

Indoor Residence Times of Semivolatile Organic Compounds: Model Estimation and Field Evaluation

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1 **ABSTRACT**

2 Indoor residence times of semivolatile organic compounds (SVOCs) are a major and
3 mostly unavailable input for residential exposure assessment. We calculated residence times for
4 a suite of SVOCs using a fugacity model applied to residential environments. Residence times
5 depend on both the mass distribution of the compound between the “mobile phase” (air and dust
6 particles settled on the carpet) and the “non-mobile phase” (carpet fibers and pad) and the
7 removal rates resulting from air exchange and cleaning. We developed dust removal rates from
8 cleaning processes using an indoor-particle mass-balance model. Chemical properties determine
9 both the mass distribution and relative importance of the two removal pathways, resulting in
10 different residence times between compounds. We conducted a field study after chlorpyrifos was
11 phased out for indoor use in the U.S. in 2001 to determine the decreases in chlorpyrifos air
12 concentrations over a one year period. A measured average decrease of 18% in chlorpyrifos air
13 concentrations indicates the residence time of chlorpyrifos is expected to be 6.9 years and
14 compares well with model predictions. The estimates from this study provide the opportunity to
15 make more reliable estimates of SVOCs exposure in the indoor environment.

1. INTRODUCTION

Semivolatile organic compounds (SVOCs) are ubiquitous in the indoor environment and include plasticizers, brominated flame retardants, pesticides, and perfluorinated compounds [2-5]. As a result, they have been measured in indoor air and dust and it is evident there is human exposure as they have also been measured in biological samples as parent compounds in blood and as metabolites in urine [6, 7]. Some SVOCs have a strong tendency to bind to organic carbon in dust or to indoor surface materials. For these compounds that are primarily partitioned to dust or surface materials, residence times of indoor SVOCs are determined by both the removal rates resulting from air exchange and cleaning and the relative fraction of compounds in the air, dust, and surface reservoirs.

Chemical transformation or degradation by photolysis has little or no effect on SVOCs removal indoors because there is little direct sunlight [8]. In general, indoor SVOCs are resistant to biodegradation and only a few plasticizers are biologically degraded under specific environmental conditions [9, 10]. Thus, the dominant removal pathways for SVOCs from the home are expected to be either air exchange processes or removal of dust through surface cleaning, depending on the chemical properties of the SVOCs.

The air and dust are considered “mobile phases”. There are also non-mobile phases such as carpet fibers that serve as reservoirs and there are no direct removal processes for a compound partitioned into surface reservoirs other than replacing surface materials. However, compounds in reservoirs can be redistributed by mass transfer to mobile phases and then removed. For example, when considering removal by cleaning, Hunt et al. found that mechanical cleaning did not completely remove SVOCs on surfaces because a significant portion of the compound was found in surface reservoirs such as carpet fibers or foam materials after a carpet vacuuming event

[11]. Therefore, we use the term “non-mobile phase” to represent the portion of the compound partitioned into surface materials as opposed to the portion in dust or air.

In addition to determining the fraction in the mobile phase, specifically dust, the residence time of dust needs to be quantified to assess the resulting residence time of SVOCs in the home. Few studies have characterized residence times of indoor dust. Qian et al. reported 81 days of residence time ($\tau = \text{loss-rate}^{-1}$ (d)) for particles with a size of 2 μm in aerodynamic diameter when vacuum cleaning was done once per week [12]. Allott et al. estimated a mean indoor residence time of 29 days from a house in the UK where vacuum cleaning occurred every day [13]. Layton and Beamer estimated a mean residence time, τ (d), of 61 days for floor dust, including a cleaning loss rate of 0.0053 d^{-1} and a resuspension rate of 0.011 d^{-1} [14].

SVOCs are fairly persistent in the indoor environment, as demonstrated by the presence of many pesticides remaining in the indoor environment for years or decades after the chemical was banned for indoor, or in some cases, outdoor use as well [15-17]. While studies have focused on the air exchange rate of homes and the residence time of dust, these factors have not been combined with the relative mass distribution between the mobile phase and the non-mobile phase to address the residence time of SVOCs partitioned to surface materials.

Distinct from previous studies, the objectives of this study are to (1) estimate the residence time of SVOCs in the indoor environment as a function of chemical partitioning and removal rates resulting from air exchange and cleaning, (2) understand the fate and transport of dust for three size ranges from an indoor dust model, (3) estimate a dust removal rate by surface cleaning, and (4) conduct a limited model evaluation. We conducted two case studies. First, based on removal rates and chemical properties, we estimated residence times for five chemicals to compare the persistence and the primary removal pathway. Second, for model evaluation, we

used chlorpyrifos air samples from two time points collected in a field experiment. The phase out of the indoor use of chlorpyrifos in the United States in 2001 [18] provides the opportunity for a unique model evaluation exercise because we can compare the average annual decrease with the modeled concentration changes.

2. METHODS

2. 1. Estimation of SVOCs residence times

For volatile organic compounds with a high vapor pressure and no indoor sources or contributions from outdoors, the mass in a one-box system decreases based on the following exponential decay equation:

$$M(t) = M_0 \cdot e^{-t/\tau} \quad (1)$$

And in the case of zero initial concentration and continuous indoor sources this mass balance takes the form:

$$M(t) = S \tau \cdot (1 - e^{-t/\tau}) \quad (2)$$

where M_0 is the initial mass in the system (mg) and τ is the residence time due only to ventilation (day) and S is an emission rate of a continuous indoor source (mg/day). In both cases τ represents the effective residence time of the system and depends on chemical and residential properties but not on M or S . However, in the case of SVOCs with low vapor pressure and high octanol-water partition coefficient (K_{ow}) values, there can be significant mass partitioned to surface compartments. Hence, at any time only a portion of the mass, including chemical desorption from surface materials, is available for ventilation removal. In this case, the mass in the system decreases more slowly:

$$M(t) = M_0 \cdot e^{-t/(\tau/\theta)} \quad (3)$$

where θ is the fraction of the compound in the mobile phase (in this case air) at equilibrium and τ/θ is the new residence time associated with θ . It is apparent that the residence time increases as the fraction in the mobile phase decreases.

Likewise, if we consider a chemical distributed between the dust and surface materials, with the dust considered the mobile phase because it can be removed by cleaning, the residence time of the compound will be longer than the residence time of the dust in the home due to the large fraction partitioned to the carpet fibers, the non-mobile phase.

A multi-compartment indoor fugacity model was used to determine the partitioning between air, carpet, vinyl, carpet dust, and vinyl dust [19]. We used a boundary layer approach to quantify diffusive transfer rates between compartments including carpet and vinyl flooring. The surface-air partition coefficients (K_{ja}) for the surface j were estimated as a function of vapor pressure derived from existing laboratory experiments. Partitioning to dust associated with surfaces was based on the relationship with the octanol-air partition coefficient (K_{oa}). SVOCs partition strongly to various indoor surfaces such as particles, flooring, furniture, walls, and ceilings. The impact of sorption to these surfaces was not included in our model due to the limited partitioning information between the surface j and air a . We summarized the model in the Supporting Information and presented the mass distribution results in Table 2. As air exchange and carpet cleaning are two primary removal pathways for various SVOCs from our preliminary mass distribution analysis, the residence time in the home depends on both the mass distribution and removal rates of two mobile phases, air and the dust settled on carpet. Then, the residence time based on equilibrium partitioning is described by the following equation:

$$\tau_{\text{sys}} = \left(\theta_1 \cdot \frac{1}{\tau_{\text{air}}} + \theta_2 \cdot \frac{1}{\tau_c} \right)^{-1} = (\theta_1 \cdot a + \theta_2 \cdot k_c)^{-1} \quad (4)$$

where τ_{sys} is the residence time of the system, τ_{air} is the residence time due to air exchange ($=1/a$), τ_c is the residence time due to dust removal from carpet ($=1/k_c$), θ_l is the fraction of the compound in the mobile phase in the air compartment [$= (M_{air} + M_{air\ particles}) / (M_{air} + M_{air\ particles} + M_{carpet} + M_{carpet\ particles})$], θ_2 is the fraction of the compound in the mobile phase in the carpet compartment [$= M_{carpet\ particles} / (M_{air} + M_{air\ particles} + M_{carpet} + M_{carpet\ particles})$], a is the air exchange rate, and k_c is the dust removal rate from carpet cleaning. As the fraction of the compound in the mobile phase decreases, its residence time in the system increases. Derivation of τ_{sys} , θ_l , and θ_2 in terms of partition coefficients, which are dependent on chemical properties, was based on an existing indoor fugacity model [19] and described in detail in the Supporting Information. Depending on the transfer rates between compartments relative to the removal rates, the system may not reach equilibrium. The residence time based on a steady-state solution can also be calculated and the equations can be found in the Supporting Information.

2.2. Particle mass balance model

We developed a particle mass balance model to describe the fate and transport of dust indoors. Within this framework, we estimated the dust removal rate from surfaces and the indoor emission rate as we identified these parameters as the least reliable values associated with particle mass flows in the literature (see Supporting Information for parameter estimation and uncertainty of these parameters). The model includes three compartments, air, carpet, and vinyl flooring, as shown in Figure 1. Carpet and vinyl flooring were treated separately because of differences in particle loading, track-in rates of outdoor soil, and removal rates through cleaning. Although particles can build-up on walls, we did not include the walls as a model compartment because the particles that land on the walls are those with an aerodynamic diameter less than 1

μm . Hence, only 0.9 % of the particles are estimated to be lost to walls (see Supporting Information for more details on particle loss to walls). Infiltration and track-in of outdoor soil (generally on shoes or by pets) were considered two primary pathways for outdoor particles to enter the house, whereas ventilation and cleaning were the main removal processes of indoor particles. Phase change, coagulation, and formation of particles in the air compartment can be treated as gains and losses from various particle size fractions and may be an important process. However, we did not have sufficient data to capture these processes independently, and rather estimated a single source term from field data. The resulting particle mass balance in the air compartment is

$$V \frac{dC_i}{dt} = C_o \cdot V \cdot a \cdot P + (C_c \cdot A_c + C_v \cdot A_v) \cdot R + S - C_i \cdot V \cdot (a + D) \quad (5)$$

where V is the volume of house (m^3), C_i is the particle concentration in indoor air (mg/m^3), C_o is the particle concentration in outdoor air (mg/m^3), a is the air exchange rate (1/day), P is the penetration efficiency of outdoor particles reaching the indoors (unitless), C_c is the particle concentration on carpet (mg/m^2), C_v is the particle concentration on vinyl flooring (mg/m^2), A_c is the area of carpet (m^2), A_v is the area of vinyl flooring (m^2), R is the resuspension rate from carpet or vinyl (1/day), S is the emission rate from indoor sources such as cooking (mg/day), and D is the deposition rate (1/day).

The particle mass balance for the surface compartment j (c for carpet and v for vinyl) is

$$A_j \frac{dC_j}{dt} = C_i \cdot f_j \cdot V \cdot D + T_j - C_j \cdot A_j \cdot R - C_j \cdot A_j \cdot k_j \quad (6)$$

where C_j is the particle concentration on the surface j (mg/m^2), A_j is the area of the surface j (m^2), f_j is the fraction of the floor that is the surface j (unitless), T_j is the particle mass brought into the

surface j through track-in (mg/day), and k_j is the dust removal rate from the surface j through cleaning (1/day).

In reality, concentrations have short-term variability to events such as cooking (increasing indoor air concentrations) or cleaning (reducing surface concentrations) at fixed intervals rather than continuously. However, we assumed that the mass of dust in any compartment would remain constant over the time scales relevant for SVOC residence times. While there is a slight increase in the particle mass in carpet over time, exemplified by the fact that carpets become dirtier over time, we assumed this rate of increase was negligible. When keeping the mass constant over time, we expected for a long-term perspective that the sum of the sources to surfaces due to deposition and track-in would be the same as the sum of the removals from surfaces due to resuspension and cleaning. The fact that the mass of dust in any compartment would remain constant over time allows us to rearrange equations (5) and (6) to estimate any parameter value in each compartment based on published estimates of other parameters in the equation. We used parameter values that were best derived from the literature along with mass balance equations to estimate the parameter with the least reliable value available from the literature for each compartment; specifically, the dust removal rates from surfaces (k_j) and the emission rate from indoor sources to air (S). Although surface cleaning including carpet vacuuming and sweeping hard floors are not actually expected to occur on a daily basis, we assumed cleaning can be represented as a continuous process.

The household properties used in the model are listed in Table S1. Both equations (5) and (6) are applicable to any particle size ranges. We used three particle size ranges including 0 – 2.5 μm , 2.5 – 10 μm , and 10 – 150 μm in aerodynamic diameter because of distinct physical properties resulting from particle size and associated health related concerns. First, the deposition

rate of particles smaller than 2.5 μm is 4 times smaller than that of particles between 2.5 and 10 μm [20]. In addition, only particles smaller than 2.5 μm may penetrate deeply into the lung [21], while any particles smaller than 10 μm are accessible for inhalation exposure [22]. Third, we restricted our analysis up to 150 μm because particles larger than 150 μm do not adhere to skin [23]. When assessing model parameter values, we also consider the relationship between those values, such as estimating that 30 – 50 % of floor dust is due to tracked soil from outdoors [24] and 20 – 43% of particles in indoor air are due to indoor sources [25]. The mean and coefficient of variation (CV) for each model parameter are listed in Table 1 and the rationale for parameter value selection and associated uncertainties are summarized extensively in the Supporting Information. We arbitrarily assigned a value of 1 for the CV of k_j , and S as there was insufficient information in the literature to determine the value of the CV and we felt the assigned mean value was reasonable considering the limited available data.

2. 3. Case study – application to a suite of SVOCs

We estimated both the equilibrium and steady-state residence times for diazinon, chlorpyrifos, permethrin, polybrominated diphenyl ethers (PBDE)-47, and PBDE-99 using both chemical partitioning between the mobile and non-mobile phases and removal rates resulting from air exchange and cleaning. We also compared the importance of the primary loss mechanisms between the compounds. Chemical properties for these compounds are listed in Table S2 in the Supporting Information. We also performed a first-order uncertainty analysis to compute the relative contribution of input variances to the resulting variance of the equilibrium residence times for each studied chemical. The first-order uncertainty analysis estimates the variance in an output variable from the first-order term of the Taylor series approximation[52].

The variance of each input variable was estimated by multiplying the sum of input variances by the square of first-order derivatives of output for each input variable. The mean and CV for each model parameter used in the uncertainty analysis are listed in Table S3 in the Supporting Information. The estimated CVs used in the uncertainty analysis included uncertainty due to the limited available data and variability due to differences between homes.

2.4. Case study – chlorpyrifos field study

We used chlorpyrifos air samples from a field study to evaluate our modeled residence time in the indoor environment. We recruited 38 families from 22 counties in Northern California and collected passive air samples at two time periods approximately one year apart in 2008 and 2009 ($\mu = 347$ days and $\sigma = 62$ days) [53]. All recruitment and data collection protocols were approved by the institutional review board at the University of California at Davis and informed consent for participation was obtained upon enrollment into the study [53]. Passive indoor air samplers were deployed for approximately 30 days ($\mu = 31$ days and $\sigma = 3.4$ days). We assumed that no chlorpyrifos was applied in the home between the two measurement points, such that the home could be considered a single system with a first-order loss rate in order to provide an observed residence time of chlorpyrifos by computing the change in air concentrations. We also assumed that chlorpyrifos concentrations in air and surface compartments were in equilibrium at the time of air sampling. Based on the K_{ow} and Henry's law constant for chlorpyrifos, the majority (>99 %) of the compound in the air compartment would be associated with the vapor phase, rather than the particle phase (see Supporting Information for more details on sampling and partitioning between air and particles) [54].

3. RESULTS AND DISCUSSION

3.1. Particle mass flows

We determined the mass flow rate to and from each compartment based both on the initial literature-based parameter values and on values obtained from balancing the emission rate of indoor sources and the floor dust removal rates using equations (5) and (6). The particle mass flow rates for each pathway and particle size range are presented in Figure 2. The five particle flow pathways include resuspension/deposition (dependent on the flow direction), track-in, infiltration/ventilation, indoor sources, and cleaning. For the smallest particle size range of 0 – 2.5 μm , the relative amount of particles resuspended from the surface compartments to the air compartment is much smaller than deposition as compared to other size ranges. For the largest size range of 10 – 150 μm , particles moving from outdoors into the air compartment through infiltration are almost negligible due to the low penetration efficiency. The mass of particles moving to the indoor air compartment through infiltration and out through ventilation are different because particle concentrations in indoor air and outdoor air are different and penetration efficiency applies only in the estimation of the particle mass flow by infiltration. We also included the particle sources to the house and the particle removals from the house to compare the contribution of each flow pathway for the house. As particle size increased, cleaning the surface compartment is a dominant removal pathway and track-in becomes a primary flow pathway into the house. The re-adjusted dust removal rates (k_j) and emission rates (S) for each particle size range are summarized in Table 1 and the re-adjusted dust removal rates for all particle size ranges for carpet and vinyl flooring from the particle mass balances are 0.008 and 0.06 day^{-1} , respectively. We carried out an uncertainty analysis on three parameters including k_c , k_v , and S (see Figure S2 of the Supporting Information). For both k_c and k_v , particle

concentrations in indoor air and on surfaces as well as the deposition rate are the three most influential parameters on the outcome uncertainties for all particle size ranges. Moreover, track-in and the area of carpet were other parameters contributing to the uncertainties of k_c . For S , as particle size increased, the contribution of the air exchange rate to the uncertainties became small and that of the deposition rate became the most significant.

3.2. Residence times of SVOCs

We computed the equilibrium mass distribution, removal rates through each pathway, percent of compounds removed in one year, and both steady-state and equilibrium overall residence times for diazinon, chlorpyrifos, permethrin, PBDE-47, and PBDE-99 (Table 2). We note that the different chemical properties of these compounds resulted in different mass distributions among the mobile phases (that include air, air particles, and carpet dust) and in the relative contribution of ventilation and carpet cleaning on total removal. For example, even though only a very small fraction of mass (0.01%) was in the air compartment compared to carpet particles (0.1%), the primary removal pathway for diazinon and chlorpyrifos was ventilation, because the ventilation rate (12.7 day^{-1}) was about three orders of magnitude larger than the dust removal rate from carpet cleaning (0.008 day^{-1}). As the amount of compound that can be eliminated through ventilation was limited by the amount of compound that can be transferred from carpet to air, the steady-state residence time was significantly longer than the equilibrium residence time.

As the mass fraction in air decreases and the mass fraction in carpet dust increases, as was the case for the other three compounds (permethrin, PBDE-47, PBDE-99), the primary removal process was through carpet cleaning. For these compounds, the percent difference

between the equilibrium and steady-state residence times was lower because the transfer rate between the carpet to the air was less limiting. Diazinon is removed most quickly, followed by permethrin, chlorpyrifos, PBDE-47, and PBDE-99.

In Figure 3, we present the concentration profiles of diazinon and PBDE-99 under two sets of removal mechanisms, carpet cleaning with ventilation and carpet cleaning only. As the gap between solid lines and dotted lines increases, the contribution of surface cleaning on SVOCs removals compared to ventilation decreases. For example, the amount of PBDE-99 removed by carpet cleaning (red dotted line) was much larger than that of diazinon by carpet cleaning (black dotted line). However, the total amount of PBDE-99 removed by both carpet cleaning and ventilation (red solid line) was smaller than that of diazinon (black solid line). These results indicate the dependence of the primary removal pathway and the residence time for SVOCs on chemical properties.

We carried out an uncertainty analysis on the residence time estimates in order to assess the sensitivity of our results to model inputs. Figure 4 illustrates results from the first-order uncertainty analysis and shows that the relative contribution to overall uncertainty in the residence time is attributable to key model inputs. For both diazinon and chlorpyrifos, a primary removal pathway was ventilation; vapor pressure and the air exchange rate were the two most influential parameters on the outcome. For the other three compounds, a primary removal pathway was carpet cleaning; therefore, the contribution of dust-related parameters including embedded dust ratio, particle loading on carpet, dust removal rate from carpet, Henry's law constant, and K_{ow} became significant to the residence time.

3.3. Comparison of field studies to model predictions

We used our field samples to evaluate the change in chlorpyrifos air concentrations collected from 38 homes after adjusting for the time difference between two air samples. This provided a basis for our model evaluation. All but one field-sample pair had levels above the method detection limit (MDL) at both time points. We assigned the half value of MDL (0.016 ng/m³) to the non-detected sample. In 25 homes, chlorpyrifos concentrations either decreased (average of 18% ($\sigma=18\%$)) or stayed relatively constant (i.e., less than 5% change). We hypothesized that although our field homes should overall have average parameter values of air exchange rate and cleaning rate, variability between homes may result in different values for the decreases in chlorpyrifos air concentrations. In Figure 5, we plot our estimated percent distribution of initial chlorpyrifos concentrations associated with different fractions in the non-mobile phase of the carpet compartment under equilibrium distribution. As the fraction of chlorpyrifos in the non-mobile phase of the carpet compartment increases, the amount removed from the house by ventilation and carpet cleaning decreases following equation (1) with the residence time defined as in equation (4).

Based on the average decrease of 18% over the year from the chlorpyrifos air samples in the majority (25/38) of the homes, we anticipated just over 98% of the chlorpyrifos to be associated with the non-mobile phase of carpet. This value compares to the fugacity-based model estimate of 98.8% of chlorpyrifos in the non-mobile phase including carpet, pad, and deeply embedded dust with only 0.1% in the mobile phase including air and carpet surface dust (Table 2). The modeled fraction of the compound in the non-mobile phase is also consistent with the measured values reported by Fortune et al. [48]. They collected pesticide samples from dust (including surface dust and more deeply embedded dust) and carpet components (including fibers and pad) from six homes in North Carolina to determine the distribution of pesticide

residues between dust, carpet fibers, and pad compartments. They found that approximately 98% of total chlorpyrifos household mass was partitioned into carpet fibers and pad.

The steady-state residence time appears to over-predict as compared to the field data. This may indicate that the model assumptions for the boundary layer resistance were not accurate, and that the true boundary layer resistance was much less than that in the model.

We note that our analysis was limited by the uncertainties associated with the increase in concentrations over time for 13 of the homes. We excluded one case where air concentrations increased dramatically from 1.9 to 35.1 ng/m³, due to an unknown chlorpyrifos source. In the remaining 12 homes chlorpyrifos concentrations increased an average of 97%. We hypothesized that increases may result from changes in experimental conditions, such as air exchange rate or indoor temperature during sampling periods. If this were the case, other compounds with long residence times would also be impacted. We compared the air sample concentrations of chlorpyrifos and simultaneously measured PBDE-47 in those 12 homes with increasing chlorpyrifos concentrations to see if PBDE-47 concentrations also increased. We found that both chlorpyrifos and PBDE-47 air concentrations increased in 6 of the 12 homes, suggesting that air exchange rate or other experimental conditions differed significantly between the two sampling times. In the other 6 homes where chlorpyrifos concentrations increased and PBDE-47 decreased, we believe measurement error may result in misinterpretation of the concentration changes. This is plausible because 5 homes had samples with concentrations under the estimated limit of quantification (ELOQ) for both chlorpyrifos and PBDE-47 and 1 additional home had concentrations under the ELOQ for chlorpyrifos. The average concentration change and the number of homes with values between the MDL and ELOQ for chlorpyrifos and PBDE-47 are summarized in the Supporting Information, Table S4.

3.4. Implications/Limitations

This study has several implications in terms of SVOCs exposure in residential and commercial areas. First, dust removal rates derived from the particle mass balance model provide insight on the residence time of SVOCs partitioned to dust, which serves as an important medium of mass transport for SVOCs in homes. Although indoor multimedia models based on fugacity have been considered useful tools to understand the behavior of SVOCs [19, 54], the role of dust removal was not well quantified in previous models. Second, the fraction of SVOCs in the non-mobile phase supports the premise that SVOCs are persistent in the indoor environment. Although chlorpyrifos was phased out for most indoor uses in 2001, the analysis of samples collected in 2008-2009 supports our determination that the equilibrium residence time (i.e., the half-life divided by $\ln(2)$) of chlorpyrifos partitioned onto the surface materials is 6.9 years. This observation is consistent with the assumption that these compounds partition primarily into the non-mobile phase of the surface materials. The steady-state residence time is much greater, indicating that chemicals may transfer more easily between phases within the indoor environment than estimated. Third, we demonstrated that the residence times and the removal rates of each pathway are strongly dependent on chemical properties. This reveals the importance of chemical properties in predicting variations of exposure relative to source levels among a range of SVOCs. As permethrin and PBDE-99 are assumed to have their removals driven by cleaning, field data is needed to evaluate the effectiveness of the model for these types of compounds.

Our approach for estimating a key set of exposure-relevant residential parameters offers the opportunity to reduce important contributions to overall uncertainties in household exposure

assessments. We note two important findings. First, the assumption that the mass of dust in any compartment will remain constant over a long period of time makes our indoor-particle mass-balance model less sensitive to input parameters that are measured during short-term experimental periods. Our concern is whether these short-term measurements reflect long-term averages appropriate to the model. Second, the use of three particle size ranges reduces the uncertainties in model prediction by avoiding the mischaracterization of particle transport properties, which vary widely by particle size.

The results here provide useful insights for future work. Based on our observation that some chlorpyrifos air samples increased over the year even though its indoor use was banned many years ago, future observational measurement studies may want to have better control over housing parameters such as air exchange rate, as the change in concentration may have also resulted from differences in the air exchange rate during the sampling period. Model predictions could be improved if the uncertain model input parameters shown in Figures 4 and S2 were measured in the same experimental conditions over a long period of time with the involvement of human activities. Chemicals with low vapor pressure and high octanol-water partition coefficient values were expected to strongly sorb to other interior surfaces including furniture, electronics, fabrics, walls, and ceilings, but the impact of sorption to these surfaces was not included in our model due to the limited partitioning information between the surface j and air (K_{ja}). Thus, accounting for sorption to these interior surfaces in the model may improve future work.

ACKNOWLEDGMENTS

The U.S. Environmental Protection Agency through its Office of Research and Development partially funded and managed the research described here. It has been subjected to Agency administrative review and approved for publication. This research was also funded by the American Chemistry Council (Grant#: 3-DBACC01).

Supporting Information Available

The fraction of the SVOCs in the mobile phases and the residence time based on chemical properties are derived in the Supporting Information. The rationale for parameter selection and estimation, chemical properties, particle loss to walls, and results of uncertainty analysis are also described in the Supporting Information. This material is available free of charge via the Internet at <http://pubs.acs.org>.

LITERATURE CITED

1. Bidleman, T. F. Atmospheric processes - wet and dry deposition of organic-compounds are controlled by their vapor particle partitioning. *Environ. Sci. Technol.* **1988**, 22(4): 361-367.
2. Allen, J. G.; McClean, M. D.; Stapleton, H. M.; Webster, T. F. Critical factors in assessing exposure to PBDEs via house dust. *Environ. Int.* **2008**, 34(8), 1085-1091.
3. Takigami, H.; Suzuki, G.; Hirai Y.; Sakai, S. Transfer of brominated flame retardants from components into dust inside television cabinets. *Chemosphere* **2008**, 73(2), 161-169.
4. Zota, A. R.; Rudel, R. A.; Morello-Forsch, R. A.; Brody, J. G. Elevated house dust and serum concentrations of PBDEs in California: unintended consequences of furniture flammability standards? *Environ. Sci. Technol.* **2008**, 42(21), 8158-8164.
5. Xu, Y.; Hubal, E. A. C.; Little, J. C. Predicting residential exposure to phthalate plasticizer emitted from vinyl flooring: sensitivity, uncertainty, and implications for biomonitoring. *Environ. Health Perspect.* **2010**, 118(2), 253-258.
6. Rudel, R. A.; Camann, D. E.; Spengler, J. D.; Korn, L. R.; Brody, J. G. Phthalates, alkylphenols, pesticides, polybrominated diphenyl ethers, and other endocrine-disrupting compounds in indoor air and dust. *Environ. Sci. Technol.* **2003**, 37(20), 4543-4553.
7. Weschler, C. J.; Nazaroff, W. W. Semivolatile organic compounds in indoor environments. *Atmos. Environ.* **2008**, 42(40), 9018-9040.
8. Weschler, C. J.; Brauer, M.; Koutrakis, P. Indoor ozone and nitrogendioxide - a potential pathway to the generation of nitrate radicals, dinitrogen pentaoxide, and nitric acid indoors. *Environ. Sci. Technol.* **1992**, 26(1), 179-184.
9. Cartwright, C. D.; Thompson, I. P.; Burns, R. G. Degradation and impact of phthalate plasticizers on soil microbial communities. *Environ. Toxicol. Chem.* **2000**, 19(5), 1253-1261.
10. Nalli, S.; Cooper, D. G.; Nicell, J. A. Metabolites from the biodegradation of diester plasticizers by *Rhodococcus rhodochrous*. *Sci. Total Environ.* **2006**, 366(1), 286-294.
11. Hunt, A.; Johnson, D. L.; Brooks, J.; Griffith, D. A. Risk remaining from fine particle contaminants after vacuum cleaning of hard floor surfaces. *Environ. Geochem. Health* **2008**, 30(6), 597-611.

12. Qian, J.; Ferro, A. R.; Fowler, K. R. Estimating the resuspension rate and residence time of indoor particles. *J. Air Waste Manage.* **2008**, 58(4), 502-516.
13. Allott, R. W.; Kelly, M.; Hewitt, C. N. A model of environmental behavior of contaminated dust and its application to determining dust fluxes and residence times. *Atmos. Environ.* **1994**, 28(4), 679-687.
14. Layton, D. W.; Beamer, P. I. Migration of contaminated soil and airborne particulates to indoor dust. *Environ. Sci. Technol.* **2009**, 43(21), 8199-8205.
15. Tolve, N. S.; Egeghy, P. P.; Fortmann, R. C.; Whitaker, D. A.; Nishioka, M. G.; Naeher, L. P.; Hilliard, A. Multimedia measurements and activity patterns in an observational pilot study of nine young children. *J. Expo. Sci. Env. Epid.* **2008**, 18(1), 31-44.
16. Wilson, N. K.; Strauss, W. J.; Iroz-Elardo, N.; Chuang, J. C. Exposures of preschool children to chlorpyrifos, diazinon, pentachlorophenol, and 2,4-dichlorophenoxyacetic acid over 3 years from 2003 to 2005: A longitudinal model. *J. Expo. Sci. Env. Epid.* **2010**, 20(6), 546-558.
17. Quiros-Alcala, L.; Bradman, A.; Nishioka, M.; Harnly, M. E.; Hubbard, A.; McKone, T. E.; Ferber, J.; Eskenazi, B. Pesticides in house dust from urban and farmworker households in California: an observational measurement study. *Environ. Health* **2011**, 10(19).
18. U.S. Environmental Protection Agency. Interim Reregistration eligibility decision for chlorpyrifos, EPA, Washington, DC, **2001**.
19. Bennett, D. H.; Furtaw, E. J. A fugacity-based indoor residential pesticide fate model. *Environ. Sci. Technol.* **2004**, 38, 2142-2152.
20. Slinn, W. G. N.; Hasse, L.; Hicks, B. B.; Hogan, A. W.; Lai, D.; Liss, P. S.; Munnich, K. O.; Sehmel, G. A.; Vittori, O. Some aspects of the transfer of atmospheric trace constituents past the air-sea interface. *Atmos. Environ.* **1978**, 12, 2055-2087.
21. U.S. Environmental Protection Agency. PM 2.5 Objectives and History, EPA, Washington, DC, **2008**.
22. U.S. Environmental Protection Agency. Particulate Matter (PM 10). EPA, Washington, DC, <http://www.epa.gov/airtrends/aqtrnd95/pm10.html>.
23. Choate, L. M.; Ranville, J. F.; Bunge, A. L.; Macalady, D. L. Dermally adhered soil: 2. Reconstruction of dry-sieve particle-size distributions from wet-sieve data. *Integr. Environ. Assess. Manag.* **2006**, 2, 385-390.
24. Fergusson, J. E.; Kim, N. D. Trace elements in street and house dusts - sources and speciation. *Sci. Total Environ.* **1991**, 100, 125-150.

25. Abt, E.; Suh, H. H.; Catalano, P. J.; Koutrakis, P. Relative contribution of outdoor and indoor particle sources to indoor concentrations. *Environ. Sci. Technol.* **2000**, 34(17), 3579-3587.
26. Offenberg, J. H.; Baker, J. E. Aerosol size distributions of elemental and organic carbon in urban and over-water atmospheres. *Atmos. Environ.* **2000**, 34(10), 1509-1517.
27. Thatcher, T. L.; Layton, D. W. Deposition, resuspension, and penetration of particles within a residence. *Atmos. Environ.* **1995**, 29(13), 1487-1497.
28. Abt, E.; Suh, H. H.; Allen, G.; Koutrakis, P. Characterization of indoor particle sources: a study conducted in the metropolitan Boston area. *Environ. Health Perspect.* **2000**, 108(1), 35-44.
29. Long, C. M.; Suh, H. H.; Koutrakis, P. Characterization of indoor particle sources using continuous mass and size monitors. *J. Air Waste Manage.* **2000**, 50(7), 1236-1250.
30. Ferro, A. R.; Kopperud, R. J.; Hildemann, L. M. Elevated personal exposure to particulate matter from human activities in a residence. *J. Expo. Sci. Env. Epid.* **2004**, 14, S34-S40.
31. Chen, Q.; Hildemann, L.M. Size-resolved concentrations of particulate matter and bioaerosols inside versus outside of homes. *Aerosol Sci. Tech.* **2009**, 43(7), 699-713.
32. Riley, W. J.; McKone, T. E.; Lai, A. C. K.; Nazaroff, W. W. Indoor particulate matter of outdoor origin: importance of size-dependent removal mechanisms. *Environ. Sci. Technol.* **2002**, 36(2), 200-207.
33. Kildeso, J.; Vallarino, J.; Spengler, J. D.; Brightman, H. S.; Schneider, T. Dust build-up on surfaces in the indoor environment. *Atmos. Environ.* **1999**, 33(5), 699-707.
34. Corsi, R. L.; Siegel, J. A.; Chiang, C. Particle resuspension during the use of vacuum cleaners on residential carpet. *J. Occ. Environ. Hyg.* **2008**, 5(4), 232-238.
35. Clougherty, J. E.; Houseman, E. A.; Levy, J. I. Source apportionment of indoor residential fine particulate matter using land use regression and constrained factor analysis. *Indoor Air* **2011**, 21(1), 53-66.
36. Leese, K. E.; Hall, R. M.; Cole, E. C.; Foarde, K. K.; Berry, M. A. US EPA and Waste Management Association's International Symposium: Measurement of Toxic and Related Air Pollutants, Durham, NC. **1993**, pp. 82-87.
37. Foarde, K.; Berry, M. Comparison of biocontaminant levels associated with hard vs. carpet floors in nonproblem schools: results of a year long study. *J. Expo. Sci. Env. Epid.* **2004**, 14, S41-S48.

38. Byrne, M. A.; Goddard, A. J. H.; Lange, C.; Roed, J. Stable tracer aerosol deposition measurements in a test chamber. *J. Aerosol Sci.* **1995**, 26(4), 645-653.
39. Wallace, L. A.; Emmerich, S. J.; Howard-Reed, C. Effect of central fans and in-duct filters on deposition rates of ultrafine and fine particles in an occupied townhouse. *Atmos. Environ.* **2004**, 38(3), 405-413.
40. Qian, J.; Ferro, A. R.; Fowler, K. R. Estimating the resuspension rate and residence time of indoor particles. *J. Air Waste Manage.* **2008**, 58(4), 502-516.
41. Chen, C.; Zhao, B. Review of relationship between indoor and outdoor particles: I/O ratio, infiltration factor and penetration factor. *Atmos. Environ.* **2011**, 45(2), 275-288.
42. Long, C. M.; Suh, H. H.; Catalano, P. J.; Koutrakis, P. Using time- and size-resolved particulate data to quantify indoor penetration and deposition behavior. *Environ. Sci. Technol.* **2001**, 35(10), 2089-2099.
43. Chao, C. Y. H.; Wan, M. P.; Cheng, E. C. K. Penetration coefficient and deposition rate as a function of particle size in non-smoking naturally ventilated residences. *Atmos. Environ.* **2003**, 37(30), 4233-4241.
44. Williams, R.; Suggs, J.; Rea, A.; Sheldon, L.; Rhodes, C.; Thornburg, J. The Research Triangle Park particulate matter panel study: modeling ambient source contribution to personal and residential PM mass concentrations. *Atmos. Environ.* **2003**, 37(38), 5365-5378.
45. Wallace, L. Indoor particles: a review. *J. Air Waste Manage.* **1996**, 46(2), 98-126.
46. Tung, T. C. W.; Chao, C. Y. H.; Burnett, J. A methodology to investigate the particulate penetration coefficient through building shell. *Atmos. Environ.* **1999**, 33(6), 881-893.
47. Nishioka, M. G.; Burkholder, H. M.; Brinkman, M. C.; Gordon, S.M.; Lewis, R.G. Measuring transport of lawn-applied herbicide acids from turf to home: correlation of dislodgeable 2,4-D turf residues with carpet dust and carpet surface residues. *Environ. Sci. Technol.* **1996**, 30(11), 3313-3320.
48. Fortune, C. R.; Blanchard, F. T.; Ellenson, W. D. Analysis of aged in-home carpeting to determine the distribution of pesticide residues between dust, carpet, and pad compartments, EPA-NERL, **2000**.
49. Ozkaynak, H.; Xue, J.; Spengler, J.; Wallace, L.; Pellizzari, E.; Jenkins, P. Personal exposure to airborne particles and metals: results from the particle team study in Riverside, California. *J. Expo. Sci. Env. Epid.* **1996**, 6(1), 57-78.

- 570 50. Fan, C. W.; Zhang, J. J. Characterization of emissions from portable household
571 combustion devices: particle size distributions, emission rates and factors, and potential
572 exposures. *Atmos. Environ.* **2001**, 35(7), 1281-1290.
- 573
- 574 51. Olson, D. A.; Burke, J. M. Distributions of PM_{2.5} source strengths for cooking from the
575 Research Triangle Park particulate matter panel study. *Environ. Sci. Technol.* **2006**, 40(1),
576 163-169.
- 577
- 578 52. Ramaswami, A.; Milford, J.B.; Small, M.J. *Integrated Environmental Modeling-Pollutant*
579 *Transport, Fate, and Risk in the Environment*, John Wiley & Sons. Inc.: Hoboken, New
580 Jersey, 2005.
- 581
- 582 53. Hertz-Picciotto, I.; Cassady, D.; Lee, K.; Bennett, D. H.; Ritz, B.; Vogt, R. Study of Use
583 of Products and Exposure-Related Behaviors (SUPERB): study design, methods, and
584 demographic characteristics of cohorts. *Environ. Health.* **2010**, 9(54).
- 585
- 586 54. Nazaroff, W. W.; Wescher C. J. Cleaning products and air fresheners: exposure to primary
587 and secondary air pollutants. *Atmos. Environ.* **2004**, 38, 2841-2865.
- 588
- 589 55. Zhang X.; Diamond M. L.; Ibarra C.; Harrad S. Multimedia modeling of polybrominated
590 diphenyl ether emissions and fate indoors. *Environ. Sci. Technol.* **2009**, 43, 2845-2850.

TABLES

Table 1. Properties of Dust in Three Size Fraction Ranges

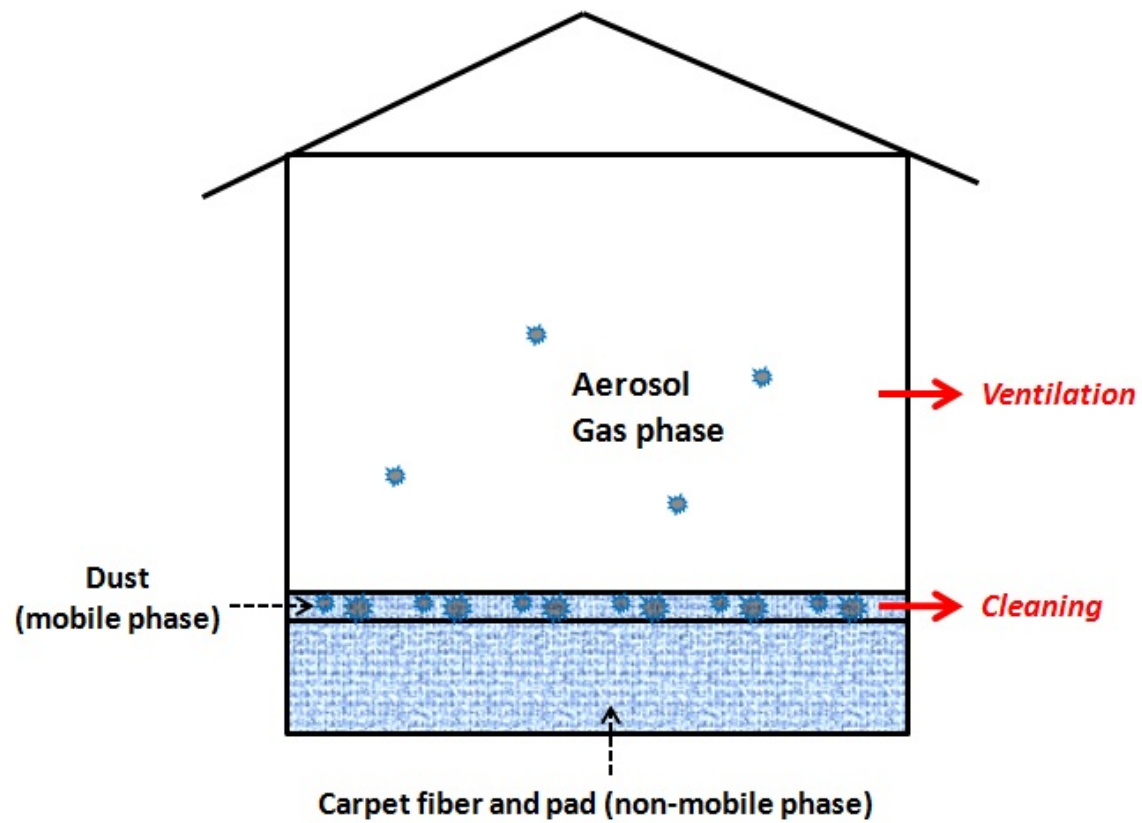
Property name (units)	symbol	Particle size ranges (μm)			References
		0 - 2.5 Mean (CV)	2.5 - 10 Mean (CV)	10 - 150 Mean (CV)	
Fraction of organic carbon (--)	f_{oc}	0.65 (0.30)	0.28 (0.30)	0.06 (0.30)	[26]
Indoor air particle concentration (mg/m ³)	C_i	1.3×10^{-2} (0.58)	1.2×10^{-2} (0.88)	4.3×10^{-3} (0.75)	[27-35]
Outdoor air particle concentration (mg/m ³)	C_o	7.3×10^{-3} (0.65)	6.9×10^{-3} (0.41)	1.6×10^{-2} (0.59)	[27-31]
Particle concentration on carpet (mg/m ²)	C_c	20 (0.30)	61 (0.30)	300 (0.30)	[27,36,37]
Particle concentration on vinyl (mg/m ²)	C_v	3 (0.30)	6 (0.30)	49 (0.30)	[27,36,37]
Deposition loss rate (1/d)	D	9 (0.47)	34 (0.36)	166 (1.00)	[27,29,32,38,39]
Resuspension rate (1/d)	R	4.3×10^{-4} (0.50)	2.7×10^{-3} (0.50)	4.8×10^{-3} (1.00)	[27,40]
Penetration efficiency (--)	P	0.82 (0.13)	0.65 (0.35)	0.10 (0.50)	[41-46]
Track-in to carpet (mg/d)	T_c	10 (0.70)	31 (0.70)	171 (0.70)	[14,27,47]
Track-in to vinyl (mg/d)	T_v	1.0 (0.70)	3.1 (0.70)	17.1 (0.70)	[14,27,47]
Initial removal rate from carpet cleaning (1/d)	k_c	5.0×10^{-3} (1.00)	5.0×10^{-3} (1.00)	5.0×10^{-3} (1.00)	[14,48]
Initial removal rate from vinyl cleaning (1/d)	k_v	2.5×10^{-2} (1.00)	2.5×10^{-2} (1.00)	2.5×10^{-2} (1.00)	[14,48]
Initial emission rate to indoor air (mg/d)	S	75 (1.00)	20 (1.00)	-	[49-51]
<i>Removal rate from carpet cleaning (1/d) ^a</i>	k_c	1.8×10^{-2} (0.69)	1.8×10^{-2} (0.91)	5.0×10^{-3} (1.64)	
<i>Removal rate from vinyl cleaning (1/d) ^a</i>	k_v	1.1×10^{-1} (0.79)	1.8×10^{-1} (0.98)	4.5×10^{-2} (1.25)	
<i>Emission rate to indoor air (mg/d) ^a</i>	S	104 (1.27)	188 (1.35)	93.4 (3.18)	

^a Re-adjusted values from particle mass balances.

Table 2. Summary of Mass Distributions, Removals, Initial Concentration Changes after 1 Year, and Residence Times for Five SVOCs

	Diazinon	Chlorpyrifos	Permethrin	PBDE-47	PBDE-99
Mass distribution (%)					
air	0.01	0.003	0.00001	0.001	0.0001
air particles	0.00002	0.00003	0.001	0.0002	0.0003
carpet + pad + deeply embedded particles	98.7	98.8	93.0	97.8	97.4
carpet particles	0.1	0.2	6.6	1.3	2.0
vinyl flooring	1.2	1.0	0.4	0.8	0.6
vinyl particles	0.001	0.001	0.04	0.01	0.01
Removal (%)					
ventilation	99.1	96.0	19.9	57.5	24.3
carpet cleaning	0.9	3.9	76.9	40.9	72.8
vinyl cleaning	0.0	0.2	3.1	1.7	3.0
Initial concentration change after 1 year (%)					
Carpet cleaning only	0.4	0.6	18.7	3.9	5.8
Ventilation only	36.5	13.0	4.2	5.2	1.9
Ventilation + carpet cleaning	36.8	13.5	22.9	9.1	7.7
Equilibrium Residence time (year)	2.2	6.9	3.8	10.5	12.5
Steady state residence time (year)	6.7	19	3.9	16.2	13.7

FIGURES



TOC and Abstract Art

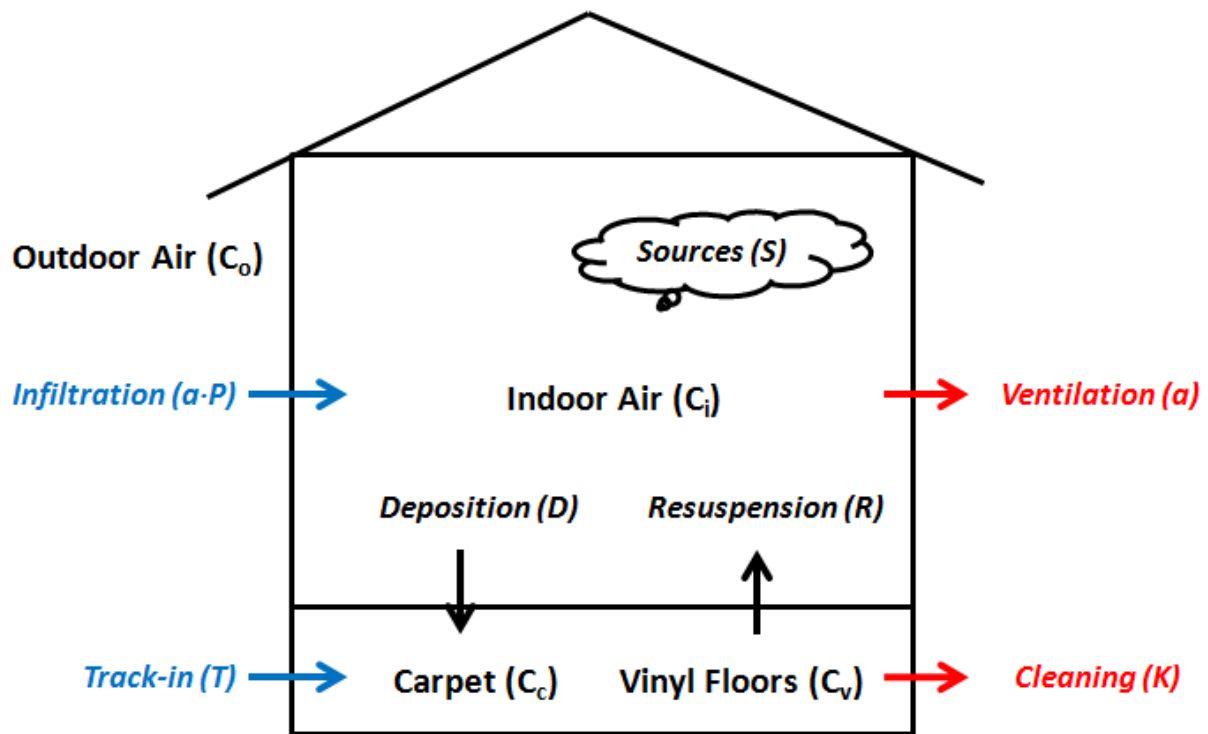


Figure 1. Model framework for indoor dust model.

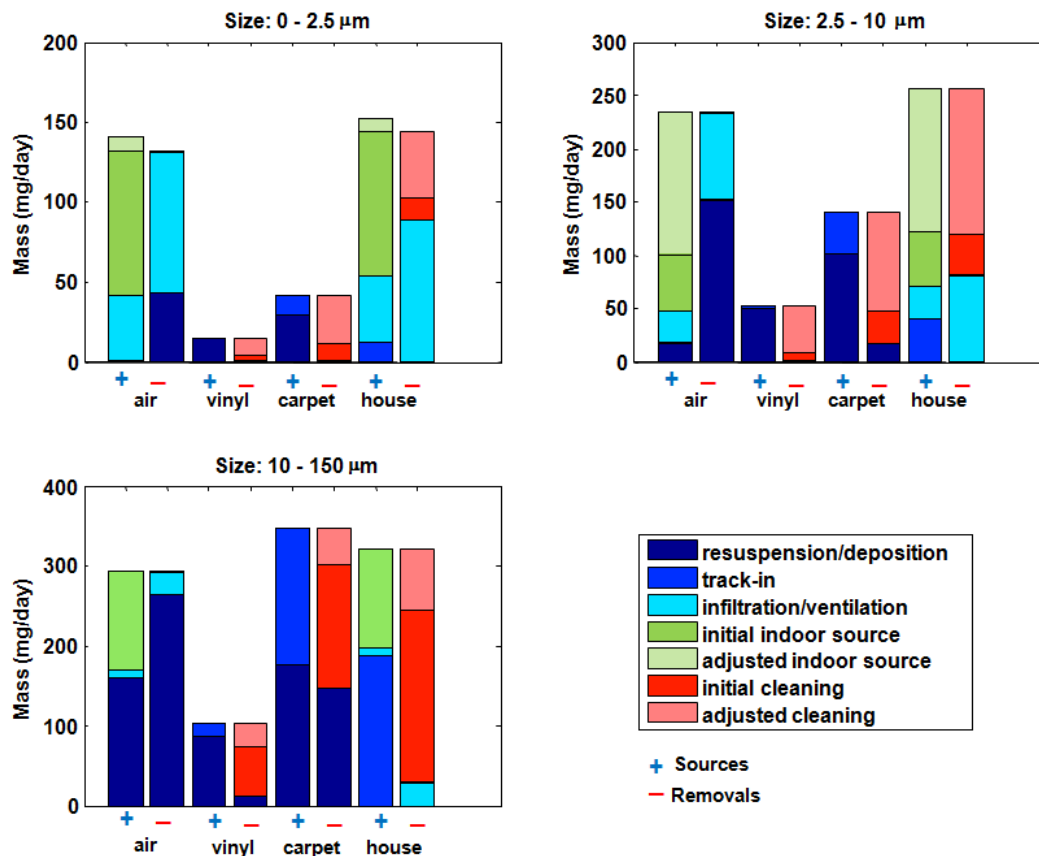


Figure 2. Summary of the average daily particle mass balance in the three size classes with (+) indicating sources and (-) removals. Indoor sources and cleaning-rate removals have been adjusted up or down relative to measurements of dust to achieve steady-state; these adjustments are indicated by lighter shading.

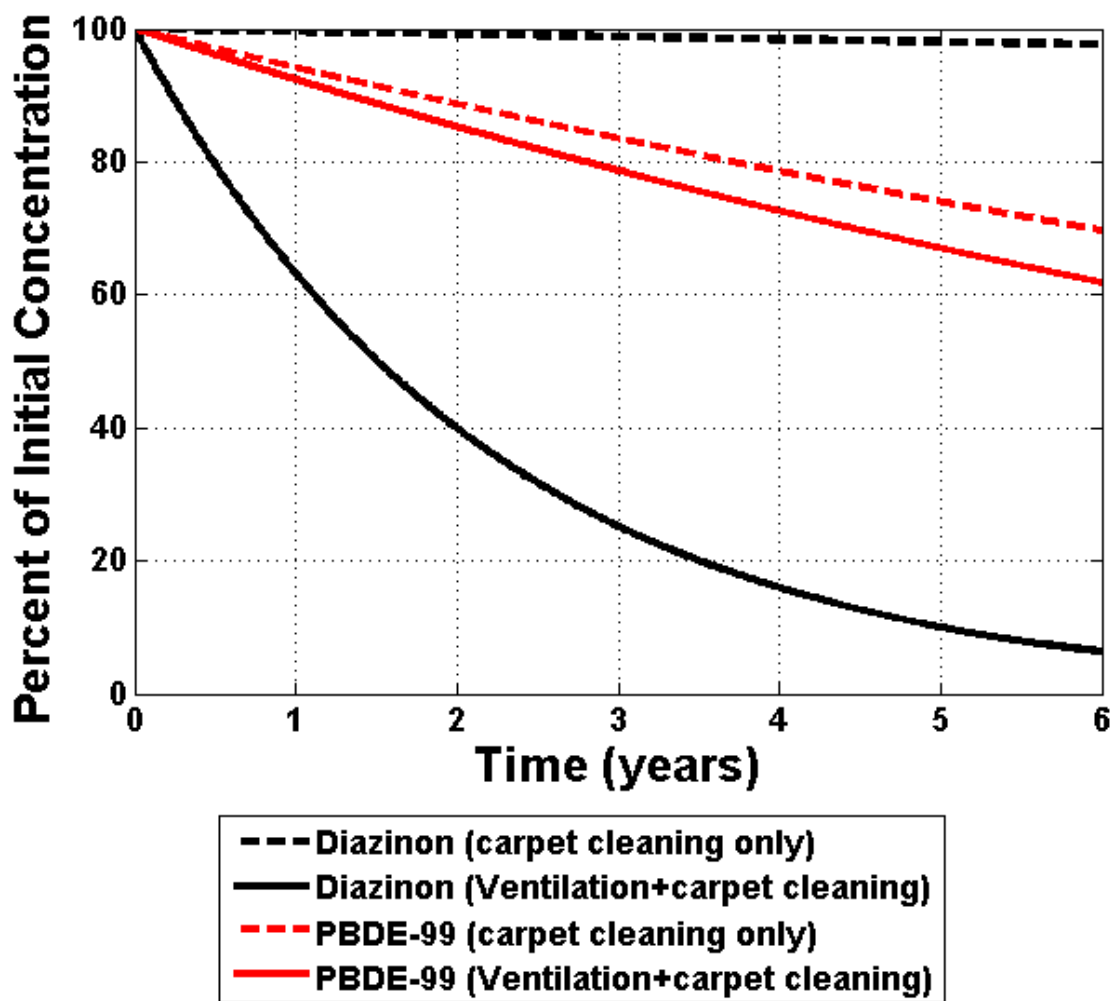


Figure 3. Concentration change of diazinon and PBDE-99 indoors as a result of different residence times from ventilation and carpet cleaning. The gaps between solid lines and dotted lines indicate the relative contribution of ventilation on SVOCs removals.

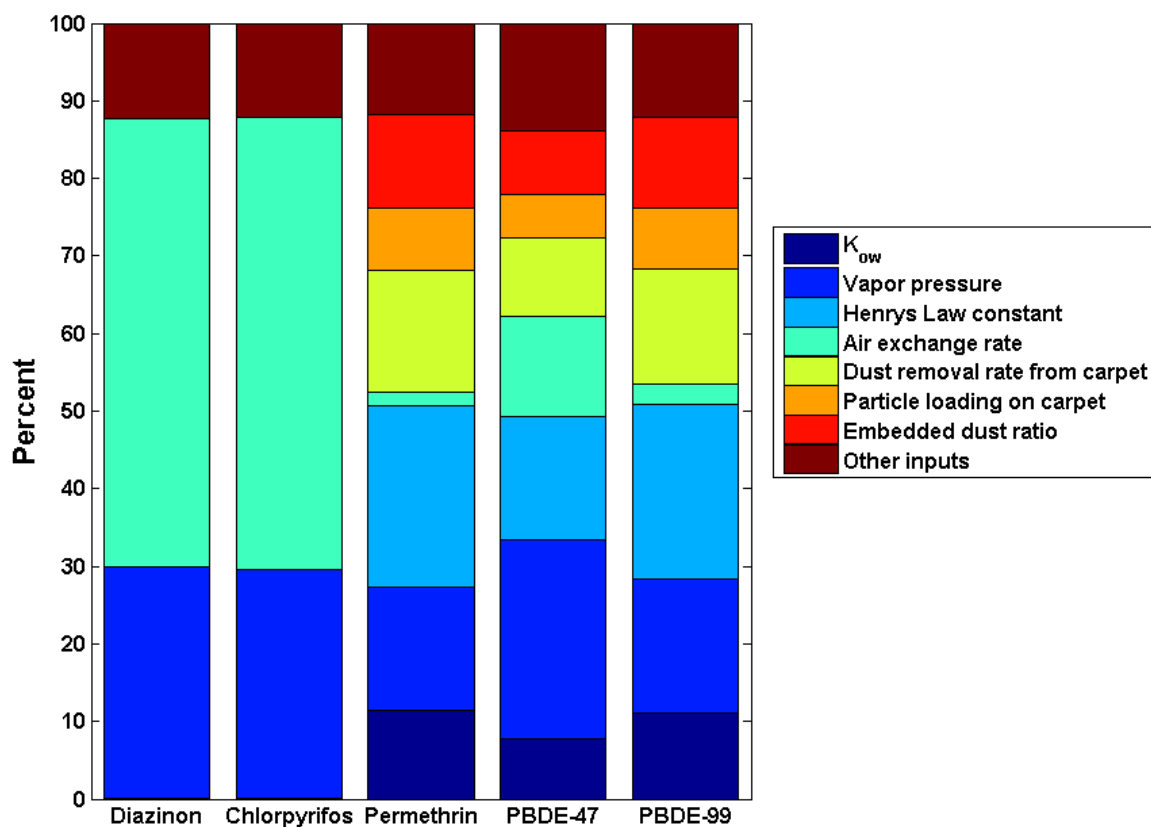


Figure 4. Percent contribution of model inputs on the uncertainty of residence times for diazinon, chlorpyrifos, permethrin, PBDE-47, and PBDE-99.

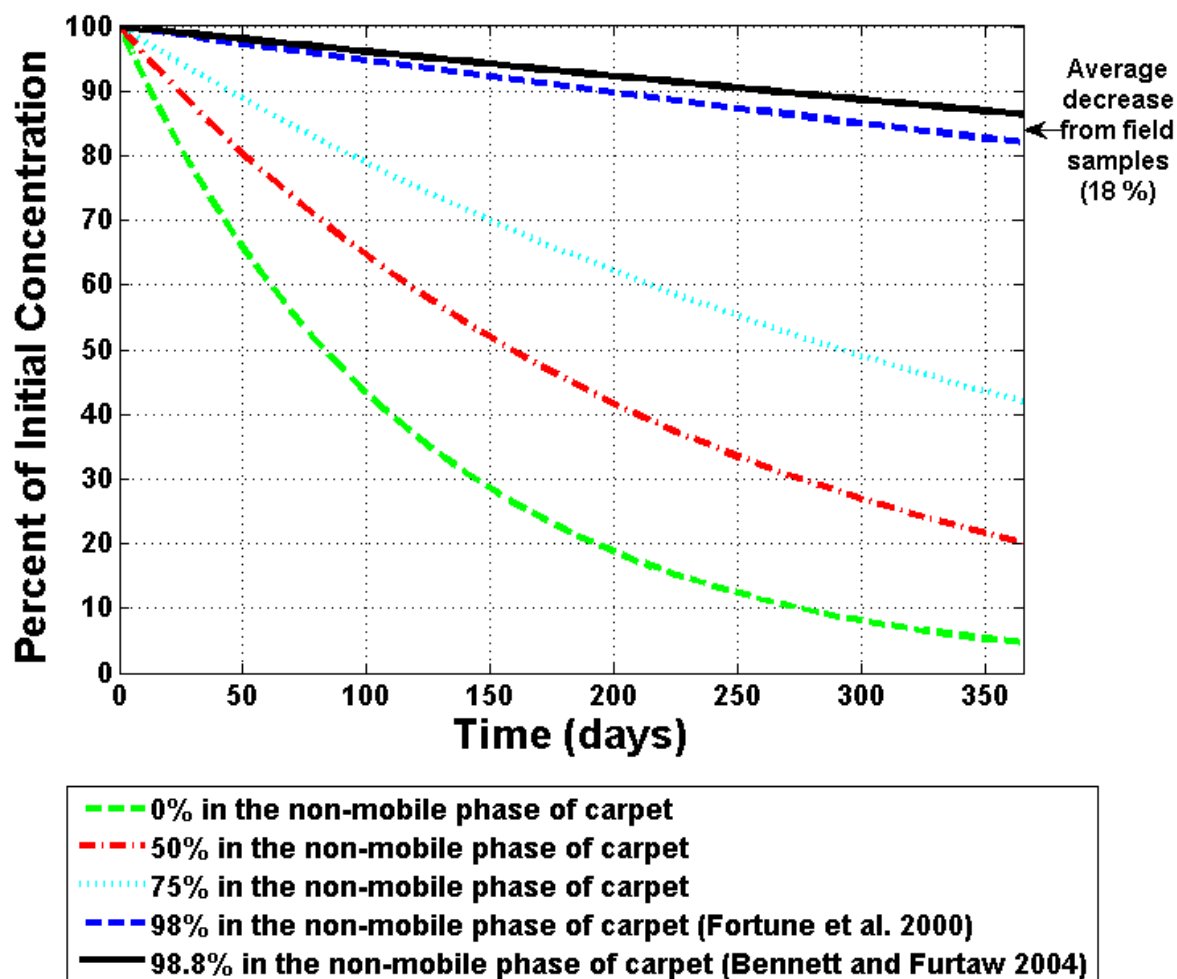


Figure 5. Chlorpyrifos concentration changes in the indoor environments by different fractions in the non-mobile phase of carpet.

Brief

Residence times are estimated by chemical partitioning between the mobile and non-mobile phases and the removal rates resulting from air exchange and cleaning.