UV Filters and Tourism: Their Impact On The Environment Charlita G. Rosal, L. Don Betowski, *Emily M. Siska

U.S. Environmental Protection Agency, Office of Research and Development, 944 E. Harmon Avenue, Las Vegas, NV 89119, rosal-charlita@epa.gov - Student Contract Services **CONTRACT**

1. Background

- Ultraviolet (UV) filters are widely used in cosmetics, sunscreens, and plastics to block UV radiation from the sun.
- · Parabens are common ingredients in sunscreen formulation and thus are included in the list of target analytes. They are used as preservatives and anti-bacterial agents.
- Studies show that some UV filters and parabens demonstrate estrogenicity and multiple hormonal activities in vitro1.
- · Potential impact of these materials on the environment and human health is of great concern.
- · Because of the high consumption volume and high lipophilicity of many UV filters, these compounds have the potential to enter and persist in the environment and to bioaccumulate in tissues of living organisms.
- People from all over the world visit Las Vegas, bringing with them their own medications and personal care products (e.g., cosmetics, sunscreens).
- Use of these materials contributes loading in the environment.
- To date, there are only 15 U.S. FDA-approved, organic UV filters, while European countries, Australia, and Japan sanction additional compounds²

2. Purpose of Current Work

- Develop methods to monitor the presence of UV filters and parabens in wastewater treatment plant effluent and other environmental compartments
- · Monitor and evaluate potential sources of and temporal variation in the occurrence of these chemicals
- · Determine if certain chemical loadings can be used as indicators of tourist influx in the Las Vegas Valley.



3. Experimental

- · Water samples are going to be collected from effluent of the Clark County Reclamation District in-situ thru SPE cartridges (Biotage Evolut® ABN and Waters Oasis® HLB, 1g)
- · Solvent elution of analytes were performed using FMS Solid Phase Extraction (SPE) system
- All HPLC-MS/MS analyses were performed using a Thermo Finnigan (San Jose, CA) HPLC Surveyor® interfaced to a Thermo Finnigan TSQ Quantum Ultra AM[®] triple quadrupole mass spectrometer (AM3QMS).
- · Analytical separation column was a Pursuit[®] diphenyl, 3.5-µm particle size, 2.1-mm x 150-mm column (Varian Inc., Walnut Creek, CA) housed in an oven at 30°C. The mobile phase at 200 µL/min consisted of 0.5% acetic acid, 100% acetonitrile, and 100% methanol in gradient elution. Methanol was introduced at constant flow rate of 10% of the total flow
- All GC-MS analyses were performed with a Varian 3800 capillary gas chromatograph and 320 mass spectrometer (Varian, Inc, Walnut Creek, CA) using electron ionization and single-ion monitoring (SIM) mode. The gas chromatograph was fitted with a 30-m x 0.25-mm ID fused silica capillary column coated with a 0.10-µm film of crossbonded 5% phenyl - 95% methyl polysiloxane (DB-5HT, J&W Scientific, Folsom, CA). The injector temperature was at 250°C. A pulsed pressure of 174 kPa (25 psi) was applied during injection and held for 0.75 min. After the initial pressure pulse, the carrier gas flow was held constant at 1 mL min-1. The initial oven temperature was 60°C held for 1 min, ramped at 30°C min-1 to 150°C, then to 290°C at 8°C min-1 and and held for 3.0 min (total chromatographic time = 24.5 min)

Instrumentation



mo Finnigan HPLC Surveyor® coupled with an TSO Õu m I lltra AM[®] triple qu

4. Results

Compound Chemical Formula (Molecular Weight)	CAS Reg. No.	Collision Energy (V)	LC-MS/MS GC-MS	LC-MS/MS GC-MS	LC-MS/MS Retention Time	GCMS Retention Time	Country Permitt in
Phenyl salicylate C ₁ :H ₂₀ , (214)	118-55-8	18 18	213 (M-H) 215 (M+H)	93 (C6H5O) 121 (C6H5COO)		8.275	
Benzylsalicylate C_H_2O,	118-58-1	20 18	227 (M-H) 229 (M+H)	93 (C6H5O) 151 [(M+H)-C6H6] ⁻		9.544	
(228) 2-Ethylbexyl salicylate C_H_O,	118-60-5	22	91 249 (M-H)	228, 92, 65 137 [(M-H)-C8H16]		8.89	EC, USA, AUS, JP, P CAN, RSA, ROK
(250)			120	138, 121, 250			
p-Aminobenzoic acid (PABA) C,H,NO ₂ (137)	150-13-0	19 19	136 (M-H) 138 (M+H)'	92 [(M-H)-CO2]' 77 (C6H5)'			EC, USA, AUS, JP, P CAN, SAf, ROK
Homosalate (3,3,5-Trimethykyclohexyl salicylate) C ₁₀ H ₂₀ ,	118-56-9	21	261 (M-H)	137 (C7H5O3)		9.484	EC, USA, AUS, JP, C PRC, RSA, ROK
(262)			138	109, 120, 262			
Uxybenzone (Benzophenone-s) (2-Hydroxy-4-methoxybenzophenone) C ₁₁ H ₁₁ O, (228)	131-57-7	20	227 (M-H) 229 (M+H) 227	211 [(M-H)-CH4] 151 [(M+H)-C6H6] 151, 228, 77	15.15		EC, USA, AUS, JP, C CHI, RSA, ROK
Ecamsule/Mexoryl SX (Terephthalylidene dicamphor sulfonic acid) C_sH_0_PS_	90457-82-2	27	280 (M-2H)2	437 (M-2H-SO, -C,H,)	10.54		EC, USA, AUS, JP, C PRC, RSA
(302) 2-phenyl-5-benzimidazole-sulfonic acid (Ensulizole) C ₁ H _a N ₂ O ₂ S	27503-81-7	28 31	273 (M-H) 275 (M+H)	193 [(M-H)-SO3] 194 [(M+H)-SO3H]'	2.94		EC, USA, AUS, JP, C PRC, RSA, ROK
(274) 4-Hydroxy benzoic acid-C6 C,H ₄ O,		16	143 (M-H)	99 [(M-H)-COO]	3.24		
(144) 4-4'-dihydroxybenzophenone C ₁ ,H ₂ ,O ₁	611-99-4	33 22	213 (M-H) 215 (M+H)	93 (C6H4OH) 121 [(M+H) – C ₄ H ₄ OH] [.]	5.38	14.362	
(214) Matheleurobae	99.76.2	20	121 151 (M H):	214, 93, 70 97 (C M O)	6.25	\$7/2	
C,H,O,	11100			74 (C 44 0)			
(152) Methyl paraben-C6		21	121 157 (M-H)	152, 93, 65 98 (C,H,O)	5.37	6.1	
(158)			127	158, 99			
Benzophenone-2 (2,2',4,4'-Tetrahydroxybenzophenone) C ₁ ,H ₁₀ O, (246)	131-55-5	19 13	245 (M-H) 247 (M+H)	135 [(M-H)-C6H6O2] 137 [(M+H) -C6H6O2]	6.8		AU, JP, RSA
Benzophenone-4	4065-45-6	32	307 (M-H)	211 [(M-H)-SO3H-CH3]	7.82		EC, USA, AUS, JP, C
(suitsobenzone) (2. Hydroxy-4-methoxybenzophenone-5- sulfonic acid) C_nH_nO_S (308)							PRC, KSA, KUK
Ethylparaben C _x H _a O _x (166)	120-47-8	21	165 (M-H)	92 (C ₄ H ₂ O) 166, 138, 93	7.25	6.253	
2,4,4'-trihydroxybenzophenone	1470-79-7	26	229 (M-H)	93 (C6H5O)	7.76	15.46	
C ₁ ,H ₁₀ O ₁ (230)		18	137	230, 121, 229			
Propylparaben	94-13-3	22	179 (M-H)	92 (C _a H ₂ O)	10.06	7.075	
(180)			121	138, 93, 180	1		
4-hydroxybenzophenone (metabolite) C ₁ ,H ₂₀ O ₂ (198)	1137-42-4	34 19	197 (M-H) 199 (M+H) 121	92 [(M-H)-C6H5CO] 121 [(M+H)-C ₄ H ₄] 198, 77, 105	10.41	10.986	
n-Butyl paraben-C6 (200)		23	199 (M-H)	98 (C ₄ H ₂ O)	12.19	8.03	
Butylparaben	94-26-8	22	127 193 (M-H)	144, 200 92 (C,H,O)	12.19	8.032	
C_H_O,			121	178 101 07	-		
Benzophenone-1 (2,4-Dihydroxybenzophenone) C ₁ ,H ₂₀ O ₁	131-56-6	28 21	213 (M-H) 215 (M+H)	91 (C6H3O) 137 [(M+H)-C6H6]	12.45	12.032	JP, RSA
(214) Benzylparaben C.,H.,O,	94-18-8	20	137 227 (M-H)	213, 214, 77 92 (C ₄ H ₂ O)	13.39	12.275	
(228) 3-(4-Methylbenzylidene) camphor C ₁₀ H ₂₀ O	36861-47-9	18	121 255 (M+H)*	91, 228, 65 212 [(M+H)-C3H7]*	18.14	11.527	EC, USA, AUS, CAN, RSA, ROK
(254) Meathyl anthranilate	134.09.8		254 276 (MaH)	128, 171, 211 138 (C6B4/COOH)NH3U	19.4	17 156	USA AUS CAN R
C,H _a NO,	1.54-0.7-0		137	119 120 275		12.150	ROK
(2:5) Octyldimethyl PABA (2-Ethylhexyl-4-dimethylaminobenzoate) C_H_NO,	21245-02-3	25	278 (M+H)'	166 [(M+H)-C _a H _a]*	19.43	13.698	EC, USA, AUS, JP, C PRC, RSA, ROK
(277) Octyl methoxycinnamate C_H.,O,	5466-77-3	9 15	165 291 (M+H) 289 (M-H)	164, 148, 277 179 [(M+H) – C ₄ H ₄₀] ⁻ 229	19.56	14.199	EC, USA, AUS, JP, C PRC, RSA, ROK
(290)			178	161, 290, 177	1		
Octocrylene (2-Ethylhexyl-2-cyano-3,3 diphenylacrylate) C ₁₀ H ₂₁ NO ₂	6197-30-4	12	362 (M+H)' 112	250 [(M+H)-C8H16-NH3] 249, 204, 360	20.84	17.54	EC, USA, AUS, JP, C PRC, RSA, ROK
(301) Bis-(2-Ethylhexyl)adipate-C6		14	377 (M+H)*	135	20.97	14.59	
(376) Ausbenzone	70356-007	20	135 311 (MeH)	153,112, 247	17 27		EC USA AUX ID C
(butyl methoxydibenzoylmethane)	10336-09-1	20	.41 (M+R)	rog (m ³ trin ³)	1/-57		PRC, RSA, ROK

[3] <u>http://www</u>

Acknowledgement - We thank FMS, Inc. for loaning us the SPE system and Biotage for the supply



· Both p-Amino benzoic acid and Mexoryl SX are problematic compounds for the current method used

· Avobenzone produced two peaks, which is a result of compound transformation due to several solvents used for dissolution. A new batch of neat standard will be analyzed for

Initial recovery experiments in DI water gave the following preliminary results:

LC-MS/MS analysis:

Benzophenones: 82 - 156% Parabens: 31 - 45% Camphor: 52 -74%

Sulfonic acids: 0% except for Benzophenone-4, which was at 44-60% Esters: 0 - 17%

· Poor recovery of esters maybe due to hydrolysis.

· GC-MS analysis of same sample extracts as above generally resulted in lower recoveries as compared to LC-MS/MS analysis.

5. Conclusions & Future Work

• LC-MS/MS and GC-MS methods have been developed to identify sunscreen agents and parabens in solvent matrix.

• Two SPE materials (Biotage Evolut® ABN and Waters Oasis® HLB) are being evaluated for optimum recovery of target analytes.

· Extraction methods need to be optimized for maximum recovery of analytes.

· Water samples from the effluent of CCWRD main facility will commence this winter and will continue thru fall of next year

· Water samples from the effluent of the City of Las Vegas Wastewater Treatment Facility will also be collected.

• Biosolids are considered as another matrix to sample in the near future.

• Research collaboration with CCWRD will be established to leverage resources.

References

[1] Schlumpf, M., Cotton, B., Conscience, M., Haller, V., Steinmann, B. and Lichtensteiger, W. (2001). "In Vitro and in Vivo Estrogenicity of UV Screens " Environmental Health Perspectives 109(3): 239-244 [2] Shaath, N. A. (2007). The Encyclopedia of Ultraviolet Filters. Shaath, N. A. Carol Stream, IL, Allured Publishing Corp.