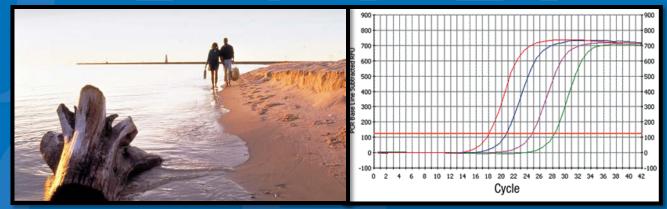


Quantitative Microbial Source Tracking in the Tillamook Basin

Xiang Li, M. Sivaganesan, C.A. Kelty, A. Zimmer-Faust, P. Clinton, J.R. Reichman, Y. Johnson, W. Matthews, Bevin Horn, S. Bailey, and O.C. Shanks



Office of Research and Development National Risk Management Research Laboratory, Water Supply and Water Resources Division





- 1. Microbial Source Tracking Overview
- 2. Tillamook Field Study
- 3. Technology Transfer



Disclaimer: The views expressed in this presentation are those of the author[s] and do not necessarily represent the views or policies of the U.S. Environmental Protection Agency. Mention of trade names or commercial products does not constitute endorsement or recommendation for use.



Fecal Pollution is a Nationwide Problem

- Fecal microbes are the most common biological contaminants in U.S. surface waters
- Public, economic and ecological health burdens









Fecal Pollution in Surface Waters:

EPA Responsibilities

Protect and Restore Waters for Recreational Use

- Clean Water Act 1972



Risk Assessment of Beach Contaminants

- BEACH Act (2000)
- Development of new or revised ambient water quality criteria (AWQC)

Management of Point and Non-Point Pollution Sources

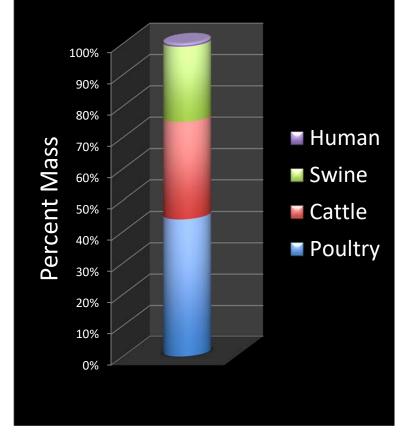
- Total Maximum Daily Load (TMDL) programs
- National Pollutant Discharge Elimination System (NPDES) programs
- National Estuary Program (NEP)
- Combined Sewer Overflow (CSO) consent decrees



Source of Fecal Pollution is Important

- Current recommended water quality criteria
 - Based on general fecal indicators
 - Measure total fecal pollution
 - Do not discriminate between sources
- Estimated 1x10⁹ tons of fecal material produced in U.S. each year
 - Human (0.01%)
- Animal sources require different management and remediation strategies

Agricultural and Human Sources of Feces in the U.S.

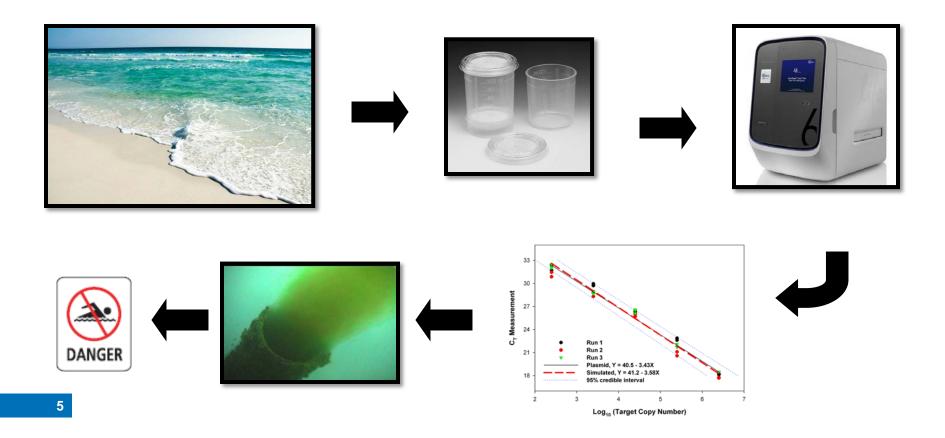


RL Kellogg, CH Lander, DC Moffitt, N Gollehon - NRCS and ERS GSA Publ. No. NPS00-0579. Washington, DC: USDA, 2000



Microbial Source Tracking: Concept Review

SOLUTION... Method designed to collect, isolate, identify, and measure a <u>host-associated identifier</u> from an environmental sample.





Microbial Source Tracking: Many Water Quality Management Applications

- Identification of non-point pollution sources
- Impaired site prioritization for remediation
- Evaluation of a best management practices
- Bacterial Total Maximum Daily Load planning
- Nutrient discharge compliance monitoring tool
- Urban stormwater management support



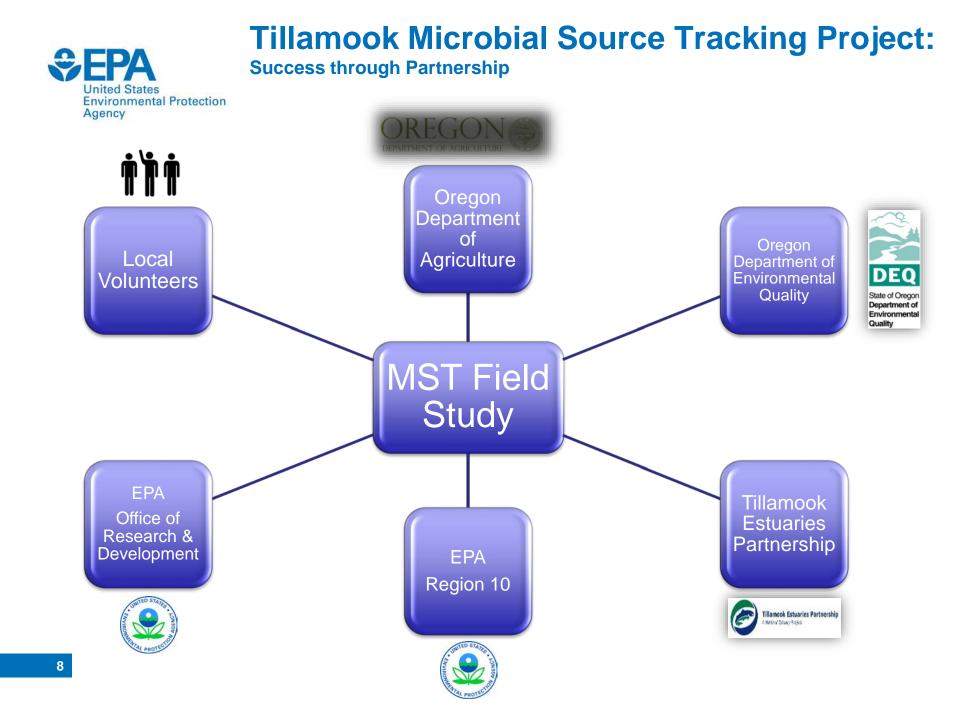


Tillamook Microbial Source Tracking Project: Background Information

Important resource

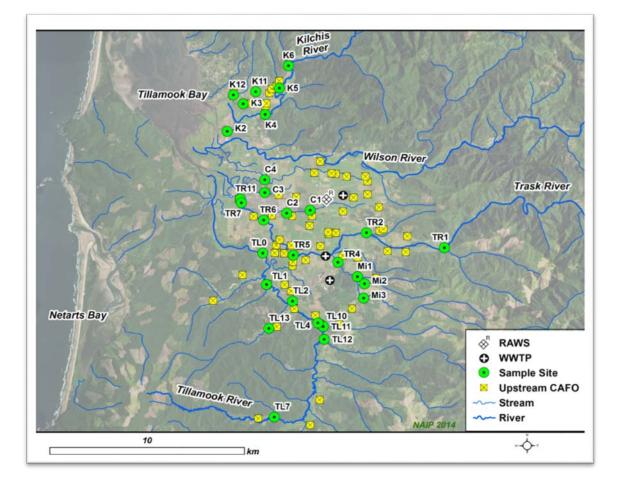
- > Dairy, cheese-making and shellfish industries
- Local human and wildlife populations
- Impact of fecal pollution
 - Economic loss
 - Poor conditions for recreational use
 - Endangerment of local wildlife
- Complex challenge
 - Multiple pollution sources
 - Limited resources for management and remediation
- How can microbial source tracking improve watershed management?







Tillamook Microbial Source Tracking Project: Sampling Approach



- Tillamook Basin (Oregon)
- Three river systems
 - Tillamook
 - Trask
 - Kilchis
- 29 sampling sites
- 12-month sampling period
- Bimonthly sampling
 696 total samples



Tillamook Microbial Source Tracking Project: Data Collection Methods

- Historical Water Quality Information
- Geographic Information System (GIS) mapping
- Weather Information
- Local Water Quality Metric
 E. coli (IDEXX Colilert)
- MST Genetic Markers of Fecal Pollution
 - Human-associated (HF183/BacR287 and HumM2)
 - Ruminant (Rum2Bac)
 - Cattle (CowM2 and CowM3)
 - Dog (DG3 and DG37)
 - Avian (GFD)











Vol. 72, No. 8

State water quality definition (E. coli 406 MPN/100mL)

- Historical E. coli exceedance trends
- Previous MST study
 - **End-point PCR** \geq

Environmental Protection

Agency

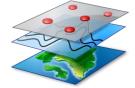
Spatial and temporal trends in ruminant and human \geq sources

APPLIED AND ENVIRONMENTAL MICROBIOLOGY, Aug. 2006, p. 5537–5546 0099-2240/06/\$08.00+0 doi:10.1128/AEM.03059-05 Copyright © 2006, American Society for Microbiology. All Rights Reserved. Basin-Wide Analysis of the Dynamics of Fecal Contamination and Fecal Source Identification in Tillamook Bay, Oregon Orin C. Shanks,¹ Christopher Nietch,¹ Michael Simonich,² Melissa Younger,² Don Reynolds,³ and Katharine G. Field²

River	Site n	<i>E. coli</i> ≥ 406 MPN/100mL			
System		% ± SE	Trend	p-value	
	K12	230	28.3 ± 3.0		
	K3	227	34.4 ± 3.2	\downarrow	0.01
Kilchis	K6	229	0.4 ± 0.4		
KIICIIIS	K5	234	0.4 ± 0.4		
	K4	234	2.1 ± 0.9	\downarrow	0.06
	K2	229	1.3 ± 0.8		
	C4	234	12.4 ± 2.2		
	C3	225	69.3 ± 3.1	\downarrow	< 0.0001
	C1	172	61.6 ± 3.7		
	C2	175	58.3 ± 3.7		
	TR1	237	0.8 ± 0.6		
	TR2	237	3.4 ± 1.2		
Trask	TR5	237	5.1 ± 1.4		
TIASK	TR6	237	59.5 ± 3.2	\downarrow	< 0.0001
	TR7	237	7.2 ± 1.7		
	TR11	237	8.0 ± 1.8		
	Mi3	236	77.5 ± 2.7		
	Mi2	236	11.4 ± 2.1		
	Mi1	237	15.2 ± 2.3		
	TR4	237	28.3 ± 2.9		
	TL7	230	37.0 ± 3.2	\downarrow	0.07
	TL12	231	5.2 ± 1.5		
	TL11	231	14.3 ± 2.3	\downarrow	0.03
	TL10	231	13.0 ± 2.2	\downarrow	0.08
Tillamook	TL4	231	11.7 ± 2.1		
	TL2	231	27.3 ± 2.9	\downarrow	< 0.0001
	TL1	231	26.4 ± 2.9	\downarrow	< 0.0001
	TL0	231	16.0 ± 2.4		
	TL13	231	12.6 ± 2.2		



Tillamook Microbial Source Tracking Project: Land Use GIS Mapping



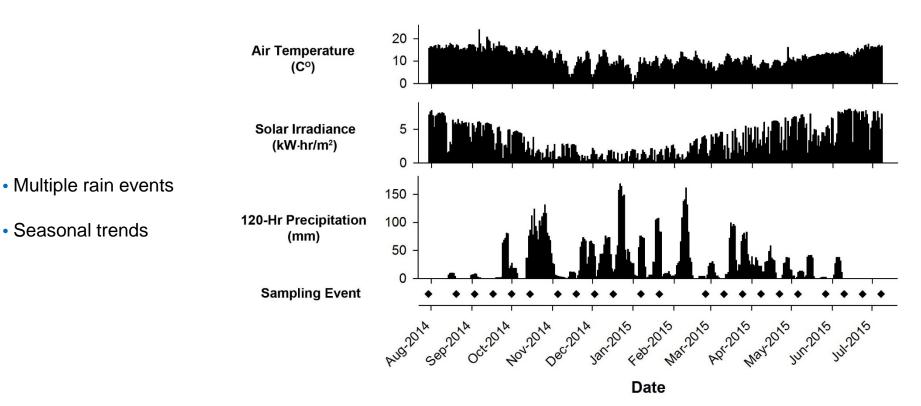
- Hydrology defined catchment area
- Cattle and human populations
- Percent non-sewer and crop land
- Investigate links to water quality data



Туре	Parameter	Mean	Median	Std.	Min.	Max.
	Human Population	1051.6	533	1727.1	2	6395
Land	Permitted Cattle					
Use	Population	2418.9	826	3472.7	0	12,371
	Non-sewer (%)	1.06	0.18	1.83	0.006	7.44
	Cropland (%)	1.97	0.53	4.89	0.08	25.7



Weather Conditions over Study Period

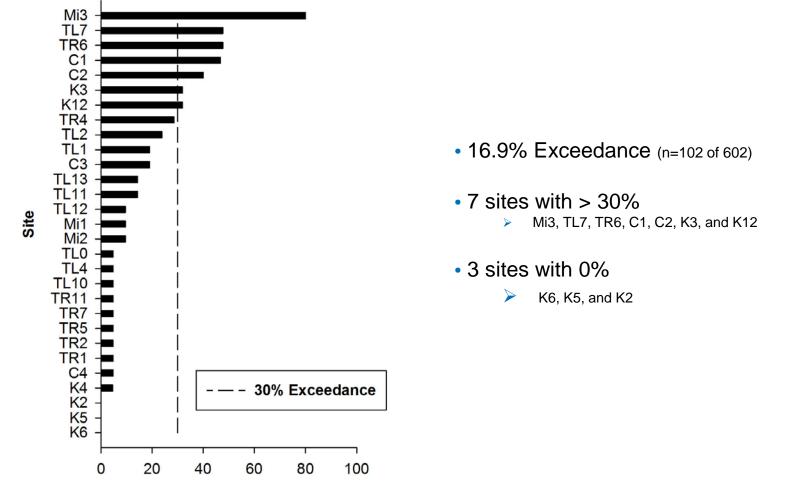


Median Std. Max. Type Parameter Mean Min. 120-hr precipitation (mm) 9.5 36.6 27.3 0 168.9 72-hr precipitation (mm) 16.4 3.8 25.9 0 144.5 Weather 24-hr precipitation (mm) 12.3 5.5 0.1 0 101.1 Condition Solar Irradiance (kW-hr/m²) 3.5 2.9 2.3 0.1 8 12 Air Temperature (°C) 11.9 3.6 0.7 23.9



The Water Quality Management Perspective

 Single-Day Maximum of 406 MPN/100mL

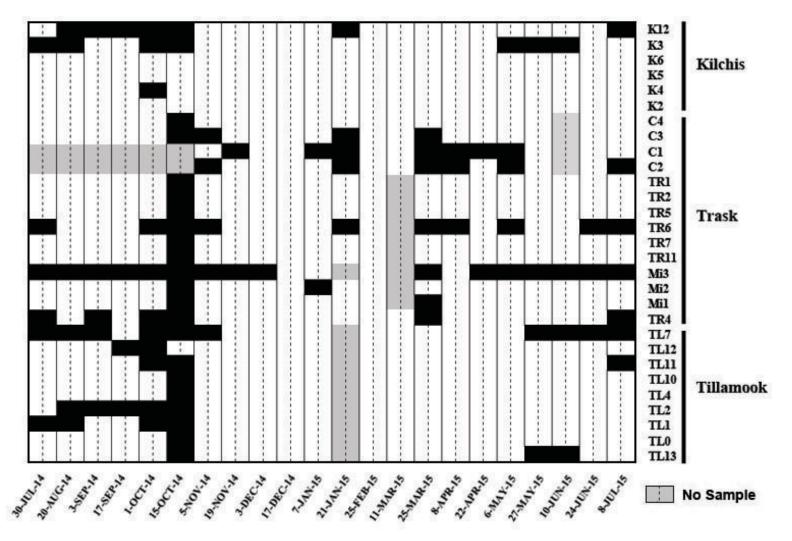


% Exceedance



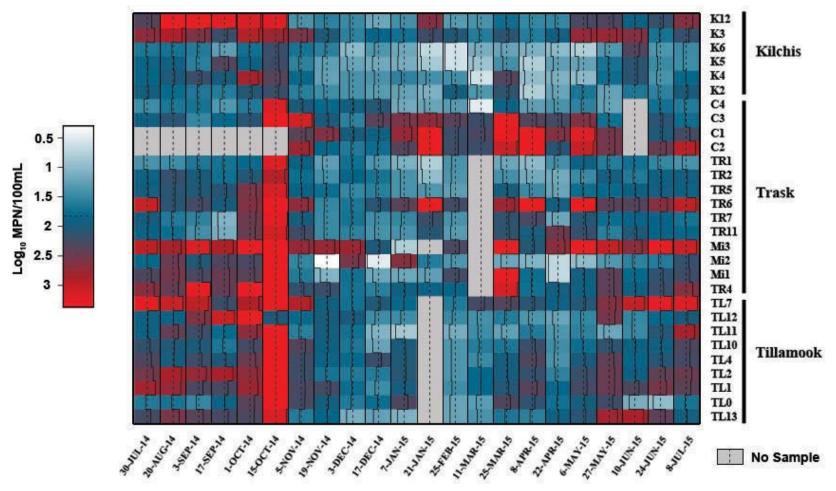
The Water Quality Management Perspective

Single-Day Maximum of ≥ 406 MPN/100mL





E. coli Spatial and Temporal Trends



 Present across all sites tested Evident temporal and spatial trends



qPCR MST Data Interpretation Considerations

- Each MST genetic marker is a discrete measurement
 - Different bacterial species
 - Different DNA sequence
- MST genetic marker shedding can vary by:
 - Animal diet, age, and health
 - Cohabitation behaviors
- Incompatibilities between cultivated *E. coli* and MST genetic markers
 - *E. coli* = live cells that can be cultivated in lab
 - MST genetic markers = any live or dead cell + free-DNA
 - Live cells and genetic material respond differently to environmental stressors
- Inconsistencies between *E. coli* and MST measurements prevent accurate source apportionment
- Recommend independent interpretation of each water quality measurement indicator/identifier data set





qPCR Data Quality Controls

Standard curve performance

- Outlier removal
- Amplification efficiency (E)
- Correlation coefficient (R²)
- Amplification inhibition testing
 - Instrument run proficiency test
 - Internal amplification control with every sample
- Contamination screening
 - Field blanks
 - Method extraction blanks
 - No template controls
- Optimal DNA recovery monitoring
 - Batch proficiency test
 - > Sample processing control with every sample





qPCR Performance in Study Area

- Pollution source reference collection (n=114)
- Determine sensitivity and specificity in Tillamook study area

Ruminant and	Accov	Sensi	Specificity	
Cattle	Assay	Adult	Juvenile	Specificity
	Rum2Bac	93%	0%	100%
	CowM2	36.5%	0%	99.4%
	CowM3	82.3%	0%	100%

Avian

n	Accov	Sens	Specificity	
	Assay	Wildlife	Chicken	Specificity
	GFD	24.4%	0%	100%
an	Assay	Sensitivity	Specificity	
	HumM2	100%	99.1%	

Huma

Assay	Sensitivity	Specificity
HumM2	100%	99.1%
HF183	100%	99.4%

Dog

Assay	Sensitivity	Specificity
DG3	97%	100%
DG37	36.4%	100%

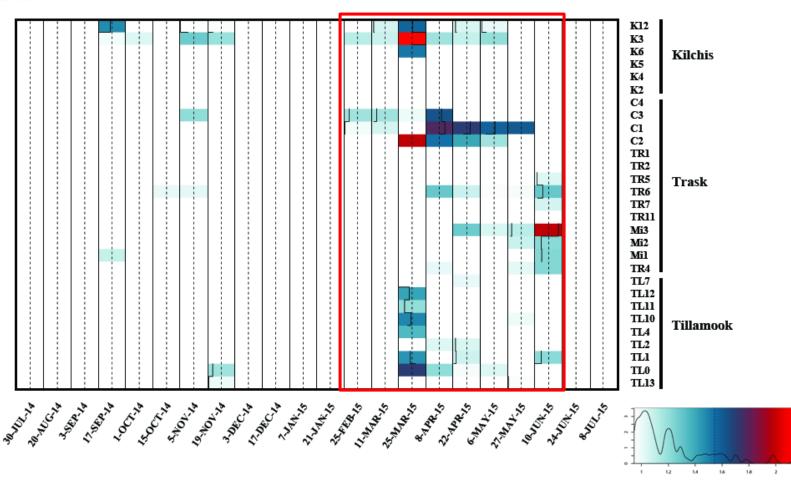
Source	n
Adult Cattle	32
Juvenile Cattle	19
Dog	11
Sewage	4
Wildlife Birds	15
Chickens	11
Elk	11
Horse	11

Key Observations:

- High specificity for all assays
- Age and diet important factors
- Avian method limitations
- DG3 superior host distribution



Watershed Spatial and Temporal Trends in Avian Pollution



Potential bird migration water quality impact



21

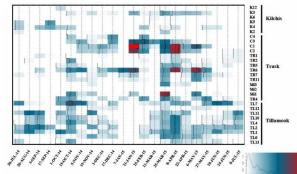
Tillamook Microbial Source Tracking Project:

Watershed Spatial and Temporal Trends in Fecal Sources

Kilchi

Trask

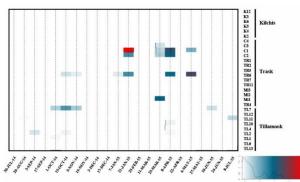
Tillamook

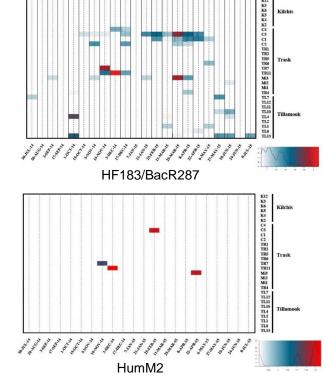




CowM2

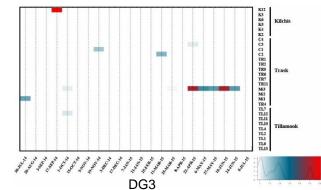
CowM3





Kilchis

Trask



Spatial trends

- > Land use
- Waste management \geq practices

• Temporal trends

- Weather conditions \triangleright
- > Agricultural practices
- Wildlife activities \geq
- Varies by MST assay

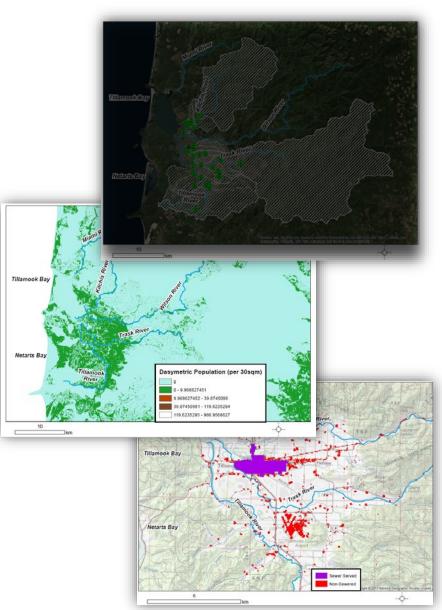


22

Tillamook Microbial Source Tracking Project:

Watershed Land Use and Water Quality Trends

- Identify pollution trends by fecal pollution metric and land use information
- Percent non-sewer most influential parameter
 - > E. coli (+, R²=0.23, p=0.081)
 - > HF183/BacR287 (+, R²=0.32, p=0.009)
 - > GFD (+, R²=0.31, p=0.014)
- Ruminant pollution closely linked to maximum number of permitted cattle
 - Rum2Bac (+, R²=0.50, p=0.001)





Watershed Weather Conditions and Water Quality Trends



Weather		Fecal Pollution Metric				
Parameter	Count	E. coli	Rum2Bac	CowM3	HF183/BacR287	GFD
24-Hr Precipitation	208	0.003	<0.0001	0.247	0.455	0.167
72-Hr Precipitation	160	0.134	<0.0001	0.157	0.354	0.114
120-Hr Precipitation	207	0.184	<0.0001	0.193	0.381	0.499
Solar Irradiance	322	<0.0001	0.002	0.127	0.151	0.218
Air Temperature	294	<0.0001	<0.0001	0.005	0.033	0.170

- *E. coli* levels significantly associated with 24-Hr precipitation, solar irradiance, and air temperature
- Ruminant pollution concentration closely linked to all weather conditions
- Avian pollution not linked to weather conditions



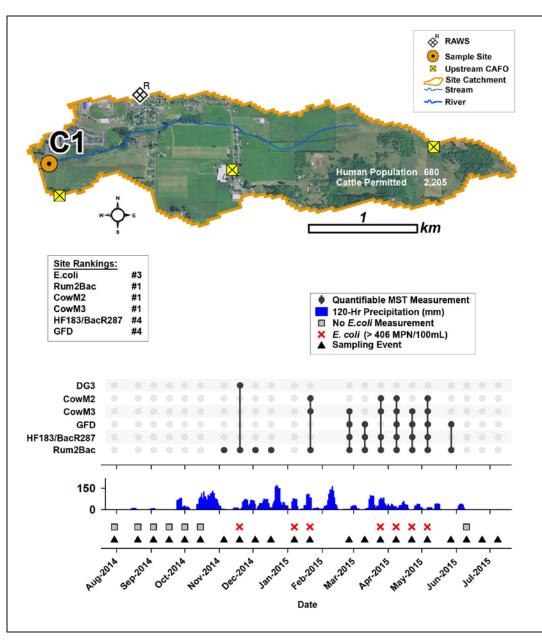
24

Tillamook Microbial Source Tracking in Action:

C1 Site Profile

- Trask River System
- E. coli exceedance (46.7%)
- Possible bird migration impact
- Human impact during spring
- Consistent ruminant impact
- Management recommendations
 - Sanitary survey in Spring
 - Prioritize by fecal source





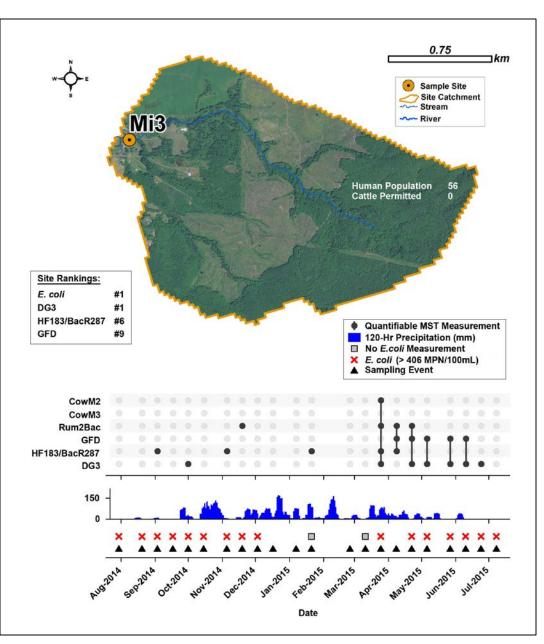
Tillamook Microbial Source Tracking in Action:



Mi3 Site Profile

- Trask River System
- E. coli exceedance (80%)
- Seasonal dog pollution, target local breeding facility
- Possible bird migration impact
- Human impact during wet season
- Ruminant in spring, likely AFO
- Management recommendations
 - Sanitary survey in Spring
 - Target AFO, septic system, and dog facility

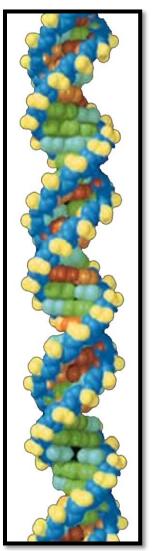






Tillamook Microbial Source Tracking Project: Conclusions

- Full-scale field study implementing quantitative MST methods
- Success via partnership
- Evident temporal, spatial, weather, and animal source pollution patterns
 - Watershed level
 - Site level
- Quantitative MST enhances water quality management
 - Site prioritization by pollution source
 - Strategic sanitary survey planning
 - Identification of non-point pollution sources
 - Evidence-based wildlife impact information
 - Increase public awareness and acceptance

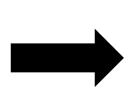




Microbial Source Tracking with qPCR: Technology Transfer

- Growing demand
- Many potential applications
- Need to transition from research method to management tool





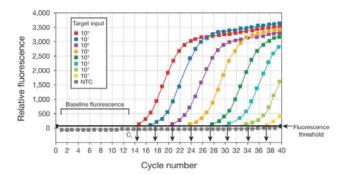
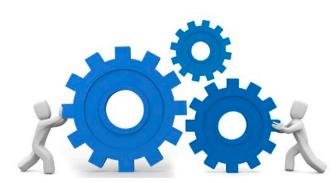


Figure 1. Relative fluorescence vs. cycle number. Amplification plots are created when the fluorescent signal from each sample is plotted against cycle number; therefore, amplification plots represent the accumulation of product over the duration of the real-time PCR experiment. The samples used to create the plots in this figure are a dilution series of the target DNA sequence.



Technology Transfer: Building an Implementation Took kit



- Publication of two EPA Methods (pending EPA OW review)
- Automated data analysis tool
- Proficiency test procedure
- Development of certified reference material
- Training opportunities



QUESTIONS?

Orin C. Shanks, Ph.D. Senior Scientist EPA Office of Research & Development Phone: (513) 569-7315 Email: <u>shanks.orin@epa.gov</u>



Disclaimer: The views expressed in this presentation are those of the author[s] and do not necessarily represent the views or policies of the U.S. Environmental Protection Agency. Mention of trade names or commercial products does not constitute endorsement or recommendation for use.