

Applications of a Chemotype-Enrichment Approach to the ToxCast data landscape and beyond: *Inverting the SAR paradigm*



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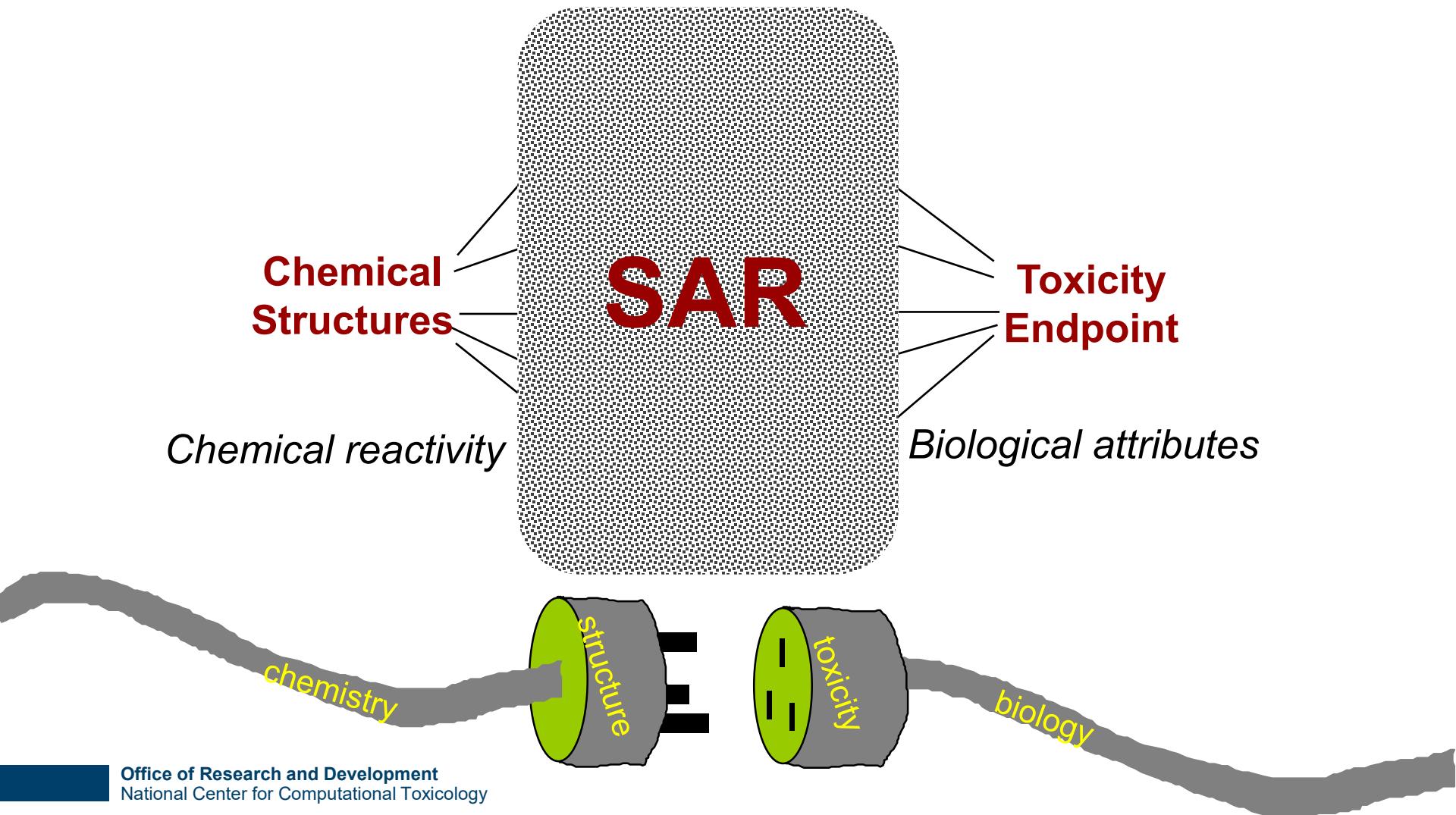
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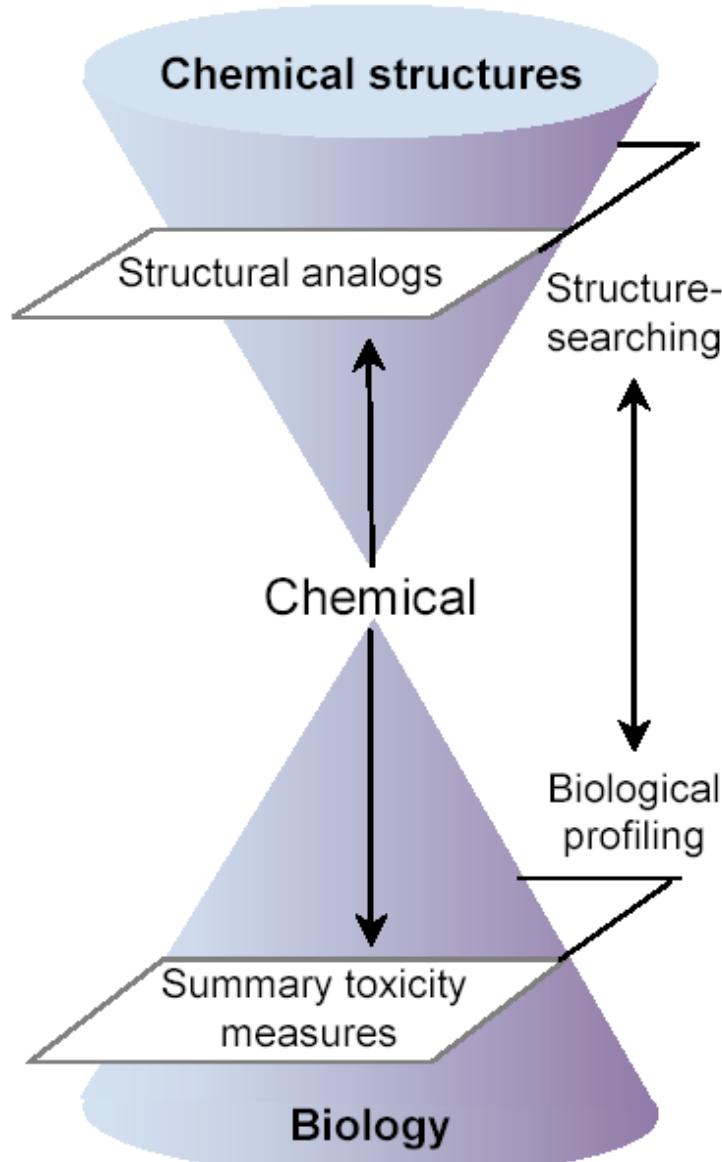
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Global vs. Local SAR models

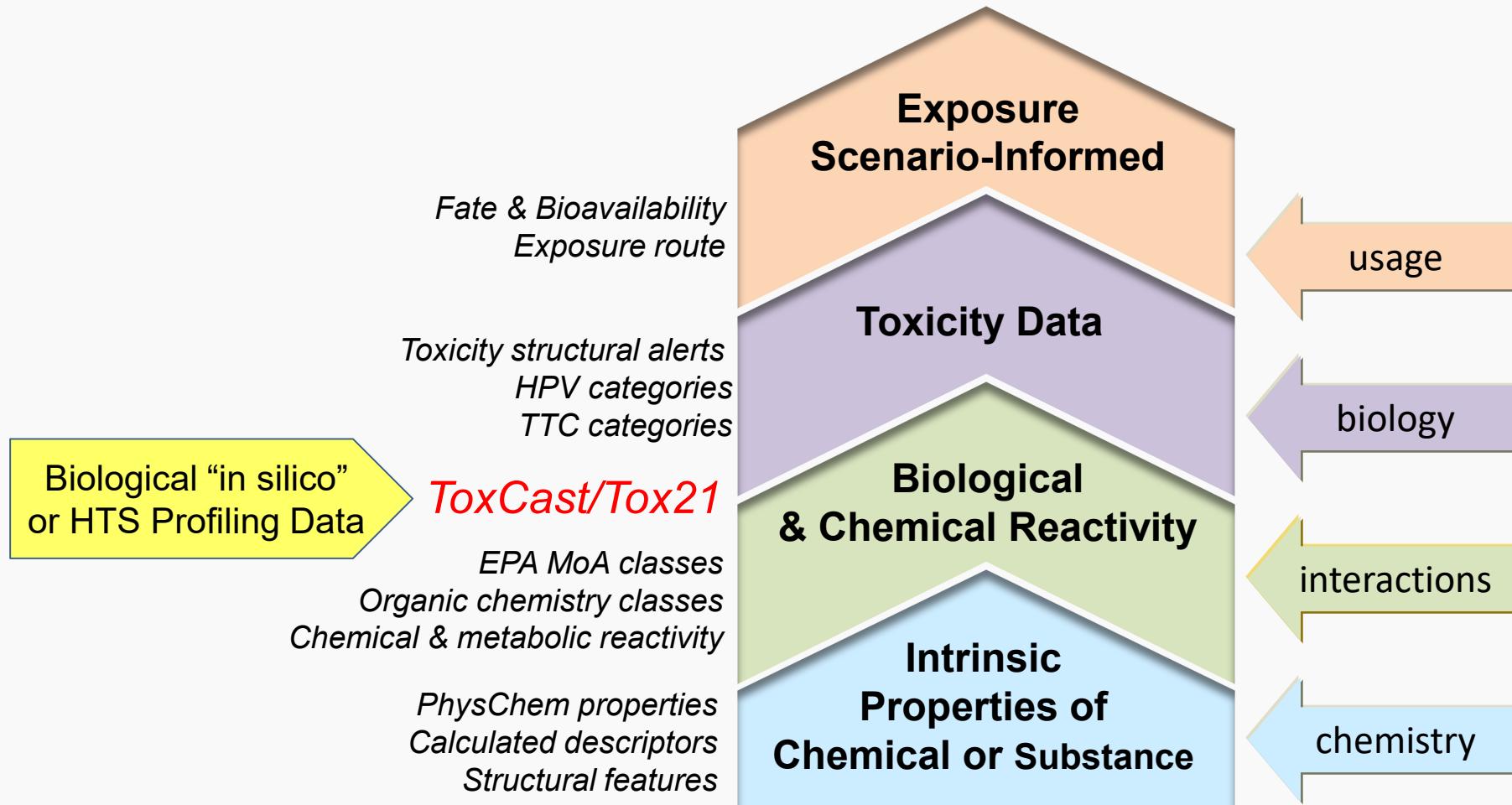


“Lensing”

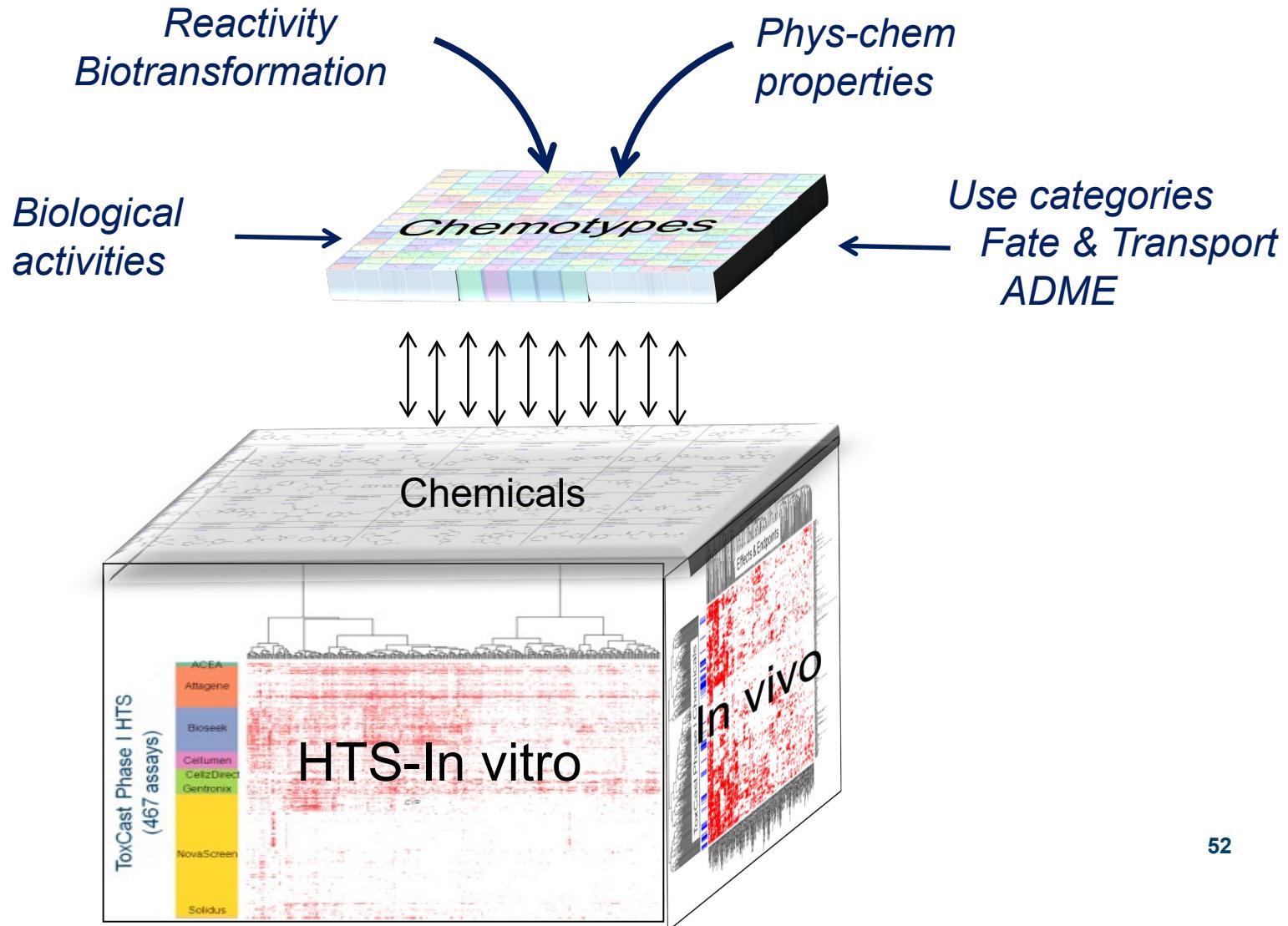
For any given chemical-assay dataset, what is the optimal level of chemical and biological specificity to discover useful SAR associations?



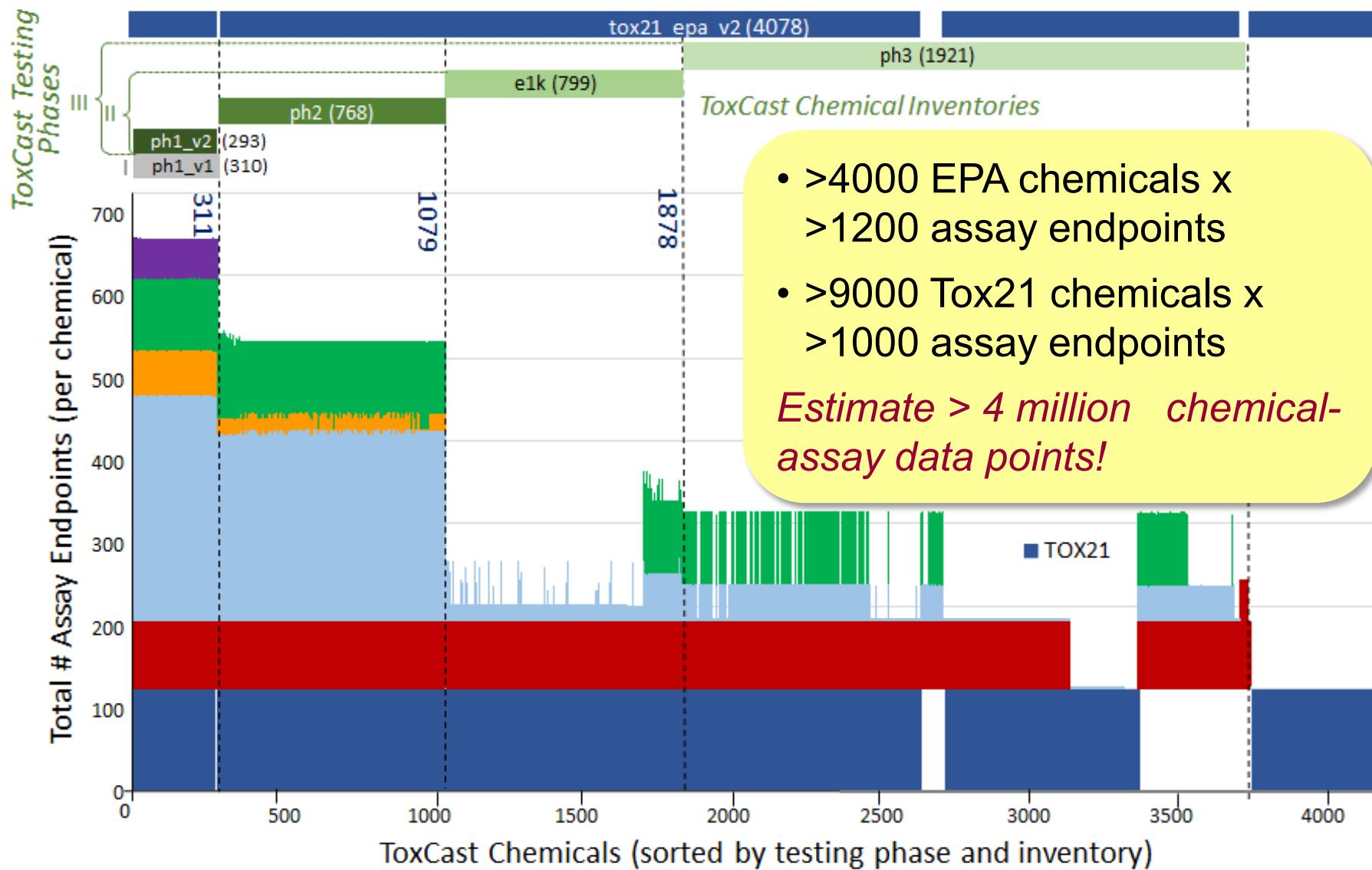
Chemical/Substance representations for hazard & risk evaluation



Building a public chemotype “knowledge- base”

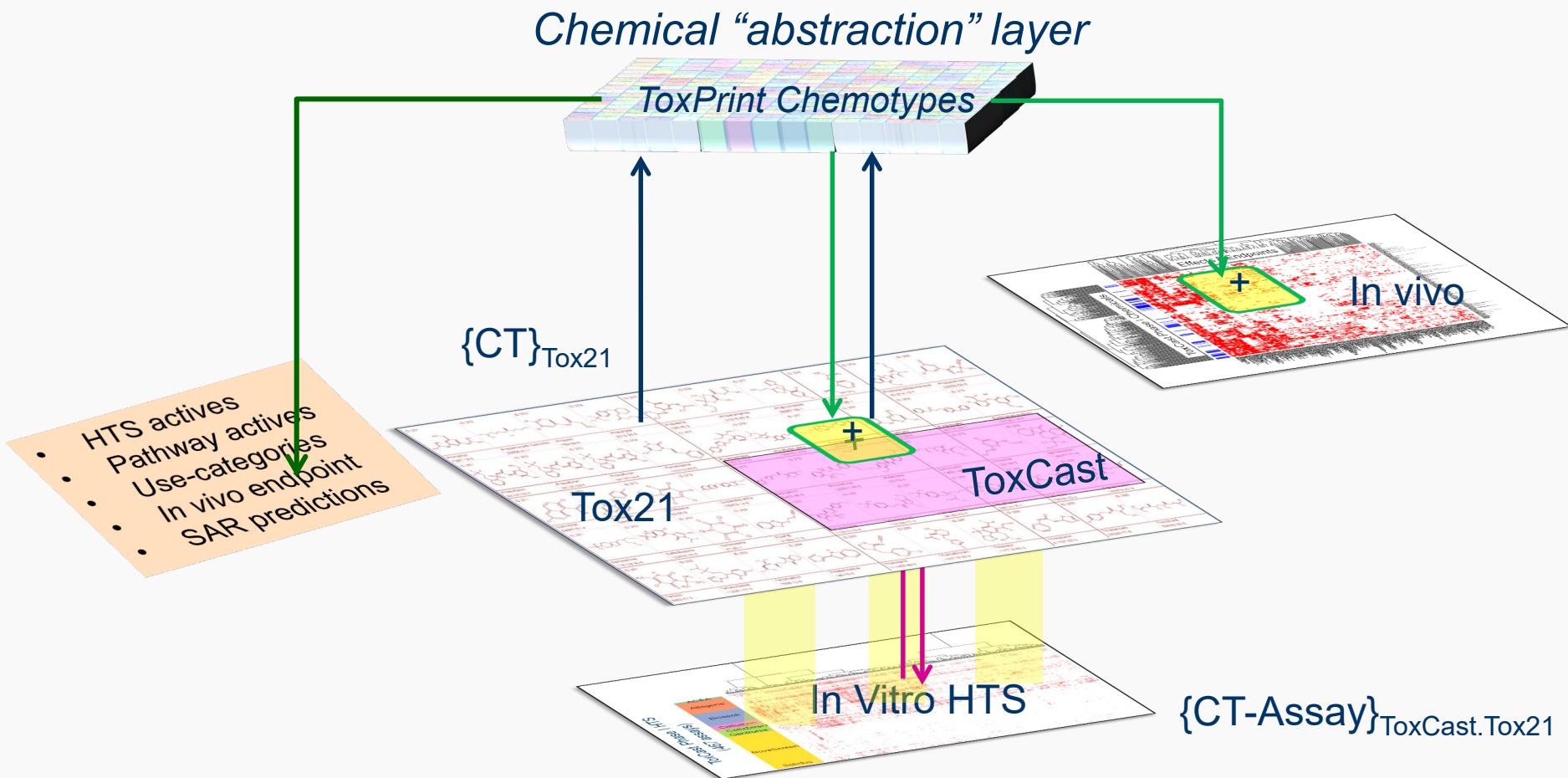


ToxCast HTS data



Chemotype-Activity Enrichments

- ✓ Create {CT-Activity} enrichment profiles for any “active” subspace of a test set
- ✓ Focus studies in local CT domains & compare enrichments across data domains

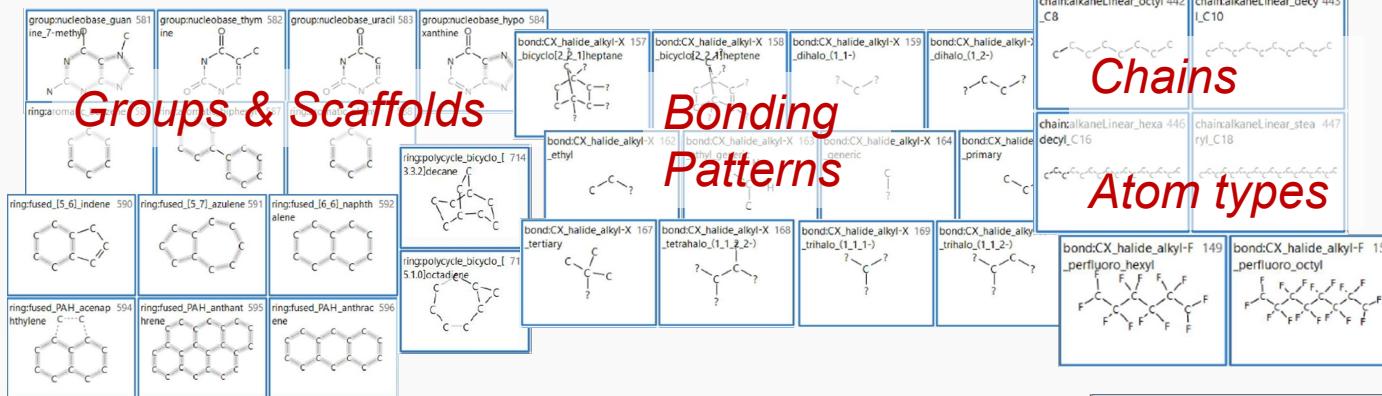


ToxPrints: A Public Set of Chemotypes

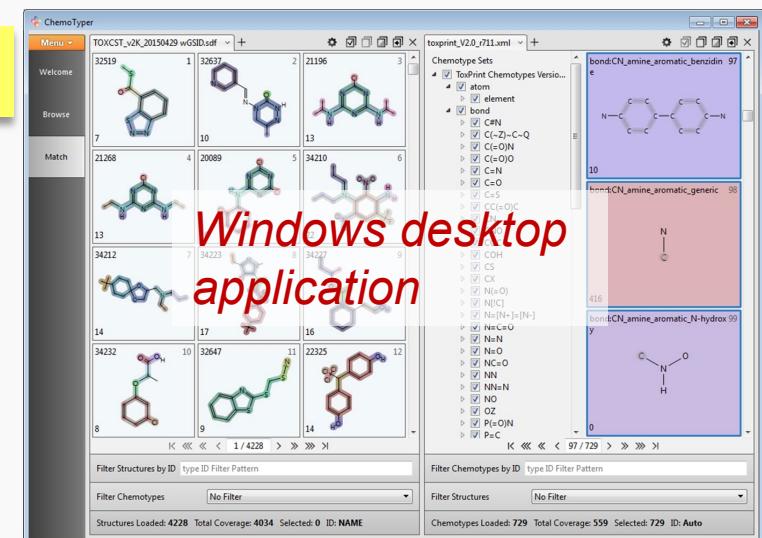
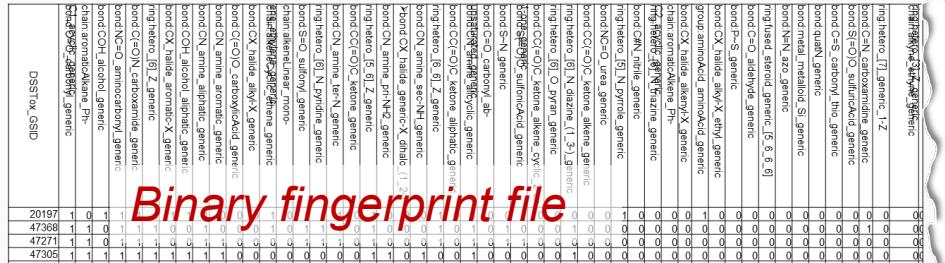
ToxPrints: <http://www.toxprint.org>

→ Clear, reproducible means for defining & visualizing local chemistry

729 features important to EPA & FDA's "chemical exposure" landscape and safety assessment workflow



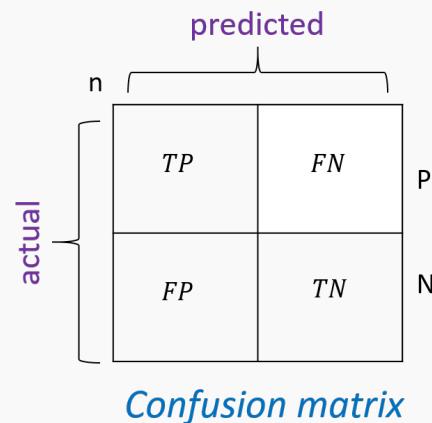
Chemotyper: <http://www.chemotyper.org>



Computing CT-Assay “Enrichments”

Set statistical thresholds & filters for significance to support data-mining objectives:

TP_ID	ToxPrint_CT_name ²	CT _{Tot}	T _{pos}	F _{pos}	F _{neg}	T _{neg}	Odd's Ratio	Fischer's pval
423	chain:alkaneBranch_t-butyl_C4	41	24	17	294	693	3.3	2.0E-04
479	chain:aromaticAlkane_Ph-C1-Ph	39	27	12	291	698	5.4	6.5E-07
303	bond:X[any_!C]_halide_inorganic	28	17	11	301	699	3.6	9.0E-04

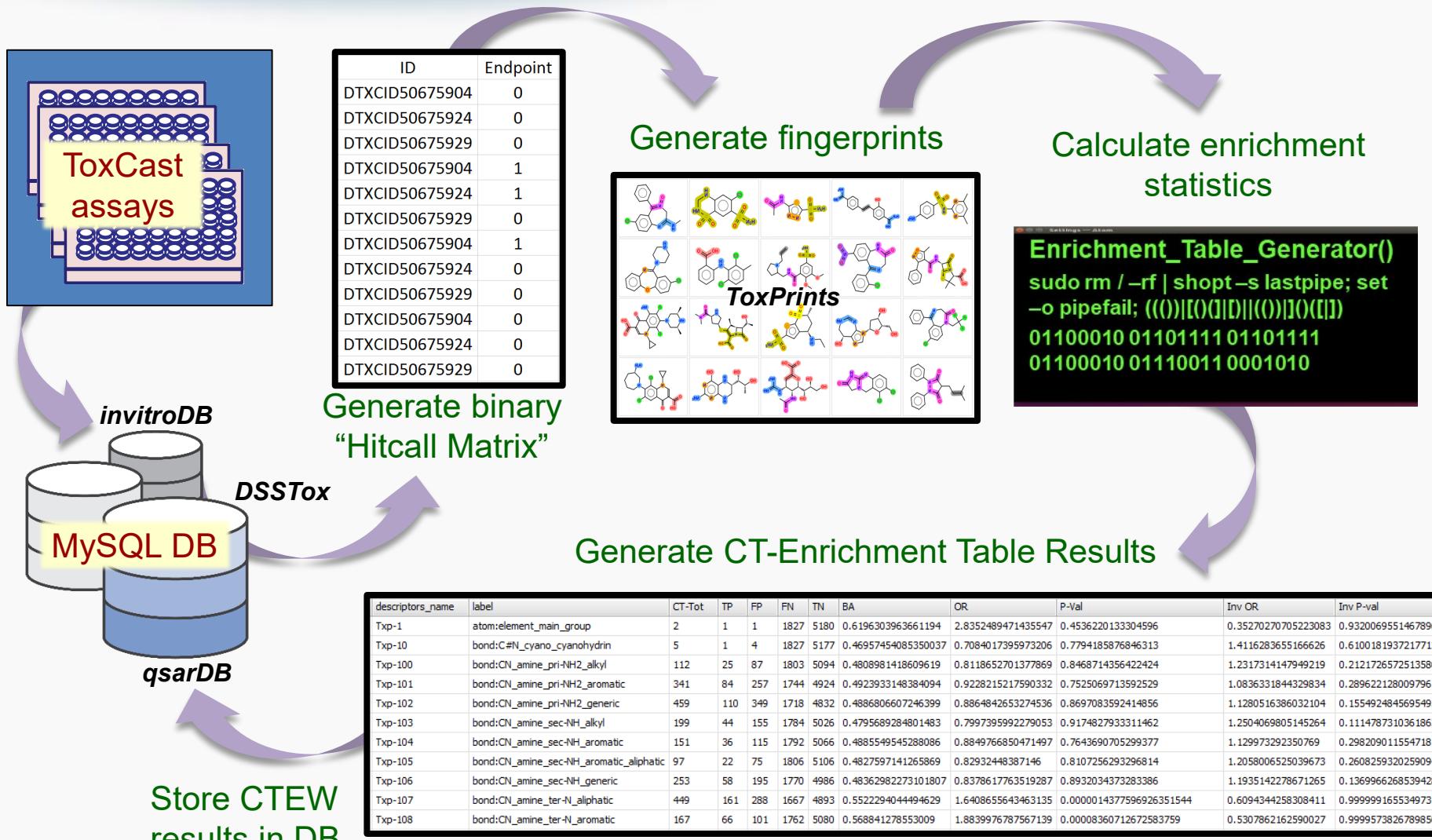


$$\text{TestSet} = \# \text{ Pos} + \# \text{ Neg} = \# \text{ chems tested}$$

CT_{Tot} = total # chems in TestSet w/ CT (Pos or Neg)
 $\text{TP} (\text{T}_{\text{pos}})$ = # Pos in TestSet w/ CT
 $\text{FP} (\text{F}_{\text{pos}})$ = # Neg in TestSet w/ CT
 $\text{FN} (\text{F}_{\text{neg}})$ = # Pos in TestSet w/o CT
 $\text{TN} (\text{T}_{\text{neg}})$ = # Neg in TestSet w/o CT

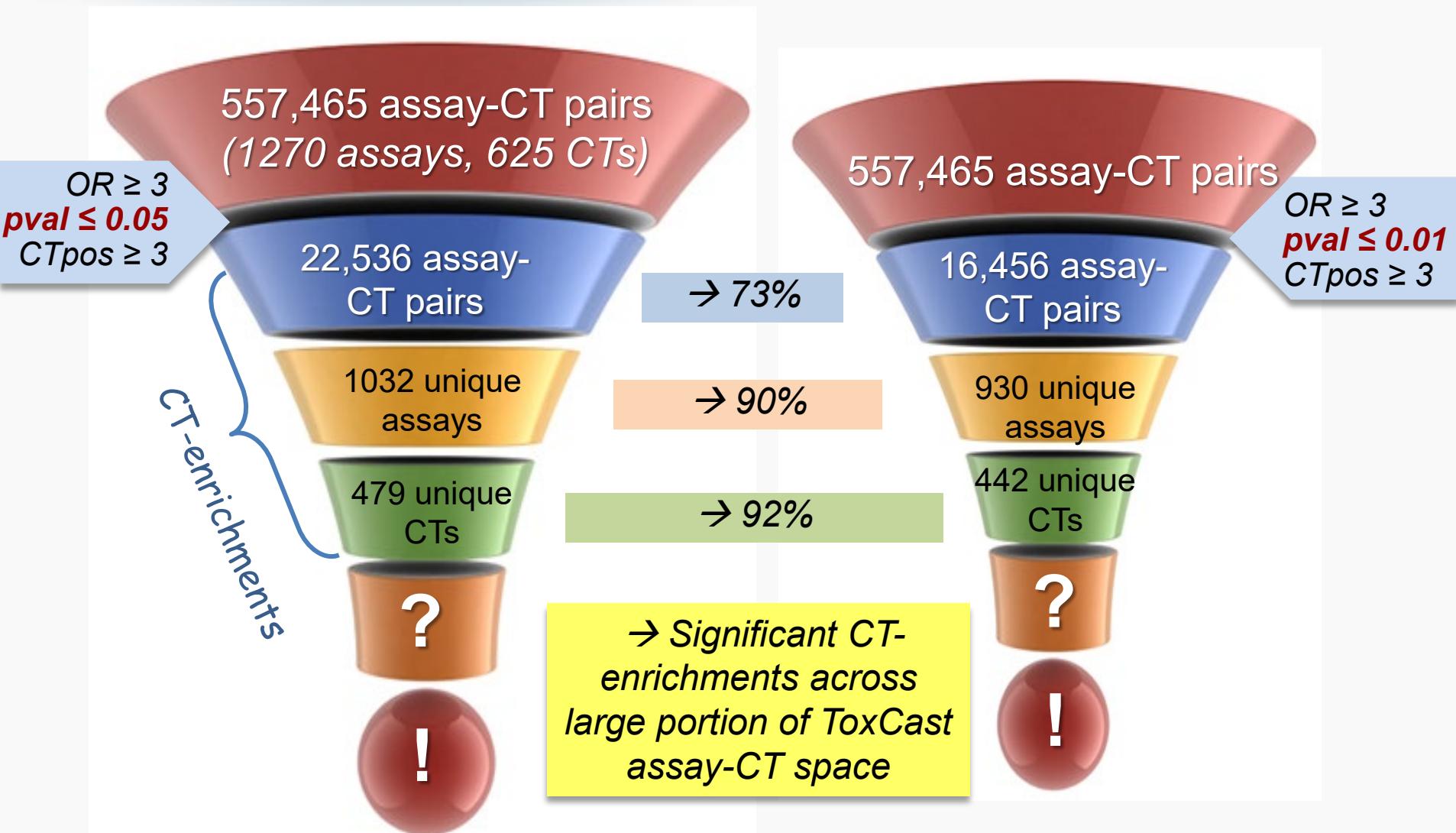
- Odds Ratio ≥ 3 , *conveys simple fractional enrichment*
- Fischer's exact p value ≤ 0.05 , *takes into account size of dataset*
- $\text{T}_{\text{pos}} (\text{TP}) \geq 3$, *require at least 3 chemicals with CT in Positives*

Automated Chemotype-Enrichment Workflow (CTEW)



Store CTEW results in DB

CTEW applied to all ToxCast assays in *invitroDB* (Dec-2017)

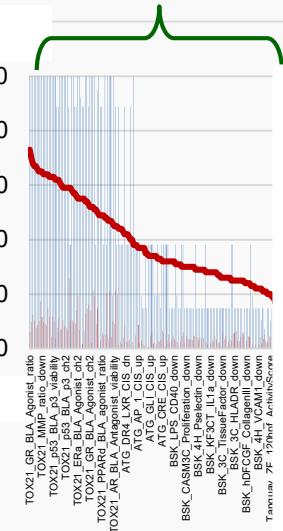


Assays x Enriched CT count

Enriched CT-counts across 1032 assays

“Global” QSAR models*
 for 167 ToxCast Assays
 Avg % Median BA = 0.67

*Random Forest models based on ToxPrint CT descriptors, validated using independent Test Set & Y-randomization, with Training (100A,100I) & Test (25A,25I) Set minimums (J. Fitzpatrick)



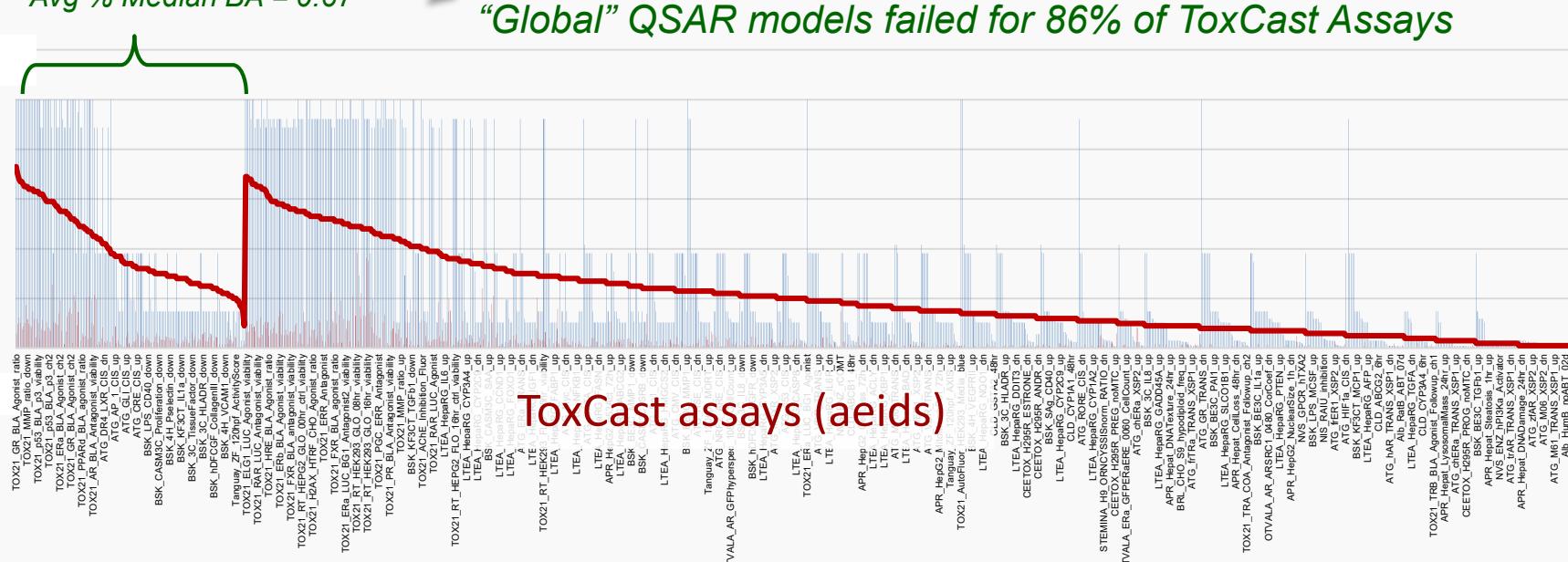
Tested Chems (scaled)

Assays x Enriched CT count

Enriched CT-counts across 1032 assays

**“Global” QSAR models*
for 167 ToxCast Assays
Avg % Median BA = 0.67**

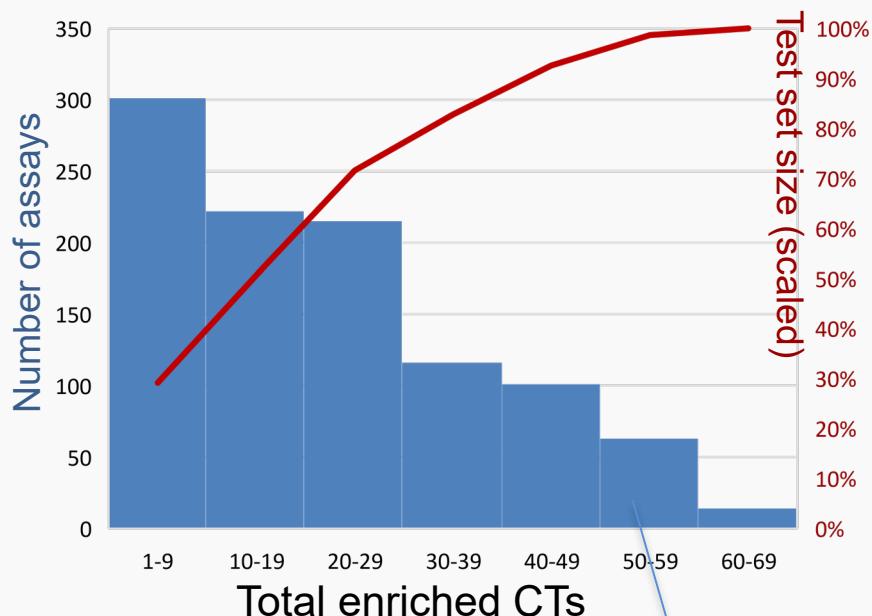
*Random Forest models based on ToxPrint CT descriptors, validated using independent Test Set & Y-randomization, with Training (100A,100I) & Test (25A,25I) Set minimums (J. Fitzpatrick)



Significant CT-enrichments across ToxCast assay space considered “unmodelable” by traditional “global” QSAR methods

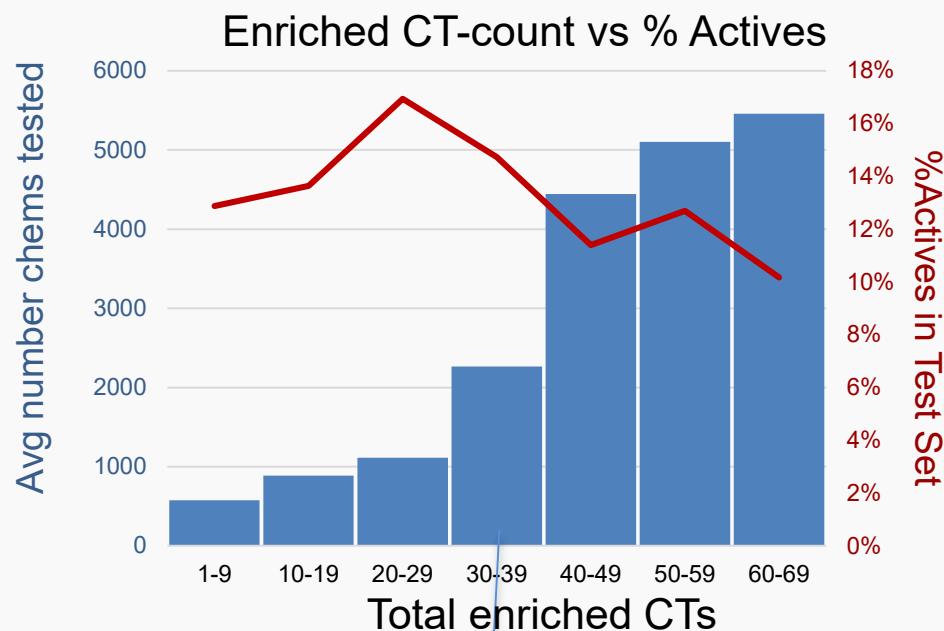
Enriched CT-count per assay trends (1032 assays)

Does # enriched CTs depend on test set size? YES!



63 assays
enriched with
50-59 CTs

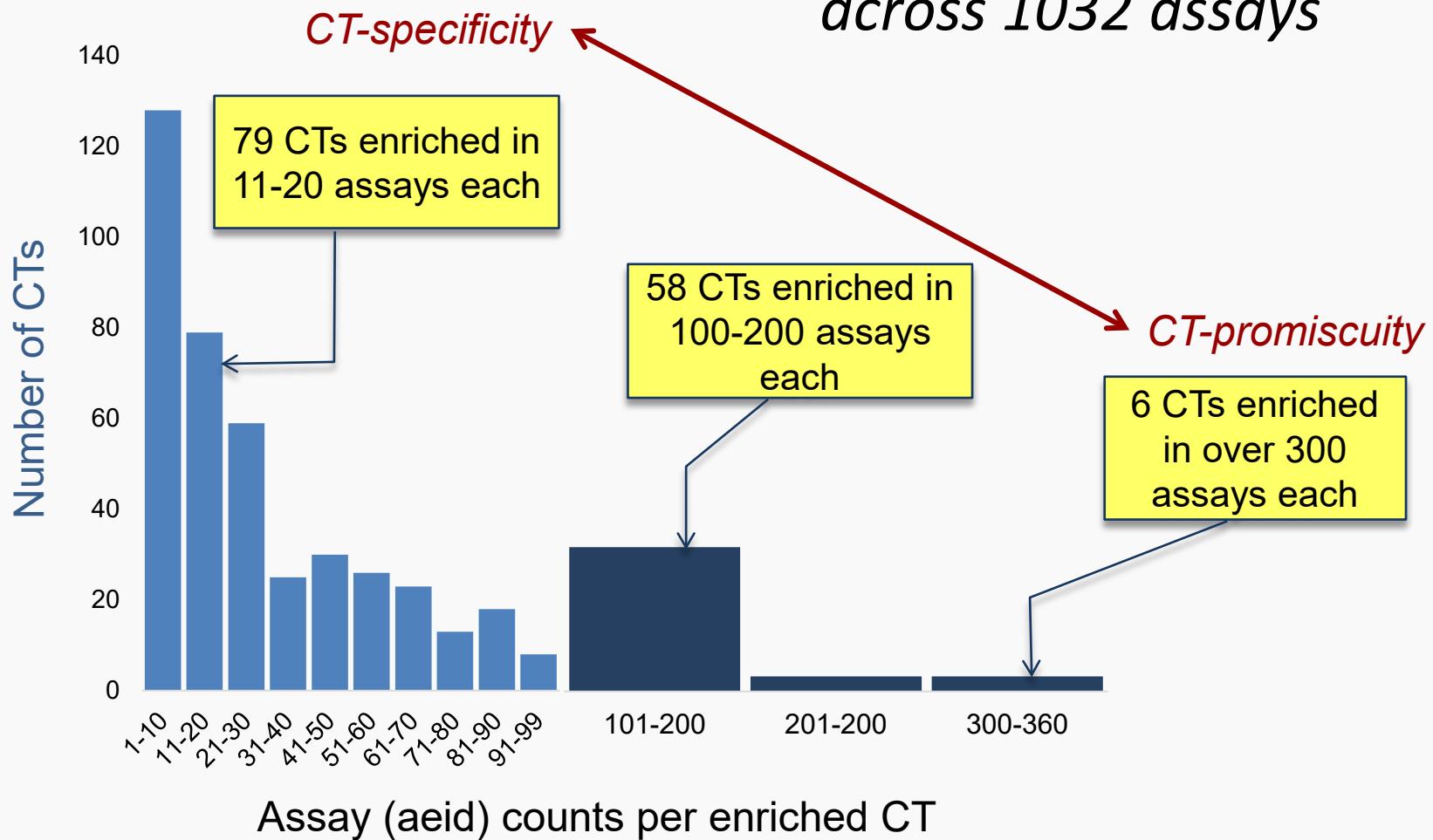
Does # enriched CTs depend on %Actives? Not so much.



30-39 CTs enriched in
assays with avg Test
Set size 2264 chems

Assay-counts per enriched CT

*Enriched CT-counts
across 1032 assays*



Tuning biology to pick up chemical “signal”

CT Enrichments of Time-series* “Assays”

aenm_870unique	TP_Count	%actives	tested.chnm
APR_Hepat_Apoptosis_1hr_up	1	3.2	310
APR_Hepat_Apoptosis_24hr_up	15	14.8	310
APR_Hepat_Apoptosis_48hr_up	6	16.8	310
APR_Hepat_DNADamage_1hr_up	8	3.9	310
APR_Hepat_DNADamage_24hr_up	7	18.4	310
APR_Hepat_DNADamage_48hr_up	14	20.0	310

* 95 Assays out of 1032 total are within time-series groups

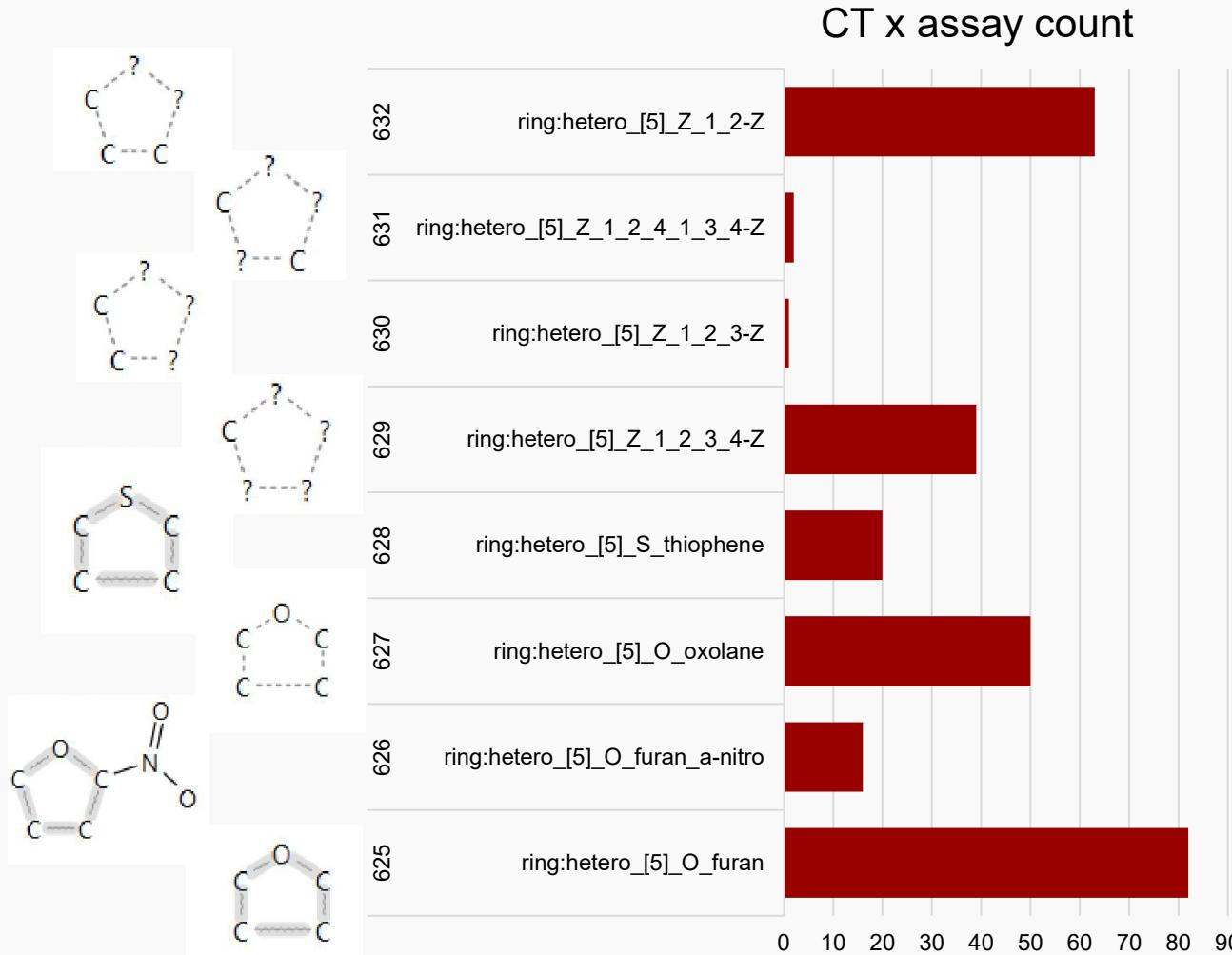


“Tuning” biology to increase CT enrichments → increases biological-chemical “signal”

- How are {CT-assay} enrichments affected by activity threshold assumptions?
- How are {CT-assay} enrichments affected by cell toxicity (“burst”) filters? (R. Lougee)

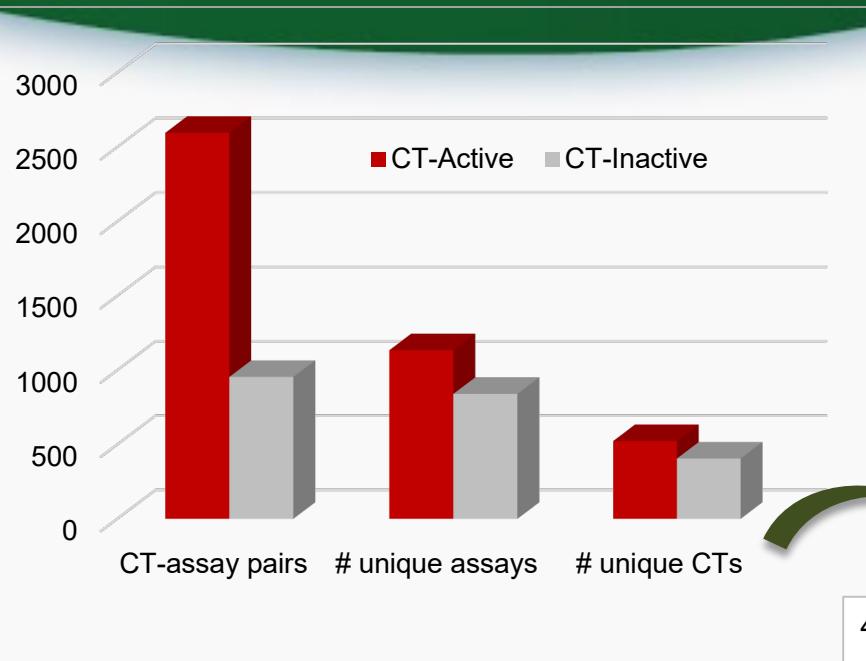
Tuning chemistry to pick up biological “signal”

Assay Enrichments of chemistry-series groups

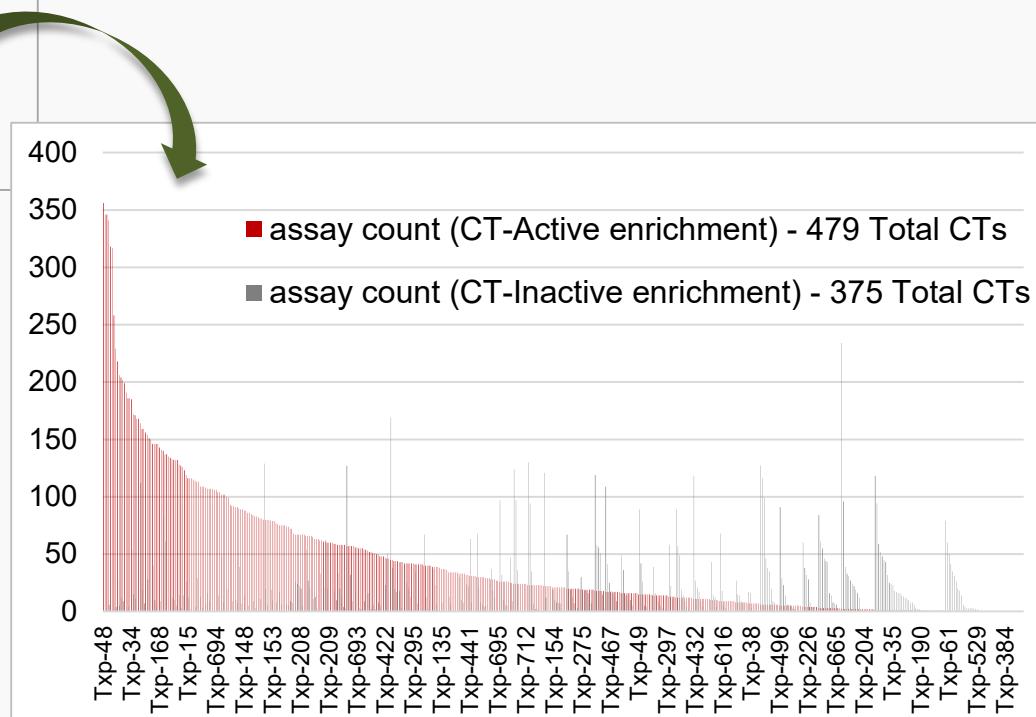


“Tuning” chemistry specificity to increase chemical-assay “signal”

What about the “Inactives”?



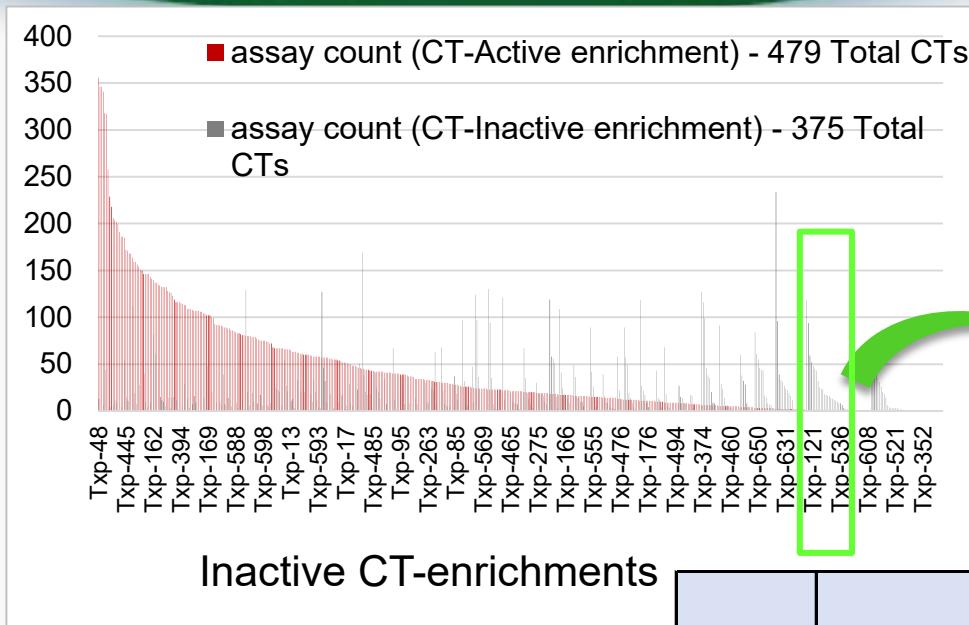
More “CT-signal” in Actives, but significant amount of signal in Inactives



Inactive CT-enrichments

- Span ToxCast assay space
- More likely to occur in assays with fewer Active CT-enrichments
- May be due to several factors:
 - True inactivity*
 - Assay artifacts*
 - QC failure*

Top 10 enriched CT-Inactives (skewed from actives)

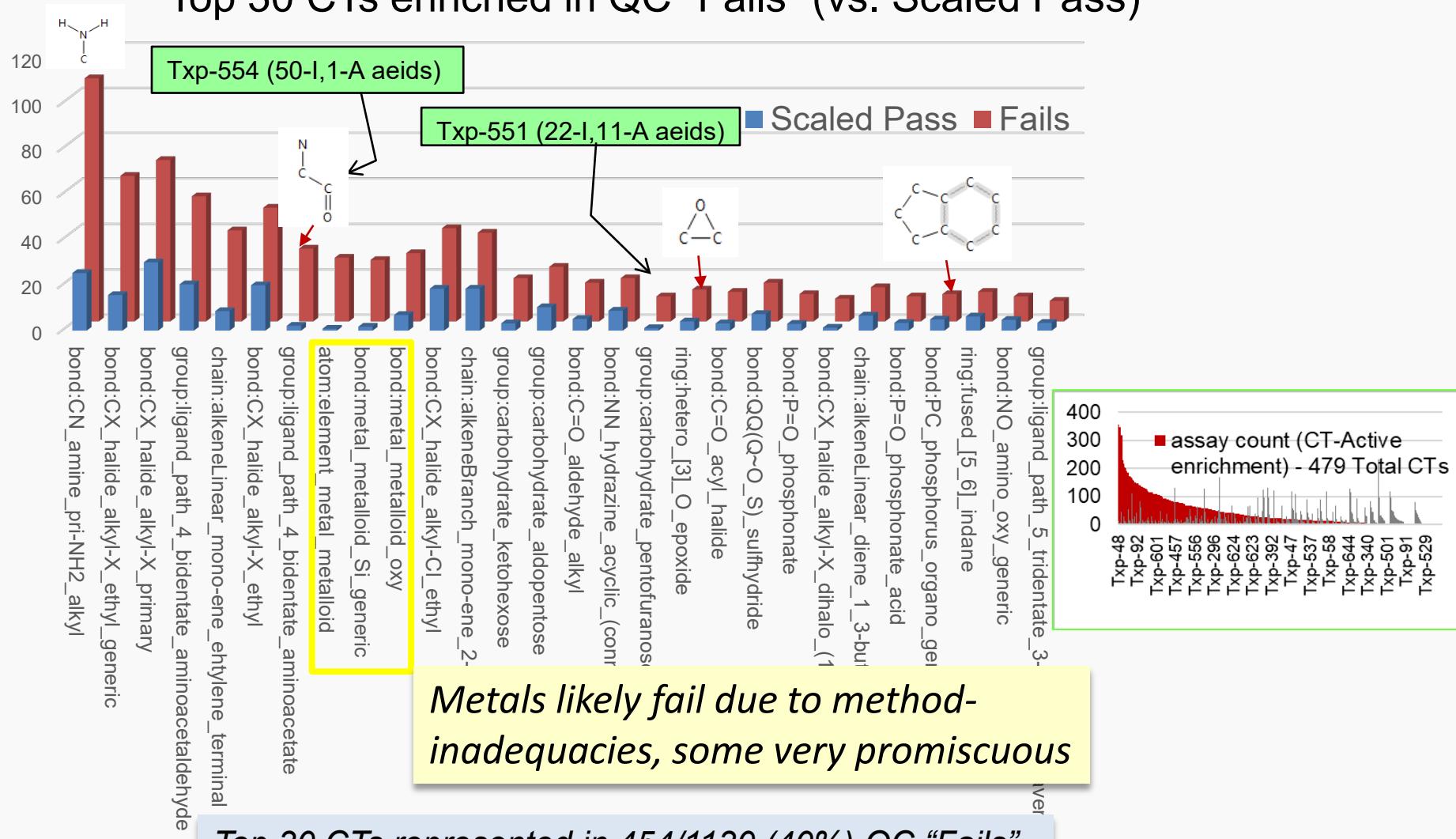


CT-Inactive >75 assays,
CT-active <10 assays

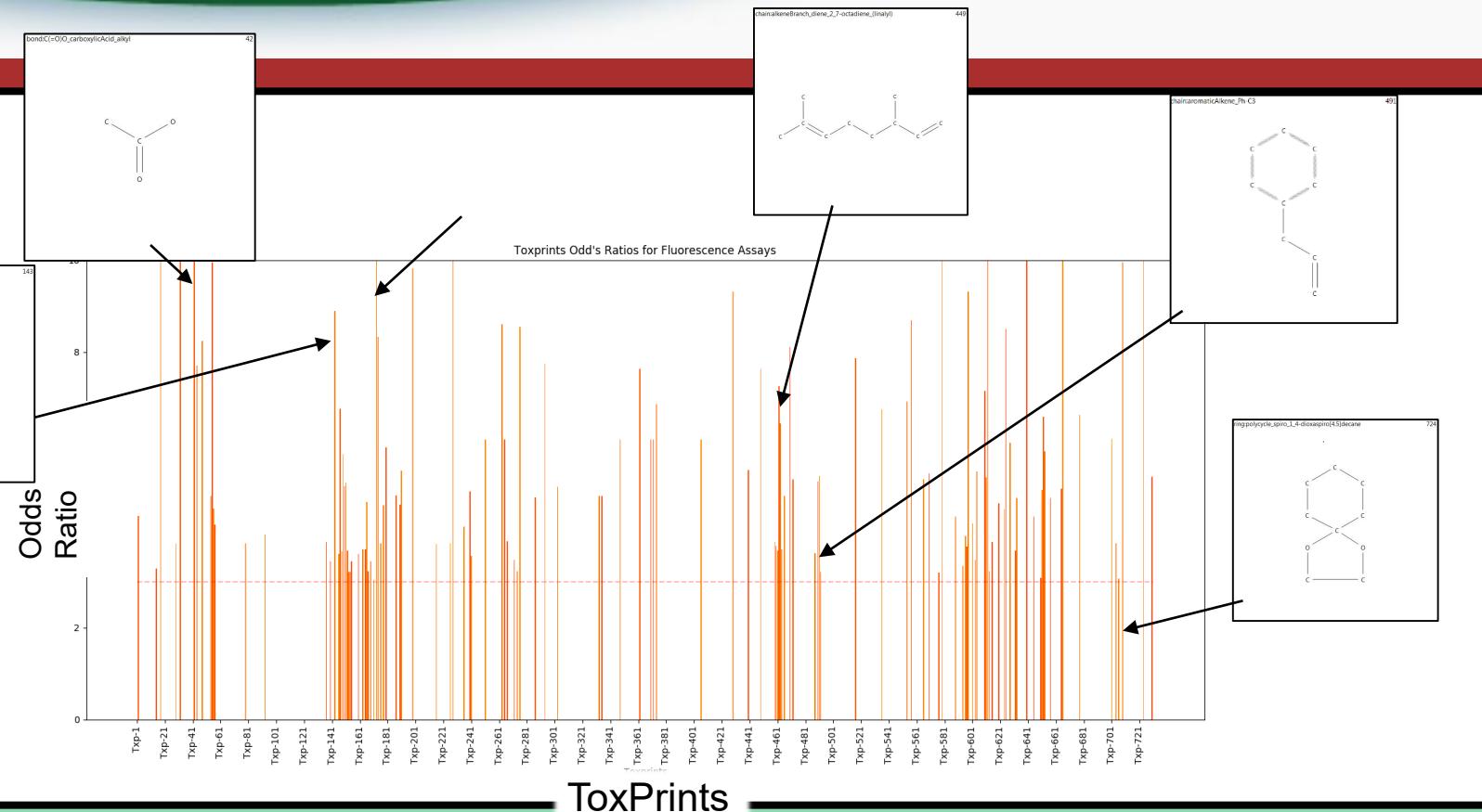
- *True inactivity?*
- *Assay artifacts?*
- **QC failure?**

ToxPrints enriched in Fails

Top 30 CTs enriched in QC “Fails” (vs. Scaled Pass)



CT Enrichments in Fluorescence Assays



CT's enriched (much more likely to be active) in “Fluorescence Assays” than in remaining assays

Multi-electrode array (MEA) neurotoxicity assay

Experimental Design

Determine Effects on Spontaneous Network Activity



1. Primary cortical neurons are cultured in 48 well MEA plates and allowed to mature for 13 Days



3. Determine firing rate in each well for 40 min prior to and after treatment with compounds

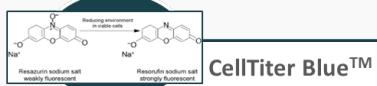
2. Plates are placed in the Axion Maestro MEA amplifier

Determine Effects on Cell Health



4. Transfer 50 µL of media from mw MEA to 96 well assay plate.

Follow Promega CytoTox® 96 Non-Radioactive Assay Kit Instructions.

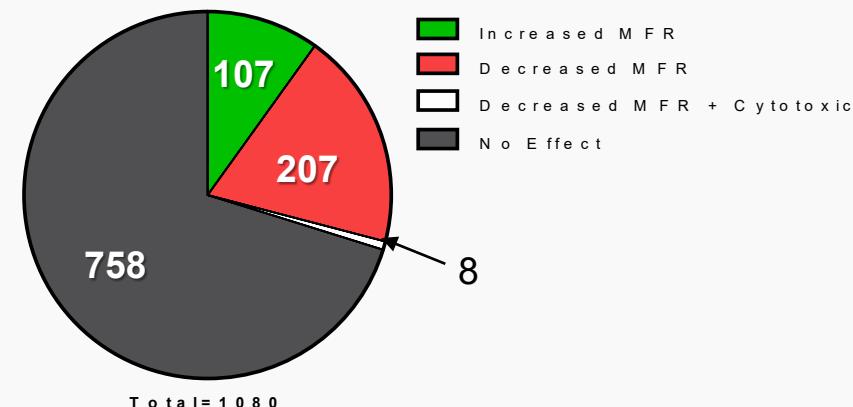


CellTiter Blue™

5. Remove remaining 450 µL from all wells of mw MEA.

- Add 200 µL of fresh media containing CellTiter Blue™ reagent (Promega; 1:6 dilution) and incubate at 37 °C for 1 h.
- Transfer 150 µL of media with reagent to an opaque 96 well assay plate and measure fluorescence at 560Ex/590Em.

- Screened 1080 ToxCast Phase II chemicals
- Activity measured as Mean Firing Rate (MFR) above or below threshold

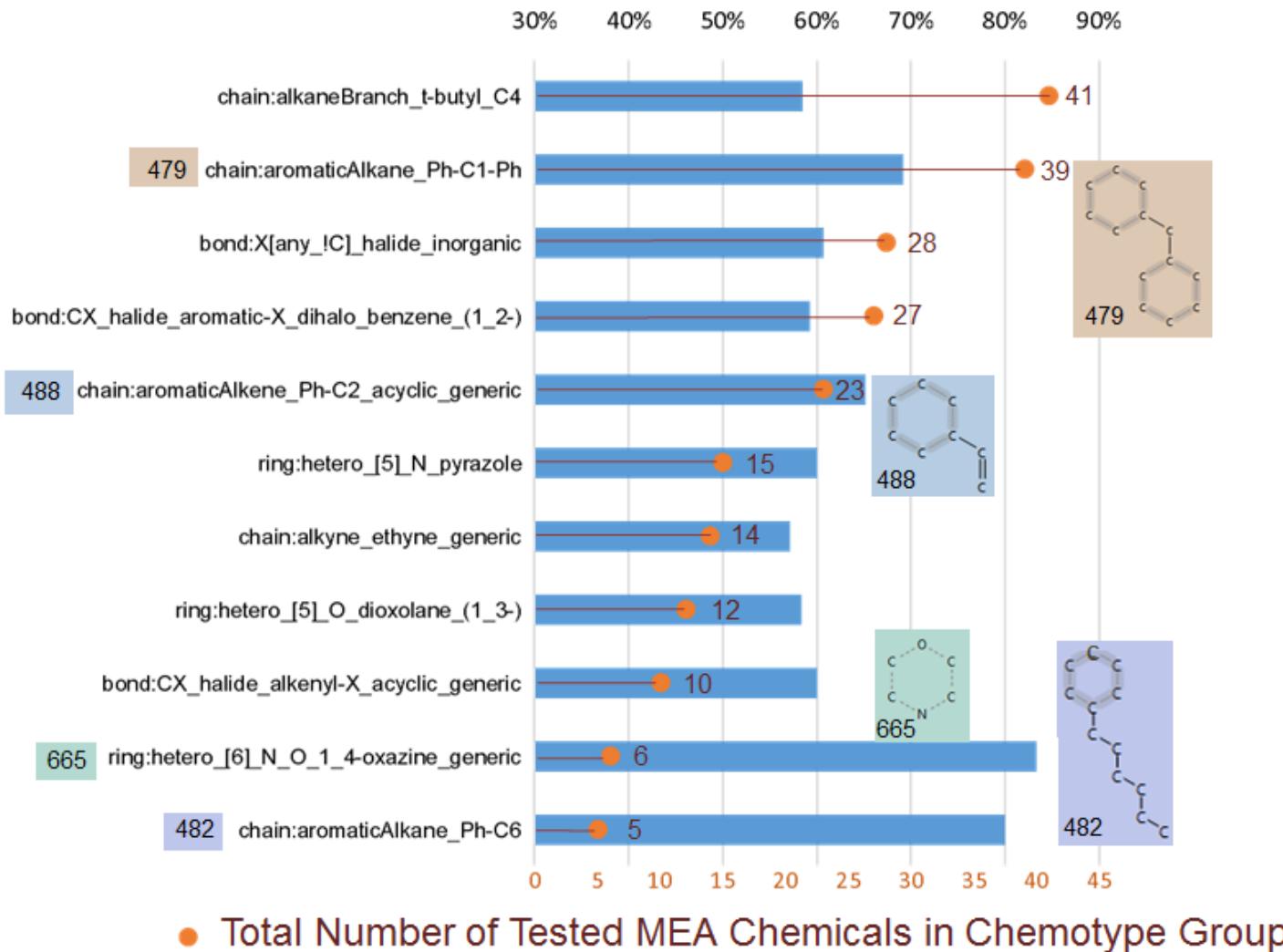


- 314 total “Actives” or Hits
- 758 “Inactives” or No-hits (8 cytotoxic)
- Total Tested = 1080 (30% hit rate)

MEA CT-Enrichment Analysis:

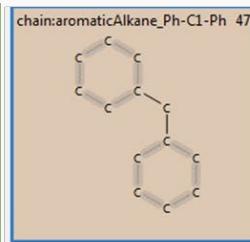
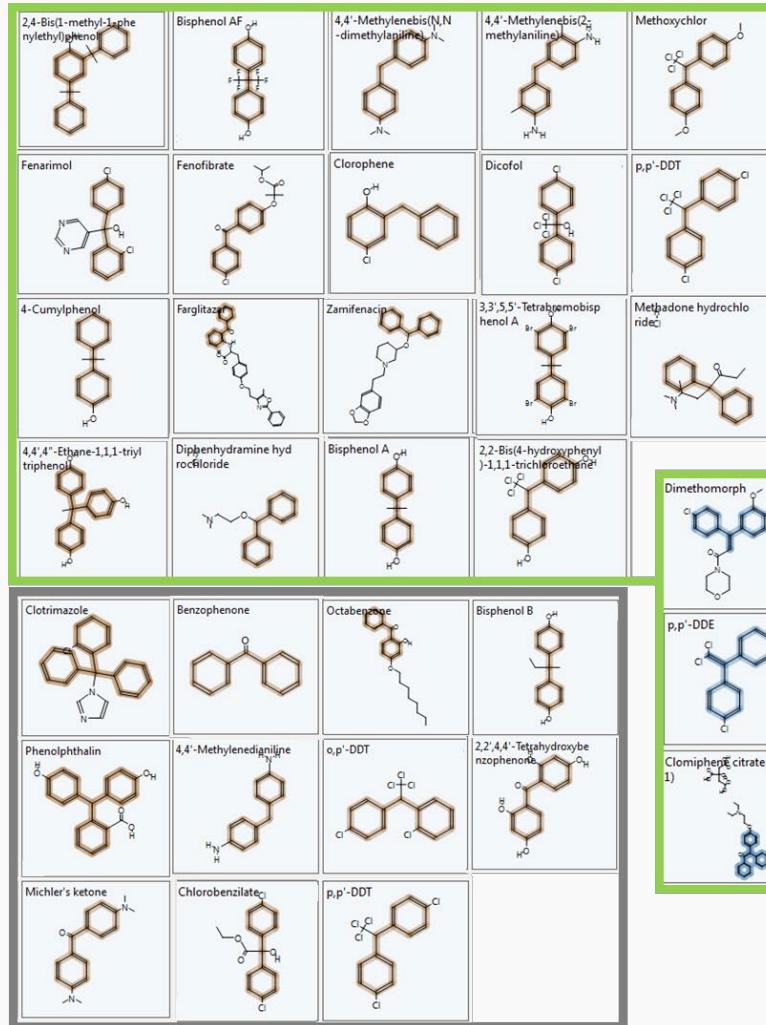
Overall coverage

■ % MEA Hits in Enriched Chemotype Group (Total MEA Hits=30%)



MEA CT-Enrichment Analysis:

Explore SAR within local CT domains

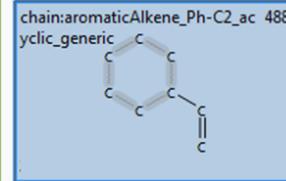
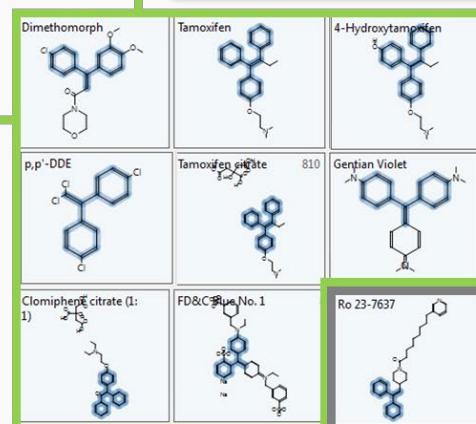


OR=5.4 (27/39 Hits)

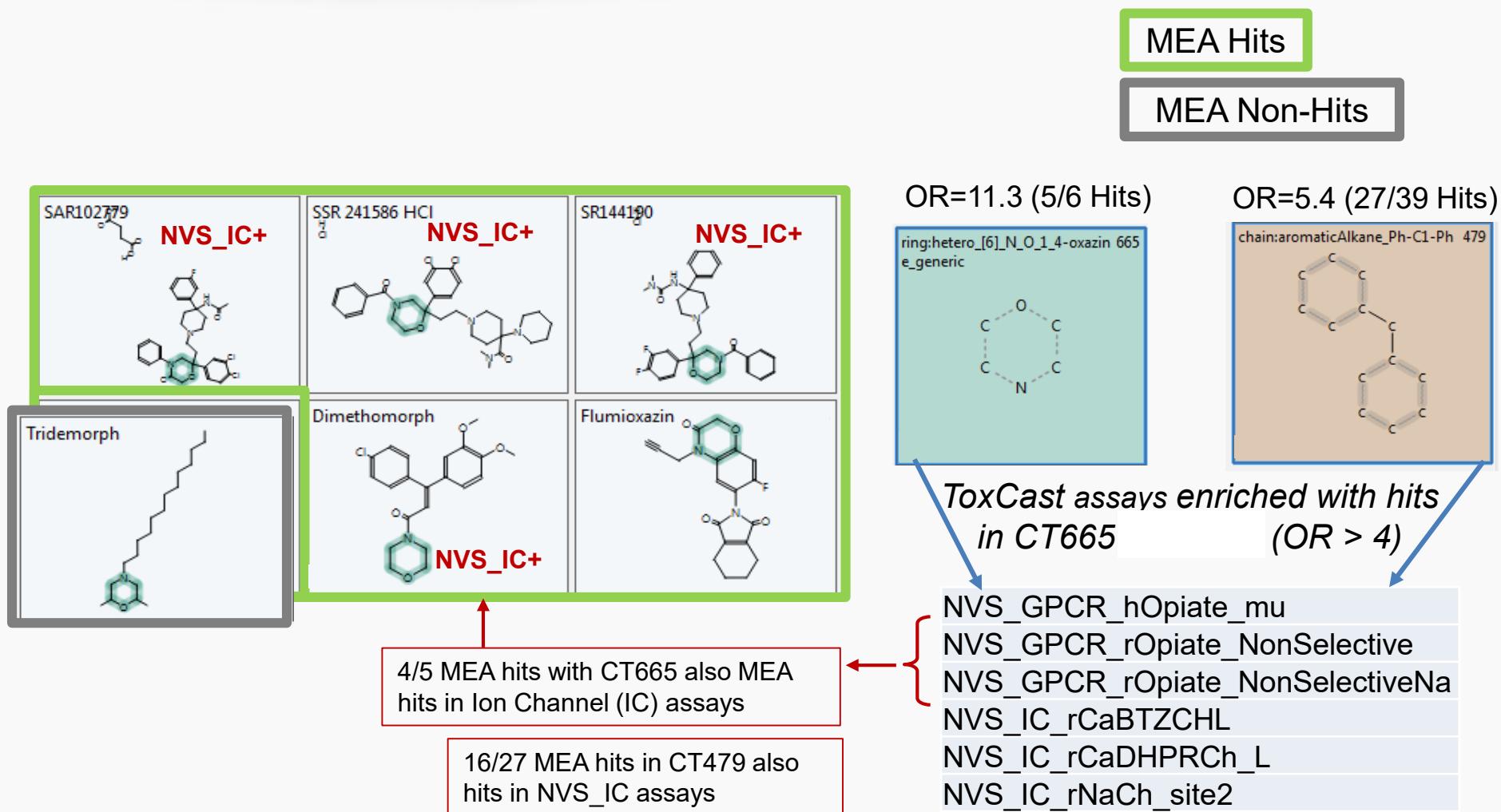
MEA Hits

MEA Non-Hits

OR=18.3 (8/9 hits)

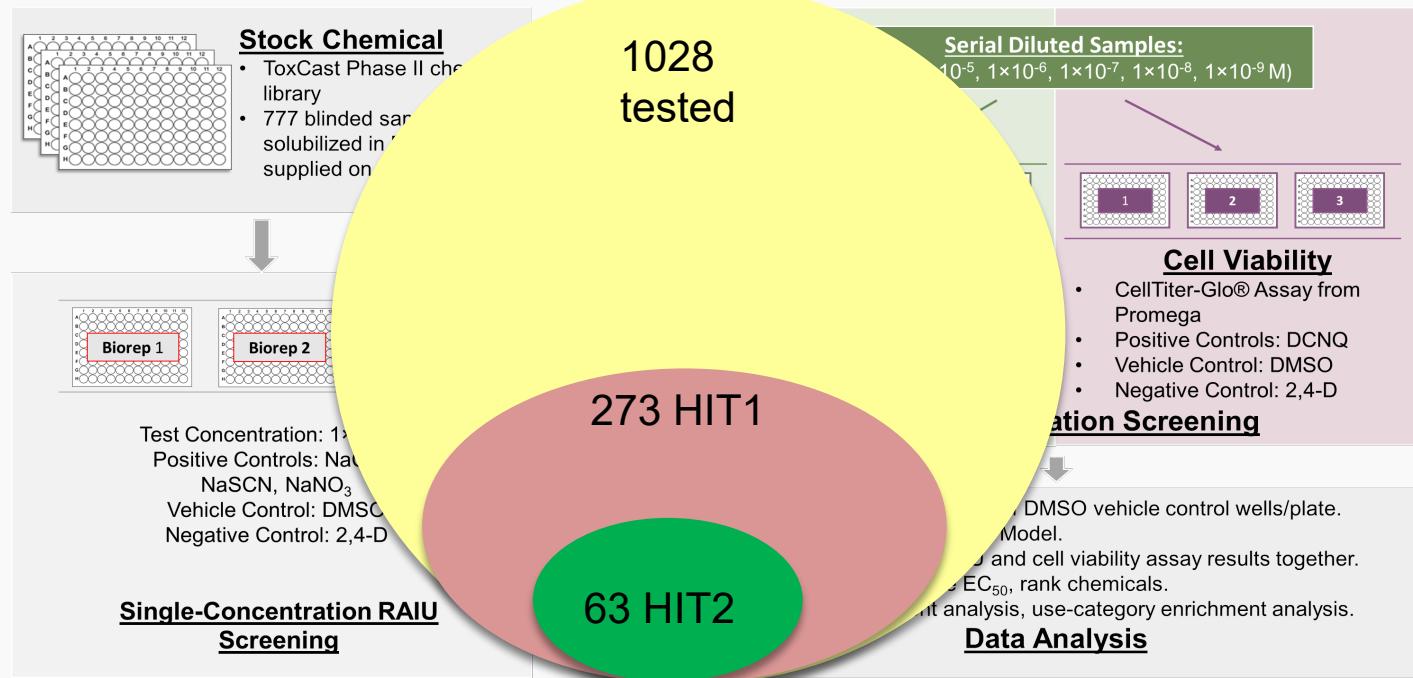


MEA CT-Enrichment Analysis: Use CTs to build biological linkages



NIS Inhibition Assay (with Jun Wang, Susan Laws et al.)

High-Throughput Screening and Chemotype-Enrichment Analysis of ToxCast Phase II Chemicals Evaluated for Human Sodium-Iodide Symporter (NIS) Inhibition



1028 ToxCast Phase II chemicals screened

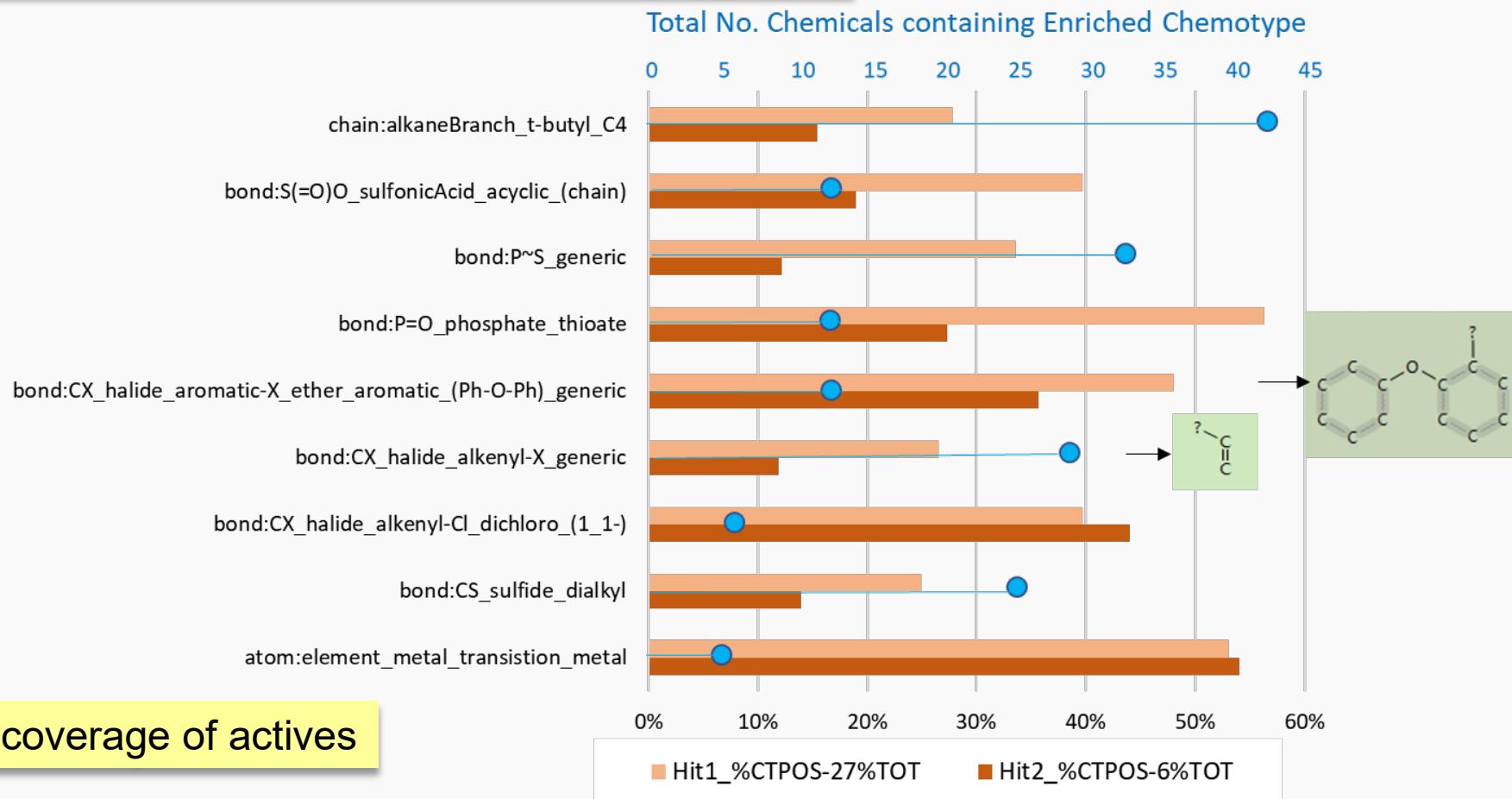
→ 27% positive at 20% threshold in single screen ... **Hit1**

→ 6% positive in multiscreen, cell viability filtered results ... **Hit2**

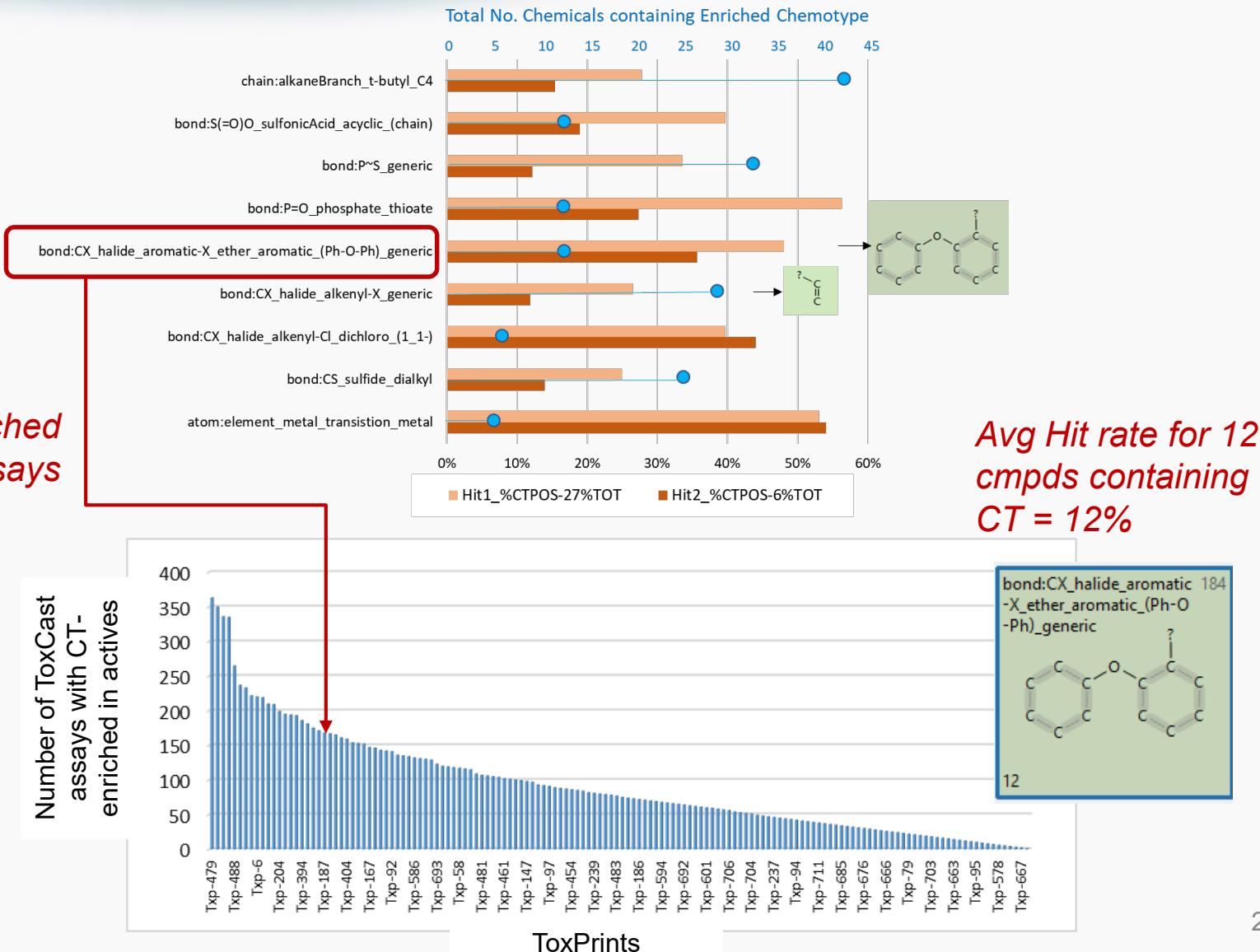
NIS Activation Assay

Overall coverage

Top 9 CT's enriched in both Hit1 and Hit2 datasets



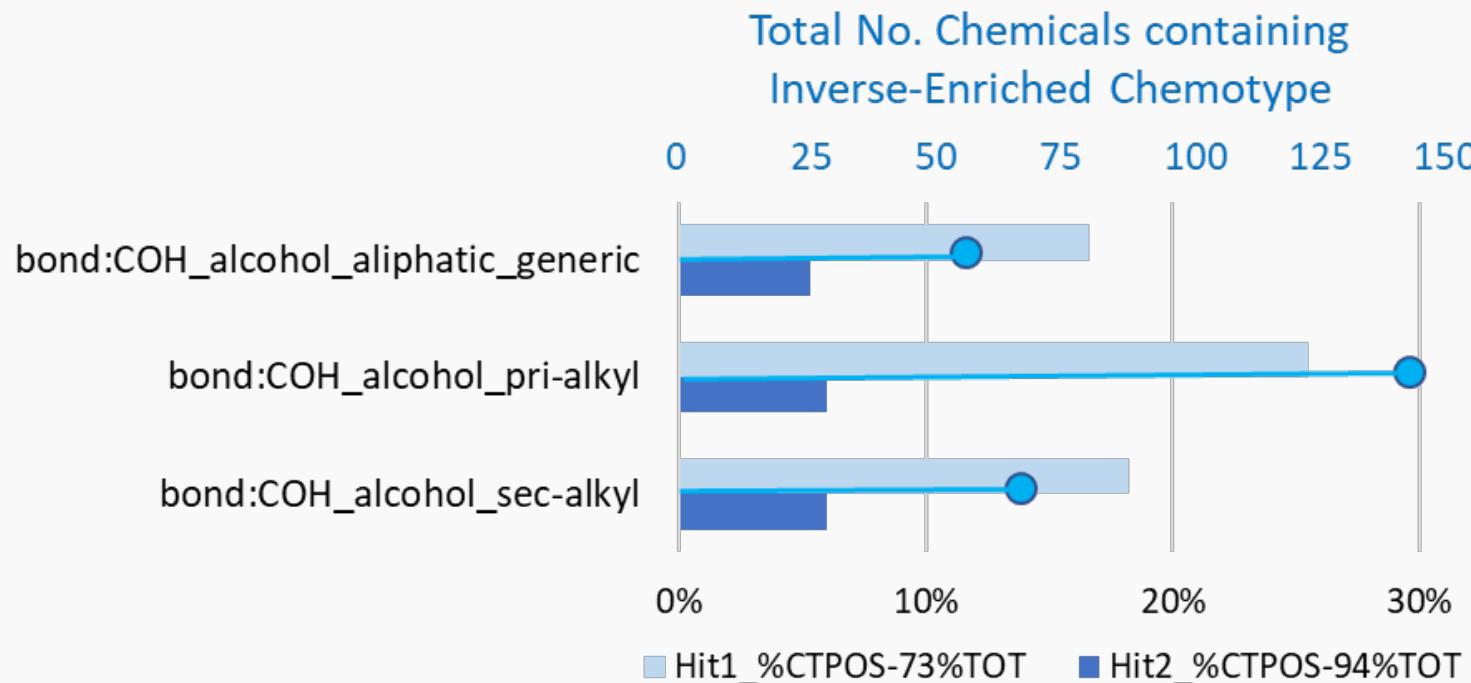
Linking global inferences to local CT enrichment results



NIS Activation Assay

Inverse activity enrichments

Top 3 CT's enriched in both Hit1 and Hit2 "Inverse" datasets

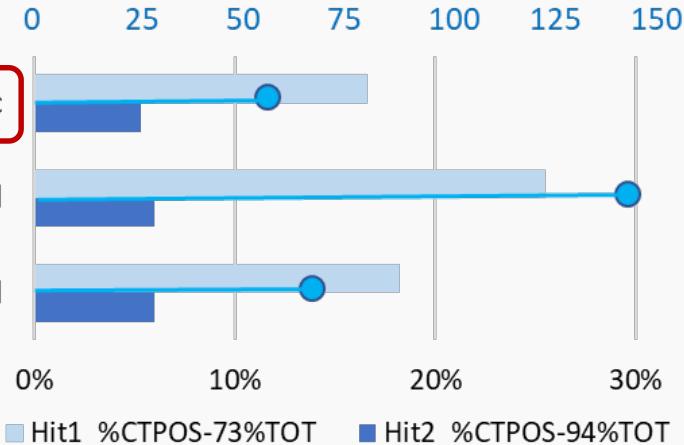


NIS Activation Assay

Inverse activity enrichments

Top 3 CT's enriched in both Hit1 and Hit2 "Inverse" datasets

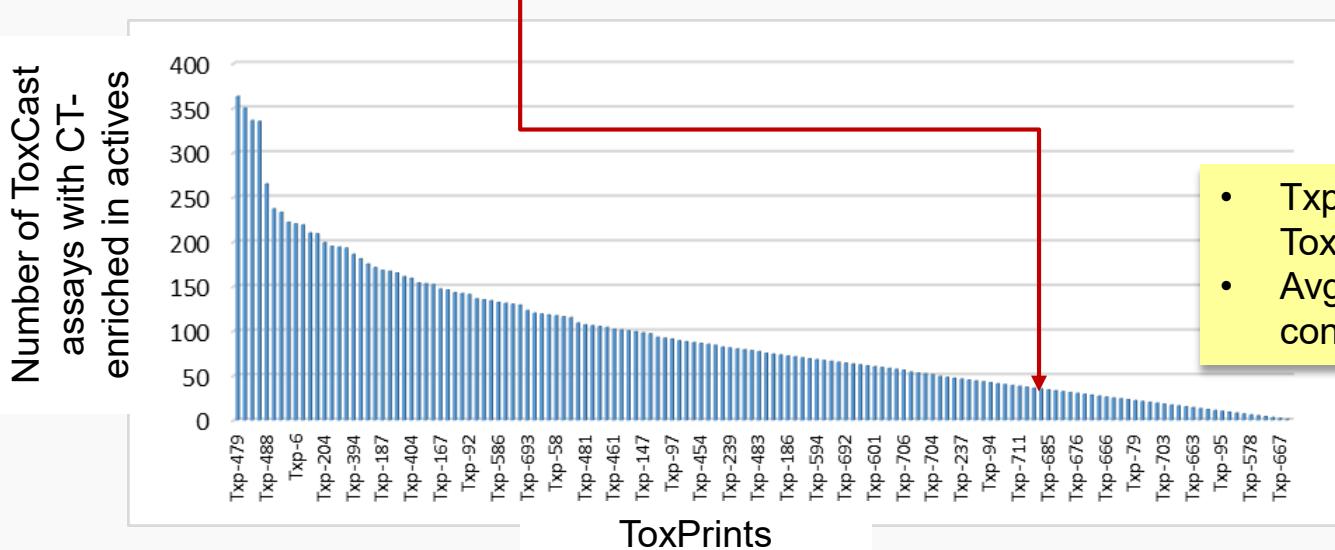
Total No. Chemicals containing Inverse-Enriched Chemotype



Txp-118 bond:COH_alcohol_aliphatic_generic

bond:COH_alcohol_pri-alkyl

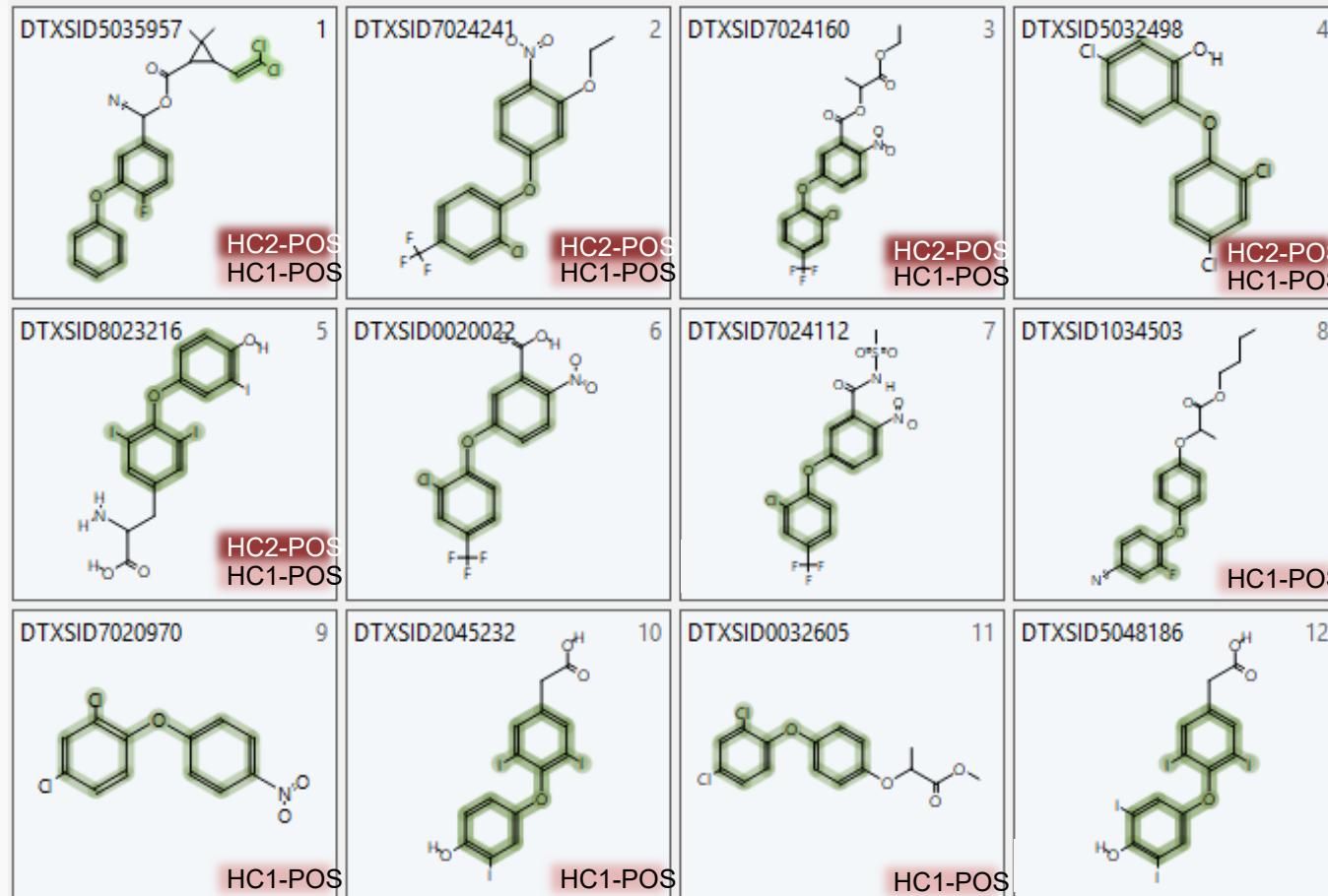
bond:COH_alcohol_sec-alkyl



- Txp-118 enriched in 42 ToxCast assays
- Avg assay hits per chemical containing Txp-118 is 8%

NIS Activation Assay

Exploring activity within CT domain



bond:CX_halide_a 142
lkenyl-X_generic

?

bond:CX_halide_aromatic 184
-X_ether_aromatic_(Ph-O
-Ph)_generic

The chemical structure of bisphenol A (BPA) is shown as a central oxygen atom bonded to two phenyl groups. Each phenyl group consists of a benzene ring with a methyl group attached at the para position.

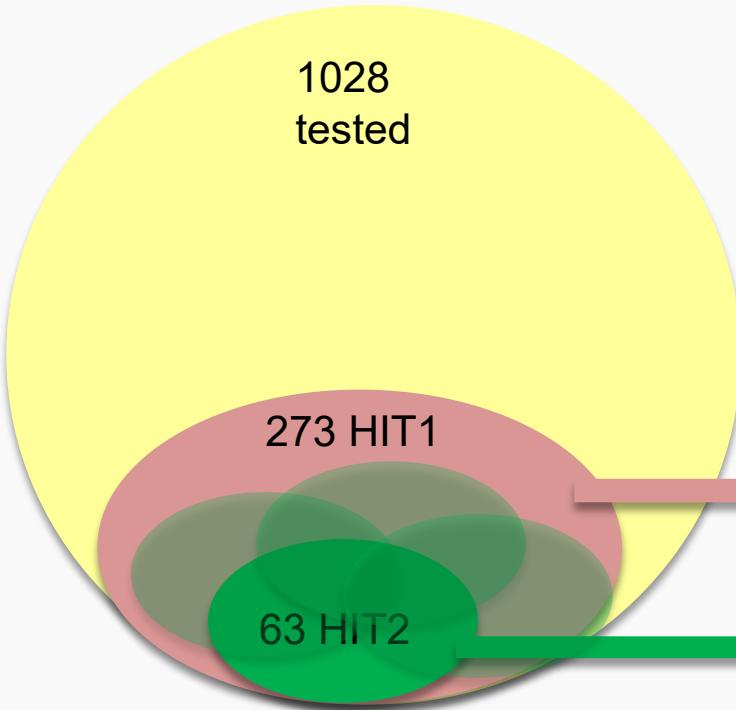
12

? (halogen) = F, Cl, I

- What distinguishes inactives in CT-subspace?
 - What distinguishes the multiscreen Hit2 actives (HC2) from the single screen Hit1 (HC1) actives?

Random folds to select Hit2 set

→ 3 Random folds to select 63 HIT2 candidates from HIT2 set



Set	Total CT-Enrich	BA (full model)
HIT1	30	0.71
HIT2	23	0.75
Fold 1	15	0.71
Fold 2	19	0.71
Fold 3	16	0.71

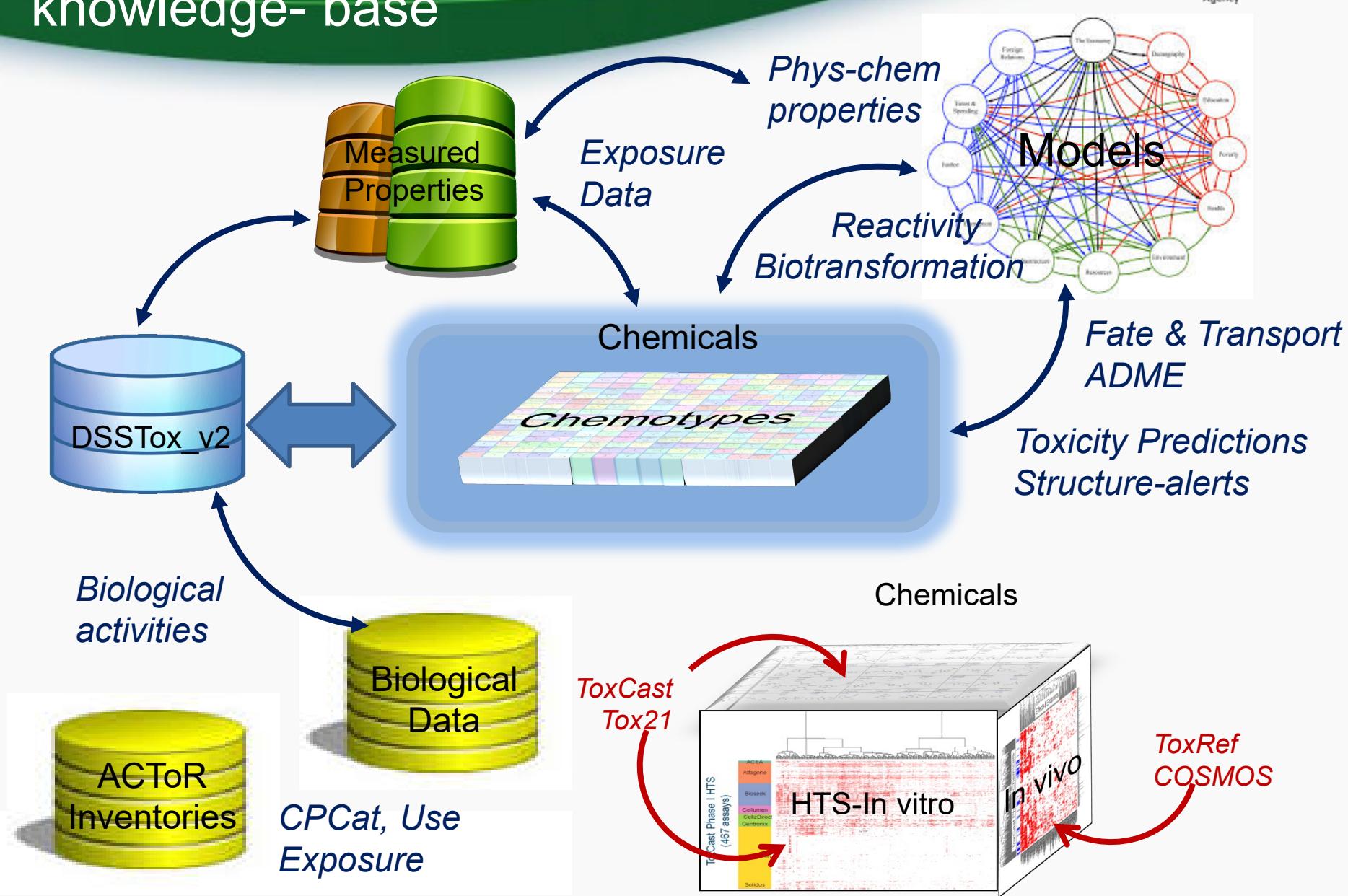
30 CTs enriched
60% coverage of actives
BA=0.71

23 CTs enriched
64% coverage of actives
BA=0.75

HIT1: Single Screen

HIT2: Multiscreen + cell viability filters

Building a public chemotype “knowledge- base”



Acknowledgements:

- ◆ EPA NCCT Chemistry Team

Ryan Lougee (CTEW workflow coding)

Chris Grulke, Antony Williams (DSSTox, Chemotypes, Dashboard)

Jeremy Fitzpatrick (Global QSARs)

- ◆ ToxCast case studies

Tim Shafer, Jenna Strickland, Susan Laws, Jun Wang, Tammy Stoker

- ◆ ToxPrint Chemotypes

Chihae Yang, Jim Rathman & colleagues, Molecular Networks & Altamira

- ◆ NCCT & ToxCast Team



Thank you for your attention



Question

OR



Comment