

St. Louis River estuary as possible *Dreissena veliger* source to western Lake Superior

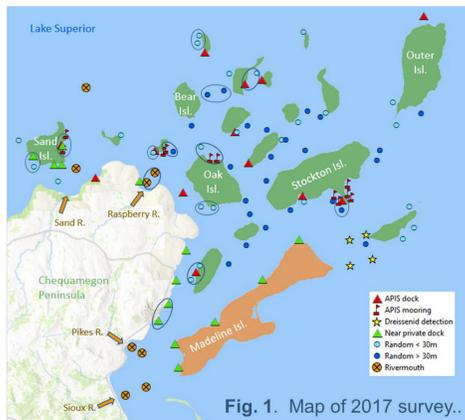
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Background (2017 survey overview & findings)

In 2017 EPA conducted early detection case study in western Lake Superior around the Apostle Islands addressing concerns over adult *Dreissena* finds on shipwrecks & native mussels by Nat'l Park Service

- What is dreissena prevalence and distribution?
- Baseline and potential impacts on zooplankton and benthos community?



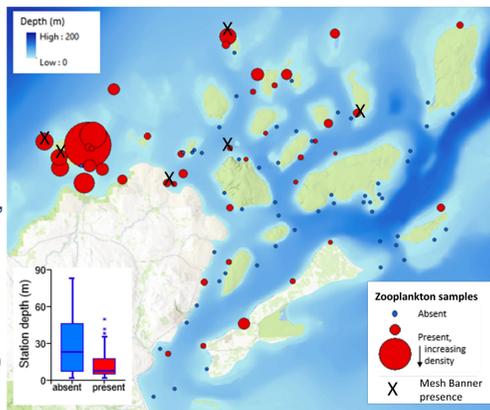
50 random depth-stratified sites & 50 sites targeting likely *Dreissena* location sampled using multiple gear types

Sample types, # samples collected

PONAR grab, benthos, N = 89
Hester Dendy (HD), benthos, N = 52
Rock bag, benthos, N = 11
Zooplankton tow, 63µ, water column, N = 98
Mesh banner (on HD lines), water column, N=37
eDNA 500mL water, 2m above benthos, N = 99
eDNA 500mL water, 1m below surface, N = 33
Video – benthos, N = 93
Ca2+, nutrients, chlorophyll A & B, N=85
Temp, conductivity, turbidity, pH, DO, N = 99

- Intensive survey found no settled juvenile or adult *Dreissena*.
- *Dreissena veligers* were present in 43 zooplankton samples
- *D. polymorph* DNA was detected using qPCR in 6 mesh banners

Fig. 2 Map of *Dreissena* detections. Veligers found at very low densities (max 39/m³, most <5/m³). DNA detections were also at very low concentrations.



Veliger spatial patterns

- Consistently present in west APIS, sporadic to east
- Densities highest around Sand Island and along north island fringe
- Finds primarily along NW (not SE) side of big Islands

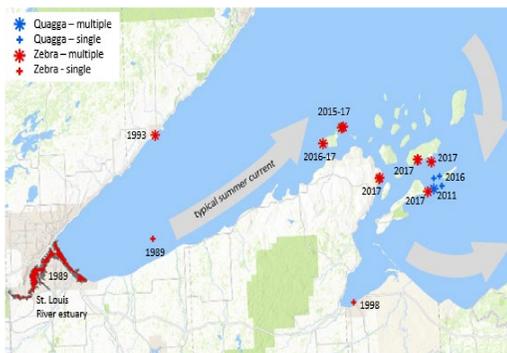


Fig. 3. Map of settled *Dreissena* known from Lake Superior (with year) and typical circulation patterns.

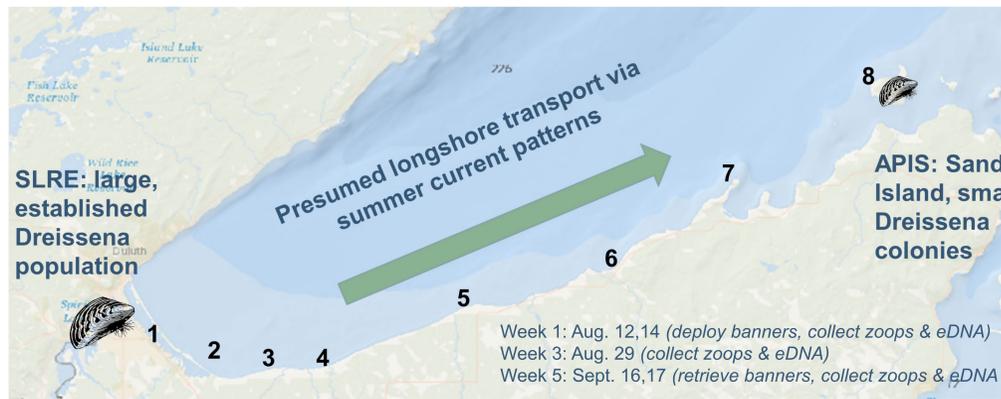
Transport of veligers from the St. Louis River to Apostle Islands???

- SLRE has largest, most established *Dreissena* population in Lake Superior
- Typical summer current patterns consistent with transport from SLRE to APIS

Survey Overview, Design, & Methods (2019 W. Lake Superior south shore survey)

Objectives: Follow up 2017 survey by sampling along SLRE to APIS gradient to evaluate if the SLRE is the veliger source to APIS *Dreissena*.

- We expect to see a pattern of veliger density declining with distance from the SLRE to APIS (and possibly size increasing over this distance)
- 2017 methods modified to increase probability of detecting low abundance *Dreissena* veligers and eDNA in Lake Superior



Study Design

8 sites (10 – 15m depth)
3 visits (2 weeks apart)
3 sampling gears

Gear	2(2'x4') mesh banners (N = 16)	Zooplankton tow 64µm mesh (N = 24)	eDNA (N = 48)
Sampling modifications	2, larger, more rigid banners. Sampled 1m below surface & 2m above benthos	Composited 4 tows per sample. Saved decanted EtOH for DNA analysis	Increased water volume sampled to 1L. Sampled 3m below surface & 2m above benthos
Processing & analysis	qPCR targeting <i>Dreissenids</i>	Zoops: full enumeration Zoop EtOH: qPCR targeting <i>Dreissenids</i>	Filtered samples & qPCR targeting <i>Dreissenids</i>

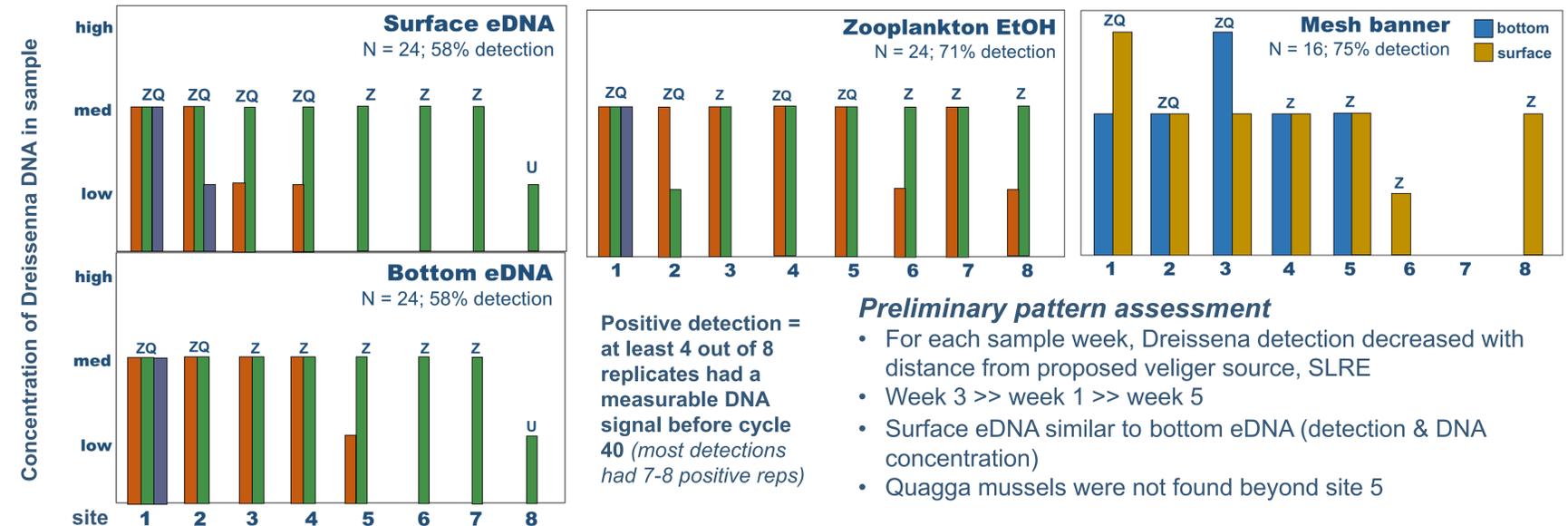
Findings (preliminary qPCR results)

- qPCR (quantitative polymerase chain reaction): Is target DNA is present?
- Fewer PCR cycles to detect target = higher DNA concentration in sample

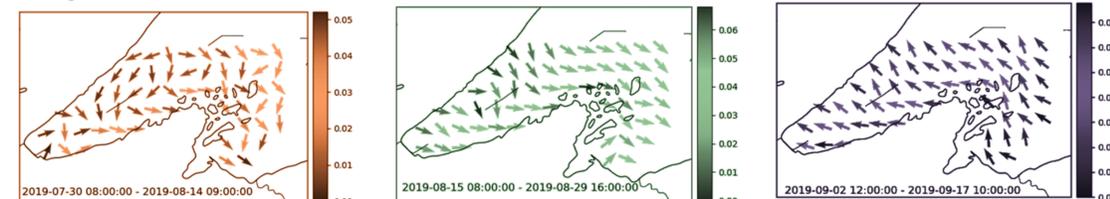
45 total cycles

DNA concentration: High=DNA detected < 29 PCR cycles; Med=DNA detected 29-38 cycles; Low=DNA detected 38-40 cycles

Legend: Week 1 (orange), Week 3 (green), Week 5 (purple). ZQ = zebra & quagga detected, Z = only zebra detected, Q = only quagga detected, U = undefined, *Dreissenid* DNA detected; species DNA may be below detectable limit



Surface current patterns leading up to each sampling week correspond to where *Dreissena* veligers & eDNA were detected (time averaged velocity; 2wks prior to sampling)



Still to come...

- More endpoints: zooplankton enumeration (veliger densities, zooplankton community gradient?)
- In depth analysis
 - qPCR data, calculate DNA copy numbers
 - longshore current patterns leading up to sampling trips