable data are  
17 available ([U.S. EPA, 2005a](#)); see Section 11.2.3 below.

18 Differences in susceptibility will be considered for use in deriving multiple slope factors or  
19 unit risks, with separate estimates for susceptible populations and lifestages ([U.S. EPA, 2005a](#)). If  
20 appropriate chemical-specific data on susceptibility from early life exposures are available, then  
21 these data will be used to develop cancer slope factors or unit risks that specifically address any  
22 potential for differential potency in early lifestages ([Farland, 2005](#); [U.S. EPA, 2005a](#)). If such data  
23 are not available, the evidence integration analyses supports a mutagenic MOA for carcinogenicity,  
24 and the extrapolation approach is linear, the dose-response assessment will indicate to decision  
25 makers that in the development of risk estimates, the default *age-dependent adjustment factors*  
26 should be used with the cancer slope factor or unit risk and age-specific estimates of exposure ([U.S.](#)  
27 [EPA, 2005a, b](#)). In this scenario, the final cancer risk value presented in the assessment(s) will  
28 reflect this adjustment, with the requisite calculations provided.

29 The derivation of an OSF and IUR for any of these five PFAS conducted as part of the current  
30 assessments will be performed in a manner consistent with EPA guidance.

### **11.2.3. Extrapolation: Reference Values**

31 Reference value derivation is EPA's most frequently used type of nonlinear extrapolation  
32 method, and it will be used in these PFAS assessments for noncancer effects. This approach will  
33 also be used for cancer effects if the available data are sufficient to ascertain the MOA and conclude  
34 that it is not linear at low doses (see Section 11.2.2). In this case, reference values for each relevant

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1 route of exposure will be developed following EPA’s established practices ([U.S. EPA, 2005a](#)); in  
2 general, the reference value will be based not on tumor incidence, but on a key precursor event in  
3 the MOA that is necessary for tumor formation. The derivation of an RfD or RfC (if feasible)  
4 conducted as part of the assessments for perfluorobutanoic acid (PFBA), PFHxA,  
5 perfluorohexanesulfonate (PFHxS), perfluorononanoic acid (PFNA), and perfluorodecanoic acid  
6 (PFDA) will be performed in a manner consistent with EPA guidance.

7 For each data set selected, reference values will be estimated by applying relevant  
8 adjustments (i.e., uncertainty factors [UFs]) to the PODs to account for the conditions of the  
9 reference value definition. These factors account for human variation, extrapolation from animals  
10 to humans, extrapolation to chronic exposure duration, extrapolation to a minimal level of risk (if  
11 not observed in the data set), and database deficiencies, as outlined below. Increasingly, data-based  
12 adjustments ([U.S. EPA, 2014c](#)), probabilistic approaches ([Chiu et al., 2018](#); [Chiu and Slob, 2015](#)),  
13 and Bayesian methods for characterizing population variability ([NAS, 2014](#)) are becoming feasible  
14 and may be distinguished from the UF considerations outlined below, if such data exist for these  
15 five PFAS. These assessments will discuss the scientific bases (or lack thereof) for each selected UF,  
16 including any data-based adjustments based on the following considerations:

- 17 • *Animal-to-human extrapolation*: If animal results are used to make inferences about  
18 humans, the reference value derivation will incorporate the potential for cross-species  
19 differences, which may arise from differences in toxicokinetics or toxicodynamics. The POD  
20 will be standardized to equivalent human terms or be based on toxicokinetic or dosimetry  
21 modeling that may range from detailed chemical-specific to default approaches ([U.S. EPA,](#)  
22 [2014c, 2011a](#)), and a factor of 10<sup>0.5</sup> (rounded to 3) will be applied to account for the  
23 remaining uncertainty involving toxicokinetic and toxicodynamic differences. Data-derived  
24 adjustments for toxicodynamic differences across species may include qualitative decisions  
25 regarding key science issues (e.g., if, during evaluation of PPAR $\alpha$ -dependency, it is  
26 concluded that humans are not more sensitive than rodents).
- 27 • *Human variation*: The assessments will account for variation in susceptibility across the  
28 human population and the possibility that the available data may not represent individuals  
29 who are most susceptible to the effect. If appropriate data or models for the effect or for  
30 characterizing the internal dose are available, the potential for data-based adjustments for  
31 toxicodynamics or toxicokinetics will also be considered ([U.S. EPA, 2014c, 2002b](#)).<sup>23, 24</sup>  
32 When sufficient data are available, an intraspecies UF either less than or greater than  
33 10-fold may be justified ([U.S. EPA, 2002b](#)). A reduction in this UF will be considered if the

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<sup>23</sup>Examples of adjusting the toxicokinetic portion of interhuman variability include the Integrated Risk Information System (IRIS) boron assessment’s use of non-chemical-specific kinetic data [e.g., glomerular filtration rate in pregnant humans as a surrogate for boron clearance ([U.S. EPA, 2004](#))] and the IRIS trichloroethylene assessment’s use of population variability in trichloroethylene metabolism, via a PBPK model, to estimate the lower 1st percentile of the dose metric distribution for each POD ([U.S. EPA, 2011b](#)).

<sup>24</sup>Note that when a PBPK model is available for relating human internal dose to environmental exposure, relevant portions of this UF may be more usefully applied prior to animal-to-human extrapolation, depending on the correspondence of any nonlinearities (e.g., saturation levels) between species.

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1           POD is derived from or adjusted specifically for susceptible individuals, but not for a general  
2           population that includes both susceptible and nonsusceptible individuals ([U.S. EPA, 2002b](#),  
3           [1998](#), [1996b](#), [1994](#), [1991a](#)). In general, when the use of such data or modeling is not  
4           supported, a UF with a default value of 10 will be used.

- 5           • *LOAEL to NOAEL*: When a POD is based on a LOAEL, the assessment will include an  
6           adjustment to an exposure level where such effects are not expected. This can be a matter  
7           of great uncertainty if no evidence is available at lower exposures. A factor of 3 or 10 will  
8           generally be applied to extrapolate to a lower exposure expected to be without appreciable  
9           effects. A factor other than 10 may also be considered, depending on the magnitude and  
10          nature of the response and the shape of the dose-response curve ([U.S. EPA, 2002b](#), [1998](#),  
11          [1996b](#), [1994](#), [1991a](#)).
- 12          • *Subchronic-to-chronic exposure*: When using studies of less-than-chronic exposure to make  
13          inferences about chronic/lifetime exposure, the assessment will consider whether lifetime  
14          exposure could reasonably be interpreted to result in effects at lower levels of exposure,  
15          including consideration of the specific health outcome(s) in question. A factor of up to 10  
16          will be considered, depending on the duration of the studies and the nature of the response  
17          ([U.S. EPA, 2002b](#), [1998](#), [1994](#)).
- 18          • *Database deficiencies*: In addition to the adjustments above, if database deficiencies raise  
19          concern that further studies might identify a more sensitive effect, organ system, or  
20          lifestage, the assessment will apply a database UF ([U.S. EPA, 2002b](#), [1998](#), [1996b](#), [1994](#),  
21          [1991a](#)). The size of the factor will depend on the nature of the database deficiency. For  
22          example, EPA typically follows the recommendation that a factor of 10 be applied if both a  
23          prenatal toxicity study and a two-generation reproduction study are missing and a factor of  
24          10<sup>0.5</sup> (i.e., 3) if either one or the other is missing ([U.S. EPA, 2002b](#)). As noted in Section 2.4.5,  
25          the evaluation of database completeness for these five PFAS will also consider existing  
26          knowledge gained through reviewing other, potentially similar, PFAS to identify data gaps.  
27          For example, there is the potential for exposure to PFAS to cause developmental effects  
28          (based on reviews of perfluorooctanoic acid [PFOA] and perfluorooctane sulfonate [PFOS])  
29          and there appears to be a lack of such studies for PFHxA. Thus, consideration of the  
30          potential for PFHxA exposure to cause developmental effects might review knowledge  
31          gained through the assessment of the other C6 PFAS, PFHxS, or the other short-chain  
32          perfluoroalkyl carboxylic acid, perfluorobutanoic acid (PFBA). In such cases, an  
33          interpretation of the relatedness between the PFAS of interest and the PFAS used for  
34          comparison will inform selection of the uncertainty factor.

35           The POD for a particular RfV will be divided by the product of these factors. As discussed in  
36           the technical document reviewing the RfD/RfC process ([U.S. EPA, 2002b](#)), any composite factor that  
37           exceeds 3,000 represents excessive uncertainty; thus, values with >3,000 UF<sub>c</sub> will not be used to  
38           derive RfVs. An RfD/RfC may be based on the POD for a single endpoint within a study, or on a  
39           collection of related PODs within or across studies, if such biological relationships are substantiated  
40           by the evidence. Confidence in any derived toxicity value(s) will be described based on three  
41           factors: confidence in the study(ies) used in the derivation of the toxicity value; confidence in the  
42           evidence base for the hazard(s) underlying the toxicity value, and confidence in the quantitative

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- 1 derivation of the toxicity value. The confidence description(s) will be separate from consideration
- 2 of the composite uncertainty factor applied to derive the toxicity value.

## 12. PROTOCOL HISTORY

1           **March 2020:** Comments on this protocol were provided in the public docket (see [Docket](#)  
 2 [ID: EPA-HQ-ORD-2019-0275](#) for detailed comments) during a 45-day public comment period from  
 3 November 8th, 2019 to December 23rd, 2019. Approximately 107 individual comments were  
 4 provided across a range of stakeholder groups. We thank the public commenters for their  
 5 constructive and informative reviews. The comments have been addressed in this update to the  
 6 protocol and were considered during development of the five draft IRIS PFAS assessments. A  
 7 summary of the comment topics and updates to the protocol is provided in Table 12-1 and below.

**Table 12-1. Topic areas of public comments on the protocol and how comments were addressed in this update (generally ordered based on descending number of comments on the topic areas)**

Topic areas raised by commenter(s)	Protocol updates and responses
<i>Toxicokinetics</i> (see Protocol Sections 2.4, 6.4, 9.2, and 11)	
Summary of comments on the use of ADME data in study selection and parameter choice for dose-response analysis: the protocol should add specificity on the approaches.	Added clarifying text to Section 11.1 on considering uncertainty in toxicokinetics across species when selecting studies for dose-response analysis, as well as how ADME information can influence selection of $C_{max}$ versus AUC as measure of risk. Also, as described below, the literature screening process for ADME data has been emphasized.
Summary of comments on the use of clearance versus half-life data: the protocol should add specificity on the approaches.	Added clarifying text to Sections 9.2.1 and 11.2.1 to explain that while clearance is preferred, these data will only be used if it is <i>not</i> based on assuming the same volume of distribution ( $V_d$ ) in the human as in a rodent. Otherwise this hides the assumption and gives the same result as the half-life ratio. Notably, the protocol maintains that analysis of the PK data will assume a single half-life estimate (half-life is assumed not to vary with dose); the text clarifies that use of a comparison across data sets will allow for an evaluation of this assumption. Specifically, if no evidence of nonlinearity is demonstrated, then it is assumed to be irrelevant at experimental levels. If evidence of nonlinearity is demonstrated, the analyses will focus on lower dose PK data.
Summary of comments on the use of allometric scaling versus data-specific adjustments: allometric scaling should be used for short-chain PFAS.	Added text to Section 11.2.1 that if half-life data are available in humans and rats but not mice, for example, then allometric scaling may be used to estimate the mouse half-life from the rat value (i.e., using two species closer in BW). This extrapolated mouse half-life can then be used with the

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	<p>measured human half-life to estimate an HED as described in the protocol (prior to this update). EPA guidance indicates a preference for data-specific adjustments over <math>BW^{3/4}</math> and says the latter should only be used in the absence of data. Hence the protocol was not revised in this regard, and the protocol still indicates use of a data-specific adjustment (e.g., a half-life ratio) when adequate data for doing so are available, irrespective of the PFAS chain length or magnitude of the half-life.</p>
<p><i>Human relevance and adversity of rodent responses (hepatic effects)</i> (see Protocol Sections 2.4.2 and 9.2.2)</p>	
<p>Summary of comments on interpreting PPAR<math>\alpha</math> responses in hepatic (and possibly other) health outcomes: comments varied, including both support for and against the human relevance of hepatic effects.</p>	<p>Expanded the discussion (and references) in Sections 2.4.2 and 9.2.2 on current information and uncertainties regarding the relative sensitivity of humans and animal models to PFAS-related PPAR<math>\alpha</math> inductions, incorporating information on the extent to which differences may relate to differing toxicokinetics as well as intrinsic variations in biological sensitivities. Examples and discussion have also been added regarding how prior evaluations of the role of PPAR<math>\alpha</math> will be considered. The updated protocol notes difficulties in applying read-across approaches related to the human relevance of rodent responses due to PFAS structural differences.</p>
<p>Summary of comments on interpreting rodent noncancer liver endpoint adversity: caution should be exercised in applying criteria developed in the context of cancer—such as the Hall et al. criteria, to noncancer endpoints.</p>	<p>Expanded the discussion on the applicability of the Hall criteria to noncancer hepatic endpoints and the development of lifetime toxicity values has been added to Section 9.2.3. This new text highlights the need to consider additional factors when applying the Hall et al. criteria during assessment development.</p>
<p><i>Study Evaluation</i> (see Protocol Sections 5 and 6).</p>	
<p>Summary of comments on overall study confidence: comments varied, including that the approach should not use the confidence ratings in a manner similar to “scoring” and that the approach should be more quantitative in the method for arriving at confidence.</p>	<p>Revised the protocol (see Section 6) to clarify that domain judgments and the specific limitations identified in the study are made available with the assessment and are carried forward to inform the synthesis; however, the overall approach was not changed. The overall study confidence ratings are not used as “scores” and are not provided without context. Text was also added to clarify that the overall study confidence is reached using expert judgment on the impact of the identified deficiencies for each specific study, and that there are no predefined weights for combining the domains.</p>
<p>Summary of comments on the use of <i>critically deficient</i> domain ratings: the approach should not exclude a study due to one <i>critical deficiency</i>.</p>	<p>Revised text in Section 6 to clarify <i>deficient</i> and <i>critically deficient</i> domain ratings; however, the overall approach was not changed. Specifically, the <i>critically deficient</i> category is used rarely and only in situations where the limitation is severe enough to warrant excluding a study as <i>uninformative</i> for the purposes of the assessment. Serious flaws that do not warrant study exclusion will be classified as <i>deficient</i>. Typically, domains rated as <i>deficient</i> are judged to reduce the</p>

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	reliability of the reported results. Studies with numerous <i>deficient</i> ratings may be excluded as <i>uninformative</i> .
Summary of comments on prioritizing epidemiology outcomes: the approach should not use one evaluator—as noted in the protocol for prioritization of some outcomes, or it should be clarified that the review of those outcomes performed with one evaluator were not systematic reviews.	Amended the approach in Section 5 to include two independent study evaluations for most outcomes. There is still a tiering system in place to prioritize outcomes, but the protocol now more clearly distinguishes the methodological rigor of “rapid reviews” from systematic reviews. Classification into the rapid review tier is now based on serious concern for reverse causality, determined a priori.
Summary of comments on study evaluation criteria: the approach should consider conflict of interest.	The approach in Section 6 was not modified to consider conflict of interest. The evaluations of risk of bias and sensitivity by subject matter experts are designed to encompass the primary aspects of methodological design that could engender concern, irrespective of the sponsoring entity.
Summary of comments on applying the exposure domain for study evaluation: caution should be exercised when assessing PFAS, in general, due to potential issues relating to analytical chemistry or physiochemical properties.	This was determined to not be a significant issue of concern for the specific PFAS being assessed. Additional support relating to this decision has been added to the protocol (see Section 6.3) based on review of data in the EPA Chemistry Dashboard.
<i>Use of Mechanistic Information</i> (see Protocol Sections 9.2, 11.2.1, and 11.2.2).	
Summary of comments on the use of mechanistic evidence and mode-of-action (MOA) understanding to inform dose-response (provided in the context of cancer): expand the discussion; note: as very few studies relevant to evaluating cancer are available for these five PFAS, these comments were viewed and addressed as more broadly applicable to any outcome.	Added clarifying text to Section 9.2.4 to emphasize that such data are considered for potential use quantitatively as well as qualitatively, and to Section 11.2.1 to indicate their potential use when a nonlinear (threshold) dose-response relationship is supported by the evidence.
Summary of comments on the use of mechanistic evidence and mode-of-action understanding to inform evaluation of key science topics: expand the discussion.	Expanded the discussions on the explicit consideration of mechanistic evidence for critical scientific topics, such as the human relevance and adversity of hepatic changes, as well as toxicokinetic interpretations, in Sections 2.4 and 9.2.
<i>Literature Identification</i> (see Protocol Sections 3.2 and 4)	
Summary of comments on PECO “outcome” criteria: ADME studies should not be “supplemental” for these PFAS.	Added clarifying text to Section 4.2 directing the reader to the separate literature identification and review process of ADME studies in Section 9.2.1.
Summary of comments on PECO “population” criteria: nonmammalian models are not included in the PECO, but they can be important.	Added a caveat to Section 4.2 clarifying the situationally increased utility of some nonmammalian models for certain outcomes.
<i>Other Comments</i>	
Summary comments on the use of the 10 key characteristics of carcinogens: the key characteristics should not be used to conduct the	The protocol (see Section 9.2) presents the key characteristics as an example approach to organize mechanistic evidence in a literature inventory, and not as a means to conduct the

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analysis of mechanistic evidence relevant to cancer.	evaluation or develop MOA judgments regarding those data; thus, this text was not revised.
Summary of comments on addressing data gaps and uncertainties using information from other, better-studied PFAS: comments varied, ranging from requests for an increased emphasis on use of information from more well-studied PFAS, up to including formal [re]assessments of PFOA and PFOS, to requests not to use data from other PFAS to influence interpretations regarding the five PFAS being assessed.	The scope of the assessments was not expanded to include PFOA and PFOS (see additional discussion in bullets below). However, as described in the protocol (see Sections 2.4.5, 9.2 and 10), the breadth of information on PFOA and PFOS (and other well-studied PFAS) is still considered potentially informative to these assessments. Examples of how these data are expected to be used include helping to identify key areas of potential concern (e.g., health outcomes associated with PFOA or PFOS exposure) that have not been examined for the PFAS of interest, and information (e.g., MOA information; studies on health outcomes of interest) on other PFAS pertinent to interpreting the reliability, adversity, and/or human relevance of effects observed in studies of the PFAS of interest. In addition, text has been added to Section 2.4 to emphasize that caution will be taken in drawing judgments for a PFAS of interest based on evidence on other PFAS due to cross-PFAS differences in toxicokinetics and toxicodynamics.
Summary of comments on evidence integration: the “weight of evidence” approach should not only be applied when integrating across evidence streams, but also in the analyses of individual streams.	The commenters misinterpreted the approach to evidence integration, as it lays out an evaluation of evidence strength within each stream of evidence as well as across evidence streams. For example: “Building from the separate syntheses of the human and animal evidence (see Section 9.1), the strength of the evidence from the available human and animal health effect studies will be summarized in parallel, but separately, using a structured evaluation of an adapted set of considerations first introduced by Sir Bradford Hill.” Additional text emphasizing this point has been added to the introductory materials in both Sections 9 and 10.
Summary of corrections and editorial suggestions: numerous comments, on a variety of topic areas.	Made edits to improve the accuracy of the protocol text; however, not all editorial suggestions were incorporated. Most notably, a number of suggestions related to Section 2.1 (Background) were not addressed. Because this section is meant to provide brief, contextual summaries and not comprehensive systematic reviews of the current information (as suggested by some of the comments), a clarifying introduction has been added regarding the purpose of this section. However, emphasis was added regarding certain aspects of exposure (e.g., the discussion of drinking water exposure) and potential susceptible population and lifestages (e.g., breast-fed infants) to improve context (see Sections 2.1.5 and 2.1.6). The section was also retitled to, “Summary of Background Information,” and the summaries of existing toxicity values for these PFAS were moved to Section 2.1.8 from Section 2.2.1) due to similar observations regarding comprehension and updating.

- 1 In addition to the changes in Table 12-1 made in direct response to public comments,
- 2 several other edits to the protocol were made during this update, specifically:

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- 1 • A technical edit was conducted. Grammatical errors were corrected, and some editorial  
2 changes were made that did not affecting the scientific approaches.
- 3 • Reference arrays providing a snapshot of existing toxicity values for these PFAS were  
4 moved from Protocol Section 2.3 (Problem Formulation) to Protocol Section 2.1  
5 (Background) and edited to remove values other than RfDs or RfD-like values (e.g., drinking  
6 water standards); these were also removed from Addendum A. These values were removed  
7 to reduce the potential for inappropriate comparisons.<sup>25</sup>
- 8 • Additional clarifications on considering CBI data for inclusion (specific to the timeliness of  
9 their availability in a publicly available form) are provided in Protocol Section 4.1.
- 10 • Text in Protocol Section 9.2.2 revised to reflect internal EPA discussions to minimize the use  
11 of single words or phrases to summarize weight-of-evidence-related judgments within the  
12 narratives for individual evidence streams (e.g., mechanistic evidence).
- 13 • Text in Protocol Sections 9.2 and 11.2.1 related to incorporating the available PBPK models  
14 (or their data) and toxicokinetics information was updated based on preliminary  
15 conclusions from a more robust evaluation of the available PBPK models and toxicokinetics  
16 data. In particular, the identified PBPK models do not appear adequate for direct  
17 application in these assessments, so alternatives are now emphasized.
- 18 • An updated version of the evidence profile table template was inserted in Section 10. The  
19 basic information included is unchanged; however, the presentation has been altered to  
20 increase transparency. In addition, text indicating that ***sufficient evidence for hazard*** can  
21 be judged based on a single epidemiology study without other supporting information has  
22 been edited to indicate that such evidence scenarios will be judged as ***insufficient evidence***.
- 23 • Text in Protocol Section 11 was updated to indicate that, when adequate data are available  
24 and it is appropriate to do so, less-than-lifetime (“subchronic”) and hazard-specific (“organ-  
25 or system-specific”) toxicity values will be derived in addition to an RfD and/or RfC. The  
26 derivation of these values is methodologically consistent with approaches already described  
27 in the protocol. During the protocol’s public comment period, EPA partners indicated that  
28 such less-than-lifetime values were potentially useful for certain decision contexts.
- 29 • Clarification was added to Section 11.2.3 that any toxicity values derived will be  
30 accompanied by a description of confidence.
- 31 • Protocol “Appendix” materials were renamed “Addendum” materials for clarity, as this  
32 protocol will be included as an Appendix to each of the five IRIS PFAS assessments.

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<sup>25</sup> IRIS does not derive drinking water standards or advisories. These values, which include information on human exposure and other considerations, are the purview of EPA programs (e.g., OW) and regional risk assessors. Although such standards or advisory levels may consider in their derivation reference values developed by IRIS, drinking water standards or advisories are not comparable to reference values, and thus they were removed so as not to convey an inappropriate comparison.

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1           In addition to the comments outlined in Table 12-1, other topics were raised which were  
2 outside of the scope of the protocol and thus did not warrant changes. The topic areas for these  
3 comments and the rationale for not updating the protocol in response to these comments are  
4 described below:

- 5           • Comments providing general support or criticism, or directing IRIS staff to other resources  
6 (e.g., other conclusions on these PFAS, or opinions on those other conclusions), did not  
7 result in changes to this protocol. Similarly, public comments recommending toxicity values  
8 that should be adopted by the EPA are not addressed by this protocol. As a reminder, IRIS  
9 assessment conclusions rely on independent evaluations of primary research studies,  
10 unless otherwise indicated as part of EPA scoping and problem formulation decisions  
11 (e.g., adopting a well-established conclusion).
- 12           • Comments relating to sites with potential PFAS contamination, recommendations for  
13 conducting PFAS exposure assessments, and requests for these assessments to include  
14 instruction on addressing coexposure to multiple PFAS are not addressed by this protocol,  
15 because such issues are the purview of other EPA programs, EPA regions, tribes, and states.
- 16           • A number of comments were related to the scope covered by these five assessments. This  
17 included recommendations that EPA should evaluate and regulate PFAS either as a class or  
18 individually (this opinion varied across commenters), or that other PFAS (e.g., PFOA; PFOS)  
19 should be (re)assessed simultaneously. In addition, some commenters wanted additional  
20 details regarding next steps for the EPA Action Plan; plans for addressing inconsistency in  
21 values developed by different federal agencies and states; or future EPA plans for  
22 monitoring, regulation, and enforcement. None of these comments are addressed by this  
23 protocol. Based on EPA program and regional needs, specific chemicals or substances  
24 (e.g., diesel exhaust) are nominated to the IRIS Program for independent, scientific  
25 assessment of potential human health hazards and dose-response analyses. The decisions  
26 on the PFAS for which an IRIS assessment would be useful, as well as the broader EPA plan  
27 for addressing PFAS, are not the purview of the IRIS Program.
- 28           • Several commenters requested details on the operating procedures used within the IRIS  
29 Program, specifically referencing the “IRIS Handbook.” These comments are not addressed  
30 by this protocol. The “IRIS Handbook” is not a public document and its development for  
31 public release are separate from the development of these assessments.
- 32           • A few commenters were interested in PFAS assessment-specific decisions rather than the  
33 methods and approaches for assessment development. As a reminder, the PFAS-specific  
34 literature screening decisions and updates will be available in HERO ([www.hero.epa.gov](http://www.hero.epa.gov));  
35 individual study evaluation decisions will be available in HAWC ([www.hawcproject.org](http://www.hawcproject.org));  
36 and decisions regarding studies and values used in support of assessment conclusions  
37 (e.g., studies and values selected to represent individual PFAS half-lives in different species;  
38 decisions regarding the human relevance of particular findings in animal studies) will be  
39 summarized and discussed in the specific PFAS assessments. As such, these comments are  
40 not further addressed in this protocol.

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## ADDENDUM A. SUMMARY OF EXISTING TOXICITY VALUE INFORMATION FOR PERFLUOROBUTANOIC ACID (PFBA), PERFLUOROHEXANOIC ACID (PFHXA), PERFLUOROHEXANESULFONIC ACID (PFHXS), PERFLUORONONANOIC ACID (PFNA), AND PERFLUORODECANOIC ACID (PFDA)

- 1 The values presented in Table A-1 through Table A-3 are current as of June 2019. Readers are referred to the individual sources
- 2 for the most up-to-date information, and more recent values from agencies not listed here may be available.

**Table A-1. Details on derivation of the available health effect reference values for inhalation exposure to selected per- and polyfluoroalkyl substances (PFAS) (current as of June 2019; please consult source references for up-to-date information)**

	Value name	Duration	PFAS	Value		Health effect	Point of departure	Qualifier	Source	Uncertainty factors	Notes on derivation	Review status
				(mg/m <sup>3</sup> )	(ppm)							
Emergency	PAC-3	1 h	PFBA	3.3 × 10 <sup>1</sup>	3.6 × 10 <sup>0</sup>	Lethality in mice	NR	NR		NR	PAC values derived via an approach developed by the Department of Energy ( <a href="#">DOE, 2016</a> )	Final ( <a href="#">DOE, 2018</a> )

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	Value name	Duration	PFAS	Value		Health effect	Point of departure	Qualifier	Source	Uncertainty factors	Notes on derivation	Review status
				(mg/m <sup>3</sup> )	(ppm)							
	PAC-2	1 h	PFBA	5.5 × 10 <sup>0</sup>	6.0 × 10 <sup>-1</sup>	Based on PAC-3	--	--	--	--	Based on PAC-3 <sup>a</sup>	
	PAC-1	1 h	PFBA	5.0 × 10 <sup>-1</sup>	5.5 × 10 <sup>-2</sup>	Based on PAC-2	--	--	--	--	Based on PAC-2 <sup>b</sup>	
General public	TCEQ RfC	Chronic	PFBA	1.0 × 10 <sup>-2</sup>	1.1 × 10 <sup>-3</sup>	Based on TCEQ RfD (see Table A-2)	--	--	--	--	Based on TCEQ RfD (route-to-route extrapolation) <sup>c</sup>	Final (TCEQ, 2016)
			PFDA	5.3 × 10 <sup>-5</sup>	2.5 × 10 <sup>-6</sup>	Based on TCEQ RfD (see Table A-2)	--	--	--	--	Based on TCEQ RfD (route-to-route extrapolation) <sup>d</sup>	
			PFNA	2.8 × 10 <sup>-5</sup>	1.4 × 10 <sup>-6</sup>	Lung noise, labored breathing, and reduced body wt. in male rats exposed for 4 h	67 mg/m <sup>3</sup> 0.83 mg/m <sup>3</sup>	NOAEL NOAEL <sub>HEC</sub>	<a href="#">Kinney et al. (1989)</a>	UF <sub>C</sub> = 30,000 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10 UF <sub>S</sub> = 100 UF <sub>D</sub> = 10	HEC adjusted <sup>e</sup>	
			PFHxS	1.3 × 10 <sup>-5</sup>	7.8 × 10 <sup>-7</sup>	Based on TCEQ RfD (see Table A-2)	--	--	--	--	Based on TCEQ RfD (route-to-route extrapolation) <sup>f</sup>	

<sup>a</sup>PAC-2 = PAC-3 ÷ 6 = 33 mg/m<sup>3</sup> ÷ 6 = 5.5 mg/m<sup>3</sup>.

<sup>b</sup>PAC-1 = PAC-1 ÷ 11 = 5.5 mg/m<sup>3</sup> ÷ 11 = 0.5 mg/m<sup>3</sup>.

<sup>c</sup>RfC = RfD × BW ÷ inhalation rate = 0.0029 mg/kg-day × 70 kg ÷ 20 m<sup>3</sup>/day = 0.01 mg/m<sup>3</sup>.

<sup>d</sup>RfC = RfD × BW ÷ inhalation rate = 0.000015 mg/kg-day × 70 kg ÷ 20 m<sup>3</sup>/day = 0.000053 mg/m<sup>3</sup>.

<sup>e</sup>NOAEL<sub>HEC</sub> = NOAEL ÷ TK adjustment factor = 67 mg/m<sup>3</sup> ÷ 81 = 0.83 mg/m<sup>3</sup>.

<sup>f</sup>RfC = RfD × BW ÷ inhalation rate = 0.0000038 mg/kg-day × 70 kg ÷ 20 m<sup>3</sup>/day = 0.000013 mg/m<sup>3</sup>.

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BW = body weight; HEC = human equivalent concentration; NOAEL = no-observed-adverse-effect level; NR = not reported; PAC = protective action criteria; PFAS = per- and polyfluoroalkyl substances; PFBA = perfluorobutanoic acid; PFDA = perfluorodecanoic acid; PFHxS = Perfluorohexanesulfonic acid; PFNA = perfluorononanoic acid; RfC = inhalation reference concentration; RfD = oral reference dose; TCEQ = Texas Commission on Environmental Quality; TK = toxicokinetic; UF<sub>A</sub> = interspecies uncertainty factor; UF<sub>C</sub> = composite uncertainty factor; UF<sub>D</sub> = database uncertainty factor; UF<sub>H</sub> = intraspecies uncertainty factor; UF<sub>L</sub> = LOAEL-to-NOAEL uncertainty factor; UF<sub>S</sub> = subchronic-to-chronic uncertainty factor.

**Table A-2. Details on derivation of the available health effect reference values for oral exposure to selected per- and polyfluoroalkyl substances (PFAS) (current as of June 2019; please consult source references for up-to-date information)**

Value name	Duration	PFAS	Value (mg/kg-d)	Health effect	Point of departure	Qualifier	Source	Uncertainty factors	Notes on derivation	Review status
MDH RfD	1–30 d	PFBA	$3.8 \times 10^{-3}$	Decreased cholesterol, serum total thyroxine, and dialysis free thyroxine and increased relative thyroid weight in rats	3.01 mg/kg-d 0.38 mg/kg-d	BMDL <sub>1SD</sub> BMDL <sub>HED</sub>	<a href="#">van Otterdijk (2007a)</a>	UF <sub>C</sub> = 100 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10 UF <sub>D</sub> = 3	HED adjusted <sup>a</sup>	Final ( <a href="#">MDH, 2018</a> )
	Subchronic		$2.9 \times 10^{-3}$	Liver-weight changes; morphological changes in the liver and thyroid gland; and decreased T4, RBCs, hematocrit, and Hb in rats	6.9 mg/kg-d 0.86 mg/kg-d	NOAEL NOAEL <sub>HED</sub>	<a href="#">van Otterdijk (2007b)</a>	UF <sub>C</sub> = 300 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10 UF <sub>D</sub> = 10	HED adjusted <sup>b</sup>	
	Chronic		$2.9 \times 10^{-3}$	Liver-weight changes; morphological changes in the liver and thyroid gland; and decreased T4, RBCs, hematocrit, and Hb in rats						
	1–30 d	PFHxS	$9.7 \times 10^{-6}$	Decreased free and total T4 and triiodothyronine (T3), changes in cholesterol levels, and increased hepatic focal necrosis in rats	32.4 mg/L serum 0.00292 mg/kg-d	BMDL <sub>20</sub> BMDL <sub>HED</sub>	<a href="#">NTP (2019)</a>	UF <sub>C</sub> = 300 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10 UF <sub>D</sub> = 10	HED adjusted <sup>c</sup>	

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Value name	Duration	PFAS	Value (mg/kg-d)	Health effect	Point of departure	Qualifier	Source	Uncertainty factors	Notes on derivation	Review status
<b>NH DES RfD</b>	Chronic	PFHxS	$4.0 \times 10^{-6}$	Reduced litter size in mice exposed for 14 d	13,900 ng/L serum  46.3 ng/mL	BMDL  Target human serum level	<a href="#">Chang et al. (2018)</a>	UF <sub>C</sub> = 300 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10 UF <sub>S</sub> = 3 UF <sub>D</sub> = 3	Target human serum level = BMDL ÷ UF  Calculated <sup>d</sup>	Final (New Hampshire DES, 2019)
		PFNA	$4.3 \times 10^{-6}$	Increased relative liver weights in mice exposed for 17 d	4,900 ng/L serum  49.0 ng/mL serum	BMDL  Target human serum level	<a href="#">Das et al. (2015)</a>	UF <sub>C</sub> = 100 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10 UF <sub>D</sub> = 3	Target human serum level = BMDL ÷ UF  Calculated <sup>e</sup>	
<b>TCEQ RfD</b>	Chronic	PFBA	$2.9 \times 10^{-3}$	Liver-weight changes; morphological changes in the liver and thyroid gland; and decreased T4, RBCs, hematocrit, and Hb in rats	6.9 mg/kg-d  0.86 mg/kg-d	NOAEL  NOAEL <sub>HED</sub>	<a href="#">van Otterdijk (2007b)</a>	UF <sub>C</sub> = 300 UF <sub>H</sub> = 10 UF <sub>S</sub> = 3 UF <sub>D</sub> = 10	HED adjusted <sup>f</sup>	Final (TCEQ, 2016)
		PFDA	$1.5 \times 10^{-5}$	Increased liver weight in rats dosed for 1 wk	1.2 mg/kg-d  0.015 mg/kg-d	NOAEL  NOAEL <sub>HED</sub>	<a href="#">Kawashima et al. (1995)</a>	UF <sub>C</sub> = 1,000 UF <sub>H</sub> = 10 UF <sub>S</sub> = 10 UF <sub>D</sub> = 10	HED adjusted <sup>g</sup>	
		PFHxS	$3.8 \times 10^{-6}$	Hematological alterations in male rats	0.3 mg/kg-d  0.0011 mg/kg-d	LOAEL  LOAEL <sub>HED</sub>	<a href="#">3M (2003)</a>	UF <sub>C</sub> = 300 UF <sub>H</sub> = 10 UF <sub>L</sub> = 3 UF <sub>D</sub> = 10	HED adjusted <sup>h</sup>	

*This document is a draft for review purposes only and does not constitute Agency policy.*

**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

Value name	Duration	PFAS	Value (mg/kg-d)	Health effect	Point of departure	Qualifier	Source	Uncertainty factors	Notes on derivation	Review status
		PFHxA	$3.8 \times 10^{-6}$	Adopted RfD for PFHxS	--	--	--	--	Adopted RfD for PFHxS	
		PFNA	$1.2 \times 10^{-5}$	Spleen cell apoptosis in rats	1 mg/kg-d 0.012 mg/kg-d	NOAEL NOAEL <sub>HED</sub>	<a href="#">Fang et al. (2010)</a>	UF <sub>C</sub> = 1,000 UF <sub>H</sub> = 10 UF <sub>S</sub> = 10 UF <sub>D</sub> = 10	HED adjusted <sup>i</sup>	
<b>Australia Dept. of Health TDI</b>	Chronic	Combined PFOS and PFHxS	$2 \times 10^{-5}$	Decreased body-weight gain in F0 female rats	0.1 mg/kg-d, 7.14 µg/mL 0.0006 mg/kg-d	NOAEL NOAEL <sub>HED</sub>	<a href="#">Luebker et al. (2005)</a>	UF <sub>C</sub> = 30 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10	HED adjusted <sup>j</sup>	Final ( <a href="#">FSANZ, 2016</a> )

<sup>a</sup>BMDL<sub>HED</sub> = BMDL<sub>1sd</sub> ÷ (t<sub>1/2</sub> Human ÷ t<sub>1/2</sub> Male Rat) = 3.01 mg/kg-day ÷ (72 h ÷ 9.22 h) = 0.38 mg/kg-day.

<sup>b</sup>NOAEL<sub>HED</sub> = NOAEL ÷ (t<sub>1/2</sub> Human ÷ t<sub>1/2</sub> Male Rat) = 6.9 mg/kg-day ÷ (72 h ÷ 9.22 h) = 0.86 mg/kg-day.

<sup>c</sup>BMDL<sub>HED</sub> = BMDL × volume of distribution × (ln2 ÷ t<sub>1/2</sub>) = 32.4 mg/L × 0.25 L/kg × (0.693 ÷ 1,935 days) = 0.00292 mg/kg-day.

<sup>d</sup>RfD = THSL × volume of distribution × (ln2 ÷ t<sub>1/2</sub>) = 46.3 ng/mL × 213 mL/kg × (0.693 ÷ 1,716 days) = 4.0 ng/kg-day.

<sup>e</sup>RfD = THSL × volume of distribution × (ln2 ÷ t<sub>1/2</sub>) = 49.0 ng/mL × 200 mL/kg × (0.693 ÷ 1,570 days) = 4.3 ng/kg-day.

<sup>f</sup>NOAEL<sub>HED</sub> = NOAEL ÷ TK adjustment factor = 6.9 mg/kg-day ÷ 8 = 0.86 mg/kg-day.

<sup>g</sup>NOAEL<sub>HED</sub> = NOAEL ÷ TK adjustment factor = 1.2 mg/kg-day ÷ 81 = 0.015 mg/kg-day.

<sup>h</sup>LOAEL<sub>HED</sub> = LOAEL ÷ TK adjustment factor = 0.3 mg/kg-day ÷ 263 = 0.0011 mg/kg-day.

<sup>i</sup>NOAEL<sub>HED</sub> = NOAEL ÷ TK adjustment factor = 1 mg/kg-day ÷ 81 = 0.012 mg/kg-day.

<sup>j</sup>TDI = NOAEL × volume of distribution × (ln2 ÷ t<sub>1/2</sub>) = 7.14 µg/mL × 0.23 L/kg × (0.693 ÷ 1,971 days) = 0.0006 mg/kg-day.

BMDL = benchmark dose lower confidence limit; HED = human equivalent dose; LOAEL = lowest-observed-adverse-effect level; MDH = Minnesota Department of Health; NH DES = New Hampshire Department of Environmental Services; NOAEL = no-observed-adverse-effect level; PFAS = per- and polyfluoroalkyl substances; PFBA = perfluorobutanoic acid; PFDA = perfluorodecanoic acid; PFHxA = perfluorohexanoic acid; PFHxS = Perfluorohexanesulfonic acid; PFNA = perfluorononanoic acid; PFOS = perfluorooctane sulfonate; RBC = red blood cell; RfD = oral reference dose; SD = standard deviation; TCEQ = Texas Commission on Environmental Quality; TDI = tolerable daily intake; THSL = target human serum level; TK = toxicokinetic; UF<sub>A</sub> = interspecies uncertainty factor; UF<sub>C</sub> = composite uncertainty factor; UF<sub>D</sub> = database uncertainty factor; UF<sub>H</sub> = intraspecies uncertainty factor; UF<sub>L</sub> = LOAEL-to-NOAEL uncertainty factor; UF<sub>S</sub> = subchronic-to-chronic uncertainty factor.

**Table A-3. Details on derivation of PFOA and PFOS reference values which served as the basis for values for the five per- and polyfluoroalkyl substances (PFAS) of interest (current as of June 2019; please consult source references for up-to-date information)**

Value name	Duration	PFAS	Value (mg/kg-d)	Health effect	Point of departure	Qualifier	Source	Uncertainty factors	Notes on derivation	Review status
EPA RfD (OW)	Chronic	PFOA	$2 \times 10^{-5}$	Decreased ossification and accelerated male puberty in F <sub>1</sub> mice	1 mg/kg-d, 38 mg/L serum  0.0053 mg/kg-d	LOAEL  LOAEL <sub>HED</sub>	<a href="#">Lau et al. (2006)</a>	UF <sub>C</sub> = 300 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10 UF <sub>L</sub> = 10 UF <sub>S</sub> = 1 UF <sub>D</sub> = 1	Average serum concentration derived using a PBPK model developed by <a href="#">Wambaugh et al. (2013)</a>  HED adjusted <sup>a</sup>	Final ( <a href="#">U.S. EPA, 2016b</a> )
		PFOS	$2 \times 10^{-5}$	Reduced body weight in F <sub>2</sub> rats	0.1 mg/kg-d, 6.26 µg/mL  0.00051 mg/kg-d	NOAEL  NOAEL <sub>HED</sub>	<a href="#">Luebker et al. (2005)</a>	UF <sub>C</sub> = 30 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10 UF <sub>L</sub> = 1 UF <sub>S</sub> = 1 UF <sub>D</sub> = 1	Average serum concentration derived using a PBPK model developed by <a href="#">Wambaugh et al. (2013)</a>  HED adjusted <sup>b</sup>	Final ( <a href="#">U.S. EPA, 2016a</a> )
Danish EPA TDI	Chronic	PFOS	$3 \times 10^{-5}$	Liver lesions in male rats	0.033 mg/kg-d  0.0008 mg/kg-d	BMDL <sub>10</sub>  BMDL <sub>HED</sub>	<a href="#">Thomford (2002)</a>	UF <sub>C</sub> = 30 UF <sub>A</sub> = 3 UF <sub>H</sub> = 10	Pharmacokinetic adjustments based on those in <a href="#">U.S. EPA (2014a)</a>  HED adjusted <sup>c</sup>	Final ( <a href="#">Danish EPA, 2015</a> )

<sup>a</sup>LOAEL<sub>HED</sub> = LOAEL × volume of distribution × (ln2 ÷ t<sub>1/2</sub>) = 38 mg/L × 0.17 L/kg × (0.693 ÷ 839.5 days) = 0.0053 mg/kg-day.

<sup>b</sup>NOAEL<sub>HED</sub> = NOAEL × volume of distribution × (ln2 ÷ t<sub>1/2</sub>) = 6.26 µg/mL × 0.23 L/kg × (0.693 ÷ 1,971 days) = 0.00051 mg/kg-day.

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$${}^c\text{BMDL}_{\text{HED}} = \text{BMDL}_{10} \div ([\text{volume of distribution} \times (\ln 2 \div t_{1/2 \text{ Rat}})] \div [\text{volume of distribution} \times (\ln 2 \div t_{1/2 \text{ Human}})]) = 0.033 \text{ mg/kg-day} \div ([0.23 \text{ L/kg} \times (0.693 \div 48 \text{ days})] \div [0.23 \text{ L/kg} \times (0.693 \div 1,971 \text{ days})]) = 0.0008 \text{ mg/kg-day}.$$

BMDL = benchmark dose lower confidence limit; EPA = Environmental Protection Agency; HED = human equivalent dose; LOAEL = lowest-observed-adverse-effect level; NOAEL = no-observed-adverse-effect level; OW = Office of Water; PBPK = physiologically based pharmacokinetic; PFAS = per- and polyfluoroalkyl substances; PFOA = perfluorooctanoic acid; PFOS = perfluorooctane sulfonate; RfD = oral reference dose; TDI = tolerable daily intake;  $\text{UF}_A$  = interspecies uncertainty factor;  $\text{UF}_C$  = composite uncertainty factor;  $\text{UF}_D$  = database uncertainty factor;  $\text{UF}_H$  = intraspecies uncertainty factor;  $\text{UF}_L$  = LOAEL-to-NOAEL uncertainty factor;  $\text{UF}_S$  = subchronic-to-chronic uncertainty factor.

## ADDENDUM B. SEARCH AND SCREENING STRATEGIES

Table B-1. Perfluorobutanoic acid (PFBA) database search strategy

Search	Search strategy	Dates of search
<b>PubMed</b>		
Search terms	375-22-4[rn] OR "Heptafluoro-1-butanoic acid"[tw] OR "Heptafluorobutanoic acid"[tw] OR "Heptafluorobutyric acid"[tw] OR "Kyselina heptafluormaselna"[tw] OR "Perfluorobutanoic acid"[tw] OR "Perfluorobutyric acid"[tw] OR "Perfluoropropanecarboxylic acid"[tw] OR "2,2,3,3,4,4,4-heptafluoro-Butanoic acid"[tw] OR "Butanoic acid, 2,2,3,3,4,4,4-heptafluoro-"[tw] OR "Butanoic acid, heptafluoro-"[tw] OR "Perfluoro-n-butanoic acid"[tw] OR "Perfluorobutanoate"[tw] OR "2,2,3,3,4,4,4-Heptafluorobutanoic acid"[tw] OR "Butyric acid, heptafluoro-"[tw] OR "Fluorad FC 23"[tw] OR "H 0024"[tw] OR "NSC 820"[tw] OR ((PFBA[tw] OR "FC 23"[tw] OR HFBA[tw]) AND (fluorocarbon*[tw] OR fluorotelomer*[tw] OR polyfluoro*[tw] OR perfluoro-*[tw] OR perfluorooa*[tw] OR perfluorob*[tw] OR perfluoroc*[tw] OR perfluorod*[tw] OR perfluoroe*[tw] OR perfluoroh*[tw] OR perfluoron*[tw] OR perfluoroo*[tw] OR perfluorop*[tw] OR perfluoros*[tw] OR perfluorou*[tw] OR perfluorinated[tw] OR fluorinated[tw] OR PFAS[tw] OR PFOS[tw] OR PFOA[tw]))	No date limit-7/19/2017
Literature update search terms	((((375-22-4[rn] OR "Heptafluoro-1-butanoic acid"[tw] OR "Heptafluorobutanoic acid"[tw] OR "Heptafluorobutyric acid"[tw] OR "Kyselina heptafluormaselna"[tw] OR "Perfluorobutanoic acid"[tw] OR "Perfluorobutyric acid"[tw] OR "Perfluoropropanecarboxylic acid"[tw] OR "2,2,3,3,4,4,4-heptafluoro-Butanoic acid"[tw] OR "Butanoic acid, 2,2,3,3,4,4,4-heptafluoro-"[tw] OR "Butanoic acid, heptafluoro-"[tw] OR "Perfluoro-n-butanoic acid"[tw] OR "Perfluorobutanoate"[tw] OR "2,2,3,3,4,4,4-Heptafluorobutanoic acid"[tw] OR "Butyric acid, heptafluoro-"[tw] OR "Fluorad FC 23"[tw] OR "H 0024"[tw] OR "NSC 820"[tw] OR ((PFBA[tw] OR "FC 23"[tw] OR HFBA[tw]) AND (fluorocarbon*[tw] OR fluorotelomer*[tw] OR polyfluoro*[tw] OR perfluoro-*[tw] OR perfluorooa*[tw] OR perfluorob*[tw] OR perfluoroc*[tw] OR perfluorod*[tw] OR perfluoroe*[tw] OR perfluoroh*[tw] OR perfluoron*[tw] OR perfluoroo*[tw] OR perfluorop*[tw] OR perfluoros*[tw] OR perfluorou*[tw] OR perfluorinated[tw] OR fluorinated[tw] OR PFAS[tw] OR PFOS[tw] OR PFOA[tw])) AND ("2017/08/01"[PDAT] : "2018/02/14"[PDAT]))	8/1/2017-2/14/2018
<b>Web of Science</b>		

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Search	Search strategy	Dates of search
Search terms	TS="Heptafluoro-1-butanoic acid" OR TS="Heptafluorobutanoic acid" OR TS="Heptafluorobutyric acid" OR TS="Kyselina heptafluormaselná" OR TS="Perfluorobutanoic acid" OR TS="Perfluorobutyric acid" OR TS="Perfluoropropanecarboxylic acid" OR TS="2,2,3,3,4,4,4-heptafluoro-Butanoic acid" OR TS="Butanoic acid, 2,2,3,3,4,4,4-heptafluoro-" OR TS="Butanoic acid, heptafluoro-" OR TS="Perfluoro-n-butanoic acid" OR TS="Perfluorobutanoate" OR TS="2,2,3,3,4,4,4-Heptafluorobutanoic acid" OR TS="Butyric acid, heptafluoro-" OR TS="Fluorad FC 23" OR TS="H 0024" OR TS="NSC 820" OR (TS=(PFBA OR "FC 23" OR HFBA) AND TS=(fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro-* OR perfluoroa* OR perfluorob* OR perfluoroc* OR perfluorod* OR perfluoroe* OR perfluoroh* OR perfluoron* OR perfluoroo* OR perfluorop* OR perfluoros* OR perfluorou* OR perfluorinated OR fluorinated OR PFAS OR PFOS OR PFOA))	No date limit–7/20/2017
Literature update search terms	((TS="Heptafluoro-1-butanoic acid" OR TS="Heptafluorobutanoic acid" OR TS="Heptafluorobutyric acid" OR TS="Kyselina heptafluormaselná" OR TS="Perfluorobutanoic acid" OR TS="Perfluorobutyric acid" OR TS="Perfluoropropanecarboxylic acid" OR TS="2,2,3,3,4,4,4-heptafluoro-Butanoic acid" OR TS="Butanoic acid, 2,2,3,3,4,4,4-heptafluoro-" OR TS="Butanoic acid, heptafluoro-" OR TS="Perfluoro-n-butanoic acid" OR TS="Perfluorobutanoate" OR TS="2,2,3,3,4,4,4-Heptafluorobutanoic acid" OR TS="Butyric acid, heptafluoro-" OR TS="Fluorad FC 23" OR TS="H 0024" OR TS="NSC 820") OR TS=(PFBA OR "FC 23" OR HFBA) AND TS=(fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro-* OR perfluoroa* OR perfluorob* OR perfluoroc* OR perfluorod* OR perfluoroe* OR perfluoroh* OR perfluoron* OR perfluoroo* OR perfluorop* OR perfluoros* OR perfluorou* OR perfluorinated OR fluorinated OR PFAS OR PFOS OR PFOA)) AND PY=2017-2018	2017–2018
<b>Toxline</b>		
Search terms	( 375-22-4 [rn] OR "heptafluoro-1-butanoic acid" OR "heptafluorobutanoic acid" OR "heptafluorobutyric acid" OR "kyselina heptafluormaselná" OR "perfluorobutanoic acid" OR "perfluorobutyric acid" OR "perfluoropropanecarboxylic acid" OR "2,2,3,3,4,4,4-heptafluoro-butanoic acid" OR "butanoic acid 2 2 3 3 4 4 4-heptafluoro-" OR "butanoic acid heptafluoro-" OR "perfluoro-n-butanoic acid" OR "perfluorobutanoate" OR "2,2,3,3,4,4,4-heptafluorobutanoic acid" OR "butyric acid heptafluoro-" OR "fluorad fc 23" OR "h 0024" OR "nsc 820" OR ( ( pfba OR "fc 23" OR hfba ) AND ( fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro* OR perfluorinated OR fluorinated OR pfas OR pfos OR pfoa ) ) ) AND ( ANEUPL [org] OR BIOSIS [org] OR CIS [org] OR DART [org] OR EMIC [org] OR EPIDEM [org] OR HEEP [org] OR HMTC [org] OR IPA [org] OR RISKLINE [org] OR MTGABS [org] OR NIOSH [org] OR NTIS [org] OR PESTAB [org] OR PPBIB [org] ) AND NOT PubMed [org] AND NOT pubdart [org]	No date limit–7/20/2017

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Search	Search strategy	Dates of search
Literature update search terms	@AND+@OR(("heptafluoro-1-butanoic acid"+"heptafluorobutanoic+acid"+"heptafluorobutyric+acid"+"kyselina+heptafuormaselná"+"perfluorobutanoic+acid"+"perfluorobutyric+acid"+"perfluoropropanecarboxylic +acid"+"2 2 3 3 4 4 4-heptafluoro-butanoic+acid"+"butanoic+acid+2 2 3 3 4 4 4-heptafluoro-"+"butanoic+acid+heptafluoro-"+"perfluoro-n-butanoic acid"+"perfluorobutanoate"+"2 2 3 3 4 4 4-heptafluorobutanoic+acid"+"butyric+acid+heptafluoro-"+"fluorad+fc+23"+"h0024"+"nsc+820"+@TERM+@rn+375-22-4("pfba"+"fc+23"+"hfba"))+(fluorocarbon*+fluorotelomer*+polyfluoro*+perfluoro*+perfluorinated+fluorinated+pfas+pfoa)+@RANGE+yr+2017+2018	2017–2018
<b>TSCATS</b>		
Search terms	375-22-4[rn] AND tscats[org]	No date limit–7/20/2017

**Table B-2. Perfluorodecanoic acid (PFDA) database search strategy**

Search	Search strategy	Dates of search
<b>PubMed</b>		
Search terms	335-76-2[rn] OR "Ndfda"[tw] OR "Nonadecafluoro-n-decanoic acid"[tw] OR "Nonadecafluorodecanoic acid"[tw] OR "Perfluoro-n-decanoic acid"[tw] OR "Perfluorodecanoic acid"[tw] OR "2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-Decanoic acid"[tw] OR "Decanoic acid, 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-"[tw] OR "Decanoic acid, nonadecafluoro-"[tw] OR "Perfluorodecanoate"[tw] OR "PFDeA"[tw] OR "PFDcA"[tw] OR ("PFDA"[tw] AND (fluorocarbon*[tw] OR fluorotelomer*[tw] OR polyfluoro*[tw] OR perfluoro-*[tw] OR perfluorooa*[tw] OR perfluorob*[tw] OR perfluoroc*[tw] OR perfluorod*[tw] OR perfluoroe*[tw] OR perfluoroh*[tw] OR perfluoron*[tw] OR perfluoroo*[tw] OR perfluorop*[tw] OR perfluoros*[tw] OR perfluorou*[tw] OR perfluorinated[tw] OR fluorinated[tw] OR PFAS[tw] OR PFOS[tw] OR PFOA[tw]))	No date limit-7/26/2017
Literature update search terms	((335-76-2[rn] OR "Ndfda"[tw] OR "Nonadecafluoro-n-decanoic acid"[tw] OR "Nonadecafluorodecanoic acid"[tw] OR "Perfluoro-n-decanoic acid"[tw] OR "Perfluorodecanoic acid"[tw] OR "2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-Decanoic acid"[tw] OR "Decanoic acid, 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-"[tw] OR "Decanoic acid, nonadecafluoro-"[tw] OR "Perfluorodecanoate"[tw] OR "PFDeA"[tw] OR "PFDcA"[tw] OR ("PFDA"[tw] AND (fluorocarbon*[tw] OR fluorotelomer*[tw] OR polyfluoro*[tw] OR perfluoro-*[tw] OR perfluorooa*[tw] OR perfluorob*[tw] OR perfluoroc*[tw] OR perfluorod*[tw] OR perfluoroe*[tw] OR perfluoroh*[tw] OR perfluoron*[tw] OR perfluoroo*[tw] OR perfluorop*[tw] OR perfluoros*[tw] OR perfluorou*[tw] OR perfluorinated[tw] OR fluorinated[tw] OR PFAS[tw] OR PFOS[tw] OR PFOA[tw])) AND ("2017/08/01"[Date - Publication] : "2018/03/01"[Date - Publication])	8/1/2017-2/14/2018
<b>Web of Science</b>		

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Search	Search strategy	Dates of search
Search terms	TS="PFDeA" OR TS="PFDcA" OR TS="Ndfda" OR TS="Nonadecafluoro-n-decanoic acid" OR TS="Nonadecafluorodecanoic acid" OR TS="Perfluoro-n-decanoic acid" OR TS="Perfluorodecanoic acid" OR TS="2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-Decanoic acid" OR TS="Decanoic acid, 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-" OR TS="Decanoic acid, nonadecafluoro-" OR TS="Perfluorodecanoate" OR (TS=PFDA AND TS=(fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro-* OR perfluoroa* OR perfluorob* OR perfluoroc* OR perfluorod* OR perfluoroe* OR perfluoroh* OR perfluoron* OR perfluoroo* OR perfluorop* OR perfluoros* OR perfluorou* OR perfluorinated OR fluorinated)) OR (TS=PFDA AND TS=(fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro-* OR perfluoroa* OR perfluorob* OR perfluoroc* OR perfluorod* OR perfluoroe* OR perfluoroh* OR perfluoron* OR perfluoroo* OR perfluorop* OR perfluoros* OR perfluorou* OR perfluorinated OR fluorinated OR PFAS OR PFOS OR PFOA	No date limit-7/26/2017
Literature update search terms	TS="PFDeA" OR TS="PFDcA" OR TS="Ndfda" OR TS="Nonadecafluoro-n-decanoic acid" OR TS="Nonadecafluorodecanoic acid" OR TS="Perfluoro-n-decanoic acid" OR TS="Perfluorodecanoic acid" OR TS="2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-Decanoic acid" OR TS="Decanoic acid, 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-" OR TS="Decanoic acid, nonadecafluoro-" OR TS="Perfluorodecanoate" OR (TS=PFDA AND TS=(fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro-* OR perfluoroa* OR perfluorob* OR perfluoroc* OR perfluorod* OR perfluoroe* OR perfluoroh* OR perfluoron* OR perfluoroo* OR perfluorop* OR perfluoros* OR perfluorou* OR perfluorinated OR fluorinated)) OR (TS=PFDA AND TS=(fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro-* OR perfluoroa* OR perfluorob* OR perfluoroc* OR perfluorod* OR perfluoroe* OR perfluoroh* OR perfluoron* OR perfluoroo* OR perfluorop* OR perfluoros* OR perfluorou* OR perfluorinated OR fluorinated OR PFAS OR PFOS OR PFOA)) AND PY=2017-2018	2017-2018
<b>Toxline</b>		
Search terms	( 335-76-2 [rn] OR "2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluorodecanoic acid" OR "2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-decanoic acid" OR "decanoic acid 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,10,10,10-nonadecafluoro-" OR "decanoic acid nonadecafluoro-" OR "nonadecafluoro-n-decanoic acid" OR "nonadecafluorodecanoic acid" OR "perfluoro-1-nonanecarboxylic acid" OR "perfluoro-n-decanoic acid" OR "perfluorocapric acid" OR "perfluorodecanoate" OR "perfluorodecanoic acid" OR "ndfda" OR "PFDeA" OR "PFDcA" OR ( pfda AND ( fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro* OR perfluorinated OR fluorinated OR pfas OR pfos OR pfoa ) ) AND ( ANEUPL [org] OR BIOSIS [org] OR CIS [org] OR DART [org] OR EMIC [org] OR EPIDEM [org] OR HEEP [org] OR HMTC [org] OR IPA [org] OR RISKLINE [org] OR MTGABS [org] OR NIOSH [org] OR NTIS [org] OR PESTAB [org] OR PPBIB [org] ) AND NOT PubMed [org] AND NOT pubdart [org]	No date limit-7/21/2017

*Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments*

<b>Search</b>	<b>Search strategy</b>	<b>Dates of search</b>
Literature update search terms		2017–2018
<b>TSCATS</b>		
Search terms	335-76-2[rn] AND TSCATS[org]	No date limit–7/21/2017

**Table B-3. Perfluorononanoic acid (PFNA) database search strategy**

Search	Search strategy	Dates of search
<b>PubMed</b>		
Search terms	"375-95-1"[rn] OR "2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluorononanoic acid"[tw] OR "Nonanoic acid, 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluoro-"[tw] OR "Nonanoic acid, heptadecafluoro-"[tw] OR "Perfluoro-n-nonanoic acid"[tw] OR "Perfluorononan-1-oic acid"[tw] OR "Perfluorononanoate"[tw] OR "Perfluorononanoic acid"[tw] OR "Perfluorononanononic acid"[tw] OR "Perfluoropelargonic acid"[tw] OR "heptadecafluorononanoic acid"[tw] OR ("PFNA"[tw] OR "C 1800"[tw]) AND (fluorocarbon*[tw] OR fluorotelomer*[tw] OR polyfluoro*[tw] OR perfluoro-*[tw] OR perfluoroa*[tw] OR perfluorob*[tw] OR perfluoroc*[tw] OR perfluorod*[tw] OR perfluoroe*[tw] OR perfluoroh*[tw] OR perfluoron*[tw] OR perfluoroo*[tw] OR perfluorop*[tw] OR perfluoros*[tw] OR perfluorou*[tw] OR perfluorinated[tw] OR fluorinated[tw] OR PFAS[tw] OR PFOS[tw] OR PFOA[tw]))	No date limit-7/26/2017
Literature update and additional PFNA synonyms search terms	(((("2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluorononanoic acid" [tw] OR "Nonanoic acid, 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluoro-" [tw] OR "Nonanoic acid, heptadecafluoro-" [tw] OR "Perfluoro-n-nonanoic acid" [tw] OR "Perfluorononan-1-oic acid" [tw] OR "Perfluorononanoate" [tw] OR "Perfluorononanoic acid" [tw] OR "Perfluorononanononic acid" [tw] OR "Perfluoropelargonic acid" [tw] OR "heptadecafluorononanoic acid" [tw] OR "PFNA" [tw] OR "C 1800" [tw] OR "Methyl-n1-Perfluorononanoic acid" [tw] OR "PFNA-n1CH3" [tw] OR "EINECS 206-801-3" [tw] OR "Heptadecafluornonansaeure" [tw] OR "Heptadekafluornonansaeure" [tw] OR "Perfluornonansaeure" [tw] OR "Perfluorononanoic acid (PFNA)" [tw] OR "UNII-5830Z6S63M" [tw] OR "perfluoro-n-nonanoic acid" [tw] OR "perfluorononan-1-oic acid" [tw] OR "perfluorononanoic acid" [tw] OR "Ammonium Perfluorononanoate" [tw] OR "Ammonium perfluorononanoate" [tw] OR "PFNA-H3N" [tw]))) AND ("2017/01/01"[Date - Publication] : "3000"[Date - Publication])	1/2017-4/2018
<b>Web of Science</b>		
Search terms	((TS=PFNA OR TS="C 1800") AND TS=(fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro-* OR perfluoroa* OR perfluorob* OR perfluoroc* OR perfluorod* OR perfluoroe* OR perfluoroh* OR perfluoron* OR perfluoroo* OR perfluorop* OR perfluoros* OR perfluorou* OR perfluorinated OR fluorinated OR PFAS OR PFOS OR PFOA)) OR TS="2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluorononanoic acid" OR TS="Nonanoic acid, 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluoro-" OR TS="Nonanoic acid, heptadecafluoro-" OR TS="Perfluoro-n-nonanoic acid" OR TS="Perfluorononan-1-oic acid" OR TS="Perfluorononanoate" OR TS="Perfluorononanoic acid" OR TS="Perfluorononanononic acid" OR TS="Perfluoropelargonic acid" OR TS="heptadecafluorononanoic acid"	No date limit-7/26/2017

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Search	Search strategy	Dates of search
Literature update and additional PFNA synonyms search terms	(TS="PFNA" OR TS="C 1800" OR TS="2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluorononanoic acid" OR TS="Nonanoic acid, 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluoro-" OR TS="Nonanoic acid, heptadecafluoro-" OR TS="Methyl-n1-Perfluorononanoic acid" OR TS="PFNA-n1CH3" OR TS="EINECS 206-801-3" OR TS="Heptadecafluorononansaeure" OR TS="Heptadecafluorononansaeure" OR TS="Perfluorononansaeure" OR TS="Perfluorononanoic acid (PFNA)" OR TS="UNII-5830Z6S63M" OR TS="perfluoro-n-nonanoic acid" OR TS="perfluorononan-1-oic acid" OR TS="perfluorononanoic acid" OR TS="Ammonium Perfluorononanoate" OR TS="Ammonium perfluorononanoate" OR TS="PFNA-H3N") AND PY=2017-2018	2017–2018
<b>Toxline</b>		
Search terms	(( pfna OR "c 1800" ) AND ( fluorocarbon* OR fluorotelomer* OR polyfluoro* OR perfluoro* OR perfluorinated OR fluorinated OR pfas OR pfos OR pfoa ) OR "375-95-1" [rn] OR "2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluorononanoic acid" OR "nonanoic acid 2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluoro-" OR "nonanoic acid heptadecafluoro-" OR "perfluoro-n-nonanoic acid" OR "perfluorononan-1-oic acid" OR "perfluorononanoate" OR "perfluorononanoic acid" OR "perfluorononanoic acid" OR "perfluoropelargonic acid" OR "heptadecafluorononanoic acid" )) AND ( ANEUPL [org] OR BIOSIS [org] OR CIS [org] OR DART [org] OR EMIC [org] OR EPIDEM [org] OR HEEP [org] OR HMTc [org] OR IPA [org] OR RISKLINE [org] OR MTGABS [org] OR NIOSH [org] OR NTIS [org] OR PESTAB [org] OR PPBIB [org] ) AND NOT PubMed [org] AND NOT pubdart [org]	No date limit–7/26/2017
Literature update and additional PFNA synonyms search terms	@AND+@OR+(pfna+"c 1800"+fluorocarbon*+"2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluorononanoic+acid"+"nonanoic+acid+2,2,3,3,4,4,5,5,6,6,7,7,8,8,9,9,9-heptadecafluoro-"+ "nonanoic+acid+heptadecafluoro-"+ "perfluoro-n-nonanoic+acid"+"perfluorononan-1-oic+acid"+perfluorononanoate+perfluorononanoic+acid"+"perfluoropelargonic+acid"+"heptadecafluorononanoic+acid"+"Methyl-n1-Perfluorononanoic+acid"+"PFNA-n1CH3"+"EINECS 206-801-3"+"Heptadecafluorononansaeure"+"Heptadecafluorononansaeure"+"Perfluorononansaeure"+"Perfluorononanoic+acid (PFNA)+"UNII-5830Z6S63M"+"perfluoro-n-nonanoic+acid"+"perfluorononan-1-oic+acid"+"perfluorononanoic+acid"+"Ammonium+Perfluorononanoate"+"Ammonium+perfluorononanoate"+"PFNA-H3N"+@TERM+@rn+375-95-1)+@RANGE+yr+2017+2018	2017–2018
<b>TSCATS</b>		
Search terms	"375-95-1" [rn] AND TSCATS[org]	No date limit–7/20/2017

***Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments***

<b>Search</b>	<b>Search strategy</b>	<b>Dates of search</b>
Literature update and additional PFNA synonyms search terms	@TERM+@rn+375-95-1+@RANGE+yr+2017+2018	2017–2018

**Table B-4. Perfluorohexanoic acid (PFHxA) database search strategy**

Search	Search strategy	Dates of search
<b>PubMed</b>		
Search terms	((307-24-4[rn] OR "2,2,3,3,4,4,5,5,6,6,6-undecafluorohexanoic acid"[tw] OR "2,2,3,3,4,4,5,5,6,6,6-undecafluoro-hexanoic acid"[tw] OR "hexanoic acid, 2,2,3,3,4,4,5,5,6,6,6-undecafluoro-"[tw] OR "hexanoic acid, undecafluoro-"[tw] OR "perfluorohexanoic acid"[tw] OR "perfluoro-1-pentanecarboxylic acid"[tw] OR "perfluorocaproic acid"[tw] OR "perfluorohexanoate"[tw] OR "perfluorohexanoic acid"[tw] OR "undecafluoro-1-hexanoic acid"[tw] OR "undecafluorocaproic acid"[tw] OR "undecafluorohexanoic acid"[tw] OR "PFHxA"[tw])) AND ("2017/08/01"[PDAT] : "2018/02/28"[PDAT])	No date limit–7/21/2017
Literature update search terms	((92612-52-7[EC/RN Number]) OR 355-38-4[EC/RN Number]) OR 2062-98-8[EC/RN Number]) OR "PFHxA_ion"[tw] OR "Perfluorohexanoate"[tw] OR "Hexanoyl fluoride, 2,2,3,3,4,4,5,5,6,6,6-undecafluoro-"[tw] OR "Hexanoyl fluoride, undecafluoro-"[tw] OR "Perfluorohexanoyl fluoride"[tw] OR "Undecafluorohexanoyl fluoride"[tw] OR "Perfluoro(2-methyl-3-oxahexanoyl) fluoride"[tw] OR "Propanoyl fluoride, 2,3,3,3-tetrafluoro-2-(1,1,2,2,3,3,3-heptafluoropropoxy)-" [tw] OR "Propanoyl fluoride, 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-" [tw] OR "Propionyl fluoride, tetrafluoro-2-(heptafluoropropoxy)-" [tw] OR "2,2,3,3,4,4,5,5,6,6,6-Undecafluorohexanoic acid"[tw] OR "EINECS 206-196-6"[tw] OR "NSC 5213"[tw] OR "Perfluoro-1-pentanecarboxylic acid"[tw] OR "Perfluoro-n-hexanoic acid"[tw] OR "UNII-ZP34Q2220R"[tw] OR "Undecafluorocaproic acid"[tw] OR "Ammonium Perfluorohexanoate"[tw] OR "PFHxA-H3N"[tw] OR "PFHxA-Na"[tw] OR "Sodium Perfluorohexanoate"[tw]))	8/1/2017–2/14/2018
<b>Web of Science</b>		
Search terms	((TS="2,2,3,3,4,4,5,5,6,6,6-undecafluorohexanoic acid" OR TS="2,2,3,3,4,4,5,5,6,6,6-undecafluoro-hexanoic acid" OR TS="hexanoic acid, 2,2,3,3,4,4,5,5,6,6,6-undecafluoro-" OR TS="hexanoic acid, undecafluoro-" OR TS="perfluorohexanoic acid" OR TS="perfluoro-1-pentanecarboxylic acid" OR TS="perfluorocaproic acid" OR TS="perfluorohexanoate" OR TS="perfluorohexanoic acid" OR TS="undecafluoro-1-hexanoic acid" OR TS="undecafluorocaproic acid" OR TS="undecafluorohexanoic acid" OR TS="PFHxA")) AND PY=2017-2018	No date limit–7/24/2017

**Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments**

Search	Search strategy	Dates of search
Literature update search terms	TS="PFHxA_ion" OR TS="Perfluorohexanoate" OR TS="Hexanoyl fluoride, 2,2,3,3,4,4,5,5,6,6,6-undecafluoro-" OR TS="Hexanoyl fluoride, undecafluoro-" OR TS="Perfluorohexanoyl fluoride" OR TS="Undecafluorohexanoyl fluoride" OR TS="Perfluoro(2-methyl-3-oxahexanoyl) fluoride" OR TS="Propanoyl fluoride, 2,3,3,3-tetrafluoro-2-(1,1,2,2,3,3,3-heptafluoropropoxy)-" OR TS="Propanoyl fluoride, 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-" OR TS="Propionyl fluoride, tetrafluoro-2-(heptafluoropropoxy)-" OR TS="2,2,3,3,4,4,5,5,6,6,6-Undecafluorohexanoic acid" OR TS="EINECS 206-196-6" OR TS="NSC 5213" OR TS="Perfluoro-1-pentanecarboxylic acid" OR TS="Perfluoro-n-hexanoic acid" OR TS="UNII-ZP34Q2220R" OR TS="Undecafluorocaproic acid" OR TS="Undecafluorohexanoic acid" OR TS="Ammonium Perfluorohexanoate" OR TS="PFHxA-H3N" OR TS="PFHxA-Na" OR TS="Sodium Perfluorohexanoate"	2017–2018
<b>Toxline</b>		
Search terms	@AND+@OR+("2,2,3,3,4,4,5,5,6,6,6-undecafluorohexanoic+acid"+"2,2,3,3,4,4,5,5,6,6,6-undecafluoro-hexanoic+acid"+"hexanoic+acid+2,2,3,3,4,4,5,5,6,6,6-undecafluoro"+"hexanoic+acid+undecafluoro"+"perfluorohexanoic+acid"+"perfluoro-1-pentanecarboxylic+acid"+"perfluorocaproic+acid"+"perfluorohexanoate"+"perfluorohexanoic acid"+"undecafluoro-1-hexanoic+acid"+"undecafluorocaproic+acid"+"undecafluorohexanoic+acid"+"pfxa"+@TERM+@rn+(307+24+4)+@RANGE+yr+2017+2018+@NOT+@org+"nih+reporter"	No date limit–7/21/2017
Literature update search terms	@AND+@OR+("PFHxA_ion"+"Perfluorohexanoate"+"Hexanoyl+fluoride,+2,2,3,3,4,4,5,5,6,6,6-undecafluoro"+"Hexanoyl+fluoride,+undecafluoro"+"Perfluorohexanoyl+fluoride"+"Undecafluorohexanoyl+fluoride"+"Perfluoro(2-methyl-3-oxahexanoyl)+fluoride"+"Propanoyl+fluoride,+2,3,3,3-tetrafluoro-2-(1,1,2,2,3,3,3-heptafluoropropoxy)-"+"Propanoyl+fluoride,+2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-"+"Propionyl+fluoride,+tetrafluoro-2-(heptafluoropropoxy)-"+"2,2,3,3,4,4,5,5,6,6,6-Undecafluorohexanoic+acid"+"EINECS+206+196+6"+"NSC+5213"+"Perfluoro-1-pentanecarboxylic+acid"+"Perfluoro-n-hexanoic+acid"+"UNII-ZP34Q2220R"+"Undecafluorocaproic+acid"+"Undecafluorohexanoic+acid"+"Ammonium+Perfluorohexanoate"+"PFHxA-H3N"+"PFHxA-Na"+"Sodium+Perfluorohexanoate")+@NOT+@org+"nih+reporter"                     @AND+@OR+(@TERM+@rn+92612+52+7+@TERM+@rn+355+38+4+@TERM+@rn+2062+98+8)+@NOT+@org+"nih+reporter"	2017–2018
<b>TSCATS</b>		
Search terms	307-24-4[rn] AND tscats[org]	No date limit–7/20/2017
Literature update search terms	@AND+@OR+(@TERM+@rn+92612+52+7+@TERM+@rn+355+38+4+@TERM+@rn+2062+98+8)+@org+tscat	

**Table B-5. Perfluorohexanesulfonate (PFHxS) database search strategy**

Search	Search strategy	Dates of search
<b>PubMed</b>		
Search terms	108427-53-8[rn] OR 355-46-4[rn] OR "1,1,2,2,3,3,4,4,5,5,6,6,6-Tridecafluorohexane-1-sulfonic acid"[tw] OR "1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-1-Hexanesulfonic acid"[tw] OR "1-Hexanesulfonic acid, 1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-"[tw] OR "1-Hexanesulfonic acid, tridecafluoro-"[tw] OR "1-Perfluorohexanesulfonic acid"[tw] OR "Perfluoro-1-hexanesulfonate"[tw] OR "Perfluorohexane sulfonic acid"[tw] OR "Perfluorohexane-1-sulphonic acid"[tw] OR "Perfluorohexanesulfonate"[tw] OR "Perfluorohexanesulfonic acid"[tw] OR "Perfluorohexylsulfonate"[tw] OR "Tridecafluorohexanesulfonic acid"[tw] OR "tridecafluoro-1-Hexanesulfonic acid"[tw] OR "PFHxS"[tw]	No date limit-7/21/2017
Literature update search terms	((108427-53-8[EC/RN Number] OR 423-50-7[EC/RN Number] OR "1-Hexanesulfonic acid, 1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-, ion(1-)"[tw]) OR "PFHxS ion(1-)"[tw]) OR "PFHxS_ion"[tw]) OR "Perfluorohexanesulfonate"[tw]) OR "Tridecafluorohexane-1-sulfonate"[tw]) OR "perfluorohexyl sulfonate"[tw]) OR "1,1,2,2,3,3,4,4,5,5,6,6,6-Tridecafluoro-1-hexanesulfonyl fluoride"[tw]) OR "1-Hexanesulfonyl fluoride, 1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-"[tw]) OR "1,1,2,2,3,3,4,4,5,5,6,6,6-Tridecafluoro-1-hexanesulfonic acid"[tw]) OR "EC 206-587-1"[tw]) OR "EINECS 206-587-1"[tw]) OR "PFHS"[tw]) OR "Perfluorhexan-1-sulfonsaure"[tw]) OR "Perfluorohexane sulfonic acid (PFHxS)"[tw]) OR "Perfluorohexane-1-sulphonic acid"[tw]) OR "acide perfluorohexane-1-sulfonique"[tw]) OR "acido perfluorohexano-1-sulfonico"[tw]) OR "perfluorohexane-1-sulphonic acid"[tw]) OR "perfluorohexanesulfonic acid"[tw]) OR "Ammonium Perfluorohexanesulfonate"[tw]) OR "Ammonium perfluorohexanesulfonate"[tw]) OR "PFHxS-H3N"[tw]) OR "PFHxS-K"[tw]) OR "Potassium Perfluorohexanesulfonate"[tw]) OR "Potassium perfluorohexanesulfonate"[tw]) OR "Lithium Perfluorohexanesulfonate"[tw]) OR "Lithium perfluorohexanesulfonate"[tw]) OR "PFHxS-Li"[tw]))	8/1/2017-2/14/2018
<b>Web of Science</b>		
Search terms	TS="1,1,2,2,3,3,4,4,5,5,6,6,6-Tridecafluorohexane-1-sulfonic acid" OR TS="1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-1-Hexanesulfonic acid" OR TS="1-Hexanesulfonic acid, 1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-" OR TS="1-Hexanesulfonic acid, tridecafluoro-" OR TS="1-Perfluorohexanesulfonic acid" OR TS="Perfluoro-1-hexanesulfonate" OR TS="Perfluorohexane sulfonic acid" OR TS="Perfluorohexane-1-sulphonic acid" OR TS="Perfluorohexanesulfonate" OR TS="Perfluorohexanesulfonic acid" OR TS="Perfluorohexylsulfonate" OR TS="Tridecafluorohexanesulfonic acid" OR TS="tridecafluoro-1-Hexanesulfonic acid" OR TS="PFHxS"	No date limit-7/24/2017

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Search	Search strategy	Dates of search
Literature update search terms	TS="1-Hexanesulfonic acid, 1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-, ion(1-)" OR TS="PFHxS ion(1-)" OR TS="PFHxS_ion" OR TS="Perfluorohexanesulfonate" OR TS="Tridecafluorohexane-1-sulfonate" OR TS="perfluorohexyl sulfonate" OR TS="1,1,2,2,3,3,4,4,5,5,6,6,6-Tridecafluoro-1-hexanesulfonyl fluoride" OR TS="1-Hexanesulfonyl fluoride, 1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-" OR TS="1,1,2,2,3,3,4,4,5,5,6,6,6-Tridecafluoro-1-hexanesulfonic acid" OR TS="EC 206-587-1" OR TS="EINECS 206-587-1" OR TS="PFHS" OR TS="Perfluorhexan-1-sulfonsaure" OR TS="Perfluorohexane sulfonic acid (PFHxS)" OR TS="Perfluorohexane-1-sulphonic acid" OR TS="acide perfluorohexane-1-sulfonique" OR TS="acido perfluorohexano-1-sulfonico" OR TS="perfluorohexane-1-sulphonic acid" OR TS="perfluorohexanesulfonic acid" OR TS="Ammonium Perfluorohexanesulfonate" OR TS="Ammonium perfluorohexanesulfonate" OR TS="PFHxS-H3N" OR TS="PFHxS-K" OR TS="Potassium Perfluorohexanesulfonate" OR TS="Potassium perfluorohexanesulfonate" OR TS="Lithium Perfluorohexanesulfonate" OR TS="Lithium perfluorohexanesulfonate" OR TS="PFHxS-Li"	2017–2018
<b>Toxline</b>		
Search terms	( 108427-53-8[rn] OR 355-46-4[rn] OR "1,1,2,2,3,3,4,4,5,5,6,6,6-Tridecafluorohexane-1-sulfonic acid" OR "1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-1-Hexanesulfonic acid" OR "1-Hexanesulfonic acid, 1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-" OR "1-Hexanesulfonic acid, tridecafluoro-" OR "1-Perfluorohexanesulfonic acid" OR "Perfluoro-1-hexanesulfonate" OR "Perfluorohexane sulfonic acid" OR "Perfluorohexane-1-sulphonic acid" OR "Perfluorohexanesulfonate" OR "Perfluorohexanesulfonic acid" OR "Perfluorohexylsulfonate" OR "Tridecafluorohexanesulfonic acid" OR "tridecafluoro-1-Hexanesulfonic acid" OR "PFHxS" ) AND ( ANEUPL [org] OR BIOSIS [org] OR CIS [org] OR DART [org] OR EMIC [org] OR EPIDEM [org] OR HEEP [org] OR HMTc [org] OR IPA [org] OR RISKLINE [org] OR MTGABS [org] OR NIOSH [org] OR NTIS [org] OR PESTAB [org] OR PPBIB [org] ) [not] PubMed [org] [not] pubdart [org]	No date limit–7/21/2017
Literature update search terms	@AND+@OR+("1-Hexanesulfonic+acid,+1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-,+ion(1-)"+"PFHxS+ion(1-)"+"PFHxS_ion"+"Perfluorohexanesulfonate"+"Tridecafluorohexane-1-sulfonate"+"perfluorohexyl+sulfonate"+"1,1,2,2,3,3,4,4,5,5,6,6,6-Tridecafluoro-1-hexanesulfonyl+fluoride"+"1-Hexanesulfonyl+fluoride,+1,1,2,2,3,3,4,4,5,5,6,6,6-tridecafluoro-"+"1,1,2,2,3,3,4,4,5,5,6,6,6-Tridecafluoro-1-hexanesulfonic+acid"+"EC+206-587-1"+"EINECS+206-587-1"+"PFHS"+"Perfluorhexan-1-sulfonsaure"+"Perfluorohexane+sulfonic+acid+(PFHxS)"+"Perfluorohexane-1-sulphonic+acid"+"acide+perfluorohexane-1-sulfonique"+"acido+perfluorohexano-1-sulfonico"+"perfluorohexane-1-sulphonic+acid"+"perfluorohexanesulfonic+acid"+"Ammonium+Perfluorohexanesulfonate"+"Ammonium+perfluorohexanesulfonate"+"PFHxS-H3N"+"PFHxS-K"+"Potassium+Perfluorohexanesulfonate"+"Potassium+perfluorohexanesulfonate"+"Lithium+Perfluorohexanesulfonate"+"Lithium+perfluorohexanesulfonate"+"PFHxS-Li")+@NOT+@org+"nih+reporter"  @OR+(@TERM+@rn+108427+53+8+@TERM+@rn+423+50+7)+@NOT+@org+"nih+reporter"	2017–2018

***Systematic Review Protocol for the PFBA, PFHxA, PFHxS, PFNA, and PFDA IRIS Assessments***

Search	Search strategy	Dates of search
<b>TSCATS</b>		
Search terms	@OR+(@term+@rn+355-46-4+@term+@rn+108427-53-8)+@AND+@org+tscats	No date limit-7/21/2017
Literature update search terms	@OR+(@TERM+@rn+"108427+53+8"+@TERM+@rn+"423+50+7")+@org+tscats	2017-2018

**Table B-6. Title/abstract-level screening criteria for the initial literature searches**

	<b>Inclusion criteria</b>	<b>Exclusion criteria</b>
Populations	<ul style="list-style-type: none"> <li>• Humans</li> <li>• Standard mammalian animal models, including rat, mouse, rabbit, guinea pig, hamster, monkey, dog</li> <li>• Alternative animal models in standard laboratory conditions (e.g., <i>Xenopus</i>, zebrafish, minipig)</li> <li>• Human or animal cells, tissues, or organs (not whole animals); bacteria, nonmammalian eukaryotes; other nonmammalian laboratory species</li> </ul>	<ul style="list-style-type: none"> <li>• Ecological species</li> </ul>
Exposures	<ul style="list-style-type: none"> <li>• Exposure is to a PFAS compound</li> <li>• Exposure via oral, inhalation, dermal, intraperitoneal, or intravenous injection routes</li> <li>• Exposure is measured in air, dust, drinking water, diet, gavage, injection or via a biomarker of exposure (PFAS levels in whole blood, serum, plasma, or breastmilk)</li> </ul>	<ul style="list-style-type: none"> <li>• Study population is not exposed to a PFAS compound</li> <li>• Exposure is to a mixture only</li> </ul>
Outcomes	<ul style="list-style-type: none"> <li>• Studies that include a measure of one or more health effect endpoints, including but not limited to, effects on reproduction, development, developmental neurotoxicity, liver, thyroid, immune system, nervous system, genotoxicity, and cancer</li> <li>• In vivo and/or in vitro studies related to toxicity mechanisms, physiological effects/adverse outcomes, and studies useful for elucidating toxic modes of action (MOAs)</li> <li>• Qualitative or quantitative description of absorption, distribution, metabolism, excretion, toxicokinetic and/or toxicodynamic models (e.g., PBPK, PBTk, PBTk/TD)</li> <li>• Studies addressing risks to infants, children, pregnant women, occupational workers, the elderly, and any other susceptible or differentially exposed populations</li> </ul>	

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	<b>Inclusion criteria</b>	<b>Exclusion criteria</b>
Other	<ul style="list-style-type: none"> <li>• Structure and physiochemical properties</li> <li>• Reviews and regulatory documents</li> </ul>	<ul style="list-style-type: none"> <li>• Not on topic, including:</li> <li>• Abstract only, inadequately reported abstract, or no abstract and not considered further because study was not potentially relevant</li> <li>• Bioremediation, biodegradation, or chemical or physical treatment of PFAS compounds, including evaluation of wastewater treatment technologies and methods for remediation or contaminated water and soil</li> <li>• Ecosystem effects</li> <li>• Studies of environmental fate and transport of PFAS compounds in environmental media</li> <li>• Analytical methods for detecting/measuring PFAS compounds in environmental media and use in sample preparations and assays</li> <li>• Studies describing the manufacture and use of PFAS compounds</li> <li>• Not chemical specific (studies that do not involve testing of PFAS compounds)</li> <li>• Studies that describe measures of exposure to PFAS compounds without data on associated health effects</li> </ul>

MOA = mode of action; PBPK = physiologically based pharmacokinetic; PBTK = physiologically based toxicokinetic; PFAS = per- and polyfluoroalkyl substance; TD = toxicodynamic.

**Table B-7. Example DistillerSR form questions to be used for title/abstract and full text-level screening for literature search updates from 2019**

	Used in title/abstract and full-text screening					Used in full text only	
Question	Source of study if not identified from database search?	Does the article meet PECO criteria?	If meets PECO, what type of evidence?	If supplemental, what type of information?	Which PFAS did the study report?	If meets PECO, which health outcome(s) apply?	If meets PECO and endocrine outcome, which endocrine tags apply?
<b>Answer options (can select multiple options)</b>	<ul style="list-style-type: none"> <li>Source other than HERO database search</li> </ul>	<ul style="list-style-type: none"> <li>Yes</li> <li>No</li> <li>Unclear</li> <li>Tag as potentially relevant supplemental information</li> </ul>	<ul style="list-style-type: none"> <li>Human</li> <li>Animal (mammalian models)</li> <li>In vitro or in silico genotoxicity</li> <li>PBPK or PK model</li> </ul>	<ul style="list-style-type: none"> <li>In vivo mechanistic or MOA studies, including non-PECO routes of exposure (e.g., injection) and populations (e.g., nonmammalian)</li> <li>In vitro or in silico studies (nongenotoxicity)</li> <li>ADME/toxicokinetic (excluding models)</li> <li>Exposure assessment or characterization (no health outcome)</li> <li>PFAS Mixture Study (no individual PFAS comparisons)</li> <li>Human case reports or case series</li> <li>Ecotoxicity studies</li> </ul>	<ul style="list-style-type: none"> <li>PFBA</li> <li>PFHxA</li> <li>PFHxS</li> <li>PFNA</li> <li>PFDA</li> </ul>	<ul style="list-style-type: none"> <li>General toxicity, including body weight, mortality, and survival</li> <li>Cancer</li> <li>Cardiovascular, including serum lipids</li> <li>Endocrine (hormone)</li> <li>Gastrointestinal</li> <li>Genotoxicity</li> <li>Growth (early life) and development</li> <li>Hematological, including nonimmune/hepatic/renal clinical chemistry measures</li> <li>Hepatic, including liver measures and serum markers (e.g., ALT; AST)</li> <li>Immune/inflammation</li> <li>Musculoskeletal</li> </ul>	<ul style="list-style-type: none"> <li>Adrenal</li> <li>Sex hormones (e.g., androgen; estrogen; progesterone)</li> <li>Neuroendocrine</li> <li>Pituitary</li> <li>Steroidogenesis</li> <li>Thyroid</li> </ul>

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			<ul style="list-style-type: none"> <li>• Environmental fate or occurrence (including food)</li> <li>• Manufacture, engineering, use, treatment, remediation, or laboratory methods</li> <li>• Other assessments or records with no original data (e.g., reviews, editorials, commentaries)</li> </ul>	<ul style="list-style-type: none"> <li>• Nervous system, including behavior and sensory function</li> <li>• Nutrition and metabolic</li> <li>• Ocular</li> <li>• PBPK or PK model</li> <li>• Renal, including urinary measures (e.g., protein)</li> <li>• Reproductive</li> <li>• Respiratory</li> <li>• Skin and connective tissue effects</li> </ul>	
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ADME = absorption, distribution, metabolism, and excretion; ALT = alanine aminotransferase; AST = aspartate aminotransferase; HERO = Health and Environmental Research Online; MOA = mode of action; PBPK = physiologically based pharmacokinetic; PECO = populations, exposures, comparators, and outcomes; PFAS = per- and polyfluoroalkyl substances; PFBA = perfluorobutanoic acid; PFDA = perfluorodecanoic acid; PFHxA = perfluorohexanoic acid; PFHxS = Perfluorohexanesulfonic acid; PFNA = Perfluorononanoic acid; PK = pharmacokinetic.