



EPA/635/R-14/194a  
www.epa.gov/iris

# **Evaluation of the Inhalation Carcinogenicity of Ethylene Oxide**

(CASRN 75-21-8)

## **In Support of Summary Information on the Integrated Risk Information System (IRIS)**

*August 2014*

### **NOTICE**

This document is a **Revised External Peer Review draft**. This information is distributed solely for the purpose of pre-dissemination peer review under applicable information quality guidelines. It has not been formally disseminated by EPA. It does not represent and should not be construed to represent any Agency determination or policy. It is being circulated for review of its technical accuracy and science policy implications.

National Center for Environmental Assessment  
Office of Research and Development  
U.S. Environmental Protection Agency  
Washington, DC

## **DISCLAIMER**

This document is a preliminary draft for review purposes only. This information is distributed solely for the purpose of pre-dissemination peer review under applicable information quality guidelines. It has not been formally disseminated by EPA. It does not represent and should not be construed to represent any Agency determination or policy. Mention of trade names or commercial products does not constitute endorsement or recommendation for use.

*This document is a draft for review purposes only and does not constitute Agency policy.*

## NOTE TO REVIEWERS

A draft of this assessment received external peer review by EPA's Science Advisory Board (SAB) and was being revised according to the peer review recommendations when the 2011 National Research Council (NRC) report was released with general recommendations for improvements to the IRIS process. As noted in the 2011 report, the NRC encouraged EPA to proceed with assessments while developing improvements to the IRIS Program. Consistent with this advice, EPA has indicated that it would not go backwards in the assessment development process, but would focus on moving forward, while also phasing in improvements.

While the ethylene oxide (EtO) assessment does not incorporate all the revisions to the IRIS assessment format and methodology recommended in the 2011 NRC recommendations (and the more recent 2014 NRC Review of the IRIS Process), such as the inclusion of a standard Preamble, systematic review and standardized approaches for evidence integration, this assessment is streamlined, and uses tables, figures, and appendices to increase transparency and clarity. In addition, the assessment is structured to have separate hazard identification and dose-response sections and the update to the literature search was conducted using systematic literature search approaches. Additionally, consistent with the goal that assessments should provide a scientifically sound and transparent evaluation of the relevant scientific literature and presentation of the analyses performed, this assessment contains an expanded discussion on the rationales for study evaluation and selection, as well as other key assessment decisions. Appendix K documents where the recommendations from Chapter 7 of the NRC 2011 report have been implemented in this assessment.

EPA obtained public comment on an external review draft in 2006 ([U.S. EPA, 2006a](#)) and completed a peer review by a panel of EPA's Science Advisory Board (SAB) in 2007 ([SAB, 2007](#)). A summary of the public and peer review comments and EPA's disposition of these comments is presented in Appendix H of the current revised draft assessment. The consensus conclusions of the SAB review supported the conclusions in the draft external review assessment regarding the cancer classification and the selection of the key dataset for quantification of cancer risk estimates. The SAB recommended that EPA examine several issues further, such as endogenous EtO production, whether an alternative dataset would add useful exposure-response information, and whether the primary epidemiology dataset could be modeled using continuous data across the full exposure range without first converting the exposure and individual occurrence data into categorical data for the derivation of unit risk estimates. The SAB also provided comments on both sides of the issue of whether or not a nonlinear low-dose

*This document is a draft for review purposes only and does not constitute Agency policy.*

extrapolation approach should be included in the analysis with two conflicting views articulated in the Appendices to the SAB's final report.

Because of the importance of this assessment, the complex issues in modeling epidemiology data, and the new modeling of epidemiology data done in response to the prior SAB peer review, EPA decided to seek additional SAB peer review. Prior to the additional external peer review, EPA released a revised draft for public comment in July 2013. The revised draft was discussed during an IRIS Bimonthly Public Science meeting in December 2013. A summary of public comments, and EPA's responses are available in Appendix L.

EPA did not formally record the details of literature search strategies employed in identifying the relevant EtO literature for the development of the 2006 external peer review draft. There were no critical studies identified as missing during public comment and peer review, but the 2007 SAB final report recommended inclusion of additional supporting studies. EPA revised the assessment according to the recommendations of the peer review panel, including extensive new modeling of epidemiologic data, and incorporated new studies that were identified through June 2010 into the revised assessment. As indicated above, EPA decided to seek additional SAB peer review primarily because of the new modeling of epidemiologic data done in response to the SAB recommendations. Therefore, in May 2013, in order to ensure that no critical studies had been missed that would warrant major revisions to the assessment, EPA conducted a well-documented systematic literature search of literature published from January 2006 through May 2013. The literature search strategy was conducted and documented following the 2011 NRC recommendations for more formal systematic literature searches. Relevant studies that could potentially impact the cancer hazard characterization or dose-response assessment were identified and considered. Appendix J provides documentation of the search methods, the bases for the judgments of the relevancy of new literature, and the disposition of studies identified in the 2013 search (Section J.1). Appendix J also includes reviews of two major studies published after June 2010 that were identified in the search and merited in-depth discussion (Section J.2). Two additional studies of potential importance to the outcome of the assessment, both published after the May 2013 search, were noted in public comments in October 2013 and are also reviewed in Appendix J (Section J.3). None of the additional studies that were identified were found to have an impact on the final conclusions of the assessment. A list of the 131 references added after the 2006 external peer review can be found in Appendix I. References considered and cited in this document, including abstracts, can be found on the Health and Environmental Research Online (HERO) website.<sup>1</sup>

---

<sup>1</sup>HERO is a database of scientific studies and other references used to develop EPA's risk assessments, which are aimed at understanding the health and environmental effects of pollutants and chemicals. HERO is developed and  
*This document is a draft for review purposes only and does not constitute Agency policy.*

Development of the hazard identification and dose-response assessments for EtO has followed the general guidelines for risk assessment as set forth by the National Research Council ([NRC, 1983](#)). United States Environmental Protection Agency (U.S. EPA) Guidelines and Risk Assessment Forum Technical Panel Reports that were used in the development of this assessment include the following: *Guidelines for Mutagenicity Risk Assessment* ([U.S. EPA, 1986](#)), *Methods for Derivation of Inhalation Reference Concentrations and Application of Inhalation Dosimetry* ([U.S. EPA, 1994](#)), *Benchmark Dose Technical Guidance* ([U.S. EPA, 2012](#)), *Science Policy Council Handbook: Risk Characterization* ([U.S. EPA, 2000](#)), *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)), *Supplemental Guidance for Assessing Susceptibility from Early-Life Exposure to Carcinogens* ([U.S. EPA, 2005b](#)), and *Science Policy Council Handbook: Peer Review* ([U.S. EPA, 2006b](#)).

---

managed in EPA's Office of Research and Development (ORD) by the National Center for Environmental Assessment (NCEA). The database includes more than 1,000,000 scientific articles from the peer-reviewed literature. New studies are added continuously to HERO.

*This document is a draft for review purposes only and does not constitute Agency policy.*

# CONTENTS

LIST OF TABLES .....	viii
LIST OF FIGURES .....	x
LIST OF ABBREVIATIONS.....	xi
AUTHORS, CONTRIBUTORS, AND REVIEWERS .....	xii
1. EXECUTIVE SUMMARY .....	1-1
2. INTRODUCTION .....	2-1
2.1. LITERATURE IDENTIFICATION.....	2-2
2.2. NRC RECOMMENDATIONS OF 2011 .....	2-3
3. HAZARD IDENTIFICATION.....	3-1
3.1. EVIDENCE OF CANCER IN HUMANS.....	3-1
3.1.1. Conclusions Regarding the Evidence of Cancer in Humans.....	3-12
3.2. EVIDENCE OF CANCER IN LABORATORY ANIMALS .....	3-19
3.2.1. Conclusions Regarding the Evidence of Cancer in Laboratory Animals.....	3-23
3.3. SUPPORTING EVIDENCE.....	3-23
3.3.1. Metabolism and Kinetics.....	3-23
3.3.2. Protein Adducts .....	3-26
3.3.3. Genotoxicity .....	3-28
3.4. MODE OF ACTION.....	3-38
3.4.1. Analysis of the Mode of Action for Ethylene Oxide Carcinogenicity under EPA’s Mode-of-Action Framework .....	3-42
3.5. HAZARD CHARACTERIZATION .....	3-45
3.5.1. Characterization of Cancer Hazard .....	3-45
3.5.2. Susceptible Life stages and Populations .....	3-49
4. CANCER DOSE-RESPONSE ASSESSMENT FOR INHALATION EXPOSURE.....	4-1
4.1. INHALATION UNIT RISK ESTIMATES DERIVED FROM HUMAN DATA.....	4-1
4.1.1. Risk Estimates for Lymphohematopoietic Cancer .....	4-4
4.1.2. Risk Estimates for Breast Cancer .....	4-22
4.1.3. Total Cancer Risk Estimates .....	4-44
4.1.4. Sources of Uncertainty in the Cancer Risk Estimates .....	4-46
4.1.5. Summary .....	4-58
4.2. INHALATION UNIT RISK DERIVED FROM EXPERIMENTAL ANIMAL DATA.....	4-59
4.2.1. Overall Approach .....	4-59
4.2.2. Cross-Species Scaling .....	4-60
4.2.3. Dose-Response Modeling Methods.....	4-62
4.2.4. Description of Experimental Animal Studies.....	4-64
4.2.5. Results of Data Analysis of Experimental Animal Studies.....	4-65

*This document is a draft for review purposes only and does not constitute Agency policy.*

## CONTENTS (continued)

4.3. SUMMARY OF INHALATION UNIT RISK ESTIMATES—NOT ACCOUNTING FOR ASSUMED INCREASED EARLY-LIFE SUSCEPTIBILITY .....	4-67
4.4. ADJUSTMENTS FOR POTENTIAL INCREASED EARLY-LIFE SUSCEPTIBILITY .....	4-68
4.5. INHALATION UNIT RISK ESTIMATES—CONCLUSIONS .....	4-74
4.6. COMPARISON WITH OTHER PUBLISHED RISK ESTIMATES .....	4-82
4.6.1. Unit Risk Estimates Based on Human Studies.....	4-82
4.6.2. Unit Risk Estimates Based on Laboratory Animal Studies.....	4-85
4.7. RISK ESTIMATES FOR SOME OCCUPATIONAL EXPOSURE SCENARIOS .....	4-85
REFERENCES .....	R-1
APPENDIX A. CRITICAL REVIEW OF EPIDEMIOLOGIC EVIDENCE .....	A-1
APPENDIX B. REFERENCES FOR FIGURE 3-3 .....	B-1
APPENDIX C. GENOTOXICITY AND MUTAGENICITY OF ETHYLENE OXIDE .....	C-1
APPENDIX D. REANALYSES AND INTERPRETATION OF ETHYLENE OXIDE EXPOSURE-RESPONSE DATA .....	D-1
APPENDIX E. LIFE-TABLE ANALYSIS .....	E-1
APPENDIX F. EQUATIONS USED FOR WEIGHTED LINEAR REGRESSION.....	F-1
APPENDIX G. MODEL PARAMETERS IN THE ANALYSIS OF ANIMAL TUMOR INCIDENCE .....	G-1
APPENDIX H. SUMMARY OF 2007 EXTERNAL PEER REVIEW AND PUBLIC COMMENTS AND DISPOSITION .....	H-1
APPENDIX I. LIST OF REFERENCES ADDED AFTER THE 2006 EXTERNAL REVIEW DRAFT .....	I-1
APPENDIX J. SUMMARY OF MAJOR NEW STUDIES SINCE THE LITERATURE CUTOFF DATE .....	J-1
APPENDIX K. DOCUMENTATION OF IMPLEMENTATIONS OF THE 2011 NATIONAL RESEARCH COUNCIL RECOMMENDATIONS .....	K-1
APPENDIX L. SUMMARY OF PUBLIC COMMENTS RECEIVED ON THE JULY 2013 PUBLIC COMMENT DRAFT AND EPA RESPONSES.....	L-1

*This document is a draft for review purposes only and does not constitute Agency policy.*

## LIST OF TABLES

Table 1-1. Summary of major findings.....	1-7
Table 3-1. Epidemiological studies of ethylene oxide and human cancer—lymphohematopoietic cancer results <sup>a</sup> .....	3-13
Table 3-2. Summary of epidemiological results on EtO and female breast cancer (all sterilizer workers) <sup>a</sup> .....	3-17
Table 3-3. Tumor incidence data in National Toxicology Program Study of B6C3F <sub>1</sub> mice NTP (1987) <sup>a</sup> and exposure-response modeling results <sup>b</sup> .....	3-21
Table 3-4. Tumor incidence data in Lynch et al. (1984a; 1984b) study of male F344 rats and exposure-response modeling results .....	3-22
Table 3-5. Tumor incidence data in Snellings et al. (1984) and Garman et al. (1985) reports on F344 rats <sup>a</sup> and exposure-response modeling results <sup>b</sup> .....	3-24
Table 3-6. Cytogenetic effects in humans.....	3-33
Table 4-1. Considerations used in this assessment for selecting epidemiology studies for quantitative risk estimation .....	4-3
Table 4-2. Cox regression results for all lymphohematopoietic cancer and lymphoid cancer mortality in both sexes in the NIOSH cohort, for the models presented by Steenland et al. (2004) .....	4-6
Table 4-3. Exposure-response modeling results for all lymphohematopoietic cancer and lymphoid cancer mortality in both sexes in the NIOSH cohort for models not presented by Steenland et al. (2004) .....	4-12
Table 4-4. Models considered for modeling the exposure-response data for lymphoid cancer mortality in both sexes in the NIOSH cohort for the derivation of unit risk estimates.....	4-14
Table 4-5. EC <sub>01</sub> , LEC <sub>01</sub> , and unit risk estimates for lymphoid cancer <sup>a</sup> .....	4-16
Table 4-6. EC <sub>01</sub> , LEC <sub>01</sub> , and unit risk estimates for all lymphohematopoietic cancer <sup>a</sup> .....	4-19
Table 4-7. Cox regression results for breast cancer mortality in females in the NIOSH cohort, <sup>a</sup> for models presented in Steenland et al. (2004) .....	4-23
Table 4-8. Exposure-response modeling results for breast cancer mortality in females in the NIOSH cohort for models not presented by Steenland et al. (2004) .....	4-28
Table 4-9. EC <sub>01</sub> , LEC <sub>01</sub> , and unit risk estimates for breast cancer mortality in females <sup>a</sup> .....	4-29

*This document is a draft for review purposes only and does not constitute Agency policy.*

Table 4-10. Cox regression results for breast cancer incidence in females from the NIOSH cohort, for the models presented by Steenland et al. (2003) <sup>a,b</sup> .....	4-31
Table 4-11. Exposure-response modeling results for breast cancer incidence in females from the NIOSH cohort for models not presented by Steenland et al. (2003).....	4-37
Table 4-12. Models considered for modeling the exposure-response data for breast cancer incidence in females in the subcohort with interviews from the NIOSH incidence study cohort for the derivation of unit risk estimates .....	4-39
Table 4-13. EC <sub>01</sub> , LEC <sub>01</sub> , and unit risk estimates for breast cancer incidence in females— invasive and in situ <sup>a</sup> .....	4-41
Table 4-14. Calculation of EC <sub>01</sub> for total cancer risk .....	4-45
Table 4-15. Calculation of total cancer unit risk estimate .....	4-45
Table 4-16. Upper-bound unit risks (per µg/m <sup>3</sup> ) obtained by combining tumor sites .....	4-64
Table 4-17. Unit risk values from multistage Weibull <sup>a</sup> time-to-tumor modeling of mouse tumor incidence in the NTP (1987) study.....	4-66
Table 4-18. Summary of unit risk estimates (per µg/m <sup>3</sup> ) in animal bioassays .....	4-67
Table 4-19. EC <sub>01</sub> , LEC <sub>01</sub> , and unit risk estimates for adult-only exposures* .....	4-70
Table 4-20. Calculation of EC <sub>01</sub> for total cancer (incidence) risk from adult-only exposure* .....	4-71
Table 4-21. Calculation of total cancer unit risk estimate from adult-only exposure*.....	4-71
Table 4-22. Adult-based unit risk estimates for use in ADAF calculations and risk estimate calculations involving less-than-lifetime exposure scenarios .....	4-73
Table 4-23. Adult-based extra risk estimates per ppm based on adult-exposure-only EC <sub>01</sub> s (0.01/EC <sub>01</sub> estimates) <sup>a</sup> .....	4-79
Table 4-24. Summary of key unit risk estimates from this assessment (see Section 4.7 for risk estimates based on occupational exposure scenarios) .....	4-81
Table 4-25. Comparison of unit risk estimates <sup>a</sup> .....	4-83
Table 4-26. Extra risk estimates for lymphoid cancer in both sexes for various occupational exposure levels <sup>a</sup> .....	4-89
Table 4-27. Extra risk estimates for breast cancer incidence in females for various occupational exposure levels <sup>a,b</sup> .....	4-92

*This document is a draft for review purposes only and does not constitute Agency policy.*

## LIST OF FIGURES

Figure 3-1. Metabolism of ethylene oxide.....	3-25
Figure 3-2. Simulated blood AUCs for EtO following a 6-hour exposure to EtO from the rat, mouse, and human PBPK models of Fennell and Brown (2001); based on data presented in Fennell and Brown (2001). (Rat1 and rat2 results use different values for pulmonary uptake.).....	3-27
Figure 3-3. Display of 203 data sets, including bacteria, fungi, plants, insects, and mammals (in vitro and in vivo), measuring the full range of genotoxic endpoints. [This is an updated version of the figure in IARC (1994b)] .....	3-29
Figure 4-1. RR estimate for lymphoid cancer vs. mean exposure (with 15-year lag, unadjusted for continuous exposure). .....	4-9
Figure 4-2. RR estimate for all lymphohematopoietic cancer vs. mean exposure (with 15-year lag, unadjusted for continuous exposure). .....	4-18
Figure 4-3. RR estimate for breast cancer mortality vs. mean exposure (with 20-year lag, unadjusted for continuous exposure). .....	4-25
Figure 4-4. RR estimate for breast cancer incidence in full cohort vs. mean exposure (with 15-year lag, unadjusted for continuous exposure).....	4-33
Figure 4-5. RR estimate for breast cancer incidence in subcohort with interviews vs. mean exposure (with 15-year lag, unadjusted for continuous exposure).....	4-34
Figure 4-6. RR estimate for breast cancer incidence in subcohort with interviews vs. mean exposure (with 15-year lag, unadjusted for continuous exposure); select models compared to deciles. ....	4-40
Figure 4-7. RR estimates for lymphoid cancer from occupational EtO exposures (with 15-year lag). .....	4-90
Figure 4-8. RR estimates for breast cancer incidence from occupational EtO exposures (with 15-year lag).....	4-95

*This document is a draft for review purposes only and does not constitute Agency policy.*

## LIST OF ABBREVIATIONS

ADAF	age-dependent adjustment factor
AIC	Akaike information criterion
AIDS	acquired immune deficiency syndrome
AML	acute myeloid leukemia
AUC	areas under the curve
BEIR	Committee on the Biological Effects of Ionizing Radiation
CI	confidence interval
DSB	double-strand breaks
EC	effective concentration
EOIC	Ethylene Oxide Industry Council
EPA	U.S. Environmental Protection Agency
EtO	ethylene oxide
FRG	Federal Republic of Germany
GST	glutathione S-transferase
HAP	hazardous air pollutants
N7-HEG	N7-(2-hydroxyethyl)guanine
IARC	International Agency for Research on Cancer
ICD	International Classification of Diseases
IRIS	Integrated Risk Information System
LEC	lower confidence limit
MLE	maximum likelihood estimate
NCEA	National Center for Environmental Assessment
NHL	non-Hodgkin lymphoma
NIOSH	National Institute for Occupational Safety and Health
NTP	National Toxicology Program
O <sup>6</sup> -HEG	O <sup>6</sup> -hydroxyethylguanine
OBS	observed number
OR	odds ratios
PBPK	physiologically based pharmacokinetic
POD	point of departure
RR	relative rate, i.e., rate ratio
SCE	sister chromatid exchanges
SE	standard error
SEER	Surveillance, Epidemiology, and End Results
SIR	standardized incidence ratio
SMR	standard mortality ratios
TWA	time-weighted average
UCC	Union Carbide Corporation
UCL	upper confidence limit
WHO	World Health Organization

*This document is a draft for review purposes only and does not constitute Agency policy.*

## **AUTHORS, CONTRIBUTORS, AND REVIEWERS**

### **ASSESSMENT AUTHORS AND CONTRIBUTORS**

Jennifer Jinot (Chemical Manager)  
National Center for Environmental Assessment  
U.S. Environmental Protection Agency  
Washington, DC

David Bayliss (retired)  
National Center for Environmental Assessment  
U.S. Environmental Protection Agency  
Washington, DC

Henry D. Kahn (retired)  
National Center for Environmental Assessment  
U.S. Environmental Protection Agency  
Washington, DC

Nagu Keshava  
National Center for Environmental Assessment  
U.S. Environmental Protection Agency  
Washington, DC

Robert McGaughy (retired)  
National Center for Environmental Assessment  
U.S. Environmental Protection Agency  
Washington, DC

Kyle Steenland (under contract to EPA)  
Rollins School of Public Health  
Emory University  
Atlanta, Georgia

Ravi Subramaniam  
National Center for Environmental Assessment  
U.S. Environmental Protection Agency  
Washington, DC

Larry Valcovic (retired)  
National Center for Environmental Assessment  
U.S. Environmental Protection Agency  
Washington, DC

*This document is a draft for review purposes only and does not constitute Agency policy.*

Suryanarayana Vulimiri  
National Center for Environmental Assessment  
U.S. Environmental Protection Agency  
Washington, DC

## **REVIEWERS**

Earlier drafts of this document were provided for review to EPA scientists, interagency reviewers from other federal agencies and the Executive Office of the President, the public, and independent scientists external to EPA. Summaries and EPA's dispositions of the comments received from the independent external peer reviewers and from the public are included in Appendices H (for the 2006 external review draft) and L (for the 2013 public comment draft).

## **INTERNAL EPA REVIEWERS**

Michele Burgess, OSWER  
Deborah Burgin, OPEI  
Carol Christensen, OPP/HED  
Kerry Dearfield, ORD/OSP (no longer with EPA)  
Joyce Donohue, OW  
Rebecca Dzubow, AO/OCHP  
Michael Firestone, AO/OCHP  
Linnea Hansen, OPP  
Karen Hogan, ORD/NCEA-IRIS  
Ray Kent, OPP/HED  
Aparna Koppikar, ORD/NCEA-W (retired)  
Tim Leighton, OPP/AD  
Tim McMahon, OPP/AD  
David Miller, OPP/HED  
Deirdre Murphy, OAR/ESD  
Steve Nesnow, ORD/NHEERL (retired)  
Marian Olsen, Region 2  
Brenda Perkovich-Foos, AO/OCHP  
Julian Preston, ORD/NHEERL (retired)  
Santhini Ramasamy, OPP/HED, OW  
Elissa Reaves, OPP/HED  
Nancy Rios-Jafolla, Region 3  
Tracey Woodruff, AO/NCEE (no longer with EPA)

The authors would like to acknowledge David Bussard, Jane Caldwell, Catherine Gibbons, Linda Phillips, Julian Preston, Cheryl Scott, Maria Spassova, and Paul White of EPA for their contributions during the draft development process.

*This document is a draft for review purposes only and does not constitute Agency policy.*

## **EXTERNAL PEER REVIEWERS**

### **SCIENCE ADVISORY BOARD ETHYLENE OXIDE REVIEW PANEL (2007)**

#### **CHAIR**

Dr. Stephen Roberts  
University of Florida

#### **OTHER SAB MEMBERS**

Dr. Timothy Buckley  
Ohio State University

Dr. Montserrat Fuentes  
North Carolina State University

Dr. Dale Hattis  
Clark University

Dr. James Kehrer  
Washington State University

Dr. Mark Miller  
California Environmental Protection Agency

Dr. Maria Morandi  
University of Texas—Houston Health Science Center

Dr. Robert Schnatter  
Exxon Biomedical Sciences, Inc.

Dr. Anne Sweeney  
TAMU System Health Science Center

*This document is a draft for review purposes only and does not constitute Agency policy.*

## **CONSULTANTS SERVING ON THE PANEL**

Dr. Steven Alan Belinsky  
University of New Mexico

Dr. Norman Drinkwater  
University of Wisconsin Medical School

Dr. Steven Heeringa  
University of Michigan

Dr. Ulrike Luderer  
University of California

Dr. James Swenberg  
University of North Carolina

Dr. Vernon Walker  
Lovelace Respiratory Research Institute

*This document is a draft for review purposes only and does not constitute Agency policy.*

## 1. EXECUTIVE SUMMARY

Ethylene oxide (EtO) is a gas at room temperature. It is manufactured from ethylene and used primarily as a chemical intermediate in the manufacture of ethylene glycol. It is also used as a sterilizing agent for medical equipment and as a fumigating agent for spices.

The DNA-damaging properties of EtO have been studied since the 1940s. EtO is known to be mutagenic in a large number of living organisms, ranging from bacteriophage to mammals, and it also induces chromosome damage. It is carcinogenic in mice and rats, inducing tumors of the lymphohematopoietic system, brain, lung, connective tissue, uterus, and mammary gland. In humans employed in EtO-manufacturing facilities and in sterilizing facilities, the greatest evidence of a cancer risk from exposure is for cancer of the lymphohematopoietic system. Increases in the risk of lymphohematopoietic cancer have been seen in most (but not all) of the epidemiological studies of EtO-exposed workers, manifested as an increase either in leukemia or in cancer of the lymphoid tissue. Of note, in one large epidemiologic study conducted by the National Institute for Occupational Safety and Health (NIOSH) of sterilizer workers that had a well-defined exposure assessment for individuals, positive exposure-response trends were reported for lymphohematopoietic cancer mortality, primarily in males and in particular for lymphoid cancer (i.e., non-Hodgkin lymphoma, myeloma, and lymphocytic leukemia), and for breast cancer mortality in females ([Steenland et al., 2004](#)). The positive exposure-response trend for female breast cancer was confirmed in an incidence study based on the same worker cohort ([Steenland et al., 2003](#)). There is supporting evidence for an association between EtO and breast cancer from other studies, but the database is more limited than that for lymphohematopoietic cancers.

Although the evidence of carcinogenicity from human studies was deemed short of conclusive on its own, EtO is characterized as “carcinogenic to humans” by the inhalation route of exposure based on the total weight of evidence, in accordance with EPA’s 2005 *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)). The lines of evidence supporting this characterization include: (1) strong, but less than conclusive on its own, epidemiological evidence of lymphohematopoietic cancers and breast cancer in EtO-exposed workers, (2) extensive evidence of carcinogenicity in laboratory animals, including lymphohematopoietic cancers in rats and mice and mammary carcinomas in mice following inhalation exposure, (3) clear evidence that EtO is genotoxic and sufficient weight of evidence to support a mutagenic mode of action for EtO carcinogenicity, and (4) strong evidence that the key precursor events are anticipated to occur in humans and progress to tumors, including evidence of chromosome

1 damage in humans exposed to EtO. Overall, there is strong confidence in the hazard  
2 characterization of EtO as “carcinogenic to humans.”

3 This document describes the derivation of inhalation unit risk estimates for cancer  
4 mortality and incidence based on the human data from the NIOSH study ([Steenland et al., 2004](#);  
5 [Steenland et al., 2003](#)). This study was selected for the derivation of risk estimates because it is  
6 a high-quality study,<sup>2</sup> it is the largest of the available studies, and it has exposure estimates for  
7 the individual workers from a high-quality exposure assessment. Multiple modeling approaches  
8 were evaluated for the exposure-response data, including modeling the cancer response as a  
9 function of either categorical exposures or continuous individual exposure levels. Preferred  
10 approaches were defined for each cancer endpoint in consideration of both the statistical  
11 properties and biological reasonableness of the resulting model forms (see Tables 4-4 and 4-12  
12 for a summary of models investigated in this assessment for lymphoid cancer and breast cancer  
13 incidence, respectively, and the considerations used in model selection).

14 Unit risk estimates based on the human data were first derived under the common  
15 assumption that relative risk is independent of age. This assumption is later superseded by an  
16 assumption of increased early-life susceptibility, and it is the unit risk estimates derived under  
17 this latter assumption that are the ultimate estimates proposed in this assessment (presented  
18 further below).

19 Under the assumption that relative risk is independent of age, an LEC<sub>01</sub> (lower 95%  
20 confidence limit on the EC<sub>01</sub>, the estimated effective concentration associated with 1% extra risk)  
21 was calculated using a life-table analysis and linear modeling of the categorical Cox regression  
22 analysis results for excess lymphoid cancer mortality [[Steenland et al. \(2004\)](#); additional results  
23 for both sexes combined provided in Appendix D] excluding the highest exposure group to  
24 mitigate the supralinearity of the exposure-response data.<sup>3</sup> Linear low-dose extrapolation below  
25 the range of observations is supported by the conclusion that a mutagenic mode of action is  
26 operative in EtO carcinogenicity. Linear low-dose extrapolation from the LEC<sub>01</sub> for lymphoid  
27 cancer mortality yielded a lifetime extra cancer unit risk estimate of  $2.2 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  (4.0

---

<sup>2</sup> The NIOSH study (Steenland et al., 2003, 2004) was judged to be a “high-quality” study based on the attributes discussed in Section 3.1 and in Section A.2.8 of Appendix A, including availability of individual worker exposure estimates from a high-quality exposure assessment, cohort study design, large size, inclusion of males and females, adequate follow-up, absence of any known confounding exposures, and use of internal comparisons. The breast cancer incidence study using the subcohort of female workers with interviews had the additional attribute of investigating and controlling for a number of breast cancer risk factors (Steenland et al., 2003).

<sup>3</sup> A variety of continuous exposure model forms were considered; however, the exposure-response data for lymphoid cancer are too supralinear (i.e., the exposure-response relationship is very steep in the lower exposure range and then attenuates at higher exposures) to obtain useful models from the best-fitting continuous exposure models (Section 4.1.1.2).

*This document is a draft for review purposes only and does not constitute Agency policy.*

1  $\times 10^{-4}$  per ppb)<sup>4</sup> of continuous EtO exposure. Applying the same linear regression coefficient  
2 and life-table analysis to background lymphoid cancer *incidence* rates and applying linear  
3 low-dose extrapolation resulted in a preferred lifetime extra lymphoid cancer unit risk estimate  
4 of  $4.8 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  ( $8.8 \times 10^{-4}$  per ppb), as cancer incidence estimates are generally  
5 preferred over mortality estimates.

6 Using the same approach,<sup>5</sup> a unit risk estimate of  $2.8 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  ( $5.1 \times 10^{-4}$  per  
7 ppb) was derived from the breast cancer mortality results of the same epidemiology study  
8 ([Steenland et al., 2004](#)). Breast cancer incidence risk estimates, on the other hand, were  
9 calculated from the data from a breast cancer incidence study of the same occupational cohort  
10 ([Steenland et al., 2003](#)), and, for these data, a two-piece linear spline model was used for the  
11 exposure-response modeling (the two-piece linear spline model was the best-fitting model of the  
12 continuous exposure models considered for the breast cancer incidence data; see Section 4.1.2.3).  
13 Using the same life-table approach and linear low-dose extrapolation, a unit risk estimate of  $9.5$   
14  $\times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  ( $1.7 \times 10^{-3}$  per ppb) was obtained for breast cancer incidence. Again, the  
15 incidence estimate is preferred over the mortality estimate. Combining the incidence risk  
16 estimates for the two cancer types resulted in a total cancer unit risk estimate of  $1.2 \times 10^{-3}$  per  
17  $\mu\text{g}/\text{m}^3$  ( $2.3 \times 10^{-3}$  per ppb).<sup>6</sup>

18 Unit risk estimates were also derived from the three chronic rodent bioassays for EtO  
19 reported in the literature. These estimates, ranging from  $2.2 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$  to  $4.6 \times 10^{-5}$  per  
20  $\mu\text{g}/\text{m}^3$ , are over an order of magnitude lower than the estimates based on human data. The  
21 Agency takes the position that human data, if adequate data are available, provide a more  
22 appropriate basis than rodent data for estimating population risks ([U.S. EPA, 2005a](#)), primarily  
23 because uncertainties in extrapolating quantitative risks from rodents to humans are avoided.  
24 Although there is a sizeable difference between the rodent-based and the human-based estimates,  
25 the human data are from a large, high-quality study, with EtO exposure estimates for the  
26 individual workers and little reported exposure to chemicals other than EtO. Therefore, the  
27 estimates based on the human data are the preferred estimates for this assessment.

28 Because the weight of evidence supports a mutagenic mode of action for EtO  
29 carcinogenicity, and as there are no chemical-specific data from which to assess early-life  
30 susceptibility, increased early-life susceptibility should be assumed, according to EPA's

---

<sup>4</sup>Conversion equation: 1 ppm = 1830  $\mu\text{g}/\text{m}^3$ .

<sup>5</sup> Several continuous exposure model forms were considered; however, the exposure-response data for breast cancer mortality were too supralinear (i.e., the exposure-response relationship is very steep in the lower exposure range and then attenuates at higher exposures) to obtain useful continuous exposure models (Section 4.1.2.2); thus, as for lymphoid cancer, a linear regression of the categorical results, excluding the highest exposure group, was used.

<sup>6</sup> The method used to derive the total cancer unit risk estimate involves estimating an upper bound on the sum of the maximum likelihood estimates of risk; see Section 4.1.3.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 *Supplemental Guidance for Assessing Susceptibility from Early-Life Exposure to*  
2 *Carcinogens*—hereinafter referred to as “EPA’s *Supplemental Guidance*” ([U.S. EPA, 2005b](#)).  
3 This mode-of-action-based assumption of increased early-life susceptibility supersedes the  
4 assumption of age independence under which the human-data-based estimates presented above  
5 were derived. Thus, using the same approach as for the estimates discussed above but initiating  
6 exposure in the life-table analysis at age 16 instead of at birth, adult-exposure-only unit risk  
7 estimates were calculated from the human data for selected exposure-response models under an  
8 alternate assumption that relative risk is independent of age for adults, which represent the life  
9 stage for which the data upon which the exposure-response modeling was conducted pertain.  
10 These adult-exposure-only unit risk estimates were then rescaled to a 70-year basis for use in the  
11 standard age-dependent adjustment factors (ADAF) calculations and risk estimate calculations  
12 involving less-than-lifetime exposure scenarios. The resulting adult-based unit risk estimates  
13 were  $4.3 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  ( $7.9 \times 10^{-4}$  per ppb) for lymphoid cancer incidence,  $8.2 \times 10^{-4}$  per  
14  $\mu\text{g}/\text{m}^3$  ( $1.5 \times 10^{-3}$  per ppb) for breast cancer incidence in females, and  $1.1 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$  ( $2.0$   
15  $\times 10^{-3}$  per ppb) for both cancer types combined. The adult-based unit risk estimates, which were  
16 derived under an assumption of increased early-life susceptibility, supersede those presented  
17 earlier that were derived under the assumption that RR is independent of age. When using the  
18 adult-based unit risk estimates to estimate extra cancer risks for a given exposure scenario, the  
19 standard ADAFs should be applied, in accordance with EPA’s *Supplemental Guidance* ([U.S.](#)  
20 [EPA, 2005b](#)). Applying the ADAFs to obtain a full lifetime total cancer unit risk estimate yields  
21  $1.8 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$  ( $3.3 \times 10^{-3}$  per ppb), and the commensurate lifetime chronic (lower-bound)  
22 exposure level of EtO corresponding to an increased cancer risk of  $10^{-6}$  is  $0.0006 \mu\text{g}/\text{m}^3$  (0.0003  
23 ppb).

24 The major sources of uncertainty in the unit risk estimates derived from the human data  
25 include the low-dose extrapolation, the retrospective exposure assessment conducted for the  
26 epidemiology study, and the exposure-response modeling of the epidemiological data (see  
27 Section 4.1.4 for a discussion of these and other sources of uncertainty in the unit risk estimates).

28 Although there are uncertainties in the unit risk estimate, confidence in the estimate is  
29 relatively high. First, there is strong confidence in the hazard characterization of EtO as  
30 “carcinogenic to humans,” which is based on strong epidemiological evidence supplemented by  
31 other lines of evidence. Second, the unit risk estimate is based on human data from a large,  
32 high-quality epidemiology study with individual worker exposures estimated using a  
33 high-quality regression model. Finally, the use of low-exposure linear extrapolation is strongly  
34 supported by the conclusion that EtO carcinogenicity has a mutagenic mode of action.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 Confidence in the unit risk estimate is particularly high for the breast cancer component,  
2 the largest contributor to the total cancer unit risk estimate, which is based on over 200 incident  
3 cases for which the investigators had information on other potential breast cancer risk factors.  
4 The selected model for the breast cancer incidence data was the best-fitting model of the models  
5 investigated as well as the model which provided the best representation of the categorical  
6 results, particularly in the lower exposure range of greatest relevance for the derivation of a unit  
7 risk estimate. Alternate estimates calculated from other reasonable models suggest that a unit  
8 risk estimate for breast cancer incidence fourfold lower (corresponding to a total cancer unit risk  
9 estimate of twofold lower) is plausible; however, unit risk estimates notably lower than that are  
10 considered unlikely from the available data (Section 4.1.2.3).

11 There is somewhat less, although still relatively high in general, confidence in the  
12 lymphoid cancer component of the unit risk estimate because it is based on fewer events (40  
13 lymphoid cancer deaths); incidence risk was estimated from mortality data; and the  
14 exposure-response relationship is exceedingly supralinear, such that the best-fitting continuous  
15 exposure models yield apparently implausibly steep low-exposure slopes. Although these  
16 continuous exposure models provided statistically significant slope coefficients, there was low  
17 confidence in such steep slopes, which, particularly for the two-piece spline model, are highly  
18 dependent on a small number of cases in the low-exposure range. Thus, a linear regression  
19 model of the categorical results for the lowest three quartiles was used to derive the unit risk  
20 estimate for lymphoid cancer, and there was greater confidence in the more moderate slope  
21 resulting from that model, although it was not statistically significant, because it was based on  
22 more data and provided a good representation of the categorical results across this larger data  
23 range in the lower exposure region. (The most suitable alternative model of those based on the  
24 continuous exposure data yielded a unit risk estimate about twofold higher than the estimate  
25 from the linear regression model of the categorical results.)<sup>7</sup> So, while there is less confidence in  
26 the lymphoid cancer unit risk estimate than in the breast cancer unit risk estimate, the lymphoid  
27 cancer estimate is considered a reasonable estimate from the available data, and overall, there is  
28 relatively high confidence in the total cancer unit risk estimate.

29 The unit risk estimate is intended to provide a reasonable upper bound on cancer risk.  
30 The estimate was developed for environmental exposure levels (it is considered valid for  
31 exposures up to 140  $\mu\text{g}/\text{m}^3$  [75 ppb]) and is not applicable to higher-level exposures, such as may

---

<sup>7</sup>The most suitable alternative model of those based on the continuous exposure data was the two-piece log-linear spline model with the knot at 1,600 ppm  $\times$  days, although it was not statistically significant and it was not the two-piece log-linear model with the maximum likelihood, which had the knot at 100 ppm  $\times$  days, but rather one based on a local maximum likelihood with respect to the changes in the value of the knot (see Section 4.1.1.2 and Figure D-3a of Appendix D).

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 occur occupationally, which appear to have a different exposure-response relationship.  
2 However, occupational exposure levels of EtO are of concern to EPA when EtO is used as a  
3 pesticide (e.g., sterilizing agent or fumigant). Therefore, this document also presents extra risk  
4 estimates for the two cancer types for a range of occupational exposure scenarios (see  
5 Section 4.7). Maximum likelihood estimates of the extra risk of lymphoid cancer and breast  
6 cancer combined for the range of occupational exposure scenarios considered (i.e., 0.1 to 1 ppm  
7 8-hour TWA for 35 years) ranged from 0.047 to 0.14. The overall uncertainty associated with  
8 the extra risk estimates for occupational exposure scenarios is less than that associated with the  
9 unit risk estimates for environmental exposures. The extra risk estimates are derived for  
10 occupational exposure scenarios that yield cumulative exposures well within the range of the  
11 exposures in the NIOSH study. Moreover, the NIOSH study is a study of sterilizer workers who  
12 used EtO for the sterilization of medical supplies or spices ([Steenland et al., 1991](#)); thus, the  
13 results are directly applicable to workers in these occupations, and these are among the  
14 occupations of primary concern to EPA.

15 Table 1-1 provides a summary of the major findings in this assessment.

**Table 1-1. Summary of major findings**

<b>Hazard Conclusions</b>	
<b>Hazard Characterization</b>	The weight of evidence from epidemiological studies and supporting information is sufficient to conclude that ethylene oxide is carcinogenic to humans.
<b>Mode of Action</b>	The weight of evidence is sufficient to conclude that ethylene oxide carcinogenicity has a mutagenic mode of action.
<b>Unit Risk Estimates (for environmental exposures)<sup>a</sup></b>	
<b>Basis</b>	<b>Inhalation unit risk estimate<sup>a</sup> (per <math>\mu\text{g}/\text{m}^3</math>)<sup>b</sup></b>
<b>Full lifetime unit risk estimate<sup>c</sup></b>	
Total cancer risk based on human data <sup>d</sup> —lymphoid cancer incidence and breast cancer incidence in females	<b><math>1.8 \times 10^{-3}</math></b>
<b>Adult-based unit risk estimates<sup>e</sup></b>	
Total cancer risk based on human data <sup>d</sup> —lymphoid cancer incidence and breast cancer incidence in females	<b><math>1.1 \times 10^{-3}</math></b>
Lymphoid cancer incidence in both sexes based on human data	$4.3 \times 10^{-4}$
Breast cancer incidence in females based on human data	$8.2 \times 10^{-4}$
Total cancer risk based on human data—range from lymphoid cancer incidence estimates from two different models and female breast cancer incidence estimates from three models	$5.6 \times 10^{-4} - 1.4 \times 10^{-3}$
Total cancer incidence risk estimate from rodent data (female mouse)	$4.6 \times 10^{-5}$
<b>Extra risk estimates for occupational exposure scenarios (see Section 4.7)</b>	
Maximum likelihood estimates of the extra risk of lymphoid cancer and breast cancer combined for the range of occupational exposure scenarios considered (i.e., 0.1 to 1 ppm 8-hr TWA for 35 yr) <sup>f</sup>	<b>0.047–0.14</b>

- 1 <sup>a</sup>These unit risk estimates are not intended for use with continuous lifetime exposure levels above about  $140 \mu\text{g}/\text{m}^3$ .  
2 See Section 4.7 for risk estimates based on occupational exposure scenarios. Preferred estimates are in bold.  
3 <sup>b</sup>To convert unit risk estimates to  $(\text{ppm})^{-1}$ , multiply the  $(\mu\text{g}/\text{m}^3)^{-1}$  estimates by  $1,830 (\mu\text{g}/\text{m}^3)/\text{ppm}$ .  
4 <sup>c</sup>Because the weight of evidence supports a mutagenic mode of action for EtO carcinogenicity, and because of the  
5 lack of chemical-specific data, EPA assumes increased early-life susceptibility and recommends the application of  
6 ADAFs, in accordance with EPA’s *Supplemental Guidance* ([U.S. EPA, 2005b](#)), for exposure scenarios that include  
7 early-life exposures. For the full lifetime (upper bound) unit risk estimate presented here, ADAFs have been  
8 applied, as described in Section 4.4.  
9 <sup>d</sup>Technically, this unit risk estimate reflects the total (upper bound) cancer risk to females and not to the general  
10 population because the breast cancer risk only applies to females. As a practical matter for regulatory purposes,  
11 however, females comprise roughly half the general population and this unit risk estimate enables risk managers to  
12 evaluate the individual risk for this substantial population group. For the purposes of estimating numbers of cancer  
13 cases attributable to specific exposure levels, e.g., for benefits analyses, it would be more appropriate to use the  
14 cancer-specific unit risk estimates (or central tendency estimates), taking sex into account.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1                   **Table 1-1. Summary of major findings (continued)**

2  
3       <sup>e</sup>These (upper bound) unit risk estimates are intended for use in ADAF calculations and less-than-lifetime adult  
4 exposure scenarios ([U.S. EPA, 2005b](#)). Note that these are not the same as the unit risk estimates derived directly  
5 from the human data in Section 4.1 under the assumption that RRs are independent of age. Under that assumption,  
6 the key unit risk estimates were  $4.8 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  for lymphoid cancer incidence,  $9.5 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  for breast  
7 cancer incidence, and  $1.2 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$  for the combined cancer incidence risk from those two cancers. See  
8 Section 4.4 for the derivation of the adult-based unit risk estimates.

9       <sup>f</sup>Technically, these sums would reflect the total cancer risk to females and not a mixed-sex workforce because the  
10 breast cancer risk only applies to females. As a practical matter for regulatory purposes, however, females typically  
11 comprise a substantial proportion of the sterilizer workforce and summing these extra risk estimates enables risk  
12 managers to evaluate the individual risk for this substantial workforce group. In a situation in which the workforce  
13 of concern is comprised predominantly of males, it might be appropriate to use a sex-weighted sum of the extra risks  
14 from the two cancer types (see Section 4.7 for the cancer-specific extra risk estimates).

## 2. INTRODUCTION

Ethylene oxide (EtO) is a gas at room temperature. It is manufactured from ethylene and used primarily as a chemical intermediate in the manufacture of ethylene glycol. It is also used as a sterilizing agent for medical equipment and certain other items and as a fumigating agent for spices. The largest sources of human exposure are in occupations involving contact with the gas in plants that manufacture or use EtO and in hospitals that sterilize medical equipment. EtO can also be inhaled by residents living near production or sterilizing/fumigating facilities. Based on EPA's 2005 National-scale Air Toxics Assessment (NATA) data, the average environmental exposure concentration from all sources (including concentrations near known sources) in the United States is 0.0062  $\mu\text{g}/\text{m}^3$ ; the average background concentration excluding concentrations near sources of EtO is 0.0044  $\mu\text{g}/\text{m}^3$  (NATA 2005 data, <http://www.epa.gov/ttn/atw/nata2005/tables.html>).

EPA offices with an interest in EtO include the Office of Air and Radiation and the Office of Pesticide Programs. The Office of Air and Radiation has an interest because EtO is 1 of the 188 hazardous air pollutants listed in the 1990 Clean Air Act Amendments. The Office of Pesticide Programs has an interest in both environmental and occupational exposures resulting from the sterilization uses of EtO because EPA is responsible for pesticide labeling and registration decisions under the Federal Insecticide, Fungicide, and Rodenticide Act (FIFRA).

The purpose of this document is to provide scientific support and rationale for the hazard and dose-response assessment in IRIS pertaining to carcinogenicity from chronic inhalation exposure to ethylene oxide (EtO) (CASRN 75-21-8). It is not intended to be a comprehensive treatise on the chemical or toxicological nature of EtO. In general, this IRIS Carcinogenicity Assessment provides information on the carcinogenic hazard potential of EtO and quantitative estimates of risk from inhalation exposure. The information includes a weight-of-evidence judgment of the likelihood that the agent is a human carcinogen and the conditions under which the carcinogenic effects may be expressed. Quantitative risk estimates for inhalation exposure (inhalation unit risks) are derived. The definition of an inhalation unit risk is a plausible upper bound on the estimate of risk per  $\mu\text{g}/\text{m}^3$  air breathed.

Development of the hazard identification and dose-response assessments for EtO has followed the general guidelines for risk assessment as set forth by the National Research Council (NRC, 1983). United States Environmental Protection Agency (U.S. EPA) Guidelines and Risk Assessment Forum Technical Panel Reports that were used in the development of this assessment include the following: *Guidelines for Mutagenicity Risk Assessment* (U.S. EPA, 1986), *Methods for Derivation of Inhalation Reference Concentrations and Application of*

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 *Inhalation Dosimetry* ([U.S. EPA, 1994](#)), *Benchmark Dose Technical Guidance* ([U.S. EPA,](#)  
2 [2012](#)), *Science Policy Council Handbook: Risk Characterization* ([U.S. EPA, 2000](#)), *Guidelines*  
3 *for Carcinogen Risk Assessment* [U.S. EPA \(2005a\)](#), *Supplemental Guidance for Assessing*  
4 *Susceptibility from Early-Life Exposure to Carcinogens* ([U.S. EPA, 2005b](#)), and *Science Policy*  
5 *Council Handbook: Peer Review* ([U.S. EPA, 2006b](#)).

6 An earlier external review draft of this carcinogenicity assessment ([U.S. EPA, 2006a](#))  
7 was peer reviewed by a panel of EPA's Science Advisory Board (SAB) in 2007 ([SAB, 2007](#)).  
8 See Appendix H for a summary and EPA's disposition of SAB and public comments on the 2006  
9 external review draft. In response to comments from that SAB review, EPA conducted extensive  
10 new exposure-response modeling of certain epidemiologic data. In addition, EPA has updated  
11 the assessment to reflect new literature through May 2013; this new literature did not  
12 substantively impact the conclusions of the assessment (Appendix J). In July 2013, EPA  
13 released a revised draft for public comment, and that draft assessment was discussed at EPA's  
14 December 2013 IRIS Bimonthly Public Meeting. EPA has also addressed the public comments  
15 that were received on the July 2013 draft (Appendix L). This newly revised 2014 external  
16 review draft is being released for additional external peer review to receive comments primarily  
17 on the expanded exposure-response modeling of the epidemiologic data.  
18

## 19 **2.1. LITERATURE IDENTIFICATION**

20 The literature search strategy first employed for this assessment was based on the  
21 Chemical Abstracts Service Registry Number (CASRN) and at least one common name. Any  
22 pertinent scientific information submitted by the public to the IRIS Submission Desk was also  
23 considered in the development of this document. References were added after the first external  
24 peer review in response to the reviewers' and public comments. In preparation for this second  
25 external peer review, a well-documented systematic literature search was conducted for the time  
26 frame from January 2006 to May 2013. This systematic literature search is described in Section  
27 J.1 of Appendix J. Based on this search, 56 references were identified as potentially relevant to  
28 the EtO assessment. None of the new studies would impact the assessment's major conclusions.  
29 Nonetheless, two new studies of high pertinence to the assessment were identified, and these  
30 studies are reviewed in Section J.2 of Appendix J for transparency and completeness. Reviews  
31 of an additional two new studies of high pertinence were added to Appendix J to address public  
32 comments received in October and December of 2013 (Section J.3); these studies similarly do not  
33 impact the assessment's major conclusions.

1 See Appendix I for a list of all 131 references added after the first external peer review.  
2 The references considered and cited in this document, including abstracts, can be found on the Health  
3 and Environmental Research Online (HERO) website.<sup>8</sup>  
4

## 5 **2.2. NRC RECOMMENDATIONS OF 2011**

6 On 23 December 2011, the Consolidated Appropriations Act, 2012, was signed into law.<sup>9</sup>  
7 The report language included direction to EPA for the IRIS Program related to recommendations  
8 provided by the National Research Council (NRC) in their 2011 review of EPA's draft IRIS  
9 assessment of formaldehyde. Consistent with the direction provided by Congress,  
10 documentation of how the recommendations from Chapter 7 of the [NRC \(2011\)](#) report have been  
11 implemented in this assessment is provided in Appendix K. This documentation also includes an  
12 explanation for why certain recommendations were not incorporated.

13 In brief, the EtO assessment was one of a group of chemical assessments that had already  
14 completed external peer review at the time the 2011 NRC recommendations were released. For  
15 this group of assessments, EPA focused on a subset of the short-term recommendations, such as  
16 streamlining documents, increasing transparency and clarity, and using more tables, figures, and  
17 appendices to present information and data in assessments. While the EtO assessment does not  
18 incorporate recent revisions to the IRIS assessment format recommended in the 2011 NRC  
19 recommendations (and the more recent 2014 NRC Review of the IRIS Process), such as the  
20 inclusion of a standard Preamble and the revised chapter structure, and does not fully implement  
21 the longer-term NRC recommendations, as discussed in Appendix K, the assessment is  
22 consistent with the goal that assessments should provide a scientifically sound and transparent  
23 evaluation of the relevant scientific literature and presentation of the analyses performed.

24 For general information about this assessment or other questions relating to IRIS, the  
25 reader is referred to EPA's IRIS Hotline at (202) 566-1676 (phone), (202) 566-1749 (fax), or  
26 [hotline.iris@epa.gov](mailto:hotline.iris@epa.gov) (email address).

---

<sup>8</sup>HERO is a database of scientific studies and other references used to develop EPA's risk assessments, which are aimed at understanding the health and environmental effects of pollutants and chemicals. HERO is developed and managed in EPA's Office of Research and Development (ORD) by the National Center for Environmental Assessment (NCEA). The database includes more than 1,000,000 scientific articles from the peer-reviewed literature. New studies are added continuously to HERO.

<sup>9</sup>Pub. L. No. 112-74, Consolidated Appropriations Act, 2012.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 **3. HAZARD IDENTIFICATION**

2 This chapter presents the evidence considered in the hazard identification of EtO  
3 carcinogenicity and the hazard characterization resulting from the weight-of-evidence evaluation.  
4 Section 3.1 summarizes the human evidence (a more detailed discussion of the human cancer  
5 studies is presented in Appendix A). Section 3.2 describes the evidence from experimental  
6 animal studies. Section 3.3 discusses supporting evidence, in particular evidence regarding the  
7 genotoxicity of EtO. Section 3.4 provides the mode-of-action analysis for EtO carcinogenicity.  
8 To conclude the chapter, Section 3.5 presents the hazard characterization for EtO carcinogenicity  
9 and a discussion of life stages and populations with potentially increased susceptibility.  
10

11 **3.1. EVIDENCE OF CANCER IN HUMANS**

12 The literature from 1988 to present contains numerous epidemiological studies of the  
13 carcinogenic effects of EtO in occupational cohorts; some of these cohorts were the subject of  
14 multiple reports. The conclusions about the human evidence of carcinogenicity in this  
15 assessment are based on the following summary of those studies, which are discussed in more  
16 detail and critically reviewed in Appendix A. Table A-5 in Appendix A provides a tabular  
17 summary of the epidemiological studies, including some study details, results, and limitations.  
18 The strengths and weaknesses of these studies were evaluated individually using standard  
19 considerations in evaluating epidemiological studies. The major areas of concern are study  
20 design, exposure assessment, and data analysis. General features of study design considered  
21 include sample size and assessment of the health endpoint. For case-control studies, design  
22 considerations include representativeness of cases, selection of controls, participation rates, use  
23 of proxy respondents, and interview approach (e.g., blinding). For cohort studies, design  
24 considerations include selection of referent population (e.g., internal comparisons are generally  
25 preferred to comparisons with an external population), loss to follow-up, and length of  
26 follow-up. Exposure assessment issues include specificity of exposure (exposure  
27 misclassification), characterization of exposure (e.g., ever exposed or quantitative estimate of  
28 exposure level), and potential confounders. Analysis considerations include adjustment for  
29 potential confounders or effect modifiers and modeling of exposure-response relationships.

30 Two primary sources of exposures to EtO are production facilities and sterilization  
31 operations. There are two types of production facilities ([IARC, 1994b](#)):  
32  
33

- 1 1. Those using the older chlorohydrin process, where ethylene is reacted with hypochlorous  
2 acid and then with calcium oxide to make EtO (this method produces unwanted  
3 byproducts, the most toxic of which is ethylene dichloride), and
- 4 2. Those producing EtO via direct oxidation of ethylene in a pressurized vessel, which  
5 involves less EtO exposure and eliminates the chemical byproducts of the chlorohydrin  
6 process.

7  
8  
9 Exposure in the sterilization of medical equipment and in the direct oxidation process is  
10 predominantly to EtO, whereas exposure in the chlorohydrin process is to EtO mixed with other  
11 chemicals.

12 [Hogstedt et al. \(1986\)](#) and [Hogstedt \(1988\)](#) summarized findings of three Swedish  
13 occupational cohorts (539 men and 170 women) exposed in a plant where hospital equipment  
14 was sterilized, in a chlorohydrin production facility, and in a direct oxidation production facility.  
15 The incidence of leukemia was elevated in all cohorts, although the risk was not statistically  
16 significant in the cohort from the direct oxidation facility. For the three cohorts combined there  
17 were statistically significantly elevated standard mortality ratios (SMRs) for leukemia  
18 (SMR = 9.2; 95% confidence interval [CI] = 3.7–19), based on 7 deaths, and for stomach cancer  
19 (SMR = 5.5; 95% CI = 2.6–10), based on 10 deaths. Although this study produced high SMRs  
20 for leukemia, stomach cancer, and total cancer, there are some limitations, such as multiple  
21 exposures to numerous other chemicals, lack of personal exposure information, and lack of  
22 latency analysis. No gender differences were separately analyzed. No dose-response  
23 calculations were possible. This study provides suggestive evidence of the carcinogenicity of  
24 EtO.

25 [Coggon et al. \(2004\)](#) reported the results of a follow-up study of a cohort originally  
26 studied by [Gardner et al. \(1989\)](#). The cohort included workers in three EtO production facilities  
27 (two using both chlorohydrin and direct oxidation processes and the third using direct oxidation  
28 only); in a fourth facility that used EtO in the manufacture of other chemicals; and in eight  
29 hospitals that used EtO in sterilizing units. The total cohort comprised 1,864 men and  
30 1,012 women. No statistically significant excesses were observed for any cancer site. Slight  
31 increases, based on small numbers, were observed for the various lymphohematopoietic cancers:  
32 Hodgkin lymphoma (2 vs. 1 expected), non-Hodgkin lymphoma (NHL) (7 vs. 4.8), multiple  
33 myeloma (3 vs. 2.5), and leukemia (5 vs. 4.6). The increases were concentrated in the  
34 1,471 chemical-manufacturing workers, of whom all but 1 were male. In the  
35 chemical-manufacturing workers with “definite” exposure, 4 leukemias were observed  
36 (1.7 expected) and 9 lymphohematopoietic cancers were observed (4.9 expected). A slight

1 deficit in the risk of breast cancer deaths (11 vs. 13.2) was observed in the cohort. No individual  
2 exposure measurements were obtained from cohort members, and no exposure measurements  
3 were available before 1977. Multiple exposures to other chemicals, small numbers of deaths,  
4 and lack of individual EtO measurements make this study only suggestive of a higher risk of  
5 leukemia from exposure to EtO.

6 A series of retrospective mortality studies of about 2,000 male workers who were  
7 assigned to operations that used or produced EtO in either of two Union Carbide Corporation  
8 (UCC) chemical production facilities in West Virginia ([Valdez-Flores et al., 2010](#); [Swaen et al.,  
9 2009](#); [Teta et al., 1999](#); [Benson and Teta, 1993](#); [Teta et al., 1993](#); [Greenberg et al., 1990](#)) have  
10 been published. EtO was produced at these facilities until 1971, after which it was imported to  
11 the facilities. For EtO production, the chlorohydrin process was used from 1925 to 1957, and the  
12 direct oxidation process was used from 1937 to 1971 (during overlapping years, both processes  
13 were in use). The cohort was observed from 1940 through 1978 in the original study ([Greenberg  
14 et al., 1990](#)), through 1988 in the [Teta et al. \(1993\)](#), [Benson and Teta \(1993\)](#), and [Teta et al.  
15 \(1999\)](#) studies, and through 2003 in the [Swaen et al. \(2009\)](#) and [Valdez-Flores et al. \(2010\)](#)  
16 studies. A large-scale industrial hygiene survey and monitoring of EtO concentrations was  
17 carried out in 1976, at which time EtO was in use at the facilities but no longer in production.

18 [Greenberg et al. \(1990\)](#) found elevated but not statistically significant risks of pancreatic  
19 cancer (SMR = 1.7) and leukemia (SMR = 2.3) (each based on seven cases) in the entire cohort;  
20 most of the cases occurred in the chlorohydrin production unit (note that the chlorohydrin  
21 production unit produced primarily ethylene chlorohydrin, which is used in chlorohydrin-based  
22 EtO production, but this unit is not where chlorohydrin-based EtO production took place).  
23 Limitations of this study included multiple exposures to many different chemicals in the facility  
24 through the years and lack of EtO exposure measurements prior to 1976. Three categories of  
25 exposure were established for analysis—low, intermediate, and high—based on a qualitative  
26 characterization of the potential for EtO exposure. The number of workers in each exposure  
27 category was not reported. No significant findings of a dose-response relationship were  
28 discernible. No quantitative estimates of individual exposure were made in this study, and no  
29 latency analysis was conducted (average follow-up was 20 years). Furthermore, EtO is not the  
30 only chemical to which the observed excesses in cancer mortality could be attributed.

31 A follow-up study ([Teta et al., 1993](#)) that extended the observation of this cohort  
32 (excluding the 278 chlorohydrin production unit workers, who reportedly had low EtO  
33 exposures) for an additional 10 years to 1988 found no significant risk of total cancer; there was  
34 a slight trend in the risk of leukemia with increasing duration of assignment to departments using  
35 or processing EtO, but it was not significant ( $p = 0.28$ ) and was based on only five cases. The

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 average follow-up was 27 years, and at least 10 years had elapsed since first exposure for all  
2 workers. The same problems of exposure ascertainment exist for this study as for that of  
3 [Greenberg et al. \(1990\)](#), and furthermore, the follow-up did not update work histories for the  
4 workers after 1978. EtO production at the plants was discontinued before 1978, as noted by [Teta](#)  
5 [et al. \(1993\)](#); however, according to [Greenberg et al. \(1990\)](#), certain nonproduction areas had  
6 “intermediate” potential for EtO exposure, although estimates of exposure levels suggest that the  
7 levels would also be lower during the update period [ $<1$  ppm 8-hour time-weighted average  
8 (TWA), according to [Teta et al. \(1993\)](#)]. It appears from the [Greenberg et al. \(1990\)](#) publication  
9 that the high potential exposure group was reserved for EtO production workers, and according  
10 to [Teta et al. \(1993\)](#), there were only 425 EtO production workers in the cohort. Of these, only  
11 118 worked in the chlorohydrin-based production process, where exposures were reportedly  
12 highest. Essentially, the study did not support the earlier studies of cancer in EtO workers;  
13 however, it was limited by low statistical power and a crude exposure assessment and, thus, is  
14 not very informative regarding whether exposure to EtO is causally related to cancer.

15 In a parallel follow-up study through 1988 of only the chlorohydrin production  
16 employees, [Benson and Teta \(1993\)](#) found that pancreatic cancer and lymphohematopoietic  
17 cancer cases continued to accumulate and that the SMRs were statistically significant for  
18 pancreatic cancer (SMR = 4.9; Obs = 8,  $p < 0.05$ ) and for lymphohematopoietic cancer  
19 (SMR = 2.9; Obs = 8,  $p < 0.05$ ). These investigators interpreted these excesses as possibly due  
20 to ethylene dichloride, a byproduct in the chlorohydrin process. Again, this small study of only  
21 278 workers was limited by the same problems as the [Greenberg et al. \(1990\)](#) study and the [Teta](#)  
22 [et al. \(1993\)](#) study. No individual estimates of exposure are available and the workers were  
23 potentially exposed to many different chemicals (see Table A-5 in Appendix A). Furthermore,  
24 the chlorohydrin production unit was reportedly considered a low potential EtO exposure  
25 department. Hence this study has little weight in determining the carcinogenicity of EtO.

26 In a later analysis, [Teta et al. \(1999\)](#) fitted Poisson regression dose-response models to  
27 the UCC data (followed through 1988 and excluding the chlorohydrin production workers) and  
28 to data (followed through 1987) from a study by the National Institute for Occupational Safety  
29 and Health (NIOSH) (described below). Because [Teta et al. \(1999\)](#) did not present risk ratios for  
30 the cumulative exposure categories used to model the dose-response relationships, the only  
31 comparison that can be made between the UCC and NIOSH data is based on the fitted models.  
32 These models are almost identical for leukemia, but for the lymphoid category, the  
33 risk—according to the fitted model for the UCC data—decreased as a function of exposure,  
34 whereas the risk for the modeled NIOSH data increased as a function of exposure. However, the  
35 models are based on small numbers of cases (16 [5 UCC, 11 NIOSH] for leukemia; 22 [3 UCC,

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 19 NIOSH] for lymphoid cancers), and no statistics are provided to assess model goodness-of-fit  
2 or to compare across models. In any event, this analysis is superseded by the more recent  
3 analysis by the same authors ([Valdez-Flores et al., 2010](#)) of the results of more recent follow-up  
4 studies of these cohorts (see below).

5 [Swaen et al. \(2009\)](#) studied the same UCC cohort identified by [Teta et al. \(1993\)](#), i.e.,  
6 without the chlorohydrin production workers, but extended the cohort enumeration period from  
7 the end of 1978 to the end of 1988, identifying 167 additional workers, and conducted mortality  
8 follow-up of the resulting cohort of 2,063 male workers through 2003. Work histories were also  
9 extended through 1988 (exposures after 1988 were considered negligible compared to earlier  
10 exposure levels). [Swaen et al. \(2009\)](#) used an exposure assessment based on the qualitative  
11 categorizations of potential EtO exposure in the different departments developed by [Greenberg et](#)  
12 [al. \(1990\)](#) and time-period exposure estimates from [Teta et al. \(1993\)](#). This exposure assessment  
13 was relatively crude, based on just a small number of department-specific and time-period-  
14 specific categories, and with exposure estimates for only a few of the categories derived from  
15 actual measurements (see Appendix A.2.20 for details).

16 At the end of the 2003 follow-up, 1,048 of the 2,063 workers had died ([Swaen et al.,](#)  
17 [2009](#)). The all-cause mortality SMR was 0.85 (95% CI = 0.80, 0.90) and the cancer SMR was  
18 0.95 (95% CI = 0.84, 1.06). None of the SMRs for specific cancer types showed any statistically  
19 significant increases. In analyses stratified by hire date [pre- (inclusive) or post-1956], the SMR  
20 for leukemia was elevated but not statistically significant (1.51; 95% CI 0.69, 2.87) in the  
21 early-hire group, based on nine deaths. In analyses stratified by duration of employment, no  
22 trends were apparent for any of the lymphohematopoietic cancers, although in the 9+ years of  
23 employment subgroup, the SMR for NHL was nonsignificantly increased (1.49; 95% CI 0.48,  
24 3.48), based on five deaths. In SMR analyses stratified by cumulative exposure, no trends were  
25 apparent for any of the lymphohematopoietic cancers and there were no notable elevations for  
26 the highest cumulative exposure category. Note that only 27 lymphohematopoietic cancer deaths  
27 (including 12 leukemias and 11 NHLs) were observed in the cohort.

28 [Swaen et al. \(2009\)](#) also did internal Cox proportional hazards modeling for some disease  
29 categories (all-cause mortality, leukemia mortality, and lymphoid cancer [NHL, lymphocytic  
30 leukemia, and myeloma] mortality [17 deaths]), using cumulative exposure as the exposure  
31 metric. These analyses showed no evidence of an exposure-response relationship. Alternate  
32 Cox proportional hazard analyses and categorical exposure-response analyses of the UCC data  
33 conducted by [Valdez-Flores et al. \(2010\)](#) for a larger set of cancer endpoints similarly reported  
34 an absence of any exposure-response relationships. Each of these cancer analyses, however,

1 relies on small numbers of cases and a crude exposure assessment, where there is a high potential  
2 for exposure misclassification.

3 In a study of 2,658 male workers at eight chemical plants where EtO is produced  
4 (manufacturing process not stated), [Kiesselbach et al. \(1990\)](#) found slightly increased SMRs for  
5 cancers of the stomach, esophagus, and lung. A latency analysis was done only for stomach  
6 cancer and total mortality. The investigators considered 71.6% of the cohort to be “weakly”  
7 exposed; only 2.6% were “strongly exposed.” No data were provided to explain how these  
8 exposure categories were derived. The workers were followed for a median 15.5 years. Without  
9 additional information on exposure to EtO, this study is of little help at this time in evaluating the  
10 carcinogenicity of EtO.

11 NIOSH conducted an industry-wide study of 18,254 workers (45% male and 55%  
12 female) in 14 plants where EtO was used ([Steenland et al., 2004](#); [Stayner et al., 1993](#); [Steenland  
13 et al., 1991](#)). Most of the workers were exposed while sterilizing medical supplies and treating  
14 spices and in the manufacture and testing of medical sterilizers. Individual exposure estimates  
15 were derived for workers from 13 of the 14 plants. The procedures for selecting the facilities and  
16 defining the cohort are described in [Steenland et al. \(1991\)](#), and the exposure model and  
17 verification procedures are described in [Greife et al. \(1988\)](#) and [Hornung et al. \(1994\)](#). Briefly, a  
18 regression model was developed, allowing the estimation of exposure levels for time periods,  
19 facilities, and operations for which industrial hygiene data were unavailable. The data for the  
20 model consisted of 2,700 individual time-weighted exposure values for workers’ personal  
21 breathing zones, acquired from 18 facilities between 1976 and 1985. The data were divided into  
22 two sets, one for developing the regression model and the second for testing it. Seven out of  
23 23 independent variables tested for inclusion in the regression model were found to be significant  
24 predictors of EtO exposure and were included in the final model. This model predicted 85% of  
25 the variation in average EtO exposure levels. (See Appendix A, Section A.2.8, for more details  
26 on the NIOSH exposure assessment and its evaluation.) Results of the original follow-up study  
27 through 1987 are presented in [Steenland et al. \(1991\)](#) and [Stayner et al. \(1993\)](#). The cohort  
28 averaged 26.8 years of follow-up in the extended follow-up study through 1998, and 16% of the  
29 cohort had died ([Steenland et al., 2004](#)).

30 The overall SMR for cancer was 0.98, based on 860 deaths ([Steenland et al., 2004](#)). The  
31 SMR for (lympho)hematopoietic cancer was 1.00, based on 79 cases. Exposure-response  
32 analyses, however, revealed exposure-related increases in hematopoietic cancer mortality risk,  
33 although the effect was primarily in males, when analyzed by sex. In categorical life-table  
34 analysis, men with >13,500 ppm-days of cumulative exposure had an SMR of 1.46 (Obs = 13).  
35 In internal Cox regression analyses (i.e., analyses in which the referent population is within the

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 cohort) with exposure as a continuous variable, statistically significant trends in males for all  
2 hematopoietic cancer ( $p = 0.02$ ) and for “lymphoid” cancers (NHL, lymphocytic leukemia, and  
3 myeloma;  $p = 0.02$ ) were observed using log cumulative exposure (ppm-days) with a 15-year  
4 lag.<sup>10</sup> In internal categorical analyses, statistically significant odds ratios (ORs) were observed  
5 in the highest cumulative exposure quartile (with a 15-year lag) in males for all hematopoietic  
6 cancer (OR = 3.42; 95% CI = 1.09–10.73) and “lymphoid” cancer (OR = 3.76; 95%  
7 CI = 1.03–13.64). The exposure metrics of duration of exposure, average concentration, and  
8 maximum (8-hour TWA) concentration did not predict the hematopoietic cancer results as well  
9 as did the cumulative exposure metric.

10 Although the overall SMR for female breast cancer was 0.99, based on 102 deaths, the  
11 NIOSH mortality follow-up study reported a significant excess of breast cancer mortality in the  
12 highest cumulative exposure quartile using a 20-year lag period compared to the U.S. population  
13 (SMR = 2.07; 95% CI = 1.10–3.54; Obs = 13). Internal exposure-response analyses also noted a  
14 significant positive trend for breast cancer mortality using the log of cumulative exposure and a  
15 20-year lag time ( $p = 0.01$ ). In internal categorical analyses, a statistically significant OR for  
16 breast cancer mortality was observed in the highest cumulative exposure quartile with a 20-year  
17 lag (OR = 3.13; 95% CI = 1.42–6.92).

18 In summary, although the overall external comparisons did not demonstrate increased  
19 risks, the NIOSH investigators found significant internal exposure-response relationships  
20 between exposure to EtO and cancers of the hematopoietic system, as well as breast cancer  
21 mortality. (Internal comparisons are considered superior to external comparisons in occupational  
22 epidemiology studies because internal comparisons help control for the healthy worker effect and  
23 other factors that might be more comparable within a study’s worker population than between  
24 the workers and the general population.) Exposures to other chemicals in the workplace were  
25 believed to be minimal or nonexistent. This study is the most useful of the epidemiologic studies  
26 in terms of carrying out a quantitative dose-response assessment. It possesses more attributes  
27 than the others for performing risk analysis (e.g., good-quality estimates of individual exposure,  
28 lack of exposure to other chemicals, and a large and diverse cohort of workers).

29 It should be noted that [Steenland et al. \(2004\)](#) used Cox regression models, which are  
30 log-linear relative rate models, thus providing some low-dose sublinear curvature for doses  
31 expressed in terms of cumulative exposure. However, the best-fitting dose-response model for  
32 both male lymphoid cancers and male all hematopoietic cancers was for dose expressed in terms  
33 of log cumulative exposure, indicating supralinearity of the low-dose data. Supralinearity of the

---

<sup>10</sup>The sex difference is not statistically significant, however, and the trends for both sexes combined are also statistically significant [ $p = 0.01$  and  $p = 0.02$ , respectively; see Tables D-3e and D-4e in Appendix D].

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 dose-response data was also indicated by the categorical exposure results. This is in contrast to  
2 the reported results of [Kirman et al. \(2004\)](#) based on the [Teta et al. \(1999\)](#) analysis combining  
3 the 1993 UCC leukemia data with the 1993 NIOSH leukemia data, which are claimed by the  
4 authors to provide empirical evidence supporting a quadratic dose-response relationship. The  
5 2004 NIOSH dose-response data for hematopoietic cancers clearly do not provide empirical  
6 evidence in support of a quadratic dose-response relationship. On the contrary, the NIOSH data  
7 suggest a supralinear dose-response relationship in the observable range.

8 [Wong and Trent \(1993\)](#) investigated the same cohort as [Steenland et al. \(1991\)](#) but added  
9 474 new unexplained subjects and increased the follow-up period by 1 year. They incremented  
10 the total number of deaths by 176 and added 392.2 more expected deaths. The only positive  
11 finding was a statistically significantly increased risk of NHL among men (SMR = 2.5; Obs = 6;  
12  $p < 0.05$ ). However, there was a deficit risk of NHL among women. For breast cancer, there  
13 was no trend of increasing risk by duration of employment or by latency. This study has major  
14 limitations, not the least of which is a lack of detailed employment histories, making it  
15 impossible to quantify individual exposures and develop dose-response relationships.  
16 Furthermore, the addition of more than twice as many expected deaths as observed deaths makes  
17 the analysis by the authors questionable.

18 [Valdez-Flores et al. \(2010\)](#) conducted alternative Cox proportional hazards modeling and  
19 categorical exposure-response analyses using data from the UCC cohort ([Swaen et al., 2009](#)), the  
20 NIOSH cohort ([Steenland et al., 2004](#)) and the two cohorts combined, analyzing the sexes both  
21 separately and together. These investigators reported that they found no evidence of  
22 exposure-response relationships for cumulative exposure with either the Cox model or  
23 categorical analyses for all of the cohort/endpoint data sets examined (endpoints included all  
24 lymphohematopoietic cancers, lymphoid cancers, and female breast cancer, the latter in the  
25 NIOSH cohort only). [Valdez-Flores et al. \(2010\)](#) did observe statistically significant increases in  
26 response rates in the highest exposure quintile relative to the lowest exposure quintile for  
27 lymphohematopoietic and lymphoid cancers in males in the NIOSH cohort, consistent with the  
28 categorical results of [Steenland et al. \(2004\)](#), as well as a statistically significant increase in the  
29 highest exposure quintile for lymphoid cancers in males and females combined in the NIOSH  
30 cohort, consistent with the results in Appendix D. Because the exposure assessment conducted  
31 for the UCC cohort is much cruder (see above and Appendix A.2.20), especially for the highest  
32 exposures, than the NIOSH exposure assessment (which was based on a validated regression  
33 model; see Appendix A.2.8), EPA considers the results of exposure-response analyses of the  
34 combined cohort data to have greater uncertainty than those from analyses of the NIOSH cohort  
35 alone, despite the additional cases contributed by the UCC cohort (e.g., the UCC cohort

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 contributes 17 cases of lymphoid cancer to the 53 from the NIOSH cohort). Furthermore,  
2 [Valdez-Flores et al. \(2010\)](#) did not use any log cumulative exposure models, and these were the  
3 models that were statistically significant in the [Steenland et al. \(2004\)](#) analyses, consistent with  
4 the apparent supralinearity of the NIOSH exposure-response data. See Appendix A.2.20 for a  
5 more detailed discussion of the [Valdez-Flores et al. \(2010\)](#) analyses and how they compared with  
6 the [Steenland et al. \(2004\)](#) analyses.

7 In a mortality study of 1,971 male chemical workers in Italy, 637 of whom were licensed  
8 to handle EtO but not other toxic gases, [Bisanti et al. \(1993\)](#) reported statistically significant  
9 excesses of hematopoietic cancers (SMR = 7.1, Obs = 5,  $p < 0.05$ ). The study was limited by the  
10 lack of exposure measurements and by the young age of the cohort. Although this study  
11 suggests that exposure to EtO leads to a significant excess of hematopoietic cancer, the lack of  
12 personal exposure measurements and the fact that members were potentially exposed to other  
13 chemicals in the workplace lessen the study's usefulness for establishing the carcinogenicity of  
14 EtO.

15 Hagmar et al. ([Hagmar et al., 1995](#); [Hagmar et al., 1991](#)) studied cancer incidence in  
16 2,170 Swedish workers (861 male and 1,309 female) in two medical sterilizing plants. They  
17 determined concentrations in six job categories and estimated cumulative exposures for each  
18 worker. They found hematopoietic cancers in 6 individuals versus 3.4 expected (SMR = 1.8) and  
19 a nonsignificant doubling in the risk when a 10-year latency period was considered. Even  
20 though the cohort was young, the follow-up time was short (median 11.8 years), and only a small  
21 fraction of the workers was highly exposed, the report is suggestive of an association between  
22 EtO exposure and hematopoietic cancers. The risk of breast cancer was less than expected,  
23 although with such short follow-up, the total numbers of cases was small (standardized incidence  
24 ratio [SIR] = 0.5, Obs = 5). In the latent category of 10 years or more, the risk was even lower  
25 (SIR = 0.4, Obs = 2).

26 In a large chemical manufacturing plant in Belgium (number of employees not stated),  
27 [Swaen et al. \(1996\)](#) performed a nested case-control study of Hodgkin lymphoma to determine  
28 whether a cluster of 10 cases in the active male work force was associated with any particular  
29 chemical. They found a significant association for benzene and EtO. This study is limited by  
30 the exclusion of inactive workers and the potential confounding effect of other chemicals besides  
31 EtO, and it is not useful for quantitative dose-response assessment.

32 [Olsen et al. \(1997\)](#) studied 1,361 male employees working in the ethylene and propylene  
33 chlorohydrin production and processing areas located within the EtO and propylene oxide  
34 production plants at four Dow Chemical Company sites in the United States. Although these  
35 investigators found a nonsignificant positive trend between duration of employment as

1 chlorohydrin workers and lymphohematopoietic cancer (Obs = 10), they concluded that there  
2 was no appreciable risk in these workers, in contrast to the findings of [Benson and Teta \(1993\)](#).  
3 The small cohort size and the lack of data on EtO exposures limit the usefulness of this study in  
4 inferring risks due to EtO.

5 [Norman et al. \(1995\)](#) studied 1,132 workers (204 male and 928 female) in a medical  
6 sterilizing plant in the United States. In the women, there was a significant excess incidence of  
7 breast cancer (SIR = 2.6, Obs = 12,  $p < 0.05$ ); no other cancer sites were elevated. The risk of  
8 breast cancer was not noted to be excessive in the few previous studies where adequate numbers  
9 of females were included and analyzed for breast cancer; however, only one of these studies was  
10 also an incidence study. The follow-up time was too short to draw meaningful conclusions at  
11 this time. This study lacks the power to determine whether risks for cancers other than breast  
12 cancer are statistically significantly elevated. It has no information regarding historical exposure  
13 and some subjects with breast cancer had worked for less than 1 month.

14 [Tompa et al. \(1999\)](#) reported a cluster of eight breast cancers and eight other cancers in  
15 98 nurses exposed to EtO in a hospital in Hungary; however, the expected number of cases  
16 cannot be identified.

17 The NIOSH investigators used the NIOSH cohort to conduct a study of breast cancer  
18 incidence and exposure to EtO ([Steenland et al., 2003](#)). The researchers identified 7,576 women  
19 from the initial cohort who had been employed in the commercial sterilization facilities for at  
20 least 1 year (76% of the original cohort). Breast cancer incidence was determined from  
21 interviews (questionnaires), death certificates, and cancer registries. Interviews were obtained  
22 for 5,139 women (68% of the study cohort). The main reason for nonresponse was inability to  
23 locate the study subject (22% of cohort). The average duration of exposure for the cohort was  
24 10.7 years. For the full study cohort, 319 incident breast cancer cases were identified, including  
25 20 cases of carcinoma in situ. Overall, the SIR was 0.87 (0.94 excluding the in situ cases) using  
26 Surveillance, Epidemiology, and End Results (SEER) reference rates for comparison. Results  
27 with the full cohort are expected to be underestimated, however, because of case  
28 under-ascertainment in the women without interviews. A significant exposure-response trend  
29 was observed for SIR across cumulative exposure quintiles, using a 15-year lag time ( $p = 0.002$ ).  
30 In internal Cox regression analyses, with exposure as a continuous variable, a significant trend  
31 for breast cancer incidence was obtained for log cumulative exposure with a 15-year lag  
32 ( $p = 0.05$ ), taking age, race, and year of birth into account. Using duration of exposure, lagged  
33 15 years, provided a slightly better fit ( $p = 0.02$ ), while models with cumulative  
34 (nontransformed), maximum or average exposure did not fit as well. In the Cox regression

1 analysis with categorical exposures and a 15-year lag, the top cumulative exposure quintile had a  
2 statistically significant OR for breast cancer incidence of 1.74 (95% CI = 1.16–2.65).

3 In the subcohort with interviews, 233 incident breast cancer cases were identified.  
4 Information on various risk factors for breast cancer was also collected in the interviews, but  
5 only parity and breast cancer in a first-degree relative turned out to be important predictors of  
6 breast cancer incidence. In internal analyses with continuous exposure variables, the model with  
7 duration of exposure (lagged 15 years) again provided the best fit ( $p = 0.006$ ). Both the  
8 cumulative exposure and log cumulative exposure models also yielded significant regression  
9 coefficients with a 15-year lag ( $p = 0.02$  and  $p = 0.03$ , respectively), taking age, race, year of  
10 birth, parity, and breast cancer in a first-degree relative into account. In the Cox regression  
11 analysis with categorical exposures and a 15-year lag, the top cumulative exposure quintile had a  
12 statistically significant OR of 1.87 (95% CI = 1.12–3.10).

13 [Steenland et al. \(2003\)](#) suggest that their findings are not conclusive of a causal  
14 association between EtO exposure and breast cancer incidence because of inconsistencies in  
15 exposure-response trends, possible biases due to nonresponse, and an incomplete cancer  
16 ascertainment. Although that conclusion seems appropriate, those concerns do not appear to be  
17 major limitations. As noted by the authors, it is not uncommon for positive exposure-response  
18 trends not to be strictly monotonically increasing, conceivably due to random fluctuations or  
19 imprecision in exposure estimates. Furthermore, the consistency of results between the full  
20 study cohort, which is less subject to nonresponse bias, and the subcohort with interviews, which  
21 should have full case ascertainment, alleviates some of the concerns about those potential biases.

22 In a study of 299 female workers employed in a hospital in Hungary where gas sterilizers  
23 were used, [Kardos et al. \(2003\)](#) observed 11 cancer deaths, including 3 breast cancer deaths,  
24 compared with slightly more than 4 expected total cancer deaths. Site-specific expected deaths  
25 are not available in this study, so RR estimates cannot be determined. However, the observation  
26 of 3 breast cancer deaths, with at most 4.4 (with Hungarian national rates as the referent) total  
27 cancer deaths expected, is indicative of an increased risk of breast cancer,<sup>11</sup> and this  
28 characterization is supported by the reference of [Major et al. \(2001\)](#) to a cluster of breast cancer  
29 cases in female nurses at the same hospital.

---

<sup>11</sup>Hungarian age-standardized female cancer mortality rates reported by the International Agency for Research on Cancer (<http://eu-cancer.iarc.fr/country-348-hungary.html,en>) suggest that the ratio of breast cancer deaths to total cancer deaths in Hungarian females is about 0.16 (28.0/100,000 breast cancer mortality rate versus 180.0/100,000 total cancer mortality rate). Although a comparison of this general population ratio with the ratio of 0.68 for breast cancer to total cancer mortality in the [Kardos et al. \(2003\)](#) study is necessarily crude because the general population ratio is not based on the age-standardized rates that would correspond to the age distribution of the person-time of the women in the study, which are unknown, the large difference between the ratios (0.68 for the study versus 0.16 for the general population) indicates an increased risk of breast cancer in the study.

*This document is a draft for review purposes only and does not constitute Agency policy.*

### 3.1.1. Conclusions Regarding the Evidence of Cancer in Humans

Most of the human studies suggest a possible increased risk of lymphohematopoietic cancers, but the total weight of the epidemiological evidence does not provide conclusive proof of causality. Of the eight relevant criteria of causality envisioned by [Hill \(1965\)](#), temporality, coherence, biological plausibility, and analogy are clearly satisfied. There is also evidence of consistency in the response, of a dose-response relationship (biological gradient), and of specificity when the loosely defined blood malignancies are combined under the rubric “cancer of the hematopoietic system.” On the other hand, most of the relative risk estimates are not large (strong) in magnitude. See Section 3.5.1 for a more detailed discussion of the Hill criteria as applied to the EtO database.

The large NIOSH study ([Steenland et al., 2004](#); [Stayner et al., 1993](#); [Steenland et al., 1991](#)) of workers at 14 chemical plants around the country provides the strongest evidence of carcinogenicity. A statistically significant positive trend was observed in the risk of lymphohematopoietic neoplasms with increasing (log) cumulative exposure to EtO, although the results for this model were reported only for males (the sex difference is not statistically significant, however, and the trend for both sexes combined is statistically significant; see Appendix D). Despite limitations in the data, most other epidemiologic studies have also found elevated risks of lymphohematopoietic cancer from exposure to EtO (summarized briefly in Section 3.1 and Table 3-1; see also Appendix A for more details, in particular Table A-5 for a summary of study results and limitations). Furthermore, when the exposure is relatively pure, such as in sterilization workers, there is an elevated risk of lymphohematopoietic cancer that cannot be attributed to the presence of confounders such as those that could potentially appear in the chlorohydrin process. Moreover, the studies that do not report a significant lymphohematopoietic cancer effect from exposure to EtO have major limitations, such as small numbers of cases and inadequate exposure information (see Table A-5 in Appendix A).

In addition, there is evidence of an increase in the risk of both breast cancer mortality and incidence in women who are exposed to EtO. Studies have reported increases in the risk of breast cancer in women employees of commercial sterilization plants ([Steenland et al., 2004](#); [Steenland et al., 2003](#); [Norman et al., 1995](#)) as well as in Hungarian hospital workers exposed to EtO ([Kardos et al., 2003](#)). In several other studies where exposure to EtO would be expected to have occurred among female employees, no elevated risks were seen ([Coggon et al., 2004](#); [Hagmar et al., 1991](#)) or breast cancer results were not reported ([Hogstedt, 1988](#); [Hogstedt et al., 1986](#)). However, these studies had far fewer cases to analyze than the NIOSH studies, and most did not have individual exposure estimates and relied on external comparisons (see Table 3-2 for a brief summary and Table A-5 in Appendix A for more details). The [Steenland et al. \(2004\)](#) and

**Table 3-1. Epidemiological studies of ethylene oxide and human cancer—lymphohematopoietic cancer results<sup>a</sup>**

Study/Population/ Industry	Number of subjects	Lymphohematopoietic cancer results				Comments
<a href="#">Hogstedt (1988)</a> and <a href="#">Hogstedt et al. (1986)</a> . Sterilizers, production workers, Sweden.	709 (539 men, 170 women)	<i>cancer deaths</i> leukemia (ICD-8 204-207; lymphohematopoietic (ICD-8 200-208)	<i>observed</i> 7 9	<i>expected</i> 0.8 2.0	<i>SMR (95% CI)</i> 9.2 (3.7, 19) <sup>b</sup> 4.6 (2.1, 8.7) <sup>b</sup>	Insufficient follow-up; 12.0% of cohort had died (85 deaths). Exposure to other chemicals.
<a href="#">Coggon et al. (2004)</a> . Update of <a href="#">Gardner et al. (1989)</a> . Sterilizing workers in 8 hospitals and users in 4 companies, Great Britain.	2,876 (1,864 men, 1,012 women)	<i>cancer deaths</i> leukemia (ICD-9 204-208) leukemia (definite or continual exposure) NHL (ICD-9 200+202) lymphohematopoietic (ICD-9 200-208)	<i>observed</i> 5 5 7 17	<i>expected</i> 4.6 2.6 4.8 12.9	<i>SMR (95% CI)</i> 1.1 (0.35, 2.5) 1.9 (0.62, 4.5) <sup>b</sup> 1.5 (0.58, 3.0) <sup>b</sup> 1.3 (0.77, 2.1) <sup>b</sup>	Short follow-up; 19.6% of cohort had died (565 deaths). Exposure to other chemicals.
<a href="#">Kiesselbach et al. (1990)</a> . Production workers (methods unspecified) from 8 chemical plants in West Germany.	2,658 men	<i>cancer deaths</i> leukemia (ICD-9 204-208) lymphohematopoietic (ICD-9 200-208)	<i>observed</i> 2 5	<i>expected</i> 2.35 5	<i>SMR (95% CI)</i> 0.85 (0.10, 3.1) 1.0 (0.32, 2.3)	Insufficient follow-up; 10.1% of cohort had died (268 deaths). Exposure to other chemicals.
<a href="#">Benson and Teta (1993)</a> . Follow-up of only the chlorohydrin-exposed employees from <a href="#">Greenberg et al. (1990)</a> cohort. Production workers at a chemical plant in West Virginia.	278 men	<i>cancer deaths</i> leukemia and aleukemia lymphosarcoma and reticulosarcoma lymphohematopoietic (ICD NS)	<i>observed</i> 4 1 8	<i>expected</i> 1.14 0.50 2.72	<i>SMR (95% CI)</i> 3.5 (0.96, 8.9) 2.0 (0.05, 11) 2.9 (1.3, 5.8)	EtO exposures reported to be low in the chlorohydrin process. Exposure to other chemicals. Very small cohort; thus, small numbers of specific cancers despite long follow-up (52.9% had died; 147 deaths).

This document is a draft for review purposes only and does not constitute Agency policy.

**Table 3-1. Epidemiological studies of ethylene oxide and human cancer—lymphohematopoietic cancer results<sup>a</sup> (continued)**

Study/Population/ Industry	Number of subjects	Lymphohematopoietic cancer results				Comments																		
<p><a href="#">Swaen et al. (2009)</a>. Update of <a href="#">Teta et al. (1993)</a> [<a href="#">Greenberg et al. (1990)</a> cohort minus all chlorohydrin-exposed employees] plus cohort enumeration extended an additional 10 years, adding 167 workers. Production workers and users at 2 chemical plants in West Virginia.</p>	2,063 men	<p><i>cancer deaths</i></p> <table border="0"> <tr> <td></td> <td style="text-align: center;"><i>observed</i></td> <td style="text-align: center;"><i>expected</i></td> <td style="text-align: center;"><i>SMR (95% CI)</i></td> </tr> <tr> <td>leukemia</td> <td style="text-align: center;">11</td> <td style="text-align: center;">11.8</td> <td style="text-align: center;">0.93 (0.47, 1.7)</td> </tr> <tr> <td>leukemia (in workers hired before 1956)</td> <td style="text-align: center;">9</td> <td style="text-align: center;">NR</td> <td style="text-align: center;">1.5 (0.69, 2.9)</td> </tr> <tr> <td>NHL</td> <td style="text-align: center;">12</td> <td style="text-align: center;">11.5</td> <td style="text-align: center;">1.05 (0.54, 1.8)</td> </tr> <tr> <td>lymphohematopoietic (ICD NS)</td> <td style="text-align: center;">27</td> <td style="text-align: center;">30.4</td> <td style="text-align: center;">0.89 (0.59, 1.3)</td> </tr> </table> <p><i>Internal Cox regression analyses:</i> No statistically significant trends were observed for lymphoid (17 deaths) or leukemia (11 deaths) cancer categories for continuous cumulative exposure.</p>		<i>observed</i>	<i>expected</i>	<i>SMR (95% CI)</i>	leukemia	11	11.8	0.93 (0.47, 1.7)	leukemia (in workers hired before 1956)	9	NR	1.5 (0.69, 2.9)	NHL	12	11.5	1.05 (0.54, 1.8)	lymphohematopoietic (ICD NS)	27	30.4	0.89 (0.59, 1.3)		<p>Small cohort; long follow-up time (50.8% had died; 1,048 deaths). Crude exposure assessment, especially for the early time periods. Exposure to other chemicals.</p>
	<i>observed</i>	<i>expected</i>	<i>SMR (95% CI)</i>																					
leukemia	11	11.8	0.93 (0.47, 1.7)																					
leukemia (in workers hired before 1956)	9	NR	1.5 (0.69, 2.9)																					
NHL	12	11.5	1.05 (0.54, 1.8)																					
lymphohematopoietic (ICD NS)	27	30.4	0.89 (0.59, 1.3)																					
<p><a href="#">Steenland et al. (2004)</a>. Update of <a href="#">Steenland et al. (1991)</a>, <a href="#">Stayner et al. (1993)</a>. Sterilizers of medical equipment and spices; and manufacturers and testers of medical sterilization equipment, in 14 plants in the United States.</p>	18,254 (45% male, 55% female)	<p><i>cancer deaths</i></p> <table border="0"> <tr> <td></td> <td style="text-align: center;"><i>observed</i></td> <td style="text-align: center;"><i>expected</i></td> <td style="text-align: center;"><i>SMR (95% CI)</i></td> </tr> <tr> <td>leukemia (ICD-9 204-208)</td> <td style="text-align: center;">29</td> <td style="text-align: center;">NR</td> <td style="text-align: center;">0.99 (0.71, 1.36)</td> </tr> <tr> <td>NHL (ICD-9 200+202)</td> <td style="text-align: center;">31</td> <td style="text-align: center;">NR</td> <td style="text-align: center;">1.00 (0.72, 1.35)</td> </tr> <tr> <td>lymphohematopoietic (ICD-9 200-208)</td> <td style="text-align: center;">79</td> <td style="text-align: center;">NR</td> <td style="text-align: center;">1.00 (0.79, 1.24)</td> </tr> </table> <p><i>Internal Cox regression analyses:</i> “lymphoid” cancers (ICD-9 200, 202, 203, 204): OR = 3.0 (<math>p = 0.046</math>) in highest cumulative exposure group, with 15-yr lag; significant regression coefficient for continuous log cumulative exposure (<math>p = 0.02</math>). lymphohematopoietic cancer (ICD-9 200-208): OR = 2.96 (<math>p = 0.03</math>) in highest cumulative exposure group, with 15-yr lag; significant regression coefficient for continuous log cumulative exposure (<math>p = 0.009</math>).</p>		<i>observed</i>	<i>expected</i>	<i>SMR (95% CI)</i>	leukemia (ICD-9 204-208)	29	NR	0.99 (0.71, 1.36)	NHL (ICD-9 200+202)	31	NR	1.00 (0.72, 1.35)	lymphohematopoietic (ICD-9 200-208)	79	NR	1.00 (0.79, 1.24)		<p>Large cohort; thus, substantial number of deaths (2,852) despite short follow-up (15.6% had died). High-quality exposure assessment. No evidence of exposure to other occupational carcinogens. No increase in lymphohematopoietic cancer risk with increase in exposure in women. Results from internal Cox regression analyses for both sexes combined from Sections D.3 and D.4 of Appendix D.</p>				
	<i>observed</i>	<i>expected</i>	<i>SMR (95% CI)</i>																					
leukemia (ICD-9 204-208)	29	NR	0.99 (0.71, 1.36)																					
NHL (ICD-9 200+202)	31	NR	1.00 (0.72, 1.35)																					
lymphohematopoietic (ICD-9 200-208)	79	NR	1.00 (0.79, 1.24)																					

This document is a draft for review purposes only and does not constitute Agency policy.

**Table 3-1. Epidemiological studies of ethylene oxide and human cancer—lymphohematopoietic cancer results<sup>a</sup>  
(continued)**

Study/Population/ Industry	Number of subjects	Lymphohematopoietic cancer results				Comments
<a href="#">Bisanti et al. (1993)</a> . Chemical workers licensed to handle EtO and other toxic chemicals, Italy.	1,971 men	<i>cancer deaths</i> leukemia (ICD-9 204-208) lymphosarcoma and reticulosarcoma (ICD-9 200) lymphohematopoietic (ICD-9 200-208) <i>in group only licensed to handle EtO (n = 637):</i> leukemia lymphosarcoma and reticulosarcoma lymphohematopoietic	<i>observed</i> 2 4 6  2 3 5	<i>expected</i> 1.0 0.6 2.4  0.3 0.2 0.7	<i>SMR (95% CI)</i> 1.9 (0.23, 7.0) 6.8 (1.9, 17) 2.5 (0.91, 5.5)  6.5 (0.79, 23) 17 (3.5, 50) 7.0 (2.3, 16)	Insufficient follow-up; 3.9% of cohort had died (76 deaths). Exposure to other chemicals.
<a href="#">Hagmar et al. (1995)</a> and <a href="#">Hagmar et al. (1991)</a> . Two plants that produced disposable medical equipment, Sweden.	2,170 (861 men, 1,309 women)	<i>cancer cases</i> leukemia (ICD-7 204-205) NHL (ICD-7 200+202) lymphohematopoietic (ICD-7 200-209) leukemia (among subjects with at least 0.14 ppm-years of cumulative exposure and 10 years latency)	<i>observed</i> 2 2 6  2	<i>expected</i> 0.82 1.25 3.37  0.28	<i>SIR (95% CI)</i> 2.4 (0.30, 8.8) 1.6 (0.19, 5.8) 1.8 (0.65, 3.9)  7.1 (0.87, 26)	Short follow-up period (only 40 cancer cases). (See Section J.2.2 and Table J-2 of Appendix J for a more recent follow- up of this cohort.)
<a href="#">Norman et al. (1995)</a> . Sterilizers of medical equipment and supplies that were assembled at this plant, New York.	1,132 (204 men, 928 women)	<i>cancer cases</i> leukemia (ICD NS)	<i>observed</i> 1	<i>expected</i> 0.54	<i>SIR (95% CI)</i> 1.85 (0.05, 10) <sup>b</sup>	Short follow-up period and small cohort (only 28 cancer cases).

This document is a draft for review purposes only and does not constitute Agency policy.

**Table 3-1. Epidemiological studies of ethylene oxide and human cancer—lymphohematopoietic cancer results<sup>a</sup> (continued)**

Study/Population/ Industry	Number of subjects	Lymphohematopoietic cancer results			Comments
<a href="#">Swaen et al. (1996).</a> Nested case-control study; cases and controls from a large chemical production plant, Belgium.	10 cases of Hodgkin lymphoma (3 exposed; 7 confirmed) and 200 controls; all male	<i>cancer</i> Hodgkin lymphoma (ICD 201)	<i>OR (95% CI)</i> 8.5 (1.4, 40)	Hypothesis-generating study to investigate a cluster of Hodgkin lymphomas observed at a chemical plant. Exposure to other chemicals.	
<a href="#">Olsen et al. (1997).</a> Four EtO production plants (chlorohydrin process) in 3 states.	1,361 men	<i>cancer deaths</i> leukemia (ICD-8 204-207) lymphosarcoma and reticulosarcoma (ICD-8 200) lymphohematopoietic (ICD-8 200-209)	<i>observed expected SMR (95% CI)</i> 2 3.0 0.67 (0.08, 2.4) 1 1.1 0.91 (0.02, 5.1) 10 7.7 1.3 (0.62, 2.4)	Short follow-up and small cohort; 22.0% had died; 300 deaths. Exposure to other chemicals.	
<a href="#">Kardos et al. (2003).</a> Female workers from pediatric clinic of hospital in Eger, Hungary.	299 women	1 lymphoid leukemia death; expected number not reported.			Short follow-up period and small cohort (11 cancer deaths). Possible exposure to natural radium, which permeates the region.

<sup>a</sup>Extracted from Table A-5 of Appendix A, with addition of some summary results (e.g., SMRs); see Table A-5 and Appendix A for more study details.

<sup>b</sup>Calculated by EPA assuming Poisson distribution.

ICD NS: ICD codes not specified; NR: not reported

**Table 3-2. Summary of epidemiological results on EtO and female breast cancer (all sterilizer workers)<sup>a</sup>**

Study	Number of Women	Breast Cancer Results	Comments																				
<a href="#">Hogstedt et al. (1986)</a> and <a href="#">Hogstedt (1988)</a> Swedish incidence and mortality study	170	not reported	8 deaths (7 from cancer) had occurred among the women; breakdown by cancer type not reported.																				
<a href="#">Coggon et al. (2004)</a> Great Britain mortality study	1,011 women hospital workers	<table border="0"> <tr> <td><i>exposure category</i></td> <td><i>observed</i></td> <td><i>expected</i></td> <td><i>SMR (95% CI)</i></td> </tr> <tr> <td>continual</td> <td>5</td> <td>7.2</td> <td></td> </tr> <tr> <td>intermittent</td> <td>0</td> <td>0.7</td> <td></td> </tr> <tr> <td>unknown</td> <td>6</td> <td>5.2</td> <td></td> </tr> <tr> <td>ALL</td> <td>11</td> <td>13.1</td> <td>1.04 (0.42, 1.51)</td> </tr> </table>	<i>exposure category</i>	<i>observed</i>	<i>expected</i>	<i>SMR (95% CI)</i>	continual	5	7.2		intermittent	0	0.7		unknown	6	5.2		ALL	11	13.1	1.04 (0.42, 1.51)	11 breast cancer deaths. 14% of the cohort of 1,405 (including males) hospital workers had died.
<i>exposure category</i>	<i>observed</i>	<i>expected</i>	<i>SMR (95% CI)</i>																				
continual	5	7.2																					
intermittent	0	0.7																					
unknown	6	5.2																					
ALL	11	13.1	1.04 (0.42, 1.51)																				
<a href="#">Steenland et al. (2004)</a> U.S. mortality study	9,908	SMR in highest quartile of cumulative exposure (with 20-yr lag) = 2.07 ( $p < 0.05$ ). significant Cox regression coefficient for log cumulative exposure (20-yr lag) ( $p = 0.01$ ).	103 breast cancer deaths.																				
<a href="#">Steenland et al. (2003)</a> U.S. breast cancer incidence study; nested within <a href="#">Steenland et al. (2004)</a> cohort	7,576 employed for $\geq 1$ yr; 5,139 with interviews	<i>full cohort results:</i> Cox regression analysis OR = 1.74 (95% CI: 1.16, 2.65) for highest cumulative exposure quintile (15-yr lag). $p = 0.05$ for regression coefficient with log cumulative exposure (15-yr lag).  <i>subcohort results:</i> Cox regression analysis OR = 1.87 (95% CI: 1.12, 3.10) for highest cumulative exposure quintile (15-yr lag). $p = 0.02$ for regression coefficient with cumulative exposure (15-yr lag); $p = 0.03$ with log cumulative exposure (15-yr lag).	319 cases in full cohort. 233 cases in subcohort with interviews.																				

This document is a draft for review purposes only and does not constitute Agency policy.

**Table 3-2. Summary of epidemiological results on EtO and female breast cancer (all sterilizer workers)<sup>a</sup>  
(continued)**

Study	Number of Women	Breast Cancer Results	Comments
<a href="#">Hagmar et al. (1995)</a> and <a href="#">Hagmar et al. (1991)</a> Swedish cancer incidence study	1,309	5 cases vs. 10.8 expected SIR = 0.46 (95% CI: 0.15, 1.08).	5 cases. (See Section J.2.2 and Table J-2 of Appendix J for a more recent follow-up of this cohort.)
<a href="#">Norman et al. (1995)</a> U.S. cancer incidence study	928	SIRs ranged from 1.72 (95% CI: 0.99, 3.00) to 2.40 (95% CI: 1.32, 4.37) depending on calendar year of follow-up, assumptions about completeness of follow-up, and reference rates used.	12 cases.
<a href="#">Kardos et al. (2003)</a> Hungarian mortality study	299	11 cancer deaths observed compared with 4.38, 4.03, or 4.28 expected ( $p < 0.01$ ), based on comparison populations of Hungary, Heves County, and city of Eger, respectively; 3 were breast cancer deaths, i.e., 3 breast cancer deaths vs. ~4.3 total deaths expected. Although the expected number of breast cancer deaths was not reported, the number of breast cancer deaths observed for the total deaths expected is indicative of an increased risk of breast cancer (see footnote #5 in Section 3.1).	3 breast cancer deaths.

<sup>a</sup>Extracted from Table A-5 of Appendix A; see Table A-5 and Appendix A for more study details, and also Table 3-1 above.

1 [Steenland et al. \(2003\)](#) studies, on the other hand, used the largest cohort of women potentially  
2 exposed to EtO and clearly show significantly increased risks of breast cancer incidence and  
3 mortality based upon internal exposure-response analyses.

4 In summary, the most compelling evidence of a cancer risk from human exposure to EtO  
5 is for cancer of the lymphohematopoietic system. Increases in the risk of lymphohematopoietic  
6 cancer are present in most of the studies, manifested as an increase in leukemia and/or cancer of  
7 the lymphoid tissue. The evidence of lymphohematopoietic cancer is strongest in the one study  
8 (the NIOSH study) that appears to possess the fewest limitations. In this large study, a  
9 significant dose-response relationship was evident with cumulative exposure to EtO. However,  
10 this effect was observed primarily in males and the magnitude of the effect was not large.  
11 Similarly, in most of the other studies, the increased risks are not great, and other chemicals in  
12 some of the workplaces cannot be ruled out as possible confounders. Thus, the findings of  
13 increased risks of lymphohematopoietic cancer in the NIOSH and other studies cannot  
14 conclusively be attributed to exposure to EtO. The few studies that fail to demonstrate any  
15 increased risks of cancer do not have those strengths of study design that give confidence to the  
16 reported lack of an exposure-related effect.

17 There is also evidence of an elevated risk of breast cancer from exposure to EtO in a few  
18 studies. The strongest evidence again comes from the large NIOSH studies, which found  
19 positive exposure-response relationships for both breast cancer incidence and mortality (319  
20 incident breast cancer cases; 103 breast cancer deaths). Of the five other studies that included  
21 females, none approached the size of the NIOSH studies in terms of breast cancer data – the  
22 study with the next largest breast cancer database had only 12 cases. Nonetheless, two of the  
23 five other studies were supportive of an increased risk of breast cancer.<sup>12</sup>

### 24 25 **3.2. EVIDENCE OF CANCER IN LABORATORY ANIMALS**

26 The International Agency for Research on Cancer (IARC) monograph ([IARC, 1994b](#)) has  
27 summarized the rodent studies of carcinogenicity, and [Health Canada \(2001\)](#) has used this  
28 information to derive levels of concern for human exposure. EPA concludes that the IARC  
29 summary of the key studies is valid and is not aware of any animal cancer bioassays that have  
30 been published since 1994. The Ethylene Oxide Industry Council ([EOIC, 2001](#)) also reviewed  
31 the same studies and did not cite additional studies. The qualitative results are described here  
32 and the incidence data are tabulated in the unit risk derivation section of this document.

---

<sup>12</sup> A more recent follow-up of a third study of the five other studies that was published after the cutoff date for literature inclusion in the main text of this assessment is also supportive of an increased risk of breast cancer (see Section J.2.2 and Table J-2 of Appendix J).

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 One study of oral administration in rats has been published; there are no oral studies in  
2 mice. [Dunkelberg \(1982\)](#) administered EtO in vegetable oil to groups of 50 female  
3 Sprague-Dawley rats by gastric intubation twice weekly for 150 weeks. There were two control  
4 groups (untreated and oil gavage) and two treated groups (7.5 and 30 mg/kg-day). A  
5 dose-dependent increase in the incidence of malignant tumors in the forestomach was observed  
6 in the treated groups (8/50 and 31/50 in the low- and high-dose groups, respectively). Of the  
7 39 tumors, 37 were squamous cell carcinomas, and metastases to other organs were common in  
8 these animals. This study was not evaluated quantitatively because oral risk estimates are  
9 beyond the scope of this document.

10 One inhalation assay was reported in mice ([NTP, 1987](#)) and two inhalation assays were  
11 reported in rats [([Lynch et al., 1984a](#); [Lynch et al., 1984b](#)) in males; ([Garman et al., 1986, 1985](#);  
12 [Snellings et al., 1984](#)) in both males and females]. In the National Toxicology Program (NTP)  
13 mouse bioassay ([NTP, 1987](#)), groups of 50 male and 50 female B6C3F<sub>1</sub> mice were exposed to  
14 EtO via inhalation at concentrations of 0, 50, and 100 ppm for 6 hours per day, 5 days per week,  
15 for 102 weeks. Mean body weights were similar for treated and control animals, and there was  
16 no decrease in survival associated with treatment. A concentration-dependent increase in the  
17 incidence of tumors at several sites was observed in both sexes. These data are summarized in  
18 Table 3-3. Males had carcinomas and adenomas in the lung. Females had carcinomas and  
19 adenomas in the lung, malignant lymphomas, adenocarcinomas in the uterus, and  
20 adenocarcinomas in the mammary glands. The NTP also reports that both sexes had dose-related  
21 increased incidences of cystadenomas of the Harderian glands, but these are benign lesions and  
22 are not considered further here.

23 In the Lynch et al. ([Lynch et al., 1984a](#); [Lynch et al., 1984b](#)) bioassay in male Fischer  
24 344 (F344) rats, groups of 80 animals were exposed to EtO via inhalation at concentrations of 0,  
25 50, and 100 ppm for 7 hours per day, 5 days per week, for 2 years. Mean body weights were  
26 statistically significantly decreased in both treated groups compared with controls ( $p < 0.05$ ).  
27 Increased mortality was observed in the treated groups, and the increase was statistically  
28 significant in the 100-ppm exposure group ( $p < 0.01$ ). [Lynch et al. \(1984a\)](#) suggest that survival  
29 was affected by a pulmonary infection alone and in combination with EtO exposure.  
30 Concentration-dependent increases in the incidence of mononuclear cell leukemia in the spleen,  
31 peritoneal mesothelioma in the testes, and glioma in the brain were observed (see Table 3-4).  
32 The fact that the increased incidence of mononuclear cell leukemia was statistically significant in  
33 the low-exposure group, but not in the high-exposure group, is probably attributable to the  
34 increased mortality in the high-exposure group. The increased incidence in just the terminal kill  
35 rats in the 100-ppm group was statistically significant compared with controls.

36 *This document is a draft for review purposes only and does not constitute Agency policy.*

1 **Table 3-3. Tumor incidence data in National Toxicology Program Study of**  
 2 **B6C3F<sub>1</sub> mice [NTP \(1987\)](#)<sup>a</sup> and exposure-response modeling results<sup>b</sup>**  
 3

Gender/tumor type	EtO concentration (time-weighted average) <sup>c</sup>			EC <sub>10</sub> (LEC <sub>10</sub> ) <sup>d</sup> , (mg/m <sup>3</sup> )	Unit risk (0.1/LEC <sub>10</sub> ) (per mg/m <sup>3</sup> )
	0 ppm	50 ppm (16.3 mg/m <sup>3</sup> )	100 ppm (32.7 mg/m <sup>3</sup> )		
<b>Males</b>					
Lung adenomas plus Carcinomas	11/49	19/49	26/49 <sup>e</sup>	6.94 (4.51)	2.22 × 10 <sup>-2</sup>
<b>Females</b>					
Lung adenomas plus Carcinomas	2/44	5/44	22/49 <sup>f</sup>	14.8 (9.12)	1.1 × 10 <sup>-2</sup>
Malignant Lymphoma	9/44	6/44	22/49 <sup>g</sup>	21.1 (13.9)	7.18 × 10 <sup>-3</sup>
Uterine Carcinoma	0/44	1/44	5/49 <sup>h</sup>	32.8 (23.1)	4.33 × 10 <sup>-3</sup>
Mammary carcinoma <sup>i</sup>	1/44	8/44 <sup>g</sup>	6/49	9.69 (5.35)	1.87 × 10 <sup>-2</sup>

4  
 5 <sup>a</sup>Incidence data were adjusted by EPA by eliminating the animals that died prior to the occurrence of the first tumor  
 6 or prior to 52 weeks, whichever was earlier.

7 <sup>b</sup>Statistical analyses and exposure-response modeling were conducted by EPA.

8 <sup>c</sup>Adjusted to continuous exposure from experimental exposure conditions of 6 hr/d, 5 d/wk; 1 ppm = 1.83 mg/m<sup>3</sup>.

9 <sup>d</sup>Calculated by EPA using Tox\_Risk program.

10 <sup>e</sup> $p < 0.01$  (pairwise Fisher's exact test).

11 <sup>f</sup> $p < 0.001$  (pairwise Fisher's exact test).

12 <sup>g</sup> $p < 0.05$  (pairwise Fisher's exact test).

13 <sup>h</sup> $p = 0.058$  by pairwise Fisher's exact test compared to concurrent controls; however, uterine carcinomas are rare  
 14 tumors in female B6C3F<sub>1</sub> mice, and  $p < 0.0001$  by pairwise Fisher's exact test compared to the NTP historical  
 15 control incidence of 1/1,077 for inhalation (air) female B6C3F<sub>1</sub> mice fed the NIH-07 diet.

16 <sup>i</sup>Highest dose was deleted in order to fit a model to the dose-response data.  
 17

1 **Table 3-4. Tumor incidence data in Lynch et al. (1984a; 1984b) study of**  
 2 **male F344 rats and exposure-response modeling results**  
 3

Tumor type	Concentration (time-weighted average) <sup>a</sup>			EC <sub>10</sub> (LEC <sub>10</sub> ) <sup>b</sup> (mg/m <sup>3</sup> )	Unit risk (0.1/LEC <sub>10</sub> ) (per mg/m <sup>3</sup> )
	0 ppm	50 ppm (19.1 mg/m <sup>3</sup> )	100 ppm (38.1 mg/m <sup>3</sup> )		
Splenic mononuclear cell leukemia <sup>c</sup>	24/77	38/79 <sup>d</sup>	30/76	7.11 (3.94)	2.54 × 10 <sup>-2</sup>
Testicular peritoneal mesothelioma	3/78	9/79	21/79 <sup>e</sup>	16.7 (11.8)	8.5 × 10 <sup>-3</sup>
Brain mixed-cell glioma	0/76	2/77	5/79 <sup>e</sup>	65.7 (37.4)	2.68 × 10 <sup>-3</sup>

4  
 5 <sup>a</sup>Adjusted by EPA to continuous exposure from experimental exposure conditions of 7 hr/d, 5 d/wk; 1 ppm = 1.83  
 6 mg/m<sup>3</sup>.

7 <sup>b</sup>Calculated by EPA using Tox\_Risk program.

8 <sup>c</sup>Highest dose deleted while fitting the dose-response data.

9 <sup>d</sup>*p* < 0.05 (pairwise Fisher's exact test).

10 <sup>e</sup>*p* < 0.01 (pairwise Fisher's exact test).

11  
 12  
 13 In the bioassay conducted by [Snellings et al. \(1984\)](#), 120 male and 120 female F344 rats  
 14 in each sex and dose group were exposed to EtO via inhalation at concentrations of 0 (2 control  
 15 groups of 120 rats of each sex were used), 10, 33, and 100 ppm for 6 hours per day, 5 days per  
 16 week, for 2 years, with scheduled kills at 6 (10 rats per group), 12 (10 rats per group), and  
 17 18 (20 rats per group) months. Significant decreases in mean body weight were observed in the  
 18 100-ppm exposure group in males and in the 100-ppm and 33-ppm exposure groups in females.  
 19 During the 15<sup>th</sup> month of exposure, an outbreak of viral sialodacryoadenitis occurred, resulting in  
 20 the deaths of 1–5 animals per group. [Snellings et al. \(1984\)](#) claim that it is unlikely that the viral  
 21 outbreak contributed to the EtO-associated tumor findings. After the outbreak, mortality rates  
 22 returned to preoutbreak levels and were similar for all groups until the 20<sup>th</sup> or 21<sup>st</sup> month, when  
 23 cumulative mortality in the 33-ppm and 100-ppm exposure groups of each sex remained above  
 24 control values. By the 22<sup>nd</sup> or 23<sup>rd</sup> months, mortality was statistically significantly increased in  
 25 the 100-ppm exposure groups of both sexes.

26 In males, concentration-dependent increases in the incidence of mononuclear cell  
 27 leukemia in the spleen and peritoneal mesothelioma in the testes were observed, and in females  
 28 an increase in mononuclear cell leukemia in the spleen was seen. These data are summarized in  
 29 Table 3-5. Note that these investigators observed the same types of tumors (splenic leukemia  
 30 and peritoneal mesothelioma) seen by [Lynch et al. \(1984a\)](#) and [Lynch et al. \(1984b\)](#). [Snellings](#)

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 [et al. \(1984\)](#) only report incidences (of incidental and nonincidental primary tumors for all  
2 exposure groups) for the 24-month (terminal) kill. However, in their paper they state that  
3 significant findings for the mononuclear cell leukemias were also obtained when all rats were  
4 included and that a mortality-adjusted trend analysis yielded positive findings for the  
5 EtO-exposed females ( $p < 0.005$ ) and males ( $p < 0.05$ ). Similarly, [Snellings et al. \(1984\)](#) report  
6 that when male rats with unscheduled deaths were included in the analysis of peritoneal  
7 mesotheliomas, it appeared that EtO exposure was associated with earlier tumor occurrence, and  
8 a mortality-adjusted trend analysis yielded a significant positive trend ( $p < 0.005$ ). In later  
9 publications describing brain tumors ([Garman et al., 1986, 1985](#)), both males and females had a  
10 concentration-dependent increased incidence of brain tumors (see Table 3-5). [Garman et al.](#)  
11 [\(1986\)](#) and [Garman et al. \(1985\)](#) report incidences including all rats from the 18- and 24-month  
12 kills and all rats found dead or killed moribund. The earliest brain tumors were observed in rats  
13 killed at 18 months.

14

### 15 **3.2.1. Conclusions Regarding the Evidence of Cancer in Laboratory Animals**

16 In conclusion, EtO causes cancer in laboratory animals. After inhalation exposure to  
17 EtO, statistically significant increased incidences of cancer have been observed in both rats and  
18 mice, in both males and females, and in multiple tissues (lung, mammary gland, uterus,  
19 lymphoid cells, brain, tunica vaginalis testis). In addition, one oral study in rats has been  
20 conducted, and a significant dose-dependent increase in carcinomas of the forestomach was  
21 reported.

22

## 23 **3.3. SUPPORTING EVIDENCE**

### 24 **3.3.1. Metabolism and Kinetics**

25 Information on the kinetics and metabolism of EtO has been derived primarily from  
26 studies conducted with laboratory animals exposed via inhalation, although some limited data  
27 from humans have been identified. Details are available in several reviews ([Fennell and Brown,](#)  
28 [2001](#); [Csanady et al., 2000](#); [Brown et al., 1998](#); [Brown et al., 1996](#)).

29 Following inhalation, EtO is absorbed efficiently into the blood and rapidly distributed to  
30 all organs and tissues. EtO is metabolized primarily by two pathways (see Figure 3-1):  
31 (1) hydrolysis to ethylene glycol (1,2-ethanediol), with subsequent conversion to oxalic acid,  
32 formic acid, and carbon dioxide; and (2) glutathione conjugation and the formation of  
33 S-(2-hydroxyethyl)cysteine and N-acetylated derivatives ([WHO, 2003](#)). From the available data,  
34 the route involving conjugation with glutathione appears to predominate in mice; in larger  
35 species (including humans), the conversion of EtO is primarily via hydrolysis through ethylene

*This document is a draft for review purposes only and does not constitute Agency policy.*

**Table 3-5. Tumor incidence data in [Snellings et al. \(1984\)](#) and [Garman et al. \(1985\)](#) reports on F344 rats<sup>a</sup> and exposure-response modeling results<sup>b</sup>**

Gender/tumor type	Concentration (time-weighted average) <sup>c</sup>				EC <sub>10</sub> (LEC <sub>10</sub> ) <sup>e</sup> (mg/m <sup>3</sup> )	Unit risk (0.1/LEC <sub>10</sub> ) (per mg/m <sup>3</sup> )
	0 ppm <sup>d</sup>	10 ppm (3.27 mg/m <sup>3</sup> )	33 ppm (10.8 mg/m <sup>3</sup> )	100 ppm (32.7 mg/m <sup>3</sup> )		
<b>Males</b>						
Splenic mononuclear cell leukemia	13/97 (13%) <sup>f</sup>	9/51 (18%)	12/39 <sup>g</sup> (32%)	9/30 <sup>g</sup> (30%)	12.3 (6.43)	1.56 × 10 <sup>-2</sup>
Testicular peritoneal mesothelioma	2/97 (2.1%)	2/51 (3.9%)	4/39 (10%)	4/30 <sup>g</sup> (13%)	22.3 (11.6)	8.66 × 10 <sup>-3</sup>
Primary brain tumors	1/181 (0.55%)	1/92 (1.1%)	5/85 <sup>g</sup> (5.9%)	7/87 <sup>h</sup> (8.1%)	36.1 (22.3)	4.5 × 10 <sup>-3</sup>
<b>Females</b>						
Splenic mononuclear cell leukemia	11/116 (9.5%)	11/54 <sup>g</sup> (21%)	14/48 <sup>h</sup> (30%)	15/26 <sup>i</sup> (58%)	4.46 (3.1)	3.23 × 10 <sup>-2</sup>
Primary brain tumors	1/188 (0.53%)	1/94 (1.1%)	3/92 (3.3%)	4/80 <sup>g</sup> (5%)	63.8 (32.6)	3.07 × 10 <sup>-3</sup>

<sup>a</sup>Denominators refer to the number of animals for which histopathological diagnosis was performed. For brain tumors, [Garman et al. \(1985\)](#) included animals in the 18-month and the 24-month sacrifice and found dead or euthanized moribund of those alive at the time of the first brain tumor, whereas for the other sites, [Snellings et al. \(1984\)](#) included animals only at the 24-month sacrifice.

<sup>b</sup>Statistical analyses and exposure-response modeling were conducted by EPA.

<sup>c</sup>Adjusted by EPA to continuous exposure from experimental exposure conditions of 6 hr/d, 5 d/wk; 1 ppm = 1.83 mg/m<sup>3</sup>.

<sup>d</sup>Results for both control groups combined.

<sup>e</sup>Calculated by EPA using Tox\_Risk program.

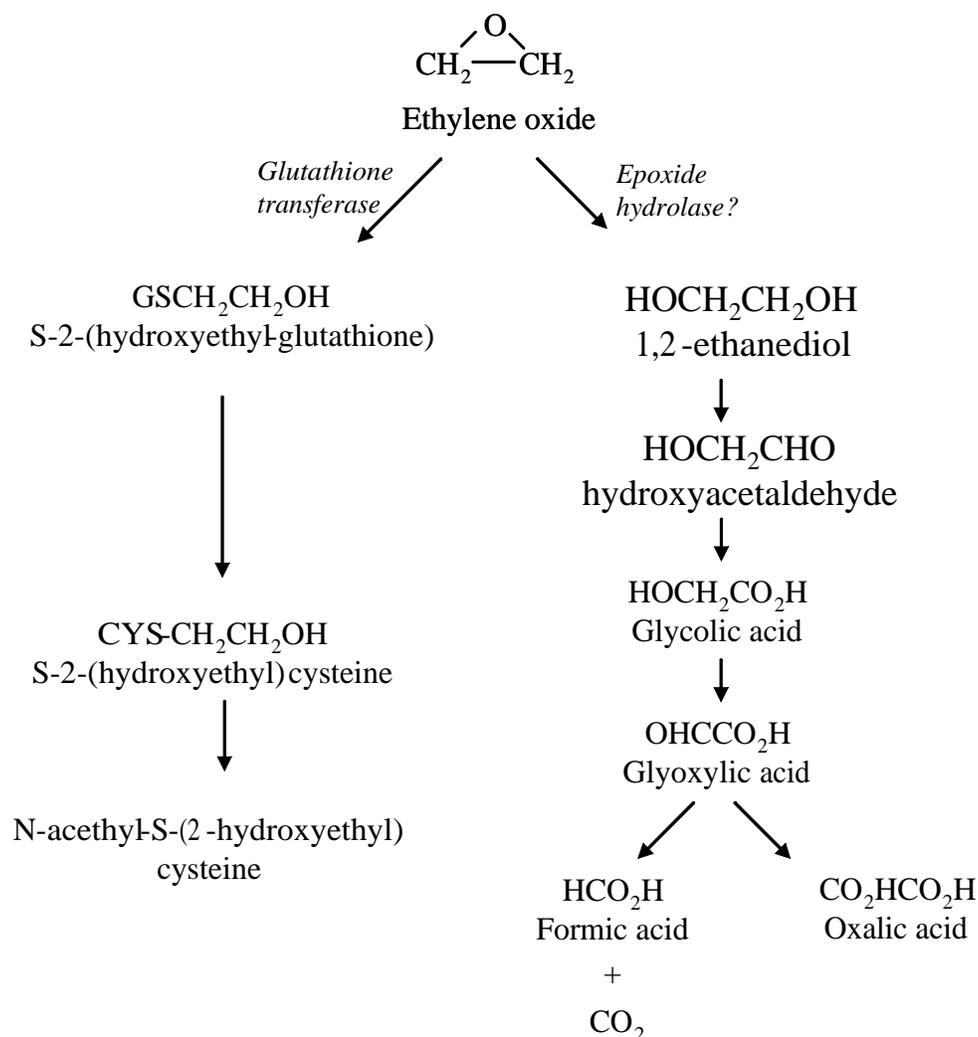
<sup>f</sup>Numbers in parentheses indicate percentage incidence values.

<sup>g</sup>*p* < 0.05 (pairwise Fisher's exact test).

<sup>h</sup>*p* < 0.01 (pairwise Fisher's exact test).

<sup>i</sup>*p* < 0.001 (pairwise Fisher's exact test).

This document is a draft for review purposes only and does not constitute Agency policy.



1      **Figure 3-1. Metabolism of ethylene oxide.**

2  
3  
4 glycol. Because EtO is an epoxide capable of reacting directly with cellular macromolecules,  
5 both pathways are considered to be detoxifying.

6      Among rodent species, there are clear quantitative differences in metabolic rates. The  
7 rate of clearance of EtO from the blood, brain, muscle, and testes was measured by [Brown et al.](#)  
8 [\(1998\)](#) and [Brown et al. \(1996\)](#). Clearance rates were nearly identical across blood and other  
9 tissues. Following a 4-hour inhalation exposure to 100 ppm EtO in mice and rats, the average

1 blood elimination half-lives ranged from 2.4 to 3.2 minutes in mice and 11 to 14 minutes in rats.  
2 The elimination half-life in humans is 42 minutes ([Filser et al., 1992](#)), and the half-life in salt  
3 water is 4 days ([IARC, 1994b](#)).

4 In a more detailed study in mice, [Brown et al. \(1998\)](#) measured EtO concentrations in  
5 mice after 4-hour inhalation exposures at 0, 50, 100, 200, 300, or 400 ppm. They found that  
6 blood EtO concentration increased linearly with inhaled concentrations of less than 200 ppm, but  
7 above 200 ppm the blood concentration increased more rapidly. In addition, glutathione levels in  
8 liver, lung, kidney, and testes decreased as exposures increased above 200 ppm. The  
9 investigators interpreted this, along with other information, to mean that at low concentrations  
10 the metabolism and disappearance of EtO is primarily a result of glutathione conjugation, but at  
11 higher concentrations, when tissue glutathione begins to be depleted, the elimination occurs via a  
12 slower nonenzymatic hydrolysis process, leading to a greater-than-linear increase in blood EtO  
13 concentration.

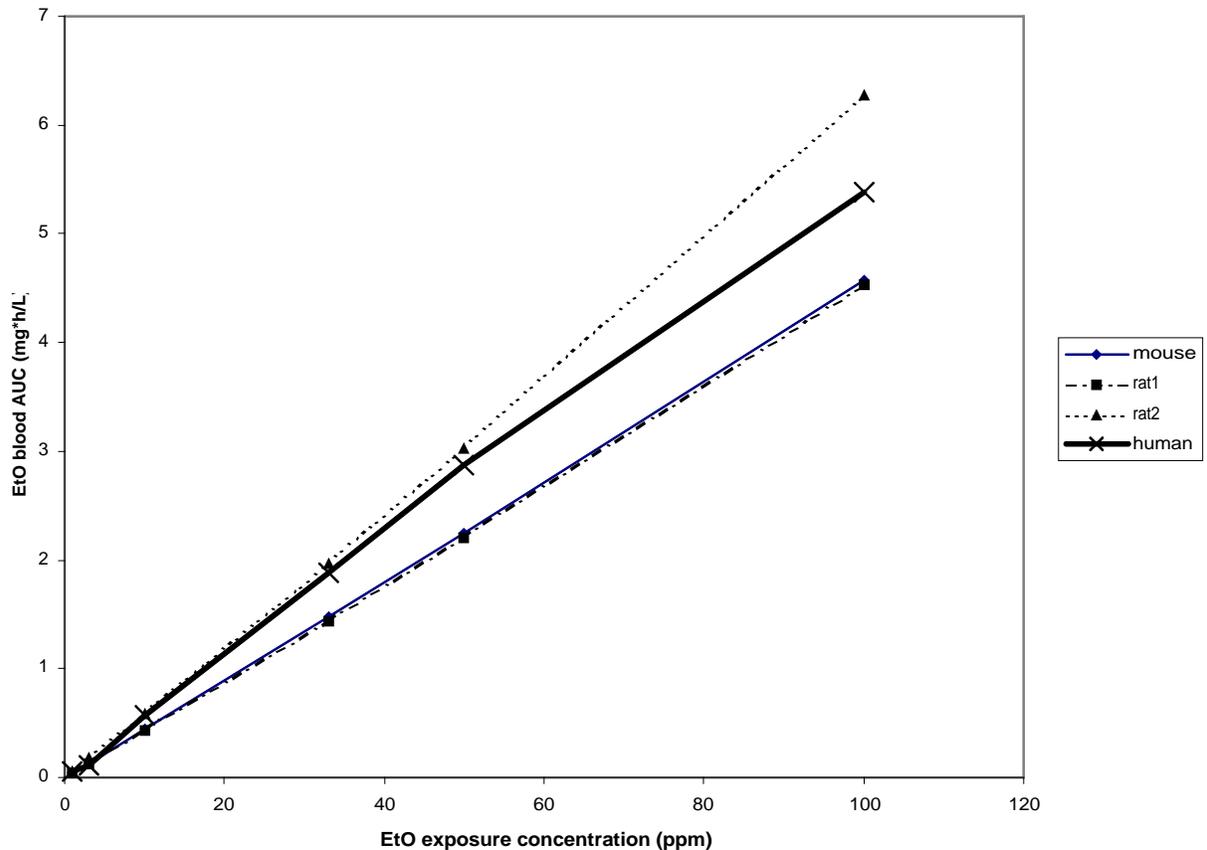
14 [Fennell and Brown \(2001\)](#) constructed physiologically based pharmacokinetic (PBPK)  
15 models of uptake and metabolism in mice, rats, and humans, based on previous studies. They  
16 reported that the models adequately predicted blood and tissue EtO concentrations in rats and  
17 mice, with the exception of the testes, and blood EtO concentrations in humans. Modeling  
18 6-hour inhalation exposures yielded simulated blood peak concentrations and areas under the  
19 curve (AUCs) that are similar for mice, rats, and humans (human levels are within about 15% of  
20 rat and mouse levels; see Figure 3-2). In other words, exposure to a given EtO concentration in  
21 air results in similar predicted blood EtO AUCs for mice, rats, and humans.

22 These studies show that tissue concentrations in mice, rats, and humans exposed to a  
23 particular air concentration of EtO are approximately equal and that they are linearly related to  
24 inhalation concentration, at least in the range of exposures used in the rodent cancer bioassays  
25 (i.e., 100 ppm and below).

### 27 **3.3.2. Protein Adducts**

28 EtO forms DNA (see Section 3.3.3.1) and hemoglobin adducts within tissues throughout  
29 the body ([Walker et al., 1992a](#); [Walker et al., 1992b](#)). Formation of hemoglobin adducts has  
30 been used as a measure of exposure to EtO. The main sites of alkylation are cysteine, histidine,  
31 and the N-terminal valine; however, for analytical reasons, the N-(2-hydroxyethyl)valine adduct  
32 is generally preferred for measurements ([Walker et al., 1990](#)). [Walker et al. \(1992b\)](#) reported  
33 measurements of this hemoglobin adduct and showed how the concentration of the adducts  
34 changes according to the dynamics of red blood cell turnover. [Walker et al. \(1992b\)](#) measured  
35 hemoglobin adduct formation in mice and rats exposed to 0, 3, 10, 33, 100, and 300 (rats only)

*This document is a draft for review purposes only and does not constitute Agency policy.*



1  
2  
3 **Figure 3-2. Simulated blood AUCs for EtO following a 6-hour exposure to**  
4 **EtO from the rat, mouse, and human PBPK models of [Fennell and Brown](#)**  
5 **[\(2001\)](#); based on data presented in [Fennell and Brown \(2001\)](#). (Rat1 and**  
6 **rat2 results use different values for pulmonary uptake.)**

7  
8  
9 ppm of EtO (6 hours/day, 5 days/week, for 4 weeks). Response was linear in both species up to  
10 33 ppm, after which the slope significantly increased. The exposure-related decrease in  
11 glutathione concentration in liver, lung, and other tissues observed by [Brown et al. \(1998\)](#) in  
12 mice is a plausible explanation for the increasing rate of hemoglobin adduct formation at higher  
13 exposures.

14 In humans, hemoglobin adducts can be used as biomarkers of recent exposure to EtO  
15 ([IARC, 2008](#); [Boogaard, 2002](#); [IARC, 1994b](#)), and several studies have reported  
16 exposure-response relationships between hemoglobin adduct levels and EtO exposure levels [e.g.  
17 ([van Sittert et al., 1993](#); [Schulte et al., 1992](#))]. Hemoglobin adducts are good general indicators  
18 of exposure because they are stable (DNA adducts, on the other hand, may be repaired

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 or fixed as mutations and hence are less reliable measures of exposure). However, [Föst et al.](#)  
2 [\(1991\)](#) noted that human erythrocytes showed marked interindividual differences in the amounts  
3 of EtO bound to hemoglobin, and [Yong et al. \(2001\)](#) reported that levels of  
4 N-(2-hydroxyethyl)valine were approximately twofold greater in persons with a *GSTT1*-null  
5 genotype than in those with positive genotypes. Endogenous ethylene oxide (see  
6 Section 3.3.3.1) also contributes to hemoglobin adduct levels, making it more difficult to detect  
7 the impacts of low levels of exogenous EtO exposure. In addition, [Walker et al. \(1993\)](#) reported  
8 that hemoglobin adducts in mice and rats were lost at a greater rate than would be predicted by  
9 the erythrocyte life span.

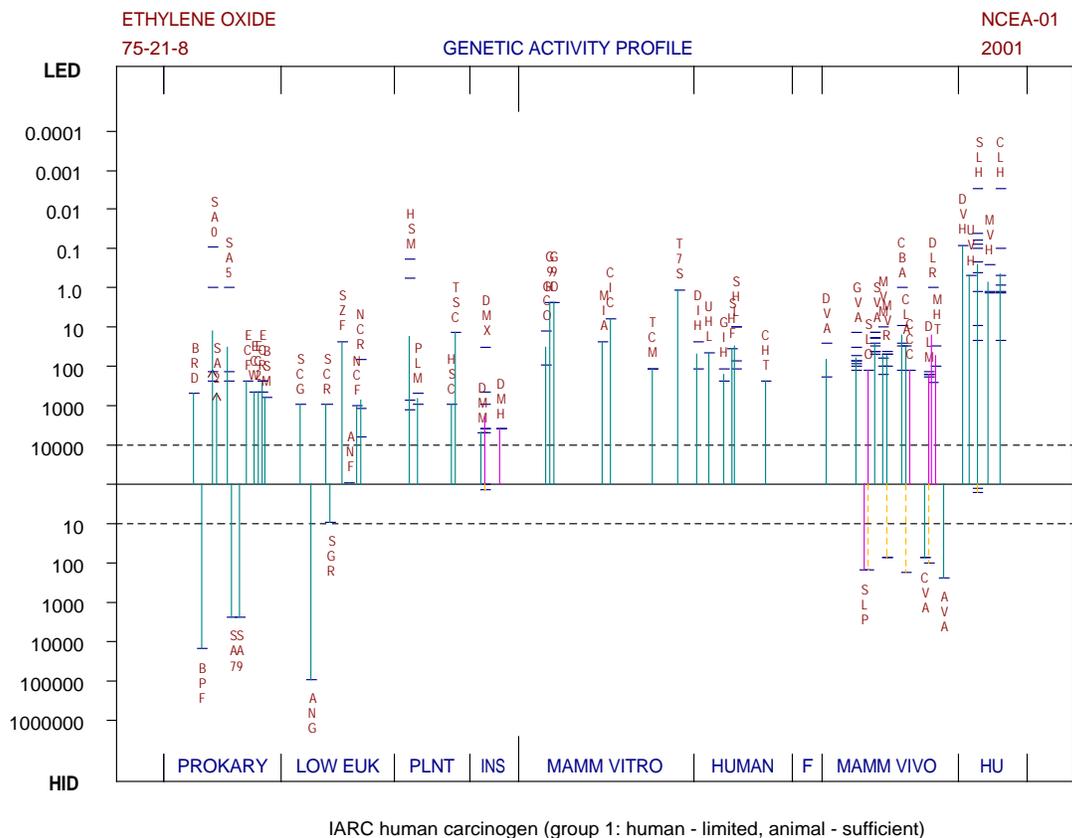
### 11 **3.3.3. Genotoxicity**

12 Since the first report of EtO induction of sex-linked recessive lethals in *Drosophila*  
13 [\(Rapoport, 1948\)](#), numerous papers have been published on the positive genotoxic activity in  
14 biological systems, spanning a wide range of assay systems, from bacteriophage to higher plants  
15 and animals. Figure 3-3 shows the 203 test entries in the EPA Genetic Activity Profile database  
16 in 2001. In prokaryotes and lower eukaryotes, EtO induced DNA damage and gene mutations in  
17 bacteria, yeast, and fungi and gene conversions in yeast. In mammalian cells (from in vitro  
18 and/or in vivo exposures), EtO-induced effects include unscheduled DNA synthesis, gene  
19 mutations, sister chromatid exchanges (SCEs), micronuclei, and chromosomal aberrations.  
20 Genotoxicity, in particular increased levels of SCEs and chromosomal aberrations, has also been  
21 observed in blood cells of workers occupationally exposed to EtO. Several publications contain  
22 details of earlier genetic toxicity studies [e.g. [\(IARC, 2008; Kolman et al., 2002; Thier and Bolt,](#)  
23 [2000; Natarajan et al., 1995; Preston et al., 1995; IARC, 1994b; Dellarco et al., 1990; Ehrenberg](#)  
24 [and Hussain, 1981\)](#)]. This review briefly summarizes the evidence of the genotoxic potential of  
25 EtO, focusing primarily on recently published studies that provide information on the mode of  
26 action of EtO (see Appendix C for more details from some individual studies).

#### 28 **3.3.3.1. DNA Adducts**

29 EtO is a direct-acting S<sub>N</sub>2 (substitution-nucleophilic-bimolecular)-type monofunctional  
30 alkylating agent that forms adducts with cellular macromolecules such as proteins (e.g.,  
31 hemoglobin, see Section 3.3.2) and DNA [\(Pauwels and Veulemans, 1998\)](#). Alkylating agents  
32 may produce a variety of different DNA alkylation products [\(Beranek, 1990\)](#) in varying  
33 proportions, depending primarily on the electrophilic properties of the agent. Reactivity of an  
34 alkylating agent is estimated by its Swain-Scott substrate constant (*s*-value), which ranges from  
35 0 to 1, and EtO has a high *s*-value of 0.96 [\(Beranek, 1990; Golberg, 1986; Warwick, 1963\)](#).

*This document is a draft for review purposes only and does not constitute Agency policy.*



IARC human carcinogen (group 1: human - limited, animal - sufficient)

1  
2 **Figure 3-3. Display of 203 data sets, including bacteria, fungi, plants, insects,**  
3 **and mammals (in vitro and in vivo), measuring the full range of genotoxic**  
4 **endpoints.** [This is an updated version of the figure in [IARC \(1994b\)](#)]  
5

6 See Appendix B for list of references.

7  
8  
9 Acting by the  $S_N2$  mechanism and having a high substrate constant both favor alkylation at the  
10 N7 position of guanine in the DNA ([Walker et al., 1990](#)). The predominant DNA adduct formed  
11 by EtO and other  $S_N2$ -type alkylating agents is N7-(2-hydroxyethyl)guanine (N7-HEG). After in  
12 vitro treatment of DNA with EtO, [Segerbäck \(1990\)](#) identified three adducts, N7-HEG,  
13 N3-hydroxyethyladenine, and O6- hydroxyethylguanine (O<sup>6</sup>-HEG), in the ratios 200:8.8:1; two  
14 other peaks, suspected of representing other adenine adducts, were also observed at levels well  
15 below that of N7-HEG.

16 Ethylene, an endogenous precursor of EtO, is produced during normal physiological  
17 processes. Such processes reportedly include oxidation of methionine and hemoglobin, lipid  
18 peroxidation of fatty acids, and metabolism of intestinal bacteria [reviewed in ([Thier and Bolt,](#)  
19 [2000](#); [IARC, 1994a](#))]. EtO is then endogenously produced through the cytochrome

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 P450-mediated conversion of ethylene ([Törnqvist, 1996](#)). This endogenous production of EtO  
2 contributes significantly to background levels of DNA adducts, making it difficult to detect the  
3 impacts of low levels of exogenous EtO exposure on DNA adduct levels. For example, in DNA  
4 extracted from the lymphocytes of unexposed individuals, mean background levels of N7-HEG  
5 ranged from 2 to 8.5 pmol/mg DNA ([Bolt, 1996](#)). Using sensitive detection techniques and an  
6 approach designed to separately quantify both endogenous N7-HEG adducts and “exogenous”  
7 N7-HEG adducts induced by EtO treatment in rats, [Marsden et al. \(2009\)](#) reported increases in  
8 exogenous adducts in DNA of spleen and liver at the lowest dose administered (0.0001 mg/kg  
9 injected i.p. daily for 3 days) and statistically significant linear dose-response relationships  
10 ( $p < 0.05$ ) for exogenous adducts in all three tissues examined (spleen, liver, and stomach),  
11 although the authors caution that some of the adduct levels induced at low EtO concentrations  
12 are below the limit of accurate quantitation. Note that the whole range of doses studied by  
13 [Marsden et al. \(2009\)](#) lies well below the dose corresponding to the lowest LOAEL from an EtO  
14 cancer bioassay (see Section C.7 of Appendix C). [Marsden et al. \(2009\)](#) also observed increases  
15 in endogenous N7-HEG adduct formation at the two highest doses (0.05 and 0.1 mg/kg),  
16 suggesting that, in addition to direct adduct formation via alkylation, EtO can induce N7-HEG  
17 adduct production indirectly. [Marsden et al. \(2009\)](#) hypothesized that this indirect adduct  
18 formation by EtO results from the induction of ethylene generation under conditions of oxidative  
19 stress.

20 In experiments with rats and mice exposed to EtO at concentrations of 0, 3, 10, 33, 100,  
21 or 300 (rats only) ppm for 6 hours per day, 5 days per week, for 4 weeks, [Walker et al. \(1992a\)](#)  
22 measured N7-HEG adducts in the DNA of lung, brain, kidney, spleen, liver, and testes. At  
23 100 ppm, the adduct levels for all tissues except testis were similar (within a factor of 3), despite  
24 the fact that not all of these tissues are targets for toxicity. The study’s data on the persistence of  
25 the DNA adducts indicate that DNA repair rates differ in different tissues. Although [Walker et](#)  
26 [al. \(1992a\)](#) suggested that N7-HEG adducts are likely to be removed by depurination forming  
27 apurinic/apyrimidinic (AP) sites in DNA, a later study from the same group showed that  
28 EtO-induced DNA damage is repaired without accumulation of AP sites or involving base  
29 excision repair ([Rusyn et al., 2005](#)). Rats exposed to high doses of EtO (300 ppm) by inhalation  
30 showed steady-state levels of O<sup>6</sup>-HEG adducts that are ~250–300 times lower than the N7-HEG  
31 levels ([Walker et al., 1992a](#)). Even though low levels of O<sup>6</sup>-HEG adducts were detected, they  
32 are more mutagenic in nature and may contribute to the tumors observed in target organs.

33 Two studies provide evidence of N7-HEG DNA adduct formation in human populations  
34 occupationally exposed to EtO, one reporting a modest increase in white blood cells ([van Delft et](#)  
35 [al., 1994](#)) and the other a four- to fivefold increase in granulocytes ([Yong et al., 2007](#)) compared

1 to unexposed controls. However, these differences were not statistically significant due to high  
2 interindividual variation in adduct levels.

### 3 4 **3.3.3.2. Point Mutations**

5 EtO has consistently yielded positive results in in vitro mutation assays from  
6 bacteriophage, bacteria, fungi, yeast, insects, plants, and mammalian cell cultures (including  
7 human cells). For example, EtO induces single base pair deletions and base substitutions in the  
8 *HPRT* gene in human diploid fibroblasts ([Kolman and Chovanec, 2000](#); [Lambert et al., 1994](#);  
9 [Bastlová et al., 1993](#)) in vitro. The results of in vivo studies on the mutagenicity of EtO have  
10 also been consistently positive following ingestion, inhalation, or injection [e.g. [Tates et al.](#)  
11 [\(1999\)](#)]. Increases in the frequency of gene mutations in T-lymphocytes (*Hprt* locus) ([Walker et](#)  
12 [al., 1997](#)) and in bone marrow and testes (*LacI* locus) ([Recio et al., 2004](#)) have been observed in  
13 transgenic mice exposed to EtO via inhalation at concentrations similar to those in  
14 carcinogenesis bioassays with this species ([NTP, 1987](#)). At somewhat higher concentrations  
15 than those used in the carcinogenesis bioassays (200 ppm, but for only 4 weeks), increases in the  
16 frequency of gene mutations have also been observed in the lungs of transgenic mice (*LacI*  
17 locus) ([Sisk et al., 1997](#)) and in T-lymphocytes of rats (*Hprt* locus) ([van Sittert et al., 2000](#); [Tates](#)  
18 [et al., 1999](#)). In in vivo studies with male mice, EtO also causes heritable mutations and other  
19 effects in germ cells ([Generoso et al., 1990](#); [Lewis et al., 1986](#)).

20 In a study of mammary gland carcinomas in EtO-exposed B6C3F<sub>1</sub> mice from the 1987  
21 NTP bioassay ([NTP, 1987](#)) and 19 mammary gland carcinomas from concurrent controls in the  
22 1987 NTP EtO bioassay and a 1986 NTP benzene bioassay, [Houle et al. \(2006\)](#) measured  
23 mutation frequencies in exons 5–8 of the *p53* tumor suppressor gene and in codon 61 of the *Hras*  
24 oncogene. Mutation frequencies in the mammary carcinomas of EtO-exposed mice were only  
25 slightly increased over frequencies in spontaneous mammary carcinomas (33% of the  
26 carcinomas in the EtO-exposed mice had *Hras* mutations versus 26% of spontaneous tumors;  
27 67% of the carcinomas in the EtO-exposed mice had *p53* mutations versus 58% of spontaneous  
28 tumors); however, the EtO-induced tumors exhibited a distinct shift in the mutational spectra of  
29 the *p53* and *Hras* genes and more commonly displayed concurrent mutations of the two genes  
30 ([Houle et al., 2006](#)). Furthermore, [Houle et al. \(2006\)](#) detected about sixfold higher levels of p53  
31 protein expression in the mammary carcinomas of EtO-exposed mice than in spontaneous  
32 mammary carcinomas, and there was an apparent dose-response relationship between EtO  
33 exposure level and both p53 protein expression and *p53* gene mutation (three of the seven tumors  
34 in the 50-ppm exposure group and all five tumors in the 100-ppm group had increased protein  
35 expression; also, three *p53* gene mutations were found in the seven tumors in the 50-ppm

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 exposure group and nine were found in the five tumors in the 100-ppm group). Some of the  
2 same investigators conducted a similar study of *Kras* mutations in lung, Harderian gland, and  
3 uterine tumors ([Hong et al., 2007](#)). Substantial increases were observed in *Kras* mutation  
4 frequencies in the tumors from the EtO-exposed mice. *Kras* mutations were reported in 100% of  
5 the lung tumors from EtO-exposed mice versus 25% of spontaneous lung tumors (108 NTP  
6 control animal tumors, including 8 from the EtO bioassay), in 86% of Harderian gland tumors  
7 from EtO-exposed mice versus 7% of spontaneous Harderian gland tumors (27 NTP control  
8 animal tumors, including 2 from the EtO bioassay), and in 83% of uterine tumors from  
9 EtO-exposed mice (there were no uterine tumors in control mice in the 1986 NTP bioassay and  
10 none were examined from other control animals). Furthermore, a specific *Kras* mutation, a  
11 G → T transversion in codon 12, was nearly universal in lung tumors from EtO-exposed mice  
12 (21/23) but rare in lung tumors from control animals (1/108). Other specific mutations were also  
13 predominant in the Harderian gland and uterine tumors, but too few *Kras* mutations were  
14 available in spontaneous Harderian gland tumors, and no spontaneous uterine tumors were  
15 examined; thus, meaningful comparisons could not be made for these sites. Overall, these data  
16 strongly suggest that EtO-induced mutations in oncogenes and tumor-suppressor genes play a  
17 role in EtO-induced carcinogenesis in multiple tissues.

18 Only a few studies have investigated gene mutations in people occupationally exposed to  
19 EtO. In one study, *HPRT* mutant frequency in peripheral blood lymphocytes was measured in a  
20 group of 9 EtO-exposed hospital workers, a group of 15 EtO-exposed factory workers, and their  
21 respective controls ([Tates et al., 1991](#)). EtO exposure scenarios suggest higher exposures in the  
22 factory workers, and this is supported by the measurement of higher hemoglobin adduct levels in  
23 those workers. *HPRT* mutant frequencies were 55% increased in the hospital workers, but the  
24 increase was not statistically significant. In the factory workers, a statistically significant  
25 increase of 60% was reported. In a study of workers in an EtO production facility ([Tates et al.,](#)  
26 [1995](#)), *HPRT* mutations were measured in three exposed groups and one unexposed group (seven  
27 workers per group). No significant differences in mutant frequencies were observed between the  
28 groups; however, the authors stated that about 50 subjects per group would have been needed to  
29 detect a 50% increase.

30 [Major et al. \(2001\)](#) measured *HPRT* mutations in female nurses employed in hospitals in  
31 Eger and Budapest, Hungary. This study and an earlier study measuring effects on chromosomes  
32 (see Table 3-6) were conducted to examine a possible causal relationship between EtO exposure  
33 and a cluster of cancers (mostly breast) in nurses exposed to EtO in the Eger hospital. The  
34 Budapest hospital was chosen because there was no apparent increase in cancer among nurses  
35 exposed to EtO. Controls were female hospital workers in the respective cities, and nurses in

**Table 3-6. Cytogenetic effects in humans**

Number exposed (number of controls)	Exposure time (years)		Ethylene oxide level in air (ppm) <sup>a</sup>		Cytogenetic observations			Reference
	Range	Mean	Range	Mean (TWA)	CA	SCE	MN	
33 (0)	1–14		±0.05–8	±0.01 <sup>b</sup>	(+)			<a href="#">Clare et al. (1985)</a>
Site I: 13 Site II: 22 Site III: 25–26 (171 total)			0.5 <sup>c</sup> 5–10 <sup>c</sup> 5–20 <sup>c</sup>		– – +	– + +		<a href="#">Stolley et al. (1984)</a> , <a href="#">Galloway et al. (1986)</a>
12 (12)			±36			+		<a href="#">Garry et al. (1979)</a>
14 (14)			<0.07–4.3 <sup>c</sup>			–		<a href="#">Hansen et al. (1984)</a>
Factory I: 18 Factory II: 10 (20 total)	0.5–8 0.5–8	3.2 1.7		<1 <1	+ +	– –	+ <sup>d</sup>	<a href="#">Högstedt et al. (1983)</a>
15 smokers (7) 10 nonsmokers (15)	0.5–10 0.5–10	5.7 4.5	20–123 20–123			+ +		<a href="#">Laurent et al. (1984)</a>
10 (10)		3	60–69 <sup>c</sup>		+	+		<a href="#">Lerda and Rizzi (1992)</a>
Low dose: 9 (48) High dose: 27 (10)		4 15	2.7–10.9 2.7–82	2.7 5.5	+ +	– +		<a href="#">Major et al. (1996)</a>
34 (23)		8 <sup>e</sup>	<0.1–2.4 <sup>c</sup>	<0.3	–	+		<a href="#">Mayer et al. (1991)</a>
11 smokers 14 nonsmokers (10 total)			0.5–417 <sup>f</sup> 0.5–208 <sup>f</sup>			– –		<a href="#">Popp et al. (1994)</a>
75 (22)	3–14	7	2–5 <sup>c</sup>		+		+	<a href="#">Ribeiro et al. (1994)</a>
56 (141)	1–10		1–40 <sup>c</sup>		+	+		<a href="#">Richmond et al. (1985)</a>

This document is a draft for review purposes only and does not constitute Agency policy.

**Table 3-6. Cytogenetic effects in humans (continued)**

Number exposed (number of controls)	Exposure time (years)		Ethylene oxide level in air (ppm) <sup>a</sup>		Cytogenetic observations			Reference
	Range	Mean	Range	Mean (TWA)	CA	SCE	MN	
22 (22) 19 (19)	0.6–4 1.5–15	3 6.8	0.2–0.5 <sup>c</sup> 3.7–20 <sup>c</sup>	0.35 10.7	(+) +	+ +		<a href="#">Sarto et al. (1984)</a>
10 (10)			0–9.3 <sup>c</sup>	1.84		+		<a href="#">Sarto et al. (1987)</a>
9 3 (27 total)	0.5–12	5	0.025–0.38 <sup>c</sup> >0.38 <sup>g</sup>				– + <sup>h</sup>	<a href="#">Sarto et al. (1990)</a>
5 5 (10 total)	0.1–4 4–12	2 8.6	<1–4.4	0.025 0.38		– +	– <sup>i</sup> – <sup>i</sup>	<a href="#">Sarto et al. (1991)</a>
32 11 (8 total)		5.1 9.5	0–0.3 <sup>c</sup> 0.1 3–0.3 <sup>c</sup>	0.04 0.16		+ +	– –	<a href="#">Schulte et al. (1992)</a>
9 hospital workers (8) 15 factory workers (15)	2–6 3–27	4 12	20–25 17–33		+ +	+ +	– +	<a href="#">Tates et al. (1991)</a>
7 7 7 (7 total)	Accidental <5 >15		28–429 <sup>c</sup> <0.005–0.02 <0.005–0.01			– – –	– – –	<a href="#">Tates et al. (1995)</a>
Low exposure: 9 High exposure: 5 (13 total)				13 <sup>j</sup> 501 <sup>j</sup>		– +		<a href="#">Yager et al. (1983)</a>
19 17 (35 total)	1–5 6–14		<0.05–8 <0.05–8	<0.05 <0.05	– –			<a href="#">Van Sittert and de Jong (1985)</a>

This document is a draft for review purposes only and does not constitute Agency policy.

### Table 3-6. Cytogenetic effects in humans (continued)

<sup>a</sup>1 ppm = 1.83 mg ethylene oxide/m<sup>3</sup>.

<sup>b</sup>Calculated by linear extrapolation.

<sup>c</sup>TWA (8-hr).

<sup>d</sup>Positive for erythroblasts and polychromatic erythrocytes (negative for lymphocytes).

<sup>e</sup>Maximum years exposed.

<sup>f</sup>Peak concentrations.

<sup>g</sup>Exposed acutely from sterilizer leakage in addition to chronic exposure.

<sup>h</sup>Nasal mucosa.

<sup>i</sup>Buccal cells.

<sup>j</sup>Average 6-month cumulative exposure (mg).

CA = chromosomal aberrations

MN = micronucleus

SCE = sister chromatid exchange

TWA = time-weighted average

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 Eger with known cancers were excluded. Mean peak levels of EtO were 5 mg/m<sup>3</sup> (2.7 ppm) in  
2 Budapest and 10 mg/m<sup>3</sup> (5.4 ppm) in Eger. *HPRT* variant frequencies in both controls and  
3 EtO-exposed workers in the Eger hospital were higher than either group in the Budapest hospital,  
4 but there was no significant increase among the EtO-exposed workers in either hospital when  
5 compared with the respective controls. The authors noted that the *HPRT* variant frequencies  
6 among smoking EtO-exposed nurses in Eger were significantly higher than among smokers in  
7 the Eger controls; however, the fact that the *HPRT* variant frequency was almost three times  
8 higher in nonsmokers than in smokers in the Eger hospital control group raises questions about  
9 the basis of the claimed EtO effect.

### 11 **3.3.3.3. Chromosomal Effects**

12 As discussed by [Preston \(1999\)](#) in an extensive review of the cytogenetic effects of EtO,  
13 a variety of cytogenetic assays can be used to measure induced chromosome damage. However,  
14 most of the assays commonly employed measure events that are detectable only in the first (or in  
15 some cases the second) metaphase after exposure and require DNA synthesis to convert DNA  
16 damage into a chromosomal aberration. In addition, DNA repair is operating in peripheral  
17 lymphocytes to repair induced DNA damage. Thus, for acute exposures, the timing of sampling  
18 is of great importance. For chronic studies, the endpoints measure only the most recent  
19 exposures, and if the time between last exposure and sampling is long, any induced DNA  
20 damage not converted to a stable genotoxic alteration is certain to be missed. The events  
21 measured include all types of chromosomal aberrations, micronuclei, SCE, and numerical  
22 chromosomal changes. Stable chromosomal aberrations include reciprocal translocations,  
23 inversions, and some fraction of insertions and deletions as well as some numerical changes.  
24 However, until the development of fluorescent in situ hybridization (FISH), chromosome  
25 banding techniques were needed to detect these types of aberrations.

26 In in vitro assays, EtO has consistently tested positive in studies for multiple types of  
27 chromosomal effects, including DNA strand breaks, SCEs, micronuclei, and chromosomal  
28 aberrations [e.g. see Table 11 of [IARC \(2008\)](#)]. Of note, [Adám et al. \(2005\)](#) measured the  
29 sensitivity of different human cell types to EtO-induced DNA damage using the comet assay,  
30 which measures direct strand breaks and/or DNA damage converted to strand breaks during  
31 alkaline treatment. [Adám et al. \(2005\)](#) reported dose-dependent increases in DNA damage in the  
32 concentration range 0–100 µM in each of the cell types examined with no notable cytotoxicity.  
33 At the lowest concentration reported (20 µM), significant increases in DNA damage were  
34 observed in lymphoblasts, lymphocytes, and breast epithelial cells, but not in keratinocytes or  
35 cervical epithelial cells, suggesting that breast epithelial cells may have increased sensitivity to

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 EtO-induced genotoxicity compared to other nonlymphohematopoietic cell types. In addition,  
2 [Godderis et al. \(2006\)](#) investigated the effects of genetic polymorphisms on DNA damage  
3 induced by EtO in peripheral blood lymphocytes of 20 nonsmoking university students. No  
4 significant increases in micronuclei were observed following EtO treatment; however,  
5 dose-related increases in DNA strand breaks were seen in the comet assay. GST polymorphisms  
6 did not have a significant impact on the EtO-induced effects; however, significant increases in  
7 DNA strand breaks were associated with low-activity alleles of two DNA repair enzymes  
8 compared to wild-type alleles.

9 In vivo, several inhalation studies in laboratory animals have demonstrated that EtO  
10 exposure levels in the range of those used in the rodent bioassays induce SCEs [see Table 11 of  
11 [IARC \(2008\)](#)]; however, evidence for micronuclei and chromosomal aberrations from these same  
12 exposure levels is less consistent. In particular, studies by [van Sittert et al. \(2000\)](#) and [Lorenti  
13 Garcia et al. \(2001\)](#) observed increases in micronuclei and chromosomal aberrations in splenic  
14 lymphocytes of rats exposed to 50, 100, or 200 ppm EtO for 6 hours/day, 5 days/week, for  
15 4 weeks compared to levels from control rats, but the increases were not statistically significant.  
16 [IARC \(2008\)](#) noted, however, that “strong conclusions cannot be drawn” from these two studies  
17 because the cytogenetic analyses “were initiated 5 days after the final day of exposure, a  
18 suboptimal time, and the power of the (FISH) studies were limited by analysis of only a single  
19 chromosome and the small numbers of rats per group examined,” which was 3 per exposure  
20 group in both of the studies, although numerous cells/rat were examined. Moreover, a recent  
21 study by [Donner et al. \(2010\)](#) showed clear, statistically significant increases in chromosomal  
22 aberrations with longer durations of exposure ( $\geq 12$  weeks) to the concentration levels used in the  
23 rodent bioassays.

24 In humans, various studies of occupationally exposed workers have reported SCEs and  
25 other chromosomal effects associated with EtO exposure, including micronuclei and  
26 chromosomal aberrations. The genotoxicity of EtO was demonstrated in humans as early as  
27 1979. Table 3-6 summarizes the cytogenetic effects of EtO on human exposures (see also  
28 Appendix C for more details on some of the studies).

29 As illustrated in Table 3-6, numerous studies observed increased SCEs in occupationally  
30 exposed workers, especially for workers with the highest exposures [e.g., ([Major et al., 1996](#);  
31 [Sarto et al., 1991](#); [Tates et al., 1991](#); [Sarto et al., 1987](#))]. Several studies of occupationally  
32 exposed workers have also reported increased micronucleus formation in lymphocytes ([Ribeiro  
33 et al., 1994](#); [Tates et al., 1991](#)), in nasal mucosal cells ([Sarto et al., 1990](#)), and in bone marrow  
34 cells ([Högstedt et al., 1983](#)), although this endpoint seems to be less sensitive than SCEs. An  
35 association between increased micronucleus frequency and cancer risk has been reported in at

1 least one large prospective general population study ([Bonassi et al., 2007](#)). In addition,  
2 chromosomal aberrations have been reported in multiple studies of workers occupationally  
3 exposed to EtO ([Ribeiro et al., 1994](#); [Tates et al., 1991](#); [Sarto et al., 1987](#)). Chromosomal  
4 aberrations have been linked to an increased risk of cancer in several large prospective general  
5 population studies [e.g., ([Boffetta et al., 2007](#); [Rossner et al., 2005](#); [Hagmar et al., 2004](#); [Liou et](#)  
6 [al., 1999](#))].

#### 8 **3.3.3.4. Summary**

9 The available data from in vitro studies, laboratory animal models, and epidemiological  
10 studies establish that EtO is a mutagenic and genotoxic agent that causes a variety of types of  
11 genetic damage.

### 13 **3.4. MODE OF ACTION**

14 EtO is an alkylating agent that has consistently been found to produce numerous  
15 genotoxic effects in a variety of biological systems ranging from bacteriophage to occupationally  
16 exposed humans. It is carcinogenic in mice and rats, inducing tumors of the  
17 lymphohematopoietic system, brain, lung, connective tissues, uterus, and mammary gland. In  
18 addition, epidemiological studies have shown an increased risk of various types of human  
19 cancers (see Table A-5 in Appendix A), in particular lymphohematopoietic and breast cancers.  
20 Target tissues for EtO carcinogenicity in laboratory animals are varied, and the cancers are not  
21 clearly attributable to any specific type of genetic alteration. Although the precise mechanisms  
22 by which the multisite carcinogenicity in mice, rats, and humans occurs are unknown, EtO is  
23 clearly a mutagenic and genotoxic agent, as discussed in Section 3.3.3, and mutagenicity and  
24 genotoxicity are well established as playing a key role in carcinogenicity.

25 Exposure of cells to DNA-reactive agents results in the formation of carcinogen-DNA  
26 adducts. The formation of DNA adducts results from a sequence of events involving absorption  
27 of the agent, distribution to different tissues, and accessibility of the molecular target ([Swenberg](#)  
28 [et al., 1990](#)). Alkylating agents may induce several different DNA alkylation products ([Beranek,](#)  
29 [1990](#)) with varying proportions, depending primarily on the electrophilic properties of the agent.  
30 The predominant DNA adduct formed by EtO is N7-HEG, although other adducts, such as  
31 N3-hydroxyethyladenine and O<sup>6</sup>-HEG, have also been observed, in much lesser amounts  
32 ([Segerbäck, 1990](#)). In addition to direct DNA adduct formation via alkylation, [Marsden et al.](#)  
33 [\(2009\)](#) observed an indirect effect of EtO exposure on endogenous N7-HEG adduct formation  
34 and hypothesized that EtO could also indirectly cause adduct formation via oxidative stress (see  
35 also Section 3.3.3.1 and Appendix C). The various adducts are processed by different repair

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 pathways, and the subsequent genotoxic responses elicited by unrepaired DNA adducts are  
2 dependent on a wide range of variables. The specific adduct(s) responsible for EtO-induced  
3 genotoxicity and the mechanism(s) by which this adduct(s) induces the genotoxic damage are  
4 unknown.

5 It had been postulated that the predominant EtO-DNA adduct, N7-HEG, although  
6 unlikely to be directly promutagenic, could be subject to depurination, resulting in an apurinic  
7 site which could be vulnerable to miscoding during cell replication [e.g., [Walker and Skopek](#)  
8 [\(1993\)](#)]. However, in a study designed to test this hypothesis, [Rusyn et al. \(2005\)](#) failed to detect  
9 an accumulation of abasic sites in brain, spleen, and liver tissues of rats exposed to EtO. [Rusyn](#)  
10 [et al. \(2005\)](#) conclude that the accumulation of abasic sites is unlikely to be a primary  
11 mechanism for EtO mutagenicity, although they note that it is also possible that their assay was  
12 not sufficiently sensitive to detect small increases in abasic sites or that abasic sites are only  
13 mutagenic under conditions of rapid cell turnover, when cell replication may occur before repair  
14 of the abasic site (the tissues examined in their study were relatively quiescent). Another  
15 potential mechanism for EtO-induced mutagenicity is the direct mutagenicity of the  
16 promutagenic adducts such as O<sup>6</sup>-HEG, although these adducts are generally considered to occur  
17 at levels too low to explain all of the observed mutagenicity ([IARC, 2008](#)). In an in vitro study,  
18 [Tompkins et al. \(2009\)](#) exposed plasmid DNA to a range of EtO concentrations in water and  
19 reported that only the N7-HEG adduct was detectable after exposure to EtO concentrations up to  
20 2,000 µM; at higher EtO concentrations (≥10 mM), N1-hydroxyethyladenine and O<sup>6</sup>-HEG  
21 adducts were also quantifiable but at much lower levels than the N7-HEG adducts. [Tompkins et](#)  
22 [al. \(2009\)](#) then examined the mutagenicity of these adducts in a *supF* forward mutation assay and  
23 reported that the relative mutation frequencies were statistically significantly elevated only for  
24 plasmids exposed to these higher EtO concentrations (increases in relative mutation frequency  
25 were observed for N7-HEG adduct levels corresponding to lower EtO concentrations, however,  
26 and biologically relevant EtO-related increases in mutation frequency at these lower  
27 concentrations cannot be ruled out given the variability of the data and the limitations of the  
28 study) (see Appendix C, Sections C.1.2 and C.2.2, for a more detailed discussion of this study).  
29 An additional mechanism that has been suggested for EtO-induced mutagenicity is the imidazole  
30 ring-opening of N7-HEG, which can result in stable, potentially mutagenic lesions; however,  
31 EtO-induced N7-HEG ring-opening has not been corroborated in vivo ([IARC, 2008](#)).

32 The events involved in the formation of chromosomal damage by EtO are similarly  
33 unknown. N-alkylated bases are removed from DNA by base excision repair pathways. A  
34 review by [Memisoglu and Samson \(2000\)](#) notes that the action of DNA glycosylase and apurinic  
35 endonuclease creates a DNA single-strand break, which can in turn lead to DNA double-strand

1 breaks (DSBs). DSBs can also be produced by normal cellular functions, such as during V(D)J  
2 recombination in the development of lymphoid cells or topoisomerase II-mediated cleavage at  
3 defined sites. A review of mechanisms of DSB repair indicates that the molecular mechanisms  
4 are not fully understood ([Pfeiffer et al., 2000](#)). This review provides a thorough discussion of  
5 both sources (endogenous and exogenous) of DSBs and the variety of repair pathways that have  
6 evolved to process the breaks. Although homology-directed repair generally restores the original  
7 sequence, during nonhomologous end-joining, the ends of the breaks are frequently modified by  
8 addition or deletion of nucleotides. The lack of accumulation of abasic sites observed in the  
9 [Rusyn et al. \(2005\)](#) study discussed above argues against a mechanism involving abasic sites as  
10 hot spots for strand breaks, although it is possible that abasic sites accumulate more readily in  
11 replicating lymphocytes, which were not examined in the study of [Rusyn et al. \(2005\)](#). Another  
12 postulated mechanism for EtO-induced strand breaks is via the formation of hydroxyethyl  
13 adducts on the phosphate backbone of the DNA, but this mechanism requires further study  
14 ([IARC, 2008](#)).

15 Lymphohematopoietic malignancies, like all other cancers, are considered to be a  
16 consequence of an accumulation of genetic and epigenetic changes involving multiple genes and  
17 chromosomal alterations. Although it is clear that chromosome translocations are common  
18 features of some hematopoietic cancers, there is evidence that mutations in *p53* or *NRAS* are  
19 involved in certain types of leukemia ([U.S. EPA, 1997](#)). It should also be noted that  
20 therapy-related leukemias exhibiting reciprocal translocations are generally only seen in patients  
21 who have previously been treated with chemotherapeutic agents that act as topoisomerase II  
22 inhibitors ([U.S. EPA, 1997](#)). In NHL, the *BCL6* gene is frequently activated by translocations  
23 ([Chaganti et al., 1998](#)) as well as by mutations within the gene coding sequence ([Lossos and  
24 Levy, 2000](#)). [Preudhomme et al. \(2000\)](#) observed point mutations in the *AML1* gene in 9 of  
25 22 patients with the M0 type (minimally differentiated acute myeloblastic leukemia) of acute  
26 myeloid leukemia (AML), and [Harada et al. \(2003\)](#) identified *AML1* point mutations in cases of  
27 radiation-associated and therapy-related myelodysplastic syndrome (MDS)/AML. In both  
28 reports, point mutations within the coding sequence were found in patients with normal  
29 karyotypes as well as some with translocations or other chromosomal abnormalities.  
30 [Zharlyganova et al. \(2008\)](#) identified *AML1* mutations in 7 of 18 radiation-exposed MDS/AML  
31 patients but in none of 13 unexposed MDS/AML cases. Other point mutations have also been  
32 identified in therapy-related MDS/AML patients, including *p53* gene mutations after exposure to  
33 alkylating agents ([Christiansen et al., 2001](#)) and mutations in *RAS* and other genes in the receptor  
34 tyrosine kinase signal transduction pathway ([Christiansen et al., 2005](#)). Several models have  
35 been developed to integrate these various types of genetic alterations. One recent model

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 suggests that the pathogenesis of MDS/AML can be subdivided into at least eight genetic  
2 pathways that have different etiologies and different biologic characteristics ([Pedersen-Bjergaard](#)  
3 [et al., 2006](#)).

4 A mode-of-action-motivated modeling approach based solely on chromosome  
5 translocations has been proposed by [Kirman et al. \(2004\)](#). The authors suggested a nonlinear  
6 dose-response relationship for EtO and leukemia, based on a consideration that “chromosomal  
7 aberrations are the characteristic initiating events in chemically induced acute leukemia and gene  
8 mutations are not characteristic initiating events.” They proposed that EtO must be responsible  
9 for two nearly simultaneous DNA adducts, yielding a dose-squared (quadratic) relationship  
10 between EtO exposure and leukemia risk. However, as discussed above, there is evidence that  
11 does not support the assumption that chromosomal aberrations represent the sole initiating event.  
12 In fact, these aberrations or translocations could be a downstream event resulting from genomic  
13 instability. In addition, it is not clear that acute leukemia is the lymphohematopoietic cancer  
14 subtype associated with EtO exposure; in the large NIOSH study, increases in  
15 lymphohematopoietic cancer risk were driven by increases in lymphoid cancer subtypes.  
16 Furthermore, even if two reactions with DNA resulting in chromosomal aberrations or  
17 translocations are early-occurring events in some EtO-induced lymphohematopoietic cancers, it  
18 is not necessary that both events be associated with EtO exposure (e.g., background error repair  
19 rates or exposure to other alkylating agents may be the cause). Moreover, EtO could also  
20 produce translocations indirectly by forming DNA or protein adducts that affect the normally  
21 occurring recombination activities of lymphocytes or the repair of spontaneous double-strand  
22 breaks. Thus, broader mode-of-action considerations were not regarded as supportive of the  
23 hypothesis that the exposure-response relationship is purely quadratic.

24 Breast cancer is similarly considered to be a consequence of an accumulation of genetic  
25 and epigenetic changes involving multiple genes and chromosomal alterations ([Ingvarsson,](#)  
26 [1999](#)). Again, the precise mechanisms by which EtO induces breast cancer are unknown. As  
27 discussed in Section 3.3.3.2, in a study of mammary gland carcinomas in EtO-exposed mice,  
28 [Houle et al. \(2006\)](#) noted that the EtO-induced tumors exhibited a distinct shift in the mutational  
29 spectra of the *p53* and *Hras* genes and more commonly displayed concurrent mutations of the  
30 two genes. The comet assay results of [Adám et al. \(2005\)](#) suggest that human breast epithelial  
31 cells may have increased sensitivity to EtO-induced genotoxicity compared to other  
32 nonlymphohematopoietic cell types (see Section 3.3.3.3); however, the basis for any increased  
33 sensitivity of breast epithelial cells is similarly unknown.

34 In summary, EtO induces a variety of types of genetic damage. It directly interacts with  
35 DNA, resulting in DNA adducts, gene mutations, and chromosome damage. Depending on a

1 number of variables, EtO-induced DNA adducts (1) may be repaired, (2) may result in a  
2 base-pair mutation during replication, or (3) may be converted to a DSB, which also may be  
3 repaired or result in unstable (micronuclei) or stable (translocation) cytogenetic damage. The  
4 available data are strongly supportive of a mutagenic mode of action involving gene mutations  
5 and chromosomal aberrations (translocations, deletions, or inversions) that critically alter the  
6 function of oncogenes or tumor suppressor genes. Although it is clear that chromosome  
7 translocations are common features of many hematopoietic cancers, there is evidence that  
8 mutations in *p53*, *AML1*, or *Nras* are also involved in some leukemias. The above scientific  
9 evidence provides support for a mutagenic mode of action.

#### 11 **3.4.1. Analysis of the Mode of Action for Ethylene Oxide Carcinogenicity under EPA's** 12 **Mode-of-Action Framework**

13 In this section, the mode of action evidence for EtO carcinogenicity is analyzed under the  
14 mode of action framework in EPA's 2005 *Guidelines for Carcinogen Risk Assessment* ([U.S.](#)  
15 [EPA \(2005a\)](#), Section 2.4.3).

16 The *hypothesis* is that EtO carcinogenicity has a mutagenic mode of action. This  
17 hypothesized mode of action is presumed to apply to all of the tumor types.

18 The *key events* in the hypothesized mutagenic mode of action are (1) DNA adduct  
19 formation by EtO, which is a direct-acting alkylating agent; (2) the resulting genetic damage,  
20 including the formation and fixation of DNA mutations, particularly in oncogenes and tumor  
21 suppressor genes, as well as chromosomal alterations; and (3) clonal expansion of mutated cells  
22 during later stages of cancer development. Mutagenicity is a well-established cause of  
23 carcinogenicity.

24  
25 1. Is the hypothesized mode of action sufficiently supported in the test animals?

26 Numerous studies have demonstrated that EtO forms protein and DNA adducts, in mice  
27 and rats (see Sections 3.3.1 and 3.4 and Figure 3-2). For example, [Walker et al. \(1992b\)](#) and  
28 [Walker et al. \(1992a\)](#) demonstrated that EtO forms protein adducts with hemoglobin in the blood  
29 and DNA adducts with tissues throughout the body, including in the lung, brain, kidney, spleen,  
30 liver, and testes.

31 In addition, there is incontrovertible evidence that EtO is mutagenic (see Section 3.3.3).  
32 The evidence is *strong* and *consistent*; EtO has invariably yielded positive results in in vitro  
33 mutation assays from bacteriophage, bacteria, fungi, yeast, insects, plants, and mammalian cell  
34 cultures. The results of in vivo studies on the mutagenicity and genotoxicity of EtO have also  
35 been consistently positive following ingestion, inhalation, or injection. Increases in the

1 frequency of gene mutations in the lung, in T-lymphocytes, in bone marrow, and in testes have  
2 been observed in transgenic mice exposed to EtO via inhalation at concentrations similar to those  
3 in the mouse carcinogenesis bioassays. Furthermore, in a study of *p53* (tumor suppressor gene)  
4 and *Hras* (oncogene) mutations in mammary gland carcinomas of EtO-exposed and control  
5 mice, [Houle et al. \(2006\)](#) noted that the EtO-induced tumors exhibited a distinct shift in the  
6 mutational spectra of the *p53* and *Hras* genes and more commonly displayed concurrent  
7 mutations of the two genes, and in a similar study of *Kras* (oncogene) mutations in lung,  
8 Harderian gland, and uterine tumors, substantial increases were observed in *Kras* mutation  
9 frequencies in the tumors from the EtO-exposed mice ([Hong et al., 2007](#)).

10 Several inhalation studies in laboratory animals have demonstrated that EtO exposure  
11 levels in the range of those used in the rodent bioassays induce SCEs. Evidence for micronuclei  
12 and chromosomal aberrations from these same exposure levels has been less consistent;  
13 however, [IARC \(2008\)](#) has noted analytical limitations with some of these analyses (see  
14 Section 3.3.3.3). Moreover, a recent study by [Donner et al. \(2010\)](#) showed clear, statistically  
15 significant increases in chromosomal aberrations with exposure durations of  $\geq 12$  weeks to the  
16 concentration levels used in the rodent bioassays.

17 Ethylene oxide induces a variety of mutagenic and genotoxic effects, including  
18 chromosome breaks, micronuclei, SCEs, and gene mutations; however, the more general effect  
19 of mutagenicity/genotoxicity is *specific* and occurs in the absence of cytotoxicity or other overt  
20 toxicity. A *temporal relationship* is also clearly evident, with adducts and mutagenicity  
21 observed in subchronic assays.

22 *Dose-response relationships* have been observed between EtO exposure in vivo and  
23 hemoglobin adducts [e.g., [Walker et al. \(1992b\)](#)], as well as DNA adducts, SCEs, and *Hprt*  
24 mutations [e.g., [van Sittert et al. \(2000\)](#)] (see also Sections 3.3 and 3.4). A mutagenic mode of  
25 action for EtO carcinogenicity also clearly comports with notions of *biological plausibility* and  
26 *coherence* because EtO is a direct-acting alkylating agent. Such agents are generally capable of  
27 forming DNA adducts, which in turn have the potential to cause genetic damage, including  
28 mutations; and mutagenicity, in its turn, is a well-established cause of carcinogenicity. This  
29 chain of key events is consistent with current understanding of the biology of cancer.

30 In addition to the clear evidence supporting a mutagenic mode of action in test animals,  
31 there are no compelling alternative or additional hypothesized modes of action for EtO  
32 carcinogenicity. For example, there is no cytotoxicity or other toxicity indicative of regenerative  
33 proliferation or some other toxicity-related mode of action. Oxidative stress has been  
34 hypothesized as a mode of action, but there is little evidentiary support for this hypothesis and

1 the role of oxidative stress in EtO-induced carcinogenicity is speculative at this time (see Section  
2 J.3.2 of Appendix J and the response to comment #7 in Appendix L).

3  
4 2. Is the hypothesized mode of action relevant to humans?

5 The evidence discussed above demonstrates that EtO is a systemic mutagen in test  
6 animals; thus, there is the presumption that it would also be a mutagen in humans. Moreover,  
7 there is human evidence directly supporting a mutagenic mode of action for EtO carcinogenicity.  
8 Several studies of humans have reported exposure-response relationships between hemoglobin  
9 adduct levels and EtO exposure levels [e.g., [van Sittert et al. \(1993\)](#); [Schulte et al. \(1992\)](#); see  
10 Section 3.3.2], demonstrating the ability of EtO to bind covalently in systemic human cells, as it  
11 does in rodent cells. DNA adducts in EtO-exposed humans have not been well studied, and the  
12 evidence of increased DNA adducts is limited.

13 In addition, EtO has yielded positive results in in vitro mutagenicity studies of human  
14 cells (see Figure 3-3). Although the studies of point mutations in EtO-exposed humans are few  
15 and insensitive and the evidence for mutations is limited, there is clear evidence from a number  
16 of human studies that EtO causes chromosomal aberrations, SCEs, and micronucleus formation  
17 in peripheral blood lymphocytes (see Section 3.3.3.3 and Table 3-6). At least one study  
18 suggested an exposure-response relationship for the formation of SCEs in peripheral blood  
19 lymphocytes ([Major et al., 1996](#)). Another study reported a statistically significant increase in  
20 micronuclei in bone marrow cells in EtO-exposed workers ([Högstedt et al., 1983](#)).

21 Finally, there is strong evidence that EtO causes cancer in humans, including cancer  
22 types observed in rodent studies (i.e., lymphohematopoietic cancers and breast cancer),  
23 providing further weight to the relevance of the aforementioned events to the development of  
24 cancer in humans (see Sections 3.1 and 3.5.1).

25 *In conclusion, the weight of evidence supports a mutagenic mode of action for EtO*  
26 *carcinogenicity.* Although oxidative stress or other processes might contribute to the  
27 development of EtO-induced cancers, the available evidence best supports a mutagenic mode of  
28 action as the primary mode for EtO carcinogenicity.

29  
30 3. Which populations or lifestages can be particularly susceptible to the hypothesized mode of  
31 action?

32 The mutagenic mode of action is considered relevant to all populations and lifestages.  
33 According to EPA's *Supplemental Guidance* ([U.S. EPA, 2005b](#)), there may be increased  
34 susceptibility to early-life exposures to carcinogens with a mutagenic mode of action. Therefore,  
35 because the weight of evidence supports a mutagenic mode of action for EtO carcinogenicity,

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 and in the absence of chemical-specific data to evaluate differences in susceptibility, increased  
2 early-life susceptibility should be assumed and, if there is early-life exposure, the age-dependent  
3 adjustment factors should be applied, in accordance with the *Supplemental Guidance* (see  
4 Section 4.4).

5 In addition, as discussed in Section 3.5.2, people with DNA repair deficiencies or genetic  
6 polymorphisms conveying a decreased efficiency in detoxifying enzymes may have increased  
7 susceptibility to EtO-induced carcinogenicity.

### 9 **3.5. HAZARD CHARACTERIZATION**

#### 10 **3.5.1. Characterization of Cancer Hazard**

11 In studies of humans there is substantial evidence that EtO exposure is causally  
12 associated with lymphohematopoietic cancer, but the evidence is not strong enough to be  
13 conclusive. There is also evidence that EtO exposure is causally associated with breast cancer,  
14 but the database for breast cancer is more limited. Of the eight relevant<sup>13</sup> Hill “criteria” (or  
15 considerations) for causality ([Hill, 1965](#)), *temporality*, *coherence*, *biological plausibility* and  
16 *analogy* are readily satisfied, and the other four criteria (*specificity*, *consistency*, *biological*  
17 *gradient*, and *strength of association*) are satisfied to varying degrees, as discussed below.

18 *Temporality*, the sole necessary criterion, is satisfied because the subjects of all the  
19 epidemiology studies of EtO were workers who were exposed to EtO before the cancers of  
20 interest were observed, i.e., exposure preceded the development of the disease.

21 The related criteria of *coherence*, *biological plausibility* and *analogy* are fulfilled by the  
22 well-established knowledge that EtO is mutagenic and genotoxic, which are common  
23 mechanistic features of many carcinogens; that EtO is carcinogenic in rodents, with  
24 lymphohematopoietic cancers being observed in both rats and mice and mammary carcinomas  
25 being observed in female mice; and that EtO is an epoxide and epoxides are capable of directly  
26 interacting with DNA and are the active metabolites of many carcinogens.

27 There is some *specificity* with respect to the lymphohematopoietic system. Most of the  
28 studies focus on examining risks associated with subcategories of the lymphohematopoietic  
29 system. These cancers include leukemia, Hodgkin lymphoma, NHL, reticulosarcoma, and  
30 myeloma. (Note that, with the exception of the [Steenland et al. \(2004\)](#) study, which includes  
31 lymphocytic leukemia in a lymphoid cancer category, the studies do not subcategorize leukemia  
32 into its distinct myeloid and lymphocytic subtypes.) In most of the studies, an enhanced risk of  
33 cancer of the lymphohematopoietic system is evident, and in some studies, it is statistically

---

<sup>13</sup>The ninth consideration is experimental evidence, which is seldom available for human populations and is not available in the case of human exposures to EtO.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 significant. There also appears to be *specificity* across the epidemiological database for an  
2 increased risk of breast cancer. It should be noted, however, that the *specificity* criterion is not  
3 expected to be strictly satisfied by agents, such as EtO, that are widely distributed in all tissues  
4 and are direct-acting chemicals.

5 As just alluded to, there is evidence of *consistency* between studies with respect to cancer  
6 of the lymphohematopoietic system as a whole. Most of the available epidemiologic studies of  
7 EtO exposure have reported elevated risks of lymphohematopoietic cancer, and the studies that  
8 do not report a significant lymphohematopoietic cancer effect have major limitations, such as  
9 small numbers of cases (from small study size and/or insufficient follow-up time), inadequate  
10 exposure information, and/or reliance on external analyses (see Table 3-1 and Table A-5 in  
11 Appendix A). Overall, about 9 of 11 studies (including only the last follow-up of independent  
12 cohorts) with adequate information to determine RR estimates reported an increased risk of  
13 lymphohematopoietic cancers or a subgroup thereof, although not all were statistically  
14 significant, possibly due to the limitations noted above (see Table 3-1 and Table A-5 in  
15 Appendix A). The large, high-quality<sup>14</sup> NIOSH study shows statistically significant  
16 exposure-response trends for lymphoid cancers and all lymphohematopoietic cancers [[Steenland  
17 et al. \(2004\)](#); see Sections D.3 and D.4 of Appendix D for results for both sexes combined]. Four  
18 other studies reported statistically significant increases in risk ([Swaen et al., 1996](#); [Benson and  
19 Teta, 1993](#); [Bisanti et al., 1993](#); [Hogstedt et al., 1986](#)), although EtO exposures were reportedly  
20 low in the [Benson and Teta \(1993\)](#) study and the increased risks may be due to other chemical  
21 exposures. Nonsignificant increases in lymphohematopoietic cancer risk were observed in four  
22 other studies, based on small numbers of cases ([Coggon et al., 2004](#); [Olsen et al., 1997](#); [Hagmar  
23 et al., 1995](#); [Norman et al., 1995](#); [Hagmar et al., 1991](#)) [e.g., [Norman et al. \(1995\)](#) had only 1  
24 case]. Only 2 of the 11 studies showed no evidence of an increase in lymphohematopoietic  
25 cancer risk ([Swaen et al., 2009](#); [Kiesselbach et al., 1990](#)).

26 Regarding *consistency* in the breast cancer studies, the large, high-quality NIOSH study  
27 shows statistically significant increased risks for both breast cancer mortality [ $n = 103$  deaths;  
28 [Steenland et al. \(2004\)](#)] and breast cancer incidence [ $n = 319$  cases; [Steenland et al. \(2003\)](#)].  
29 Two other studies suggest an increased risk of breast cancer despite their small size [[Norman et](#)

---

<sup>14</sup>The NIOSH study ([ENREF 125](#); [ENREF 127](#)) was judged to be a “high-quality” study based on the attributes discussed (Section 3.1 and Section A.2.8 of Appendix A), including availability of individual worker exposure estimates from a high-quality exposure assessment (which used a comprehensive job-exposure matrix with exposure-level estimates from a regression model that incorporated a variety of plant and production variables and that explained 85% of the variability in an independent data set of EtO measurements), cohort study design, large size, inclusion of males and females, adequate follow-up, absence of any known confounding exposures, and use of internal comparisons. The breast cancer incidence study using the subcohort with interviews had the additional attribute of investigating and controlling for several breast cancer risk factors ([ENREF 127](#)).

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 [al. \(1995\)](#),  $n = 12$  cases; [Kardos et al. \(2003\)](#),  $n = 3$  deaths]. No elevated risks were seen in the  
2 only other two studies reporting breast cancer results; however these studies had few cases,  
3 owing to their small size and/or inadequate follow-up time [[Hagmar et al. \(1991\)](#),  $n = 5$  cases;  
4 [Coggon et al. \(2004\)](#),  $n = 11$  deaths] (see Table 3-2 and Table A-5 in Appendix A).

5 There is also some evidence of dose-response relationships (*biological gradient*). In the  
6 large, high-quality NIOSH study, a statistically significant positive trend was observed in the risk  
7 of lymphohematopoietic cancers with increasing (log) cumulative exposure to EtO, although  
8 results for this model were reported only for males ([Steenland et al., 2004](#)) (the sex difference is  
9 not statistically significant, however, and the trend for both sexes combined is also statistically  
10 significant; see Tables D-3e and D-4e in Appendix D). For only two other cohorts were results  
11 for exposure-response analyses reported, probably because most cohorts had too few cases  
12 and/or lacked adequate exposure information. In the [Swaen et al. \(2009\)](#) study of the UCC  
13 cohort, no statistically significant trends were observed for leukemia or lymphoid cancer using a  
14 Cox proportional hazards model with cumulative exposure, a model which notably did not yield  
15 statistically significant trends in the NIOSH study, either. In the small study of [Hagmar et al.](#)  
16 [\(1995\)](#), an SIR for leukemia of 7.14 was reported for subjects with at least 0.14 ppm  $\times$  years of  
17 cumulative exposure and 10 years latency, but this result was based on only two cases and was  
18 not statistically significant. For breast cancer, exposure-response analyses were reported only for  
19 the NIOSH cohort, again presumably because most cohorts had too few cases and/or lacked  
20 adequate exposure information. These analyses yielded clear, statistically significant trends for  
21 both breast cancer mortality ([Steenland et al., 2004](#)) and breast cancer incidence ([Steenland et al.,](#)  
22 [2003](#)) for a variety of models.

23 Whereas most of the considerations are largely satisfied, as discussed above, there is little  
24 *strength* in the associations, as reflected by the modest magnitude of most of the RR estimates.  
25 For example, in the large NIOSH study, the RR estimate for lymphoid cancer mortality in the  
26 highest exposure quartile is about 3.0 and the RR estimate for breast cancer incidence in the  
27 highest exposure quintile in the subcohort with interviews is on the order of 1.9. While large RR  
28 estimates increase the confidence that an observed association is not likely due to chance, bias,  
29 or confounding, modest RR estimates, such as those observed with EtO, do not preclude a causal  
30 association ([U.S. EPA, 2005a](#)). With EtO, the modest RR estimates may, in part, reflect the  
31 relatively high background rates of these cancers, particularly of breast cancer incidence.

32 In addition to the Hill criteria, other factors such as *chance, bias, and confounding* are  
33 considered in analyzing the weight of epidemiological evidence. Given the consistency of the  
34 findings across studies and the exposure-response relationships observed in the largest study,  
35 none of these factors is likely to explain the associations between these cancers and EtO

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 exposure. Coexposures to other chemicals are expected to have occurred for workers in the  
2 chemical industry cohorts but would have been much less likely in the sterilizer worker cohorts,  
3 such as the NIOSH cohort, which reported no evidence of confounding exposures to other  
4 occupational carcinogens ([Steenland et al., 1991](#)). For breast cancer in the NIOSH subcohort  
5 with interviews ([Steenland et al., 2003](#)), other risk factors for breast cancer were assessed, and  
6 statistically significant factors were included in the exposure-response models.

7 In conclusion, the overall epidemiological evidence for a causal association between EtO  
8 exposure and lymphohematopoietic cancer was judged to be strong but less than conclusive. For  
9 breast cancer, the existing evidence was strong based on the large, high-quality NIOSH study,  
10 but other studies lacked sufficient breast cancer data and, thus, overall, the epidemiological  
11 evidence was judged to be more limited.

12 There is inadequate evidence for other cancer types (e.g., stomach cancer and pancreatic  
13 cancer) in the epidemiology studies.

14 The experimental animal evidence for carcinogenicity is concluded to be “sufficient”  
15 based on findings of tumors at multiple sites, by both oral and inhalation routes of exposure, and  
16 in both sexes of both rats and mice. Tumor types resulting from inhalation exposure included  
17 mononuclear cell leukemia in male and female rats and malignant lymphoma and mammary  
18 carcinoma in female mice, suggesting some site concordance with the lymphohematopoietic and  
19 breast cancers observed in humans, also exposed by inhalation.

20 The evidence of EtO genotoxicity and mutagenicity is unequivocal. EtO is a  
21 direct-acting alkylating agent and has invariably tested positive in in vitro mutation assays from  
22 bacteriophage, bacteria, fungi, yeast, insects, plants, and mammalian cell cultures (including  
23 human cells). In mammalian cells (including human cells), EtO-induced genotoxic effects  
24 include unscheduled DNA synthesis, gene mutations, SCEs, and chromosomal aberrations. The  
25 results of in vivo genotoxicity studies of EtO have also been largely positive, following  
26 ingestion, inhalation, or injection. Increases in frequencies of gene mutations have been reported  
27 in the lung, T-lymphocytes, bone marrow, and testes of EtO-exposed mice. In particular,  
28 increases in frequencies of oncogene mutations have been observed in several tumor types from  
29 EtO-exposed mice compared to spontaneous mouse tumors of the same types. Several inhalation  
30 studies in laboratory animals have demonstrated that EtO exposure levels in the range of those  
31 used in the rodent bioassays (i.e., 10–100 ppm, 6–7 hours/day, 5 days/week) induce SCEs.  
32 Evidence for micronuclei and chromosomal aberrations from these same exposure levels in  
33 short-term studies (4 weeks or less) is less consistent, although concerns have been raised about  
34 some of the negative studies. A recent study showed clear, statistically significant increases in  
35 chromosomal aberrations with longer durations of exposure ( $\geq 12$  weeks) to the concentration

1 levels used in the rodent bioassays. The studies of point mutations in EtO-exposed humans are  
2 few and insensitive and the evidence for mutations is limited; however, there is clear evidence  
3 from a number of human studies that EtO causes chromosomal aberrations, SCEs, and  
4 micronucleus formation in peripheral blood lymphocytes, and one study has reported increased  
5 levels of micronuclei in bone marrow cells in EtO-exposed workers.

6 Under EPA's 2005 *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)), the  
7 conclusion can be made that EtO is "carcinogenic to humans." In general, the descriptor  
8 "carcinogenic to humans" is appropriate when there is convincing epidemiologic evidence of a  
9 causal association between human exposure and cancer. This descriptor is also appropriate when  
10 there is a lesser weight of epidemiologic evidence that is strengthened by specific lines of  
11 evidence set forth in the *Guidelines* ([U.S. EPA, 2005a](#)), which are satisfied for EtO. The lines of  
12 evidence supporting the characterization of "carcinogenic to humans" include the following:  
13 (1) there is strong, although less than conclusive on its own, evidence of cancer in humans  
14 associated with EtO exposure via inhalation, specifically, evidence of lymphohematopoietic  
15 cancers and female breast cancer in EtO-exposed workers; (2) there is extensive evidence of  
16 EtO-induced carcinogenicity in laboratory animals, including lymphohematopoietic cancers in  
17 rats and mice and mammary carcinomas in mice following inhalation exposure; (3) EtO is a  
18 direct-acting alkylating agent whose mutagenic and genotoxic capabilities have been well  
19 established in a variety of experimental systems, and a mutagenic mode of carcinogenic action  
20 has been identified in animals involving the key precursor events of DNA adduct formation and  
21 subsequent DNA damage, including point mutations and chromosomal effects; and (4) there is  
22 strong evidence that the key precursor events are anticipated to occur in humans and progress to  
23 tumors, including evidence of chromosome damage, such as chromosomal aberrations, SCEs,  
24 and micronuclei in EtO-exposed workers.

### 26 **3.5.2. Susceptible Life stages and Populations**

27 There are no data on the relative susceptibility of children and the elderly when compared  
28 with adult workers, in whom the evidence of hazard has been gathered, but because EtO does not  
29 have to be metabolized before binding to DNA and proteins, the maturing of enzyme systems in  
30 very young children is thought not to be a predominant factor in its hazard, at least for activation.  
31 However, the immaturity of *detoxifying* enzymes in very young children may increase children's  
32 susceptibility because children may clear EtO at a slower rate than adults. As discussed in  
33 Section 3.3.1, EtO is metabolized (i.e., detoxified) primarily by hydrolysis in humans but also by  
34 glutathione conjugation. Both hydrolytic activity and glutathione-S-transferase activity  
35 apparently develop after birth ([Clewell et al., 2002](#)); thus, very young children might have a

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 decreased capacity to detoxify EtO compared to adults. In the absence of data on the relative  
2 susceptibility associated with EtO exposure in early life, increased early-life susceptibility is  
3 assumed, in accordance with EPA's *Supplemental Guidance* ([U.S. EPA, 2005b](#)), because the  
4 weight of evidence supports the conclusion of a mutagenic mode of action for EtO  
5 carcinogenicity (see Section 3.4).

6 Other than the occurrence of sex-specific cancers (e.g., breast cancer in human females,  
7 mammary and uterine carcinomas in female mice, and testicular peritoneal mesotheliomas in  
8 male rats; see Section 3.2), there is no clear sex difference in EtO-induced carcinogenicity. With  
9 the exception of the sex-specific cancers and the observation of malignant lymphomas in female  
10 but not male mice, there is no sex difference in EtO-induced cancer types in the rat and mouse  
11 bioassays. Cancer potency estimates for females are roughly 50% higher than those for males  
12 for both mice and rats (see Table 4-18 in Section 4.2.5). In humans, in the large NIOSH study  
13 ([Steenland et al., 2004](#)), the association between lymphoid cancers and EtO exposure was seen  
14 primarily in males, but the sex difference was not statistically significant (see Section D.3.b in  
15 Appendix D), and the SAB panel that reviewed an earlier draft of this assessment recommended  
16 that data from both sexes be combined for the derivation of quantitative risk estimates for the  
17 lymphohematopoietic cancers ([SAB, 2007](#)).

18 [Brown et al. \(1996\)](#) reported that sex differences in EtO toxicokinetics were observed in  
19 mice but not in rats; female mice had a significantly higher steady-state blood EtO concentration  
20 after 4 hours of exposure to either 100 or 330 ppm than male mice. As noted above and  
21 discussed in Section 3.3.1, EtO is metabolized primarily by hydrolysis in humans. [Mertes et al.](#)  
22 [\(1985\)](#) reported no sex difference in microsomal or cytosolic epoxide hydrolase activities in  
23 human liver in vitro using benzo[*a*]pyrene 4,5-oxide or *trans*-stilbene oxide, respectively, as  
24 substrates. Using EtO as a substrate, but with far fewer subjects, [Fennell and Brown \(2001\)](#)  
25 reported similar values for males and females for epoxide hydrolase activity in human liver  
26 microsomes and for GSH transferase in human liver cytosolic fractions.

27 Because EtO is detoxified by glutathione conjugation or hydrolysis, people with  
28 genotypes conveying deficiencies in glutathione-S-transferase or epoxide hydrolase activities  
29 may be at increased risk of cancer from EtO exposure. [Yong et al. \(2001\)](#) measured  
30 approximately twofold greater EtO-hemoglobin adduct levels in occupationally exposed persons  
31 with a null GSTT1 genotype than in those with positive genotypes. Similarly, in a study of  
32 hospital workers, [Haufroid et al. \(2007\)](#) reported increased urinary excretion of a glutathione  
33 conjugate of EtO, reflecting increased detoxification of EtO, associated with a nonnull GSTT1  
34 genotype, although the increase was not statistically significant in all the regression models

1 tested; associations were less clear for other glutathione-S-transferase or epoxide hydrolase  
2 polymorphisms.

3 In addition, people with DNA repair deficiencies such as xeroderma pigmentosum,  
4 Bloom's syndrome, Fanconi anemia, and ataxia telangiectasia ([Gelehrter et al., 1990](#)) are  
5 expected to be especially sensitive to the damaging effects of EtO exposure. [Paz-y-Miño et al.](#)  
6 [\(2002\)](#) have recently identified a specific polymorphism in the excision repair pathway gene  
7 *hMSH2*. The polymorphism was present in 7.5% of normal individuals and in 22.7% of NHL  
8 patients, suggesting that this polymorphism may be associated with an increased risk of  
9 developing NHL.

## 4. CANCER DOSE-RESPONSE ASSESSMENT FOR INHALATION EXPOSURE

This chapter presents the derivation of cancer unit risk estimates from human and rodent data. Section 4.1 discusses the derivation of unit risk estimates for lymphohematopoietic cancers, breast cancer, and total cancer from human data, as well as sources of uncertainty in these estimates. (Note that the estimates in Section 4.1 were derived under the common assumption that relative risk is independent of age. This assumption is later superseded by an assumption of increased early-life susceptibility, and it is the unit risk estimates derived under this latter assumption, which are developed in Section 4.4, that are the ultimate estimates proposed in this assessment.) Section 4.2 presents the derivation of unit risk estimates from rodent data. Section 4.3 summarizes the unit risk estimates derived from the different data sets. Section 4.4 discusses adjustments for assumed increased early-life susceptibility, based on recommendations from EPA's *Supplemental Guidance* ([U.S. EPA, 2005b](#)), because the weight of evidence supports the conclusion of a mutagenic mode of action for EtO carcinogenicity (see Section 3.4). Section 4.5 presents conclusions about the unit risk estimates. Section 4.6 compares the unit risk estimates derived in this EPA assessment to those derived in other assessments. Finally, Section 4.7 provides risk estimates derived for some general occupational exposure scenarios.

### 4.1. INHALATION UNIT RISK ESTIMATES DERIVED FROM HUMAN DATA

The NIOSH retrospective cohort study of more than 18,000 workers in 13 sterilizing facilities [most recent update by [Steenland et al. \(2004\)](#) and [Steenland et al. \(2003\)](#)] provides the most appropriate data sets for deriving quantitative cancer risk estimates in humans for several reasons: (1) exposure estimates were derived for the individual workers using a comprehensive exposure assessment, (2) the cohort was large and diverse (e.g., 55% female), and (3) there was little reported exposure to chemicals other than EtO. Exposure estimates, including estimates for early exposures for which no measurements were available, were determined using a regression model that estimated exposures to each individual as a function of facility, exposure category, and time period. The regression model was based on extensive personal monitoring data from 18 facilities spanning a number of years as well as information on factors influencing exposure, such as engineering controls [[Hornung et al. \(1994\)](#); see also Section A.2.8 in Appendix A]. When evaluated against test data, the model accounted for 85% of the variation in average EtO exposure levels. The investigators were then able to estimate the cumulative exposure (ppm × days) for each individual worker by multiplying the estimated exposure for each job (exposure category) held by the worker by the number of days spent in that job and summing

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 over all the jobs held by the worker. [Steenland et al. \(2004\)](#) present follow-up results for the  
2 cohort mortality study previously discussed by [Steenland et al. \(1991\)](#) and [Stayner et al. \(1993\)](#).  
3 Positive findings in the current follow-up include increased rates of (lympho)hematopoietic  
4 cancer mortality and of breast cancer mortality in females. [Steenland et al. \(2003\)](#) present results  
5 of a breast cancer incidence study of a subcohort of 7,576 women from the NIOSH cohort.

6 The other major occupational study [most recent update by [Swaen et al. \(2009\)](#)]  
7 described risks to Union Carbide workers exposed to EtO at two chemical plants in West  
8 Virginia, but this study is less useful for estimating quantitative cancer risks for a number of  
9 reasons. First, the exposure assessment is much less extensive than that used for the NIOSH  
10 cohort, with greater likelihood for exposure misclassification, especially in the earlier time  
11 periods when no measurements were available (1925–1973). Exposure estimation for the  
12 individual workers was based on a relatively crude exposure matrix which cross-classified 3  
13 levels of exposure intensity with 4 time periods. The exposure estimates for 1974–1988 were  
14 based on measurements from air sampling at the West Virginia plants since 1976. The exposure  
15 estimates for 1957–1973 were based on measurements in a similar plant in Texas. The exposure  
16 estimates for 1940–1956 were based loosely on “rough” estimates reported for  
17 chlorohydrin-based EtO production in a Swedish facility in the 1940s. The exposure estimates  
18 for 1925–1939 were essentially guesses. Thus, for the two earliest time periods (1925–1939 and  
19 1940–1956) at least, the exposure estimates are highly uncertain. (See Section A.2.20 of  
20 Appendix A for a more detailed discussion of the exposure assessment for the Union Carbide  
21 cohort.) This is in contrast to the NIOSH exposure assessment in which exposure estimates were  
22 based on extensive sampling data and regression modeling. In addition, the sterilization  
23 processes used by the NIOSH cohort workers were fairly constant back in time, unlike chemical  
24 production processes, which likely involved much higher and more variable exposure levels in  
25 the past. Furthermore, the Union Carbide cohort is of much smaller size and has far fewer deaths  
26 than the NIOSH cohort, it is restricted to males and so cannot be used to investigate breast cancer  
27 risk in females, and there are coexposures to other chemicals.

28 A third study ([Hagmar et al., 1995](#); [Hagmar et al., 1991](#)) estimated cumulative exposures  
29 for individual workers; however, insufficient exposure-response data are presented for the  
30 derivation of unit risk estimates. Exposure-response results for specific cancers are provided  
31 only in the 1991 paper and then only for two lymphohematopoietic cancers across two  
32 categorical exposure groups.

1 Table 4-1 provides a summary of the considerations taken into account in selecting the  
 2 NIOSH study as the basis for the derivation of unit risk estimates. The NIOSH EtO cohort  
 3 mortality data can be obtained from the Industrywide Studies Branch of NIOSH.<sup>15</sup>  
 4  
 5

6 **Table 4-1. Considerations used in this assessment for selecting epidemiology**  
 7 **studies for quantitative risk estimation**  
 8

Consideration	Studies	Comments
Availability of quantitative exposure estimates	1. <a href="#">Hagmar et al. (1995)</a> and <a href="#">Hagmar et al. (1991)</a> [Swedish sterilizer cohort] 2. <a href="#">Swaen et al. (2009)</a> [latest follow-up of Union Carbide Corporation (UCC) cohort] 3. <a href="#">Steenland et al. (2004)</a> and <a href="#">Steenland et al. (2003)</a> [latest follow-up of NIOSH cohort]	These are the only 3 studies with quantitative exposure estimates, which is an essential criterion for quantitative risk estimation.
Availability of exposure-response information	1. <a href="#">Swaen et al. (2009)</a> 2. <a href="#">Steenland et al. (2004)</a> and <a href="#">Steenland et al. (2003)</a>	<a href="#">Hagmar et al. (1995)</a> and <a href="#">Hagmar et al. (1991)</a> did not present sufficient exposure-response results, presumably because they had a short follow-up time and thus few cases of specific cancers (5 breast cancer cases; 6 lymphohematopoietic cancer cases).
Other factors affecting the utility of epidemiology studies for quantitative risk estimation	<a href="#">Steenland et al. (2004)</a> and <a href="#">Steenland et al. (2003)</a>	The NIOSH study [ <a href="#">Steenland et al. (2004)</a> and <a href="#">Steenland et al. (2003)</a> ] alone was selected for quantitative risk estimation, as it was judged to be substantially superior to the UCC study ( <a href="#">Swaen et al., 2009</a> ) with respect to a number of key considerations [in particular, in order of importance: (1) quality of the exposure estimates, (2) cohort size, and (3) the absence of coexposures and the inclusion of women].

9  
10  
11  
12  
13  
14  
15  
16

---

<sup>15</sup>Industrywide Studies Branch; Division of Surveillance, Hazard Evaluations and Field Studies: NIOSH; Centers for Disease Control and Prevention, 4676 Columbia Parkway MS R-13, Cincinnati, Ohio 45226, telephone: 513-841-4203.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 The derivation of unit risk estimates, defined as the lifetime risk of cancer from chronic  
2 inhalation of EtO per unit of air concentration, for lymphohematopoietic cancer mortality and  
3 incidence and for breast cancer mortality and incidence in females, based on results of the recent  
4 analyses of the NIOSH cohort, is presented in the following subsections.

5 The exposure-response models used to fit the epidemiological data are empirical  
6 “curve-fitting” models. Considerations used in the selection of the exposure-response models  
7 upon which to base the derivation of unit risk estimates included statistical fit (as reflected by  
8 *p*-values),<sup>16</sup> visual fit of the model shape to the categorical results, and biological plausibility.  
9 When multiple models were deemed to be reasonable candidates for selection based on those  
10 considerations, AIC<sup>17</sup> was also considered in selecting the “preferred” model. (Note that all the  
11 models discussed in Chapter 4 are fitted treating exposure as a continuous variable except for the  
12 categorical models and the linear regressions of categorical results, which are explicitly  
13 described as such.)  
14

#### 15 **4.1.1. Risk Estimates for Lymphohematopoietic Cancer**

##### 16 **4.1.1.1. *Lymphohematopoietic Cancer Results From the NIOSH Study***

17 [Steenland et al. \(2004\)](#) investigated the relationship between (any) EtO exposure and  
18 mortality from cancer at a number of sites using life-table analyses with the U.S. population as  
19 the comparison population. Categorical SMR analyses were also done by quartiles of cumulative  
20 exposure. Then, to further investigate apparent exposure-response relationships observed for  
21 (lympho)hematopoietic cancer and breast cancer, internal exposure-response analyses were  
22 conducted using Cox proportional hazards models, which have the form

$$23 \text{Relative rate (RR)} = e^{\beta X}, \quad (4-1)$$

24  
25  
26  
27  
28 where  $\beta$  represents the regression coefficient and  $X$  is the exposure (or some function of  
29 exposure, e.g., the natural log of exposure). Internal analyses were done two ways—with  
30 exposure as a categorical variable and with exposure as a continuous variable. A nested

---

<sup>16</sup> *p*-values generally were obtained from the likelihood ratio test. Small *p*-values (conventionally,  $p < 0.05$ ) indicate a statistically significant fit, e.g., rejection of the null hypothesis that the regression coefficient on the exposure term ( $\beta$ ) is 0.

<sup>17</sup> Akaike Information Criteria. The AIC is a measure of information loss from a dose-response model that can be used to compare a specified set of models. The AIC is defined as  $2p - 2\ln(L)$ , where  $p$  is the number of estimated parameters included in the model and  $L$  is the maximized value of the likelihood function. Among a set of specified models, the model with the lowest AIC is the preferred model.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 case-control approach was used, with age as the time variable used to form the risk sets. Risk  
2 sets were constructed with 100 controls randomly selected for each case from the pool of those  
3 surviving to at least the age of the index case. According to the authors, use of 100 controls per  
4 case has been shown to result in ORs virtually identical to the RR estimates obtained with full  
5 cohorts. Cases and controls were matched on race (white/nonwhite), sex, and date of birth  
6 (within 5 years). Exposure was the only covariate in the model, so the  $p$ -value for the model also  
7 serves as a  $p$ -value for the regression coefficient,  $\beta$ , as well as for a test of exposure-response  
8 trend.

9 For lymphohematopoietic cancer mortality, [Steenland et al. \(2004\)](#) analyzed both all  
10 lymphohematopoietic cancers combined and a subcategory of lymphohematopoietic cancers that  
11 they called “lymphoid” cancers; these included NHL, myeloma, and lymphocytic leukemia.  
12 Their exposure-response analyses focused on cumulative exposure and (natural) log cumulative  
13 exposure, with various lag periods. Other EtO exposure metrics (duration of exposure, average  
14 exposure, and peak exposure) were also examined, but models using these metrics did not  
15 generally predict lymphohematopoietic cancer as well as models using cumulative exposure. A  
16 lag period defines an interval before death, or end of follow-up, during which any exposure is  
17 disregarded because it is not considered relevant to the outcome under investigation. For  
18 lymphohematopoietic (and lymphoid) cancer mortality, a 15-year lag provided the best fit to the  
19 data, based on the likelihood ratio test. One ppm  $\times$  day was added to cumulative exposures in  
20 lagged analyses to avoid taking the log of 0. For both all lymphohematopoietic and lymphoid  
21 cancers, [Steenland et al. \(2004\)](#) found stronger positive exposure-response trends in males and so  
22 presented the results for some of the regression models separately by sex. The apparent sex  
23 difference was not statistically significant (see Appendix D), however, and results for both sexes  
24 combined were subsequently obtained from Dr. Steenland (see Appendix D; Section 3 for  
25 lymphoid cancer, Section 4 for all lymphohematopoietic cancer). These results are presented in  
26 Table 4-2. For additional details and discussion of the [Steenland et al. \(2004\)](#) study, see  
27 Appendix A.  
28

1 **Table 4-2. Cox regression results for all lymphohematopoietic cancer and**  
 2 **lymphoid cancer mortality in both sexes in the NIOSH cohort, for the models**  
 3 **presented by [Steenland et al. \(2004\)](#)**  
 4

Exposure variable <sup>a</sup>	p-value <sup>b</sup>	Coefficient (SE) (per ppm × day)	ORs by category <sup>c</sup> (95% CI)
<b>All lymphohematopoietic cancer<sup>d</sup></b>			
Cumulative exposure, 15-yr lag	0.40	0.00000326 (0.00000349)	
Log cumulative exposure, 15-yr lag	0.009	0.107 (0.0418)	
Categorical cumulative exposure, 15-yr lag	0.10		1.00, 2.33 (0.93–5.86), 3.46 (1.33–8.95), 3.02 (1.16–7.89), 2.96 (1.12–7.81)
<b>Lymphoid cancer<sup>e</sup></b>			
Cumulative exposure, 15-yr lag	0.22	0.00000474 (0.00000335)	
Log cumulative exposure, 15-yr lag	0.02	0.112 (0.0486)	
Categorical cumulative exposure, 15-yr lag	0.21		1.00, 1.75 (0.59–5.25), 3.15 (1.04–9.49), 2.44 (0.80–7.50), 3.00 (1.02–8.45)

5  
6 <sup>a</sup>Cumulative exposure is in ppm × days.

7 <sup>b</sup>p-values from likelihood ratio test.

8 <sup>c</sup>Exposure categories are 0, >0–1,199, 1,200–3,679, 3,680–13,499, ≥13,500 ppm × days.

9 <sup>d</sup>9<sup>th</sup> revision ICD codes 200–208; results based on 74 cases.

10 <sup>e</sup>NHL, myeloma, and lymphocytic leukemia (9<sup>th</sup> revision ICD codes 200, 202, 203, 204); results based on 53 cases.

11 Source: Additional analyses performed by Dr. Steenland (see Sections D.3 and D.4 of Appendix D).  
 12  
 13

14 **4.1.1.2. Prediction of Lifetime Extra Risk of Lymphohematopoietic Cancer Mortality**

15 The exposure-response trends for lymphohematopoietic cancers observed by [Steenland et](#)  
 16 [al. \(2004\)](#) appear to be driven largely by the lymphoid cancers; therefore, the primary risk  
 17 analyses for lymphohematopoietic cancer are based on the lymphoid cancer results.

18 Lymphohematopoietic cancers are a diverse group of diseases with diverse etiologies, and  
 19 myeloid and lymphoid cells develop from different progenitor cells; thus, there is stronger  
 20 support for an etiologic role of EtO in the development of lymphoid cancers than in the  
 21 development of the cancers in the aggregate all lymphohematopoietic cancer category. The  
 22 consideration of NHL, (plasma cell) myeloma, and lymphocytic leukemia together as  
 23 “lymphoid” cancers is consistent with the current World Health Organization classification of  
 24 such cancers based on their derivation from B-cells, T-cells, and NK-cells rather than previous  
 25 distinctions ([Harris et al., 1999](#)). Nonetheless, for comprehensiveness and for the reasons listed  
 26 below, risk estimates based on the all lymphohematopoietic cancer results are presented for

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 comparison. Judging roughly from the *p*-values, the model fits do not appear notably better for  
2 lymphoid cancers than for all lymphohematopoietic cancers (see Table 4-2, *p*-values for log  
3 cumulative exposure models), and the “lymphoid” category did not include Hodgkin lymphoma,  
4 which also exhibited evidence of exposure-response trends, although based on few cases  
5 ([Steenland et al., 2004](#)). In addition, misclassification or nonclassification of tumor type is more  
6 likely to occur for subcategories of lymphohematopoietic cancer (e.g., 4 of the 25 leukemias in  
7 the analyses were classified as “not specified” and so could not be considered for the lymphoid  
8 cancer analysis).

9 The results of internal exposure-response analyses of lymphoid cancer in the NIOSH  
10 cohort (Cox regression analyses, summarized in Table 4-2) were used for predicting the extra  
11 risks of lymphoid cancer mortality from continuous environmental exposure to EtO. Extra risk  
12 is defined as

$$15 \text{ Extra risk} = (R_x - R_o)/(1 - R_o), \quad (4-2)$$

16  
17  
18 where  $R_x$  is the lifetime risk in the exposed population and  $R_o$  is the lifetime risk in an  
19 unexposed population (i.e., the background risk). These risk estimates were calculated using the  
20  $\beta$  regression coefficients and an actuarial program (life-table analysis) that accounts for  
21 competing causes of death.<sup>18</sup> An inherent assumption in the Cox regression model and its  
22 application in the life-table analyses is that RR is independent of age. (An alternate assumption  
23 of increased susceptibility from early-life exposure to EtO, as recommended in EPA's  
24 *Supplemental Guidance* ([U.S. EPA, 2005b](#)) for chemicals, such as EtO [see Section 3.4], with a  
25 mutagenic mode of action, is considered in Section 4.4. This alternate assumption is the  
26 prevailing assumption in this assessment, based on the recommendations in the *Supplemental*  
27 *Guidance*. Risk estimates are first developed under the assumption of age independence,  
28 however, because that is the standard approach in the absence of evidence to the contrary or of  
29 sufficient evidence of a mutagenic mode of action to invoke the divergent assumption of  
30 increased early-life susceptibility.)

31 U.S. age-specific all-cause mortality rates for 2004 for both sexes of all race groups  
32 combined ([Arias, 2007](#)) were used to specify the all-cause background mortality rates in the  
33 actuarial program. For the cause-specific background mortality rates for lymphoid cancers,

---

<sup>18</sup>This program is an adaptation of the approach previously used by the Committee on the Biological Effects of Ionizing Radiation ([BEIR, 1988](#)). A spreadsheet illustrating the extra risk calculation for the derivation of the LEC<sub>01</sub> for lymphoid cancer incidence (see Section 4.1.1.3) is presented in Appendix E.

*This document is a draft for review purposes only and does not constitute Agency policy.*

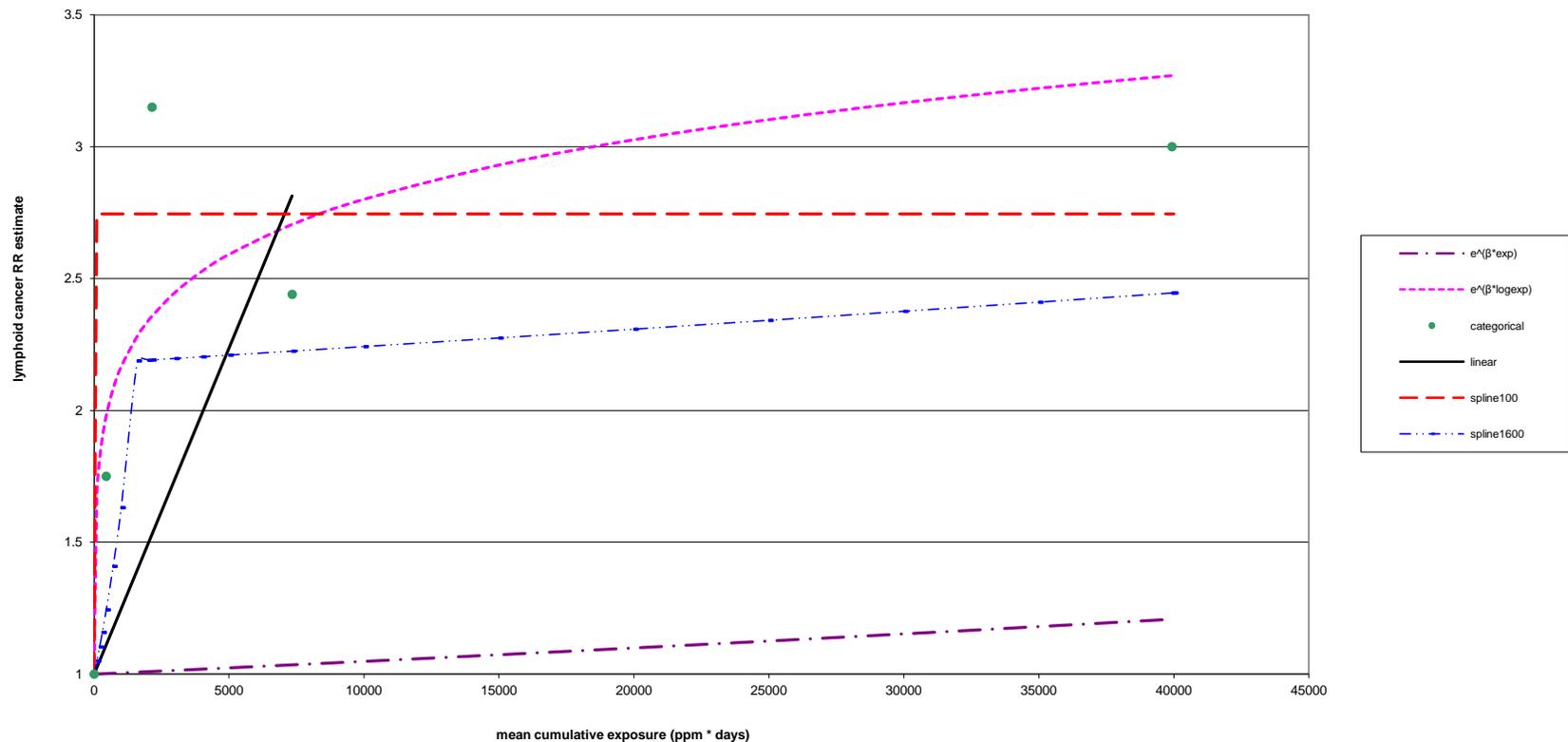
1 age-specific mortality rates for the relevant subcategories of lymphohematopoietic cancer (NHL  
2 [C82-C85 of 10<sup>th</sup> revision of the International Classification of Diseases (ICD)], multiple  
3 myeloma [C88, C90], and lymphoid leukemia [C91]) for the year 2004 were obtained from the  
4 National Center for Health Statistics Data Warehouse website  
5 (<http://www.cdc.gov/nchs/datawh/statab/unpubd/mortabs.htm>). The risks were computed up to  
6 age 85 for continuous exposures to EtO beginning at birth.<sup>19</sup> Conversions between occupational  
7 EtO exposures and continuous environmental exposures were made to account for differences in  
8 the number of days exposed per year (240 vs. 365 days) and in the amount of EtO-contaminated  
9 air inhaled per day [10 vs. 20 m<sup>3</sup>; [U.S. EPA \(1994\)](#)]. An adjustment was also made for the lag  
10 period. The reported standard errors for the regression coefficients from Table 4-2 were used to  
11 compute the 95% upper confidence limits (UCLs) for the relative rates, based on a normal  
12 approximation.

13 The only statistically significant Cox regression model presented by [Steenland et al.](#)  
14 [\(2004\)](#) for lymphoid cancer mortality was for log cumulative exposure with a 15-year lag in  
15 males ( $p = 0.02$ ). This was similarly true for the analyses of lymphoid cancer using the data for  
16 both sexes (see Table 4-2). However, using the log cumulative exposure model to estimate the  
17 risks from low environmental exposures is problematic because this model, which is intended to  
18 fit the full range of occupational exposures in the study, is inherently supralinear (i.e., risk  
19 increases steeply with increasing exposures in the low exposure range and then plateaus), and  
20 results are unstable for low exposures (i.e., small changes in exposure correspond to large  
21 changes in risk; see Figure 4-1). Some consideration was thus given to the cumulative exposure  
22 model, which is typically used and which is stable at low exposures, although the fit to these data  
23 was not statistically significant ( $p = 0.22$ ). However, the Cox regression model with cumulative  
24 exposure is inherently sublinear (i.e., risk increases gradually in the low exposure range and then  
25 with increasing steepness as exposure increases) and does not reflect the apparent supralinearity  
26 of the data demonstrated by the categorical results and the superior fit of the log cumulative  
27 exposure model; thus, this model was not considered further.

---

<sup>19</sup>Rates above age 85 years are not included because cause-specific disease rates are less stable for those ages. Note that 85 years is not employed here as an average lifespan but, rather, as a cutoff point for the life-table analysis, which uses actual age-specific mortality rates. The average lifespan for males and females combined in a life-table analysis truncated at age 85 years is about 75 years.

*This document is a draft for review purposes only and does not constitute Agency policy.*



**Figure 4-1. RR estimate for lymphoid cancer vs. mean exposure (with 15-year lag, unadjusted for continuous exposure).**

$e^{(\beta \times \text{exp})}$ : Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$ ;  $e^{(\beta \times \ln(\text{exposure}))}$ : Cox regression results for  $RR = e^{(\beta \times \ln(\text{exposure}))}$ ; categorical: Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$  with categorical exposures; linear: weighted linear regression of categorical results, excluding highest exposure group (see text); spline100(1600): 2-piece log-linear spline model with knot at 100 (1,600) ppm  $\times$  days (see text). (Note that, with the exception of the categorical results and the linear regression of the categorical results, the different models have different implicitly estimated baseline risks; thus, they are not strictly comparable to each other in terms of RR values, i.e., along the y-axis. They are, however, comparable in terms of general shape.)

Source: Steenland reanalyses for males and females combined; see Appendix D (except for linear regression of categorical results, which was done by EPA).

1 In a 2006 external review draft of this assessment ([U.S. EPA, 2006a](#)), which relied on the  
2 original published results of [Steenland et al. \(2004\)](#), EPA proposed that the best way to represent  
3 the exposure-response relationship in the lower exposure region, which is the region of interest  
4 for low-exposure extrapolation, was through the use of a weighted linear regression of the results  
5 from the Cox regression model with categorical cumulative exposure and a 15-year lag [for  
6 males only, as this was the significant finding in the published paper of [Steenland et al. \(2004\)](#)].  
7 In addition, the highest exposure group was not included in the regression to alleviate some of  
8 the “plateauing” in the exposure-response relationship at higher exposure levels and to provide a  
9 better fit to the lower exposure data. Linear modeling of categorical (i.e., grouped)  
10 epidemiologic data and elimination of the highest exposure group(s) under certain circumstances  
11 to obtain a better fit of low-exposure data are both standard techniques used in EPA  
12 dose-response assessments ([U.S. EPA, 2012, 2005a](#)). An established methodology was  
13 employed for the weighted linear regression of the categorical epidemiologic data, as described  
14 by [Rothman \(1986\)](#) and used by others [e.g., [van Wijngaarden and Hertz-Picciotto \(2004\)](#)].  
15 However, the SAB panel that reviewed the draft assessment recommended that EPA employ  
16 models using the individual continuous exposure data as an alternative to modeling the published  
17 grouped data. The SAB also recommended that both males and females be included in the  
18 modeling of lymphohematopoietic cancer mortality ([SAB, 2007](#)).

19 In response to these recommendations and in consultation with Dr. Steenland, one of the  
20 investigators from the NIOSH cohort studies, EPA determined that, using the full continuous  
21 exposure data set, an alternative way to address the supralinearity of the data (while avoiding the  
22 extreme low-exposure curvature obtained with the log cumulative exposure model) might be to  
23 use a two-piece log-linear spline model. Spline models have been used previously for  
24 exposure-response analyses of epidemiological data ([Steenland and Deddens, 2004](#); [Steenland et  
25 al., 2001](#)). These models are particularly useful for exposure-response data such as the EtO  
26 lymphoid cancer data, for which RR initially increases with increasing exposure but then tends to  
27 plateau, or attenuate, at higher exposures. Such plateauing exposure-response relationships have  
28 been seen with other occupational carcinogens and may occur for various reasons, including the  
29 depletion of susceptible subpopulations at high exposures, mismeasurement of high exposures,  
30 or a healthy worker survivor effect ([Stayner et al., 2003](#)). No other traditional exposure-response  
31 models for continuous exposure data that might suitably fit the observed exposure-response  
32 pattern were apparent. Dr. Steenland was commissioned to do the spline analyses using the full  
33 data set with cumulative exposure as a continuous variable, and his findings are included in  
34 Appendix D (see Section D.3 for lymphoid cancer, Section D.4 for all lymphohematopoietic  
35 cancer). The results of the spline analyses are presented below.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 For the two-piece log-linear spline modeling approach, the Cox regression model  
2 (Equation 4-1) was the underlying basis for the splines which were fit to the lymphoid cancer  
3 exposure-response data.<sup>20</sup> Taking the log of both sides of Equation 4-1, log RR is a linear  
4 function of exposure (cumulative exposure is used here), and, with the two-piece log-linear  
5 spline approach, log RR is a function of two lines which join at a single point of inflection, called  
6 a “knot.” The shape of the two-piece log-linear spline model, in particular the slope in the  
7 low-exposure region, depends on the location of the knot. For this assessment, the knot was  
8 generally selected by evaluating different knots in increments of 100 ppm × days over some  
9 range of cumulative exposures starting at 0 and then choosing the one that resulted in the best  
10 (largest) model likelihood. The model likelihood did not change much across the different trial  
11 knots for any of the data sets, but it did change slightly, and the largest calculated likelihood was  
12 used as the basis for knot selection. For more discussion of the two-piece spline approach, see  
13 Appendix D.

14 For the lymphoid cancer data, the range examined for knot selection was from 0 to  
15 15,000 ppm × days, and the largest model likelihood was observed with the knot at 100 ppm ×  
16 days, although, as noted above, the model likelihood did not actually change much across the  
17 different trial knots (see Figure D-3a of Appendix D). The overall fit of this two-piece spline  
18 model was statistically significant ( $p = 0.048$ ); however, this model yielded a very steep slope in  
19 the exposure range below the knot of 100 ppm × days (see Figure 4-1),<sup>21</sup> and there was low  
20 confidence in the slope, given that it is based primarily on a relatively small number of cases in  
21 the low-exposure region. Consideration was also given to the two-piece log-linear spline model  
22 with a knot at 1,600 ppm × days, which provided a local maximum for the likelihood (see  
23 Figure D-3a of Appendix D; see Table 4-3 and Section D.3 of Appendix D for parameter  
24 estimates and fit statistics for the two spline models). However, the two-piece spline model with  
25 the knot at 1,600 ppm × days was not the model with the maximum likelihood; the  $p$ -value  
26 exceeded 0.05, although minimally ( $p = 0.072$ ); and the slope in the low-exposure range (i.e.,  
27 below the knot), although not as steep as the low-exposure slope from the two-piece log-linear  
28 spline model with the knot at 100 ppm × days, was still based primarily on a relatively small  
29 number of cases in the low-exposure region and, thus, there was less confidence in this slope  
30 than in that of the linear regression of the categorical results discussed above. Therefore, after  
31 examining the new modeling analyses, it was determined that the weighted linear regression of  
32 the categorical results still provided the best available approach for risk estimates for lymphoid

---

<sup>20</sup>As parameterized in Appendix D, for cumulative exposures less than the value of the knot,  $RR = e^{(\beta_1 \times \text{exposure})}$ ; for cumulative exposures greater than the value of the knot,  $RR = e^{(\beta_1 \times \text{exposure} + \beta_2 \times (\text{exposure} - \text{knot}))}$ .

<sup>21</sup>Although the log-linear spline segments appear fairly linear in the plotted range, they are not strictly linear.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 cancer.<sup>22</sup> Nonetheless, the two-piece log-linear spline model with the knot at 1,600 ppm × days  
 2 might be considered a reasonable alternative based on the continuous exposure data, and thus,  
 3 unit risk calculations using this model are included in the assessment for comparison purposes.  
 4

5  
 6 **Table 4-3. Exposure-response modeling results for all lymphohematopoietic**  
 7 **cancer and lymphoid cancer mortality in both sexes in the NIOSH cohort for**  
 8 **models not presented by [Steenland et al. \(2004\)](#)**  
 9

Model <sup>a</sup>	<i>p</i> -value <sup>b</sup>	Coefficient (SE) (per ppm × day)
<b>All lymphohematopoietic cancer<sup>c</sup></b>		
2-piece log-linear spline (knot at 500 ppm × days)	0.02	low-exposure spline segment: B1 = 0.00201 (0.0007731)
Linear regression of categorical results, excluding highest exposure group	0.08	0.0003459 (0.0001944)
<b>Lymphoid cancer<sup>d</sup></b>		
2-piece log-linear spline with maximum likelihood (knot at 100 ppm × days)	0.048	low-exposure spline segment: B1 = 0.01010 (0.00493)
Alternative 2-piece log-linear spline (knot at 1,600 ppm × days)	0.07	low-exposure spline segment: B1 = 0.0004893 (0.0002554)
Linear regression of categorical results, excluding the highest exposure quartile	0.18	0.000247 (0.000185)

10  
 11 <sup>a</sup>All with cumulative exposure in ppm × days as the exposure variable and with a 15-yr lag.

12 <sup>b</sup>*p*-values from likelihood ratio test, except for linear regressions of categorical results, where Wald *p*-values are reported.

13 <sup>c</sup>9<sup>th</sup> revision ICD codes 200–208; results based on 74 cases.

14 <sup>d</sup>NHL, myeloma, and lymphocytic leukemia (9<sup>th</sup> revision ICD codes 200, 202, 203, 204); results based on 53 cases.

15 Source: Additional analyses performed by Dr. Steenland (see Sections D.3 and D.4 of Appendix D), except for the  
 16 linear regression of the categorical results, which was performed by EPA.  
 17  
 18

19  
 20 For the weighted linear regression, the Cox regression results from the model with  
 21 categorical cumulative exposure and a 15-year lag (see Table 4-2) was used, excluding the

---

<sup>22</sup>When this assessment was largely complete, a two-piece linear spline model (with a linear model, i.e.,  $RR = 1 + \beta \times \text{exposure}$ , as the underlying basis for the spline pieces) was attempted, using the then just-published approach of [Langholz and Richardson \(2010\)](#) to model the individual data with cumulative exposure as a continuous variable; however, this model did not alleviate the problem of the excessively steep low-exposure spline segment (see Figure D-3c in Appendix D) and was not pursued further for the lymphoid cancer data. The [Langholz and Richardson \(2010\)](#) approach was also employed to model the lymphoid cancer data using linear RR models with cumulative exposure and log cumulative exposure as continuous variables; however, these linear models similarly did not alleviate the problems of the corresponding log-linear RR models (see Figure D-3c in Appendix D).

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 highest exposure group, as discussed above.<sup>23</sup> The weights used for the ORs were the inverses  
2 of the variances, which were calculated from the confidence intervals.<sup>24</sup> Mean and median  
3 exposures for the cumulative exposure groups were provided by Dr. Steenland (see Table D-3a  
4 of Appendix D).<sup>25</sup> The mean values were used for the weighted regression analysis because the  
5 cancer response is presumed to be a function of cumulative exposure, which is expected to be  
6 best represented by mean exposures. If the median values had been used, a slightly larger  
7 regression coefficient would have been obtained, resulting in slightly larger risk estimates. See  
8 Table 4-3 for the results obtained from the weighted linear regression and Figure 4-1 for a  
9 depiction of the resulting model.

10 As the lymphoid cancer data set is the primary data set used for the derivation of unit risk  
11 estimates for lymphohematopoietic cancers, a summary of all the models considered for  
12 modeling the lymphoid cancer exposure-response data and the judgments made about model  
13 selection is provided in Table 4-4. See Figures 4-1 and D-3c in Appendix D for visual  
14 representations of the models. See Tables 4-2 and 4-3 and Section D.3 of Appendix D for other  
15 information about the models.

---

<sup>23</sup>Concerns have been raised that this approach of dropping high-dose data appears arbitrary. It should be noted, however, that only the highest exposure group was omitted from the linear regression, and the exposure groupings were derived a priori by the NIOSH investigators and not by U.S. EPA in the course of its analyses.

<sup>24</sup>Equations for this weighted linear regression approach are presented in [Rothman \(1986\)](#) and summarized in Appendix F.

<sup>25</sup>Mean exposures for both sexes combined with a 15-year lag for the categorical exposure quartiles in Table 4-1 were 446; 2,143; 7,335; and 39,927 ppm × days. Median values were 374; 1,985; 6,755; and 26,373 ppm × days. These values are for the full cohort, not just the risk sets.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 **Table 4-4. Models considered for modeling the exposure-response data for**  
 2 **lymphoid cancer mortality in both sexes in the NIOSH cohort for the**  
 3 **derivation of unit risk estimates**  
 4

Model <sup>a</sup>	Comments
Cox regression (log-linear) model	Inadequate overall statistical fit ( $p = 0.22$ ) and poor visual fit
Cox regression model with log cumulative exposure	Good overall statistical fit ( $p = 0.02$ ) but too steep in the low-exposure region
2-piece log-linear spline with maximum likelihood (knot at 100 ppm × days)	Good overall statistical fit ( $p = 0.048$ ) but too steep in the low-exposure region
Alternative 2-piece log-linear spline (knot at 1,600 ppm × days)	ALTERNATIVE. Not 2-piece spline model with maximum likelihood and not statistically significant fit ( $p = 0.07$ ) but reasonable alternative model (see text above)
linear model ( $RR = 1 + \beta \times \text{exposure}$ ) <sup>b</sup>	Inadequate overall statistical fit ( $p = 0.13$ ) and poor visual fit
linear model with log cumulative exposure	Good overall statistical fit ( $p = 0.02$ ) but too steep in the low-exposure region
2-piece linear spline model (knot at 100 ppm × days)	Good overall statistical fit ( $p = 0.04$ ) but too steep in the low-exposure region
Linear regression of categorical results, excluding the highest exposure quartile	<b>SELECTED.</b> The continuous exposure supralinear models (e.g., the log-cumulative-exposure models and the 2-piece log-linear spline model with the maximum likelihood) are statistically significant for lymphoid cancer mortality; however, they are too steep in the low-exposure region for the derivation of stable unit risk estimates. Thus, the linear regression model of categorical results, excluding the highest exposure quartile, was used for the derivation of unit risk estimates, despite the lack of statistical significance ( $p = 0.18$ ), as it was considered a better representation of the data in the low-exposure region. Lack of statistical significance is not critical given the low statistical power with categorical data and the statistical significance of the continuous exposure supralinear models, which establishes the significance of the exposure-response correlation for the underlying data.

5  
 6 <sup>a</sup>All with cumulative exposure as the exposure variable, except where noted, and with a 15-yr lag.

7 <sup>b</sup>The linear models are discussed in footnote #14 above.  
 8  
 9

10 The linear regression of the categorical results for males and females combined and the  
 11 actuarial program (life-table analysis) were used to estimate the exposure level ( $EC_x$ ; “effective  
 12 concentration”) and the associated 95% lower confidence limit ( $LEC_x$ ) corresponding to an extra  
 13 risk of 1% ( $x = 0.01$ ). A 1% extra risk level is commonly used for the determination of the point  
 14 of departure (POD) for low-exposure extrapolation from epidemiological cancer data (except for  
 15 rare cancers); higher extra risk levels, such as 10%, would be an upward extrapolation for these

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 data. Thus, 1% extra risk was selected for determination of the POD, and, consistent with EPA's  
2 *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)), the LEC value corresponding to  
3 that risk level was used as the POD to derive the cancer unit risk estimates.

4 Because EtO is DNA-reactive and has direct mutagenic activity (see Section 3.3.3),  
5 which is one of the cases cited by EPA's *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA,](#)  
6 [2005a](#)) for the use of linear low-dose extrapolation, a linear low-exposure extrapolation was  
7 performed. The EC<sub>01</sub>, LEC<sub>01</sub>, and inhalation unit risk estimate calculated for lymphoid cancer  
8 mortality from the linear regression of the categorical results are presented in Table 4-5 (the  
9 incidence results also presented in Table 4-5 are discussed in Section 4.1.1.3 below). The  
10 resulting unit risk estimate for lymphoid cancer mortality based on the linear regression of the  
11 categorical results for both sexes using cumulative exposure with a 15-year lag is 0.397 per ppm.  
12 The unit risk estimate from the alternative model, the two-piece log-linear spline model with the  
13 knot at 1,600 ppm × days, is 0.917 per ppm, about 2.3-times higher than the estimate from the  
14 preferred model. EC<sub>01</sub> and LEC<sub>01</sub> estimates from the other models considered are presented for  
15 comparison only, to illustrate the differences in model behavior at the low end of the  
16 exposure-response range. Unit risk estimates are not presented for these other models because,  
17 as discussed above, these models were deemed unsuitable for the derivation of risks from (low)  
18 environmental exposure levels. The log cumulative exposure model, with its extreme  
19 supralinearity in the lower exposure region, and the two-piece log-linear spline model with the  
20 maximum likelihood (knot at 100 ppm × days), with its very steep low-exposure slope, yield  
21 substantially lower EC<sub>01</sub> estimates (0.00441 ppm and 0.000982 ppm, respectively). Converting  
22 the units, the resulting unit risk estimate of 0.397 per ppm from the linear regression model of  
23 the categorical results corresponds to a unit risk estimate of  $2.17 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  for lymphoid  
24 cancer mortality.<sup>26</sup>

25 As discussed above, risk estimates based on the all lymphohematopoietic cancer results  
26 are also derived for comparison. The same methodology presented above for the lymphoid  
27 cancer results was used for the all lymphohematopoietic cancer risk estimates. Age-specific  
28 background mortality rates for all lymphohematopoietic cancers for the year 2004 were obtained  
29 from the NCHS Data Warehouse website ([http://www.cdc.gov/nchs/datawh/statab/unpubd/  
30 mortabs.htm](http://www.cdc.gov/nchs/datawh/statab/unpubd/mortabs.htm)). The results of Dr. Steenland's reanalyses using the Cox regression models  
31 presented in the [Steenland et al. \(2004\)](#) paper with data for males and females combined are  
32 presented in Table 4-2. As for lymphoid cancer and for all hematopoietic cancer in males  
33 presented in the [Steenland et al. \(2004\)](#) paper, the only statistically significant Cox regression

---

<sup>26</sup>Conversion equation: 1 ppm = 1,830  $\mu\text{g}/\text{m}^3$ .

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 **Table 4-5. EC<sub>01</sub>, LEC<sub>01</sub>, and unit risk estimates for lymphoid cancer<sup>a</sup>**  
 2

Model <sup>c</sup>	Mortality			Incidence <sup>b</sup>		
	EC <sub>01</sub> (ppm)	LEC <sub>01</sub> (ppm)	Unit risk (per ppm)	EC <sub>01</sub> (ppm)	LEC <sub>01</sub> (ppm)	Unit risk (per ppm)
Cox regression model, log cumulative exposure, 15-yr lag	0.00441	0.000428	-- <sup>d</sup>	0.000288	0.0000898	-- <sup>d</sup>
low-exposure log-linear spline from 2-piece spline model with maximum likelihood (knot at 100 ppm × days), <sup>e</sup> cumulative exposure, 15-yr lag	0.000982	0.000545	-- <sup>d</sup>	0.000525	0.000291	-- <sup>d</sup>
Alternative low-exposure log-linear spline (knot at 1,600 ppm × days), <sup>f</sup> cumulative exposure, 15-yr lag	0.0203	0.0109	0.917	0.0108	0.00583	1.72 <sup>g</sup>
Linear regression of categorical results, cumulative exposure, 15-yr lag <sup>h</sup>	0.0564	0.0252	0.397	0.0254	0.0114	<b>0.877</b>

3  
 4 <sup>a</sup>From lifetime continuous exposure. Unit risk = 0.01/LEC<sub>01</sub>.

5 <sup>b</sup>Incidence estimates are presented here for comparison; they are derived in Section 4.1.1.3.

6 <sup>c</sup>From Dr. Steenland's analyses for males and females combined (see Section D.3 of Appendix D), Cox regression  
 7 models. Note that the EC<sub>01</sub> and LEC<sub>01</sub> results presented here will not exactly match those presented in Appendix D  
 8 because, although EPA used the regression coefficients reported by Dr. Steenland in Appendix D, the life-table  
 9 analyses using 2004 all-cause mortality rates were redone to be more up-to-date and consistent with the  
 10 cause-specific mortality rates; the results presented in Appendix D were based on life-table analyses using 2000  
 11 all-cause mortality rates.

12 <sup>d</sup>Unit risk estimates are not presented for these models because these models were deemed unsuitable for the  
 13 derivation of risks from (low) environmental exposure levels (see text).

14 <sup>e</sup>Using regression coefficient from low-exposure segment of two-piece log-linear spline model with largest  
 15 likelihood (knot at 100 ppm × days); see text and Appendix D. Each of the EC<sub>01</sub> values is below the value of 0.0013  
 16 ppm roughly corresponding to the knot of 100 ppm × days [(100 ppm × days) × (10 m<sup>3</sup>/20 m<sup>3</sup>) × (240 d/365 d) ×  
 17 (365 d/yr)/70 yr = 0.0013 ppm] and, thus, appropriately in the range of the low-exposure segment.

18 <sup>f</sup>Using regression coefficient from low-exposure segment of alternative two-piece log-linear spline model (local  
 19 maximum likelihood) with a knot at 1,600 ppm × days. Each of the EC<sub>01</sub> values is below the value of 0.021 ppm  
 20 roughly corresponding to the knot of 1,600 ppm × days (see footnote e for calculation) and, thus, appropriately in  
 21 the range of the low-exposure segment.

22 <sup>g</sup>To obtain unit risk estimates less than 1, convert to risk per ppb (e.g., 1.72 per ppm = 1.72 × 10<sup>-3</sup> per ppb).

23 <sup>h</sup>Regression coefficient derived from linear regression of categorical Cox regression results from Table 4-2, as  
 24 described in Section 4.1.1.2. Each of the EC<sub>01</sub> values is appropriately below the value of 0.090 ppm roughly  
 25 corresponding to the value of about 7,000 ppm × days (see footnote e for calculation) above which the linear  
 26 regression model of the categorical results does not apply (see Figure 4-1).  
 27

1 model was for log cumulative exposure with a 15-year lag ( $p = 0.01$ ). The cumulative exposure  
2 model did not provide an adequate fit to the data and is not considered further here ( $p = 0.35$ ).

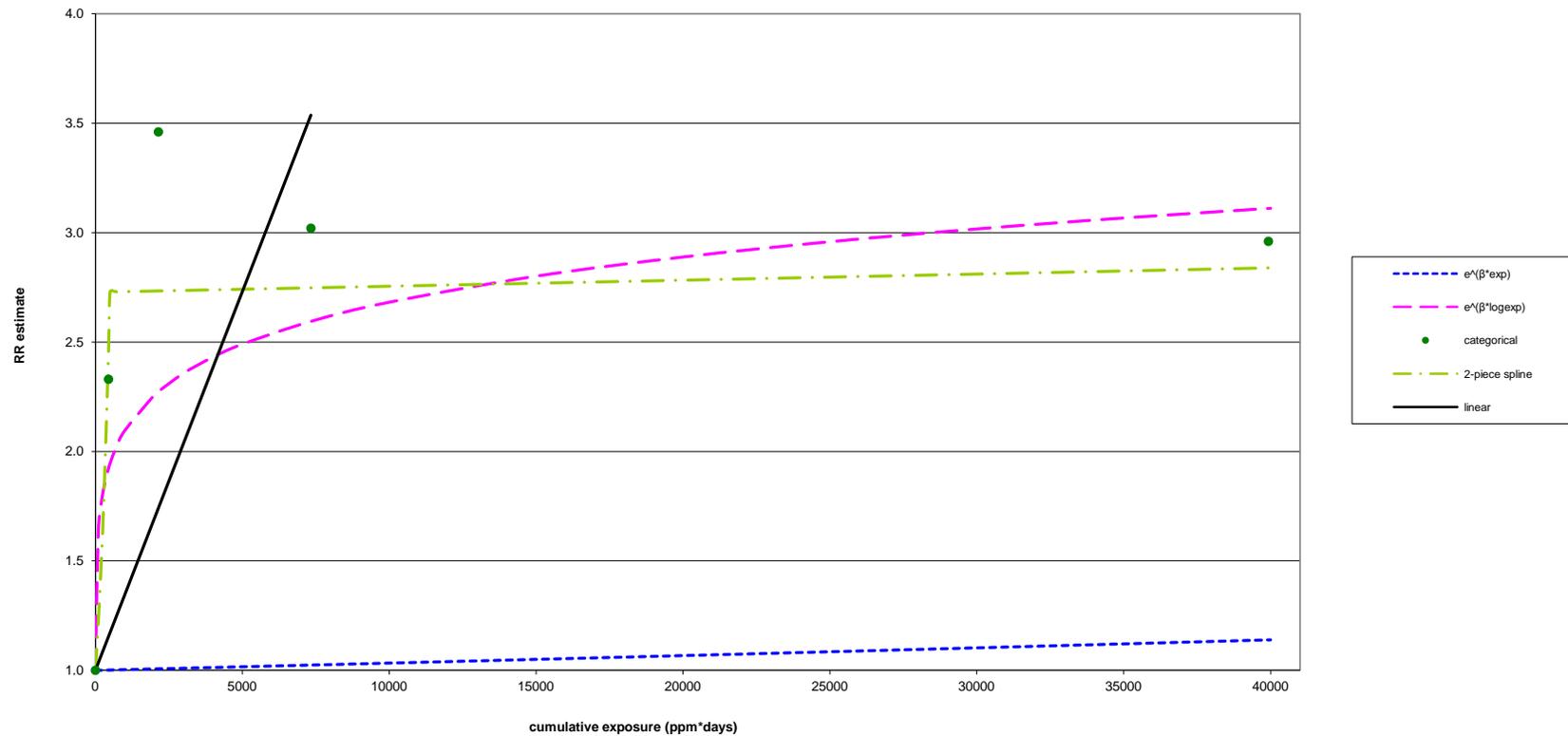
3 Because of the problems with the supralinear log cumulative exposure model which are  
4 discussed for the lymphoid cancers above, EPA again investigated the use of a two-piece  
5 log-linear spline model to attempt to address the supralinearity of the data while avoiding the  
6 extreme low-exposure curvature obtained with the log cumulative exposure model. For the all  
7 lymphohematopoietic cancer mortality data, the range examined for knot selection was from 0 to  
8 7,000 ppm  $\times$  days, and the largest model likelihood was obtained with the knot at 500 ppm  $\times$   
9 days (see Figure D-4a of Appendix D). See Table 4-3 and Section D.4 of Appendix D for  
10 parameter estimates and fit statistics for the two-piece spline model. As with the lymphoid  
11 cancer mortality results, however, this model resulted in an apparently excessively steep  
12 low-exposure spline (see Figure 4-2), so, again, the linear regression model of the categorical  
13 results was used to derive the cancer unit risk estimate for this data set.<sup>27</sup>

14 For the weighted linear regression, the results from the Cox regression model with  
15 categorical cumulative exposure and a 15-year lag (see Table 4-2) were used, excluding the  
16 highest exposure group, and the approach discussed above for lymphoid cancer mortality. See  
17 Table 4-3 for the results obtained from the weighted linear regression and Figure 4-2 for a  
18 graphical presentation of the resulting linear regression model. As discussed above, this linear  
19 regression model was used to derive the unit risk estimates for all lymphohematopoietic cancer.  
20

---

<sup>27</sup>When this assessment was largely complete, a two-piece linear spline model (with a linear model, i.e.,  $RR = 1 + \beta \times \text{exposure}$ , as the underlying basis for the spline pieces) was attempted, using the then just-published approach of [Langholz and Richardson \(2010\)](#) to model the individual data with cumulative exposure as a continuous variable; however, this model did not alleviate the problem of the excessively steep low-exposure spline segment (see Figure D-4c in Appendix D) and was not pursued further for the all lymphohematopoietic cancer data. The [Langholz and Richardson \(2010\)](#) approach was also employed to model the all lymphohematopoietic cancer data using linear RR models with cumulative exposure and log cumulative exposure as continuous variables; however, these linear models similarly did not alleviate the problems of the corresponding log-linear RR models (see Figure D-4c in Appendix D).

*This document is a draft for review purposes only and does not constitute Agency policy.*



**Figure 4-2. RR estimate for all lymphohematopoietic cancer vs. mean exposure (with 15-year lag, unadjusted for continuous exposure).**

$e^{(\beta \times \text{exp})}$ : Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$ ;  $e^{(\beta \times \ln(\text{exposure}))}$ : Cox regression results for  $RR = e^{(\beta \times \ln(\text{exposure}))}$ ; categorical: Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$  with categorical exposures; linear: weighted linear regression of categorical results, excluding highest exposure group (see text); 2-piece spline: 2-piece log-linear spline model with knot at 500 ppm × days (see text). (Note that, with the exception of the categorical results and the linear regression of the categorical results, the different models have different implicitly estimated baseline risks; thus, they are not strictly comparable to each other in terms of RR values, i.e., along the y-axis. They are, however, comparable in terms of general shape.)

Source: Steenland reanalyses for males and females combined; see Appendix D (except for linear regression of the categorical results, which was done by EPA).

1 The EC<sub>01</sub>, LEC<sub>01</sub>, and inhalation unit risk estimate calculated for all  
 2 lymphohematopoietic cancer mortality from the linear regression model of the categorical results  
 3 are presented in Table 4-6 (the incidence results also presented in Table 4-6 are discussed in  
 4 Section 4.1.1.3 below). The resulting unit risk estimate for all lymphohematopoietic cancer  
 5 mortality based on the linear regression of the categorical results for both sexes using cumulative  
 6 exposure with a 15-year lag is 0.680 per ppm. EC<sub>01</sub> and LEC<sub>01</sub> estimates from the other models  
 7 considered are presented for comparison only, to illustrate the differences in model behavior at  
 8 the low end of the exposure-response range. Unit risk estimates are not presented for these other  
 9 models because, as discussed above, these models were deemed unsuitable for the derivation of  
 10 risks from (low) environmental exposure levels. The resulting unit risk estimate for all  
 11 lymphohematopoietic cancer mortality from the linear regression model of the categorical results  
 12 is similar to that for lymphoid cancer mortality (70% higher; see Table 4-5). Converting the  
 13 units, the resulting unit risk estimate of 0.680 per ppm corresponds to a unit risk estimate of  
 14  $3.72 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  for all lymphohematopoietic cancer mortality.

15  
 16  
 17 **Table 4-6. EC<sub>01</sub>, LEC<sub>01</sub>, and unit risk estimates for all lymphohematopoietic**  
 18 **cancer<sup>a</sup>**  
 19

Model <sup>c</sup>	Mortality			Incidence <sup>b</sup>		
	EC <sub>01</sub> (ppm)	LEC <sub>01</sub> (ppm)	Unit risk (per ppm)	EC <sub>01</sub> (ppm)	LEC <sub>01</sub> (ppm)	Unit risk (per ppm)
Log cumulative exposure, 15-yr lag	0.00140	0.000245	-- <sup>d</sup>	0.000190	0.0000753	-- <sup>d</sup>
Low-exposure log-linear spline; <sup>e</sup> cumulative exposure, 15-yr lag	0.00377	0.00231	-- <sup>d</sup>	0.00216	0.00132	-- <sup>d</sup>
Linear regression of categorical results, cumulative exposure, 15-yr lag <sup>f</sup>	0.0283	0.0147	0.680	0.0144	0.00746	<b>1.34<sup>g</sup></b>

20  
 21 <sup>a</sup>From lifetime continuous exposure. Unit risk = 0.01/LEC<sub>01</sub>.

22 <sup>b</sup>Incidence estimates presented here for comparison; they are derived in Section 4.1.1.3.  
 23

1 **Table 4 6. EC01, LEC01, and unit risk estimates for all**  
2 **lymphohematopoietic cancer<sup>a</sup> (continued)**  
3  
4

5 <sup>c</sup>From Dr. Steenland's analyses for males and females combined (see Appendix D), Cox regression models. Note  
6 that the EC<sub>01</sub> and LEC<sub>01</sub> results presented here will not exactly match those presented in Appendix D because,  
7 although EPA used the regression coefficients reported by Dr. Steenland in Appendix D, the life-table analyses  
8 using 2004 all-cause mortality rates were redone to be more up-to-date and consistent with the cause-specific  
9 mortality rates; the results presented in Appendix D were based on life-table analyses using 2000 all-cause mortality  
10 rates.

11 <sup>d</sup>Unit risk estimates are not presented for these models because these models were deemed unsuitable for the  
12 derivation of risks from (low) environmental exposure levels (see text).

13 <sup>e</sup>Using regression coefficient from low-exposure segment of two-piece log-linear spline model with knot at 500 ppm  
14 × days; see text and Appendix D. Each of the EC<sub>01</sub> values is below the value of 0.0064 ppm roughly corresponding  
15 to the knot of 500 ppm × days [(500 ppm × days) × (10 m<sup>3</sup>/20 m<sup>3</sup>) × (240 d/365 d) × (365 d/yr)/70 yr = 0.0064 ppm]  
16 and, thus, appropriately in the range of the low-exposure segment.

17 <sup>f</sup>Regression coefficient derived from linear regression of categorical Cox regression results from Table 4-2, as  
18 described in Section 4.1.1.2. Each of the EC<sub>01</sub> values is appropriately below the value of 0.064 ppm roughly  
19 corresponding to the value of about 5,000 ppm × days (see footnote d for calculation) above which the linear  
20 regression model of the categorical results does not apply (see Figure 4-2).

21 <sup>g</sup>For unit risk estimates below 1, convert to risk per ppb (e.g., 1.34 per ppm = 1.34 × 10<sup>-3</sup> per ppb).  
22  
23

24 **4.1.1.3. Prediction of Lifetime Extra Risk of Lymphohematopoietic Cancer Incidence**

25 EPA cancer risk estimates are typically derived to represent an upper bound on increased  
26 risk of cancer *incidence*, as from experimental animal incidence data. Cancer data from  
27 epidemiologic studies are commonly mortality data, as is the case in the [Steenland et al. \(2004\)](#)  
28 study. For tumor sites with low survival rates, mortality-based estimates are reasonable  
29 approximations of cancer incidence risk; however, for many lymphohematopoietic cancers, the  
30 survival rate is substantial, and incidence risk estimates are preferred by EPA ([U.S. EPA, 2005a](#)).

31 Therefore, another calculation was done using the same regression coefficients presented  
32 above (see Section 4.1.1.2), but with age-specific lymphoid cancer incidence rates for the  
33 relevant subcategories of lymphohematopoietic cancer (NHL, myeloma, and lymphocytic  
34 leukemia) for 2000–2004 from SEER ([Ries et al., 2007](#)); Tables XIX, XVIII, XIII: both sexes,  
35 all races) in place of the lymphoid cancer mortality rates in the actuarial program. SEER collects  
36 good-quality cancer incidence data from a variety of geographical areas in the United States.  
37 The incidence data used here are from “SEER 17,” a registry of seventeen states, regions, and  
38 cities covering about 26% of the U.S. population.

39 The incidence risk calculation assumes that (1) lymphoid cancer incidence and mortality  
40 have the same exposure-response relationship for the relative rate of effect from EtO exposure  
41 and that (2) the incidence data are for first occurrences of primary lymphoid cancer or that  
42 relapses and secondary lymphoid cancers provide a negligible contribution. (The latter  
43 assumption is probably sound; the former assumption is more potentially problematic. Because

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 various lymphoid cancer subtypes with different survival rates are included in the categorization  
2 of lymphoid cancers, if the EtO-associated relative rates of the subtypes differ, a bias could  
3 occur, resulting in either an underestimation or overestimation of the extra risk for lymphoid  
4 cancer incidence.)<sup>28</sup> Potential concern that the incidence risk estimates might be overestimated  
5 would come primarily from the inclusion of multiple myeloma, because that subtype has the  
6 lowest incidence:mortality ratios (and, thus, if that subtype were driving the increased mortality  
7 observed for the lymphoid cancer grouping, then including the incidence rates for the other  
8 subtypes, which have higher incidence:mortality ratios, might inflate the incidence risk  
9 estimates). Multiple myelomas, however, constitute only 25% of the lymphoid cancer cases in  
10 the cohort, and there is no evidence that multiple myeloma is driving the EtO-induced excess in  
11 lymphoid cancer mortality.<sup>29</sup> Thus, using the total lymphoid cancer incidence rates is not  
12 expected to result in an overestimation of the incidence risk estimates; if anything, the incidence  
13 risks would likely be diluted with the inclusion of the multiple myeloma rates. The incidence  
14 risk calculation also relies on the fact that the lymphoid cancer incidence rates (more  
15 specifically, the differential rates obtained by subtracting the mortality rates from the incidence  
16 rates) are small when compared with the all-cause mortality rates.<sup>30</sup> The resulting EC<sub>01</sub> and  
17 LEC<sub>01</sub> estimates for lymphoid cancer incidence from the various models examined are presented  
18 in Table 4-5. The unit risk estimate for lymphoid cancer incidence from the selected linear  
19 regression model of the categorical results is 0.877 per ppm. The unit risk estimate for lymphoid  
20 cancer incidence from the alternative model, the two-piece log-linear spline model with the knot

---

<sup>28</sup>[Sielken and Valdez-Flores \(2009\)](#) reject the assumption that lymphohematopoietic cancer incidence and mortality have the same exposure-response relationship, reporting that, except at high exposure levels, the exposure-response data in the male workers in the NIOSH cohort are consistent with a decreased survival time and suggesting that this could explain the observed increases in mortality. However, they do not establish that this is what is occurring, and the mechanistic data support an exposure-related increase in incident cancers. See Appendix A.2.20 for a more detailed discussion of this issue.

<sup>29</sup>According to data from SEER ([www.seer.cancer.gov](http://www.seer.cancer.gov)), 25% is below the proportion of multiple myeloma deaths one would expect based on age-adjusted U.S. background mortality rates of multiple myeloma, NHL, and chronic lymphocytic leukemia, and these 3 subtypes have the same pattern for mortality rates increasing as a function of age mostly above age 50, so the comparison with lifetime background rates is reasonable. In addition, the low proportion of multiple myeloma deaths in the lymphoid cancer subgrouping cannot be attributed to an underrepresentation of blacks, who have incidence rates of multiple myeloma more than twice those of whites (<http://seer.cancer.gov/statfacts/html/mulmy.html>), in the cohort because blacks comprise 16% of the cohort versus 12.3% in the U.S. population.

<sup>30</sup>[Sielken and Valdez-Flores \(2009\)](#) suggest that the methods used by EPA to calculate incidence risk estimates in the life-table analysis are inappropriate; however, as explained in more detail in Appendix A.2.20, we disagree. For the situation where the cause-specific incidence rates are small compared to the all-cause mortality rates, as with lymphoid cancer, there is no problem, as [Sielken and Valdez-Flores \(2009\)](#) themselves demonstrate, and, for the situation where the cause-specific incidence rates are not negligible compared to the all-cause mortality rates, as with breast cancer, an adjustment was made in the analysis to remove those with incident cases from the population at risk, i.e., those "surviving" each interval without a diagnosis of breast cancer (see Section 4.1.2.3). See Appendix A.2.20 for a more detailed discussion of this issue.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 at 1,600 ppm × days, is 1.72 per ppm, about 2.0 times higher than the estimate from the preferred  
2 model.

3 Overall, as discussed above, the preferred estimate for the unit risk for lymphoid cancer is  
4 the estimate of **0.877 per ppm** ( $4.79 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$ ) derived, using incidence rates for the  
5 cause-specific background rates, from the weighted linear regression of the categorical results,  
6 dropping the highest exposure group. The incidence unit risk estimate from the linear regression  
7 model of the categorical results is about 120% higher than (i.e., 2.2 times) the mortality-based  
8 estimate from the same model.

9 As discussed in Section 4.1.1.2, risk estimates based on the results of Dr. Steenland's  
10 reanalyses of the all lymphohematopoietic cancer data (see Appendix D and Table 4-2) are also  
11 derived for comparison. The same methodology presented above for the lymphoid cancer  
12 incidence results was used for the all lymphohematopoietic cancer incidence risk estimates, and  
13 the same assumptions apply. Age-specific SEER incidence rates for all lymphohematopoietic  
14 cancer for the years 2000–2004 were used ([Ries et al., 2007](#)); Tables XIX, IX, XVIII, and XIII:  
15 both sexes, all races). The  $\text{EC}_{01}$  and  $\text{LEC}_{01}$  estimates for all lymphohematopoietic cancer  
16 incidence from the different all lymphohematopoietic cancer mortality models examined are  
17 presented in Table 4-6. The resulting unit risk estimate for all lymphohematopoietic cancer  
18 incidence from the linear regression of the categorical results is about 2.0-times the  
19 mortality-based estimate and about 1.5-times the lymphoid cancer incidence estimate (see  
20 Table 4-5).

## 21 22 **4.1.2. Risk Estimates for Breast Cancer**

### 23 **4.1.2.1. Breast Cancer Results From the NIOSH Study**

24 The [Steenland et al. \(2004\)](#) study discussed above in Section 4.1.1.1 also presents results  
25 from exposure-response analyses for breast cancer mortality in female workers. [Steenland et al.](#)  
26 [\(2003\)](#) present results of a breast cancer incidence study of a subcohort of the female workers  
27 from the NIOSH cohort. In addition to the analyses presented in the [Steenland et al. \(2003\)](#) and  
28 [Steenland et al. \(2004\)](#) papers, Dr. Steenland did subsequent analyses of the breast cancer  
29 incidence and mortality data sets for EPA; these are discussed below and reported in  
30 Sections D.1 and D.2 of Appendix D, respectively.

### 31 32 **4.1.2.2. Prediction of Lifetime Extra Risk of Breast Cancer Mortality**

33 Results from the Cox regression models presented by [Steenland et al. \(2004\)](#), with some  
34 reanalyses reported by Dr. Steenland in Appendix D (see Section D.2), are summarized in  
35 Table 4-7. These models were considered for the derivation of unit risk estimates for breast

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 cancer mortality in females from continuous environmental exposure to EtO, applying the  
2 methodologies described in Section 4.1.1.2.

3  
4  
5 **Table 4-7. Cox regression results for breast cancer mortality in females in**  
6 **the NIOSH cohort,<sup>a</sup> for models presented in [Steenland et al. \(2004\)](#)**  
7

Exposure variable <sup>b</sup>	p-value <sup>c</sup>	Coefficient (SE) (per ppm × day)	ORs by category <sup>d</sup> (95% CI)
Cumulative exposure, 20-yr lag <sup>e</sup>	0.06	0.0000122 (0.00000641)	
Log cumulative exposure, 20-yr lag <sup>f</sup>	0.01	0.084 (0.035)	
Categorical cumulative exposure, 20-yr lag <sup>f</sup>	0.07		1.00, 1.76 (0.91–3.43), 1.77 (0.88–3.56), 1.97 (0.94–4.06), 3.13 (1.42–6.92)

8  
9 <sup>a</sup>Based on 103 breast cancer (ICD-9 174,175) deaths.

10 <sup>b</sup>Cumulative exposure is in ppm × days.

11 <sup>c</sup>p-values reported by [Steenland et al. \(2004\)](#).

12 <sup>d</sup>Exposure categories are 0; >0–646; 647–2,779; 2,780–12,321; ≥12,322 ppm × days.

13 <sup>e</sup>From reanalyses in Section D.2 of Appendix D; [Steenland et al. \(2004\)](#) reported the Cox regression results for  
14 cumulative exposure with no lag.

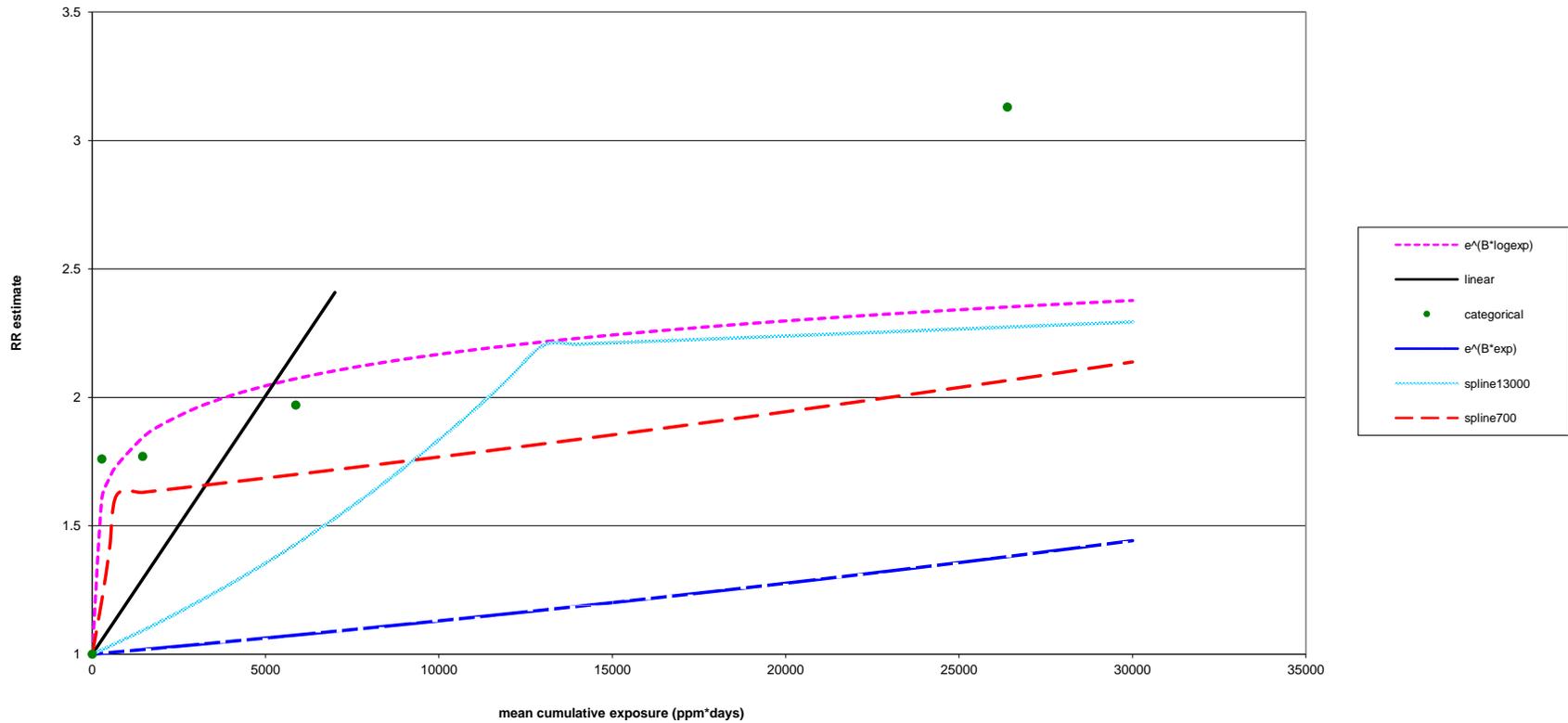
15 <sup>f</sup>From Table 8 of [Steenland et al. \(2004\)](#).

16  
17  
18 U.S. age-specific all-cause mortality rates for 2000 for females of all race groups  
19 combined ([Miniño et al., 2002](#)) were used to specify the all-cause background mortality rates in  
20 the actuarial program (life-table analysis). The National Center for Health Statistics 1997–2001  
21 cause-specific background mortality rates for invasive breast cancers in females were obtained  
22 from a SEER report ([Ries et al., 2004](#)). The risks were computed up to age 85 for continuous  
23 exposures to EtO, conversions were made between occupational EtO exposures and continuous  
24 environmental exposures, and 95% UCLs were calculated for the relative rates, as described  
25 above.

26 The only statistically significant Cox regression model presented by [Steenland et al.](#)  
27 [\(2004\)](#) for breast cancer mortality in females was for log cumulative exposure with a 20-year lag  
28 ( $p = 0.01$ ). However, as for the lymphohematopoietic cancers in Section 4.1.1, using the log  
29 cumulative exposure model to estimate the risks from low environmental exposures is  
30 problematic because this model is highly supralinear and results are unstable for low exposures  
31 (see Figure 4-3). The cumulative exposure model, which is typically used and which is stable at  
32 low exposures, was nearly statistically significant ( $p = 0.06$  with a 20-year lag; see Section D.2  
33 of Appendix D) in terms of the global fit to the data; however, the Cox regression model with

1 cumulative exposure is sublinear and does not reflect the apparent supralinearity of the breast  
2 cancer mortality data (see Figure 4-3).

3 In a 2006 external review draft of this assessment ([U.S. EPA, 2006a](#)), which relied on the  
4 original published results of [Steenland et al. \(2004\)](#), EPA proposed that the best way to reflect  
5 the exposure-response relationship in the lower exposure region, which is the region of interest  
6 for low-exposure extrapolation, was to do a weighted linear regression of the results from the  
7 Cox regression model with categorical cumulative exposure and a 20-year lag. In addition, the  
8 highest exposure group was not included in the regression to alleviate some of the “plateauing”  
9 in the exposure-response relationship at higher exposure levels and to provide a better fit to the  
10 lower exposure data. Linear modeling of categorical epidemiologic data and elimination of the  
11 highest exposure group(s) in certain circumstances to obtain a better fit of low-exposure data are  
12 both standard techniques used in EPA dose-response assessments ([U.S. EPA, 2005a](#)). However,  
13 as discussed in Section 4.1.1.2 for the similarly supralinear lymphohematopoietic cancer data,  
14 the SAB panel that reviewed the draft assessment recommended that EPA employ models using  
15 the individual exposure data as an alternative to modeling the published grouped data ([SAB,](#)  
16 [2007](#)). Consequently, it was determined that, using the full data set, an alternative way to  
17 address the supralinearity of the data (while avoiding the extreme low-exposure curvature  
18 obtained with the log cumulative exposure model) might be to use a two-piece spline model, and  
19 Dr. Steenland was commissioned to do the spline analyses using the full data set with cumulative  
20 exposure as a continuous variable. His findings are reported in Section D.2 of Appendix D, and  
21 the results for the breast cancer mortality analyses are summarized below.



**Figure 4-3. RR estimate for breast cancer mortality vs. mean exposure (with 20-year lag, unadjusted for continuous exposure).**

$e^{(B \cdot \text{exp})}$ : Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$ ;  $e^{(B \cdot \log \text{exp})}$ : Cox regression results for  $RR = e^{(\beta \times \ln(\text{exposure}))}$ ; categorical: Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$  with categorical exposures; linear: weighted linear regression of categorical results, excluding highest exposure group (see text); spline700(13000): 2-piece log-linear spline model with knot at 700 (13,000) ppm × days (see text). (Note that, with the exception of the categorical results and the linear regression of the categorical results, the different models have different implicitly estimated baseline risks; thus, they are not strictly comparable to each other in terms of RR values, i.e., along the y-axis. They are, however, comparable in terms of general shape.)

Source: Steenland reanalyses with 20-year lag; see Section D.2 of Appendix D (except for linear regression of the categorical results, which was done by EPA).

1 For the two-piece log-linear spline modeling approach, as described in Section 4.1.1.2  
2 and discussed more fully in Section D.2 of Appendix D, the Cox regression model was the  
3 underlying basis for the splines which were fit to the breast cancer mortality exposure-response  
4 data (cumulative exposure, with a 20-year lag), and thus, log RR is a function of two lines that  
5 join at a single point of inflection, called a “knot.” The shape of the two-piece log-linear spline  
6 model, in particular the slope in the low-exposure region, depends on the location of the knot.  
7 Knot selection was made by trying different knots over a reasonable range and choosing the one  
8 that resulted in the largest model likelihood. For the breast cancer mortality data, the range  
9 examined for knot selection was from 0 to 25,000 ppm × days, using increments of  
10 100 ppm × days to 7,000 ppm × days and increments of 1,000 ppm × days above  
11 7,000 ppm × days. The largest model likelihood was observed with the knot at 700 ppm × days,  
12 although, as noted above, the model likelihood did not change much across the various trial  
13 knots (see Figure D-2a of Appendix D).<sup>31</sup> The overall fit of this two-piece spline model was not  
14 statistically significant ( $p = 0.067$ ). Of greater concern, this model yielded a very steep slope in  
15 the exposure range below the knot of 700 ppm × days (see Figure 4-3), and confidence in the  
16 slope was low, given that it is based primarily on relatively few cases in the low-exposure region.

17 Consideration was also given to the two-piece log-linear spline model with a knot at  
18 13,000 ppm × days, which provided a local maximum for the likelihood (see Figure D-2a' of  
19 Appendix D; see Table 4-8 and Section D.2 of Appendix D for parameter estimates and fit  
20 statistics for the two spline models).<sup>32</sup> However, the two-piece spline model with the knot at  
21 13,000 ppm × days was not the model with the maximum likelihood; its  $p$ -value similarly  
22 exceeded 0.05, although minimally ( $p = 0.074$ ); and the large difference between the knots in the  
23 maximum-likelihood and alternative two-piece spline models decreased confidence in this  
24 alternative two-piece spline model. Therefore, after examining the new modeling analyses, it  
25 was determined that the weighted linear regression of the categorical results still provided the  
26 best available approach for risk estimates for lymphoid cancer.<sup>33</sup> Nonetheless, the two-piece  
27 log-linear spline model with the knot at 13,000 ppm × days might be considered a reasonable

---

<sup>31</sup>Using the two-piece log-linear spline model with the maximum likelihood (knot at 700 ppm × days), a regression coefficient of 0.0006877 per ppm × day (SE = 0.0004171 per ppm × day) was obtained for the low-exposure spline segment (see Appendix D).

<sup>32</sup>Using the alternative two-piece log-linear spline model with the knot at 13,000 ppm × days, a regression coefficient of 0.000607 per ppm × day (SE = 0.0000309 per ppm × day) was obtained for the low-exposure spline segment (see Appendix D).

<sup>33</sup>When this assessment was largely complete, some linear models (i.e.,  $RR = 1 + \beta \times \text{exposure}$ ) were investigated, using the then just-published approach of [Langholz and Richardson \(2010\)](#), to model the individual data with cumulative exposure as a continuous variable; however, these linear models did not alleviate the problems of the corresponding log-linear RR models and the results are not presented here (see Section D.2.c and Figure D-2d in Appendix D).

1 alternative based on the continuous exposure data, and thus, unit risk calculations using this  
2 model are included in the assessment for comparison purposes. (For more discussion of the  
3 breast cancer mortality exposure-response modeling using the continuous exposure data, see  
4 Section D.2 of Appendix D.)

5 For the weighted linear regression, the results from the Cox regression model with  
6 categorical cumulative exposure (and a 20-year lag) presented in Table 4-7 were used, excluding  
7 the highest exposure group, and the approach discussed above for the lymphoid cancers (see  
8 Section 4.1.1.2).<sup>34</sup> Mean and median exposures for the cumulative exposure groups were  
9 provided by Dr. Steenland (see Appendix D).<sup>35</sup> See Table 4-8 for the results obtained from the  
10 weighted linear regression of the categorical results and mean exposures and Figure 4-3 for a  
11 depiction of the resulting linear regression model.

12 The linear regression of the categorical results and the actuarial program (life-table  
13 analysis) were used to estimate the exposure level ( $EC_x$ ) and the associated 95% lower  
14 confidence limit ( $LEC_x$ ) corresponding to an extra risk of 1% ( $x = 0.01$ ). As discussed in  
15 Section 4.1.1.2, a 1% extra risk level is a more reasonable response level for defining the POD  
16 for these epidemiologic data than 10%.

---

<sup>34</sup>Equations for this weighted linear regression approach are presented in [Rothman \(1986\)](#) and summarized in Appendix F.

<sup>35</sup>Mean exposures for females with a 20-year lag for the categorical exposure quartiles in Table 8 of [Steenland et al. \(2004\)](#) were 276; 1,453; 5,869; and 26,391 ppm × days. Median values were 250; 1,340; 5,300; and 26,676 ppm × days. These values are for the risk sets but should provide a good approximation to the full cohort values.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 **Table 4-8. Exposure-response modeling results for breast cancer mortality**  
 2 **in females in the NIOSH cohort for models not presented by [Steenland et al.](#)**  
 3 **[\(2004\)](#)**  
 4

Model <sup>a</sup>	<i>p</i> value	Coefficient (SE) (per ppm × day)
2-piece log-linear spline with maximum likelihood (knot at 700 ppm × days)	0.067	low-exposure spline segment: B1 = 0.000688 (0.000417)
Alternative 2-piece log-linear spline (knot at 13,000 ppm × days)	0.074	low-exposure spline segment: B1 = 0.0000607 (0.0000309)
Linear regression of categorical results, excluding the highest exposure quartile	0.09	0.000201 (0.000120)

5  
 6 <sup>a</sup>All with cumulative exposure in ppm × days as the exposure variable and with a 20-yr lag; based on 103 breast cancer deaths.

7  
 8 <sup>b</sup>*p*-values from likelihood ratio test, except for linear regression of categorical results, where Wald *p*-values are reported.

9  
 10 Source: Additional analyses performed by Dr. Steenland (see Section D.2 of Appendix D), except for the linear regression of the categorical results, which was performed by EPA.

11  
 12  
 13  
 14 Because EtO is DNA-reactive and has direct mutagenic activity (see Section 3.3.3),  
 15 which is one of the cases cited by EPA's *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA,](#)  
 16 [2005a](#)) for the use of linear low-dose extrapolation, a linear low-exposure extrapolation was  
 17 performed. The EC<sub>01</sub>, LEC<sub>01</sub>, and inhalation unit risk estimate calculated for breast cancer  
 18 mortality from the linear regression model of the categorical results are presented in Table 4-9.  
 19 The resulting unit risk estimate for breast cancer mortality based on the linear regression of the  
 20 categorical results using cumulative exposure with a 20-year lag is 0.513 per ppm. The unit risk  
 21 estimate from the alternative model, the two-piece log-linear spline model with the knot at  
 22 13,000 ppm × days, is 0.172 per ppm, about 34% of the estimate from the preferred model. EC<sub>01</sub>  
 23 and LEC<sub>01</sub> estimates from the other models considered are presented for comparison only, to  
 24 illustrate the differences in model behavior at the low end of the exposure-response range. Unit  
 25 risk estimates are not presented for these other models because, as discussed above, these models  
 26 were deemed unsuitable for the derivation of risks from (low) environmental exposure levels.  
 27 As one can see, the standard Cox regression cumulative exposure model, with its extreme  
 28 sublinearity in the lower exposure region, yields a substantially higher EC<sub>01</sub> estimate  
 29 (0.530 ppm) than the EC<sub>01</sub> estimate of 0.0387 ppm from the linear regression of the categorical  
 30 results, while the log cumulative exposure Cox regression model, with its extreme supralinearity  
 31 in the lower exposure region, yields a substantially lower EC<sub>01</sub> estimates (0.00112 ppm). The  
 32 estimates from the two-piece log-linear spline models flank the result from the linear regression  
 33 of the categorical results more closely. The steep low-exposure segment of the two-piece

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 log-linear spline model with the maximum likelihood (knot at 700 ppm × days) yields an EC<sub>01</sub>  
 2 estimate of 0.00941 ppm, whereas the shallower low-exposure slope from the two-piece  
 3 log-linear spline model with the local maximum likelihood suggesting a knot at 13,000 ppm ×  
 4 days yields an EC<sub>01</sub> estimate of 0.107 ppm. Converting the units, the unit risk estimate of 0.513  
 5 per ppm for breast cancer mortality from the linear regression model of the categorical results  
 6 corresponds to a unit risk estimate of 2.80 × 10<sup>-4</sup> per µg/m<sup>3</sup>.

7  
 8 **Table 4-9. EC<sub>01</sub>, LEC<sub>01</sub>, and unit risk estimates for breast cancer mortality**  
 9 **in females<sup>a</sup>**

Model	EC <sub>01</sub> (ppm)	LEC <sub>01</sub> (ppm)	Unit risk (per ppm)
Log cumulative exposure, 20-yr lag <sup>b</sup>	0.00112	0.000219	-- <sup>c</sup>
Cumulative exposure, 20-yr lag <sup>d</sup>	0.530	0.285	-- <sup>c</sup>
Low-exposure log-linear spline, cumulative exposure with knot at 700 ppm × days, 20-yr lag <sup>e</sup>	0.00941	0.00471	-- <sup>c</sup>
Low-exposure log-linear spline, cumulative exposure with knot at 13,000 ppm × days, 20-yr lag <sup>f</sup>	0.107	0.0580	0.172
Categorical; cumulative exposure, 20-yr lag <sup>g</sup>	0.0387	0.0195	<b>0.513</b>

11  
 12 <sup>a</sup>From lifetime continuous exposure. Unit risk = 0.01/LEC<sub>01</sub>.

13 <sup>b</sup>From Table 8 of [Steenland et al. \(2004\)](#), Cox regression model.

14 <sup>c</sup>Unit risk estimates are not presented for these models because these models were deemed unsuitable for the  
 15 derivation of risks from (low) environmental exposure levels (see text).

16 <sup>d</sup>From Dr. Steenland's reanalyses (see Table D-2d of Appendix D), Cox regression model.

17 <sup>e</sup>From low-exposure segment of two-piece log-linear spline model with largest model likelihood and a knot at  
 18 700 ppm × days; see text and Table D-2c of Appendix D. The EC<sub>01</sub> value is below the value of 0.009 ppm roughly  
 19 corresponding to the knot of 700 ppm × days [(700 ppm × days) × (10 m<sup>3</sup>/20 m<sup>3</sup>) × (240 d/365 d) × (365 d/yr)/70 yr  
 20 = 0.0013 ppm] and, thus, appropriately in the range of the low-exposure segment.

21 <sup>f</sup>From low-exposure segment of two-piece log-linear spline model with a local largest likelihood for knot at  
 22 13,000 ppm × days; see text and Table D-2f of Appendix D. The EC<sub>01</sub> value is below the value of 0.17 ppm roughly  
 23 corresponding to the knot of 13,000 ppm × days (see calculation in footnote e) and, thus, appropriately in the range  
 24 of the low-exposure segment.

25 <sup>g</sup>Regression coefficient derived from linear regression of categorical Cox regression results from Table 8 of  
 26 [Steenland et al. \(2004\)](#), as described in Section 4.1.2.2. The EC<sub>01</sub> value is appropriately below the value of 0.064  
 27 ppm roughly corresponding to the value of about 5,000 ppm × days (see footnote e for calculation) above which the  
 28 linear regression model of the categorical results does not apply (see Figure 4-3).

### 1 **4.1.2.3. Prediction of Lifetime Extra Risk of Breast Cancer Incidence**

2 As discussed in Section 4.1.1.3, risk estimates for cancer incidence are preferred to  
3 estimates for cancer mortality, especially for cancer types with good survival rates, such as breast  
4 cancer. In the case of female breast cancer in the NIOSH cohort, there is a corresponding  
5 incidence study ([Steenland et al., 2003](#)) with exposure-response results for breast cancer  
6 incidence, so one can estimate cancer incidence risks directly rather than estimate them from  
7 mortality data. The incidence study used a (sub)cohort of 7,576 (76%) of the female workers  
8 from the original cohort. Cohort eligibility for the incidence study was restricted to the female  
9 workers who had been employed at 1 of the 14 plants for at least 1 year, owing to cost  
10 considerations and the greater difficulties in locating workers with short-term employment.  
11 Interviews were sought from all the women in the incidence study cohort or their next-of-kin  
12 (18% of the cohort had died). Completed interviews were obtained for 5,139 (68%) of the  
13 7,576 women in the cohort. The investigators also attempted to acquire breast cancer incidence  
14 data for the cohort from cancer registries (available for 9 of the 11 states in which the plants were  
15 located) and death certificates; thus, results are presented for both the full cohort ( $n = 7,576$ ) and  
16 the subcohort of women with interviews ( $n = 5,139$ ). For additional details and discussion of the  
17 [Steenland et al. \(2003\)](#) study, see Section A.2.16 of Appendix A.

18 [Steenland et al. \(2003\)](#) identified 319 incident cases of breast cancer in the cohort through  
19 1998. Interview (questionnaire) data were available for 73% (233 cases). Six percent were  
20 carcinoma in situ (20 cases). [Steenland et al. \(2003\)](#) performed internal exposure-response  
21 analyses similar to those described in their 2004 paper and in Section 4.1.1.1 above. Controls for  
22 each case were selected from the cohort members without breast cancer at the age of diagnosis of  
23 the case. Cases and controls were matched on race. Of the potential confounders evaluated for  
24 those with interviews, only parity and breast cancer in a first-degree relative were important  
25 predictors of breast cancer, and only these variables were included in the final models for the  
26 subcohort analyses. In situ cases were included with invasive breast cancer cases in the analyses;  
27 however, the in situ cases represent just 6% of the total, and excluding them reportedly did not  
28 greatly affect the results.

29 From the [Steenland et al. \(2003\)](#) internal analyses (Cox regression) using the full cohort,  
30 the best-fitting model with exposure as a continuous variable was for (natural) log cumulative  
31 exposure, lagged 15 years ( $p = 0.05$ ). Duration of exposure, lagged 15 years, provided a slightly  
32 better fitting model. Models using maximum or average exposure did not fit as well. In  
33 addition, use of a threshold model did not provide a statistically significant improvement in fit.  
34 For internal analyses using the subcohort with interviews, the cumulative exposure and log  
35 cumulative exposure models, both lagged 15 years, and the log cumulative exposure model with

1 no lag all fit almost equally well, and the duration of exposure (also lagged 15 years) model fit  
 2 slightly better. Results of the Cox regression analyses for the cumulative and log cumulative  
 3 exposure models, with 15-year lags, are shown in Table 4-10, and these are the results  
 4 considered for the unit risk calculations. The models using duration of exposure are less useful  
 5 for estimating exposure-related risks, duration of exposure and cumulative exposure are  
 6 correlated, and the fits for these models are only marginally better than those with cumulative  
 7 exposure. The log cumulative exposure model with no lag was considered less biologically  
 8

9 **Table 4-10. Cox regression results for breast cancer incidence in females**  
 10 **from the NIOSH cohort, for the models presented by [Steenland et al.](#)**  
 11 **[\(2003\)](#)<sup>a,b</sup>**  
 12

Cohort	Exposure variable <sup>c</sup>	Coefficient (SE) (per ppm × day), p-value <sup>d</sup>	ORs by category <sup>e</sup> (95% CI)
Full incidence study cohort n = 7,576 319 cases	Cumulative exposure, 15-yr lag	0.0000054 (0.0000035), p = 0.12	
	Log cumulative exposure, 15-yr lag	0.037 (0.019), p = 0.05	
	Categorical cumulative exposure, 15-yr lag		1.00, 1.07 (0.72–1.59), 1.00 (0.67–1.50), 1.24 (0.85–1.90), 1.17 (0.78–1.78), 1.74 (1.16–2.65)
Subcohort with interviews n = 5,139 233 cases	Cumulative exposure, 15-yr lag	0.0000095 (0.0000041), p = 0.02	
	Log cumulative exposure, 15-yr lag	0.050 (0.023), p = 0.03	
	Categorical cumulative exposure, 15-yr lag	-- <sup>f</sup>	1.00, 1.06 (0.66–1.71), 0.99 (0.61–1.60), 1.24 (0.76–2.00), 1.42 (0.88–2.29), 1.87 (1.12–3.10)

13 <sup>a</sup>Invasive breast cancer (ICD-9 174) and carcinoma in situ (ICD-9 233.0).

14 <sup>b</sup>Cases and controls matched on age and race (white/nonwhite). Full cohort models include cumulative exposure  
 15 and categorical variable for yr of birth (quartiles). Subcohort models include cumulative exposure, categorical  
 16 variables for yr of birth (quartiles), breast cancer in first-degree relative, and parity.

17 <sup>c</sup>Cumulative exposure is in ppm × days.

18 <sup>d</sup>p-values for exposure variable from Wald test, as reported by [Steenland et al. \(2003\)](#).

19 <sup>e</sup>Exposure categories are 0, >0–647, 647–2,026, 2,026–4,919, 4,919–14,620, >14,620 ppm × days.

20 <sup>f</sup>p-value for the addition of the categorical exposure variables = 0.11 (email dated 5 March 2010 from Kyle  
 21 Steenland, Emory University, to Jennifer Jinot, EPA).

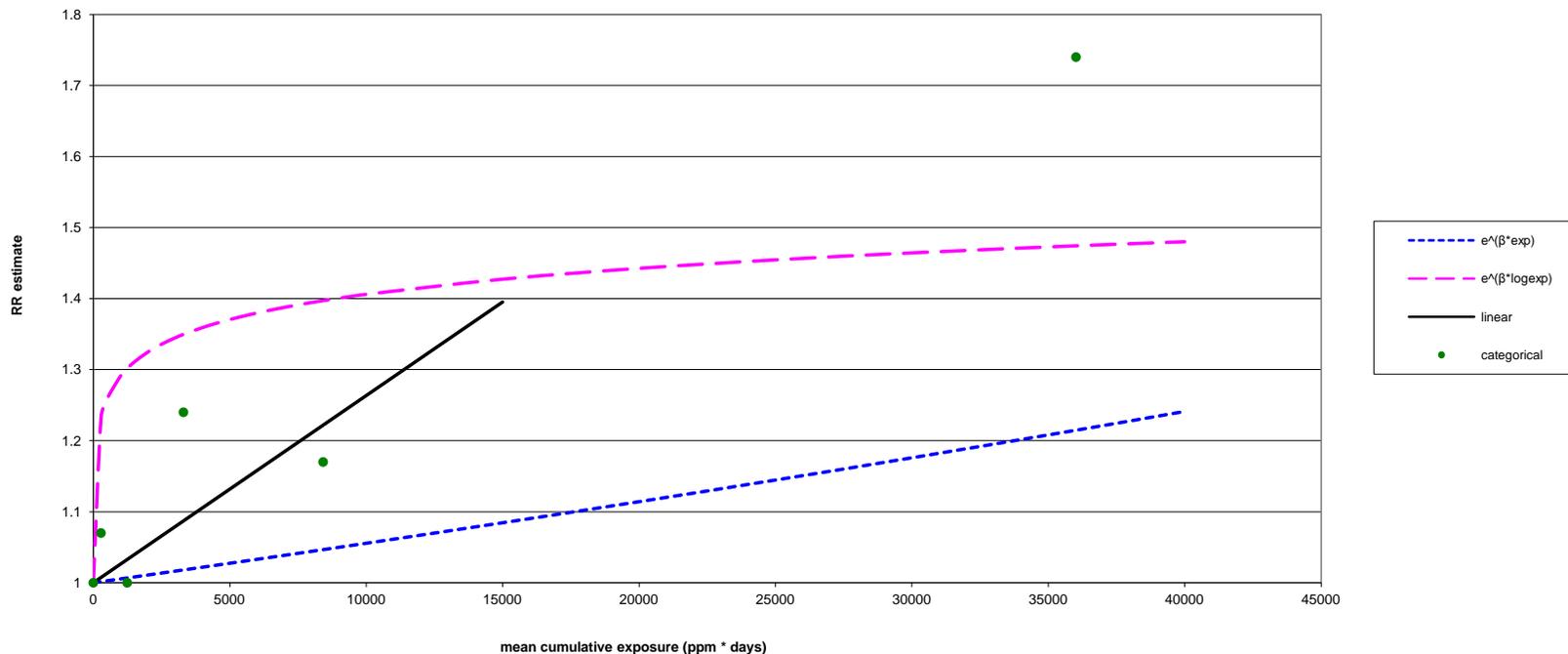
22 Source: Tables 4 and 5 of [Steenland et al. \(2003\)](#).  
 23  
 24  
 25  
 26

1 realistic than the corresponding model with a 15-year lag because some lag period would be  
2 expected for the development of breast cancer. Furthermore, although initial risk estimates  
3 based on the full cohort results are calculated for comparison, the preferred estimates are those  
4 based on the subcohort with interviews because the subcohort should have more complete case  
5 ascertainment and has additional information available on potential breast cancer confounders.

6 For the actuarial program (life-table analysis), U.S. age-specific all-cause mortality rates  
7 for 2004 for females of all race groups combined ([Arias, 2007](#)) were used to specify the all-cause  
8 background mortality rates. Because breast cancer incidence rates (more specifically, the  
9 differential rates obtained by subtracting the mortality rates from the incidence rates) are not  
10 negligible compared to all-cause mortality rates, the all-cause mortality rates in the life-table  
11 analysis were adjusted to reflect women dying *or* being diagnosed with breast cancer in a given  
12 age interval. All-cause mortality rates and breast cancer incidence rates were summed, and  
13 breast cancer mortality rates were subtracted so that those dying of breast cancer were not  
14 counted twice (i.e., as deaths and as incident cases of breast cancer). The National Center for  
15 Health Statistics 2002–2006 mortality rates for invasive breast cancer in females were obtained  
16 from a SEER report ([Horner et al., 2009](#)). The SEER report also provided SEER-17 incidence  
17 rates for invasive and in situ breast cancer. The Cox regression results reported by [Steenland et  
18 al. \(2003\)](#) are for invasive and in situ breast cancers combined. It is consistent with EPA’s  
19 *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)) to combine these two tumor  
20 types because the in situ tumors can progress to invasive tumors. Thus, the primary risk  
21 calculations in this assessment use the sum of invasive and in situ breast cancer incidence rates  
22 for the cause-specific background rates. Comparison calculations were performed using just the  
23 invasive breast cancer incidence rates for the cause-specific rates; this issue is further discussed  
24 in Section 4.1.3 on sources of uncertainty. The risks were computed up to age 85 for continuous  
25 exposures to EtO, conversions were made between occupational EtO exposures and continuous  
26 environmental exposures, and 95% UCLs were calculated for the relative rates, as described in  
27 Section 4.1.1.2 above.

28 For breast cancer incidence in both the full cohort (see Figure 4-4) and the subcohort with  
29 interviews (see Figure 4-5), the low-exposure categorical results suggest a more linear  
30 low-exposure exposure-response relationship than that obtained with either the continuous  
31 variable log cumulative exposure (supralinear) or cumulative exposure (sublinear) Cox  
32 regression models. Thus, as with the lymphohematopoietic cancer and the breast cancer  
33 mortality results above, EPA proposed in the 2006 Draft Assessment ([U.S. EPA, 2006a](#)), which  
34 relied on the original published results of [Steenland et al. \(2003\)](#), that the best way to reflect the  
35 data in the lower exposure region, which is the region of interest for low-exposure extrapolation,

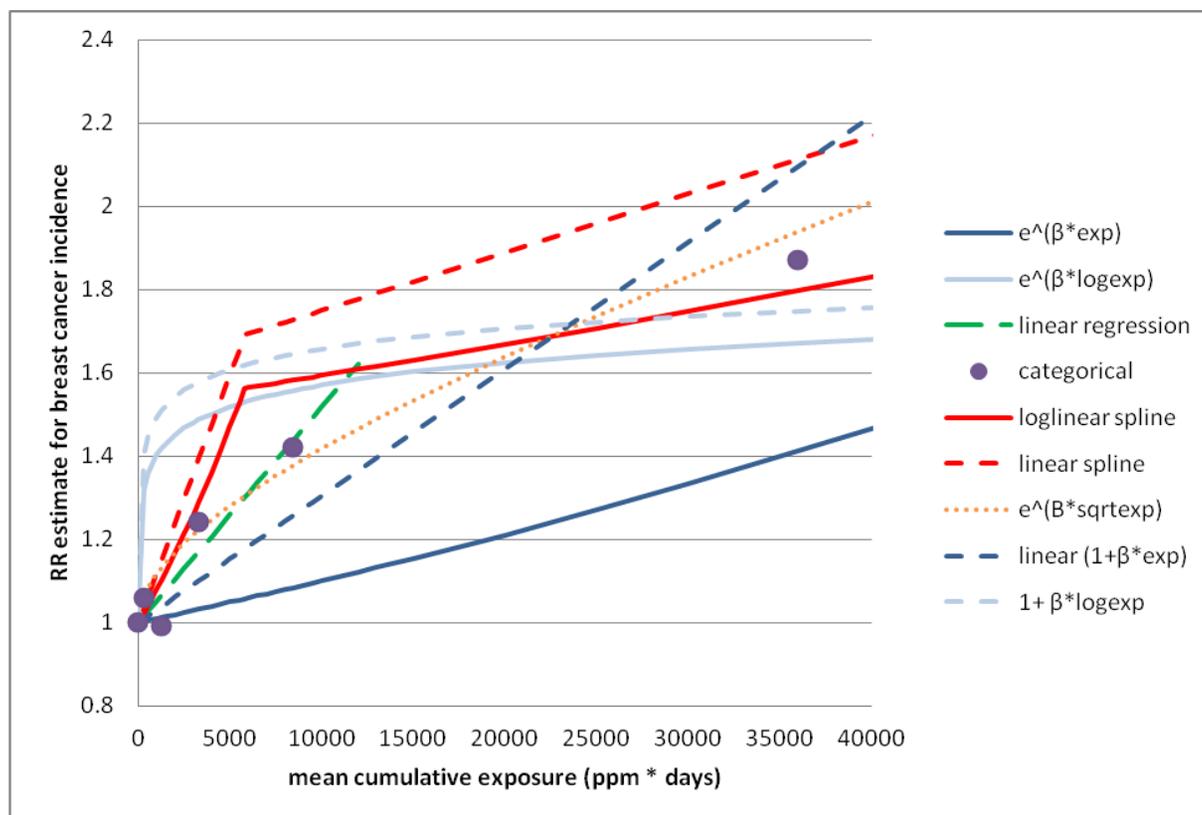
*This document is a draft for review purposes only and does not constitute Agency policy.*



**Figure 4-4. RR estimate for breast cancer incidence in full cohort vs. mean exposure (with 15-year lag, unadjusted for continuous exposure).**

$e^{(\beta \cdot \text{exp})}$ : Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$ ;  $e^{(\beta \cdot \log \text{exp})}$ : Cox regression results for  $RR = e^{(\beta \times \ln(\text{exposure}))}$ ; categorical: Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$  with categorical exposures; linear: weighted linear regression of categorical results, excluding highest exposure group (see text). (Note that, with the exception of the categorical results and the linear regression of the categorical results, the various models have different implicitly estimated baseline risks; thus, they are not strictly comparable to each other in terms of RR values, i.e., along the y-axis. They are, however, comparable in terms of general shape.)

Source: [Steenland et al. \(2003\)](#) (except for linear regression of the categorical results, which was done by EPA).



**Figure 4-5. RR estimate for breast cancer incidence in subcohort with interviews vs. mean exposure (with 15-year lag, unadjusted for continuous exposure).**

$e^{(\beta \times \text{exp})}$ : Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$ ;  $e^{(\beta \times \ln(\text{exposure}))}$ : Cox regression results for  $RR = e^{(\beta \times \ln(\text{exposure}))}$ ; categorical: Cox regression results for  $RR = e^{(\beta \times \text{exposure})}$  with categorical exposures;  $e^{(\beta \times \sqrt{\text{exposure}})}$ : Cox regression results for  $RR = e^{(\beta \times \sqrt{\text{exposure}})}$ ; linear regression: weighted linear regression of categorical results, excluding highest exposure group (see text); log-linear and linear spline: 2-piece spline models, both with knots at 5,800 ppm × days (see text); linear:  $RR = 1 + \beta \times \text{exposure}$ , with exposure as a continuous variable;  $1 + \beta \times \ln(\text{exposure})$ :  $RR = 1 + \beta \times \ln(\text{exposure})$ , with exposure as a continuous variable. (Note that, with the exception of the categorical results and the linear regression of the categorical results, the various models have different implicitly estimated baseline risks; thus, they are not strictly comparable to each other in terms of RR values, i.e., along the y-axis. They are, however, comparable in terms of general shape.)

Sources: [Steenland et al. \(2003\)](#) except for Steenland 2-piece spline models (see Appendix D) and linear regression of the categorical results, which was done by EPA.

1 was to do a weighted linear regression of the results from the model with categorical cumulative  
2 exposure (with a 15-year lag). In addition, the highest exposure group was not included in the  
3 regression to provide a better fit to the lower-exposure data (The RR estimates for the highest  
4 exposure quintiles suggest somewhat supralinear exposure-response relationships for both the  
5 full cohort and the subcohort with interviews, and supralinearity is evidenced in the subcohort  
6 with interviews by the strong influence of the top 5% of cumulative exposures on dampening the  
7 slope of the [cumulative exposure] Cox regression model [see Section D.1 and Figure D-1d of  
8 Appendix D]. Moreover, there is more uncertainty in using the mean cumulative exposure to  
9 represent the range of exposures in a highest exposure categorical group because such groups  
10 contain a wider range of exposures; for example, for the subcohort with interviews, the highest  
11 exposure quintile contains exposures ranging from about 14,500 ppm × days to over  
12 250,000 ppm × days). Linear modeling of categorical (i.e., grouped) epidemiologic data and  
13 elimination of the highest exposure group(s) under certain circumstances to obtain a better fit of  
14 low-exposure data are both standard techniques used in EPA dose-response assessments ([U.S.  
15 EPA, 2012, 2005a](#)). However, as discussed in Section 4.1.1.2 for the lymphohematopoietic  
16 cancer data, the SAB panel that reviewed the draft assessment recommended that EPA not rely  
17 on the published grouped data but, rather, do additional analyses using the individual data ([SAB,  
18 2007](#)).

19 Consequently, it was determined that using the individual data, a better way to address  
20 the apparent supralinearity of the data (while avoiding the extreme low-exposure curvature  
21 obtained with the log cumulative exposure Cox regression model) might be to use a two-piece  
22 spline model, and Dr. Steenland was commissioned to do the spline analyses. His findings are  
23 reported in Appendix D (see Section D.1), and the results for the breast cancer incidence  
24 analyses are summarized below. Note that, for the two-piece spline analyses, only the data from  
25 the subcohort with interviews and for the invasive and in situ breast cancers combined were  
26 analyzed, because this was the preferred data set, as discussed above. (Dr. Steenland also  
27 employed a cubic spline model as a semiparametric approach to visualize the underlying  
28 exposure-response relationship; however, this approach produces an overly complicated function  
29 for an empirical model, as opposed to a biologically based model, and was not used for risk  
30 assessment purposes. In addition, Dr. Steenland investigated the use of a Cox regression model  
31 with a square-root transformation of cumulative exposure; however, this approach, though less  
32 extreme than using the log transformation of cumulative exposure, also yields a notably  
33 supralinear model [see Figure 4-5], which can result in unstable low-exposure risk estimates.  
34 The model results for both the cubic spline and square-root transformation models are included  
35 in Appendix D, Section D.1, but are not considered further here. EPA chose to pursue the

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 development of two-piece spline models to attempt to avoid the problem of unstable risk  
2 estimates from supralinear curvature in the low-exposure region because these models provide a  
3 more general and systematic approach to modeling supralinear exposure-response data, as  
4 opposed to using random, arbitrary power-transformations of the exposure variable.)

5 For the two-piece log-linear spline modeling approach, as described in Section 4.1.1.2  
6 and discussed more fully in Appendix D, the Cox regression model was the underlying basis for  
7 the splines which were fit to the breast cancer incidence exposure-response data (cumulative  
8 exposure is used here, with a 15-year lag), and, thus, log RR is a function of two lines which join  
9 at a single point of inflection, called a “knot.” The shape of the two-piece spline model, in  
10 particular the slope in the low-exposure region, depends on the location of the knot. For breast  
11 cancer incidence, the knot was selected by evaluating different knots from 100 to  
12 15,000 ppm × days in increments of 100 ppm × days and then choosing the one that resulted in  
13 the best (largest) model likelihood. The model likelihood did not actually change much across  
14 the different trial knots (see Figure D-1a of Appendix D), but it did change slightly, and a knot of  
15 5,800 ppm × days was chosen for the breast cancer incidence data based on the largest  
16 likelihood. The two-piece log-linear spline model with this knot provided a statistically  
17 significant fit to the data ( $p = 0.01$  for the addition of the exposure terms; see Table D-1g in  
18 Appendix D), as well as a good visual fit (see Figure 4-5).

19 A two-piece linear spline model was also fitted, using the just-published approach of  
20 [Langholz and Richardson \(2010\)](#). This model is similar to the log-linear spline model discussed  
21 above; however, for the linear spline model, the underlying basis for the splines is a linear model  
22 (i.e.,  $RR = 1 + \beta \times z$ , where  $z$  represents the covariate data, including exposure, and  $\beta$  are the  
23 parameters being estimated). The knot was selected as for the log-linear spline model, and the  
24 same knot of 5,800 ppm × days yielded the largest likelihood (see Figure D-1h of Appendix D)  
25 and was also chosen for the two-piece linear spline model. The two-piece linear spline model  
26 with this knot provided a statistically significant fit to the data ( $p = 0.002$  for the addition of the  
27 exposure terms), as well as a good visual fit (see Figure 4-5). Because this model provided a  
28 better fit than the log-linear spline model, i.e., it had a lower AIC, the two-piece linear spline  
29 model was selected as the preferred model for the unit risk estimates for breast cancer incidence.  
30 See Table 4-11 and Section D.1 of Appendix D for parameter estimates and fit statistics for the  
31 two spline models.

32 Linear RR models with cumulative exposure and log cumulative exposure as continuous  
33 variables were also investigated using the approach of [Langholz and Richardson \(2010\)](#), and  
34 these models fit better than the corresponding log RR models (see Table 4-11 and Section D.1 of  
35 Appendix D) although not as well as the two-piece linear spline model, which had the lowest

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 AIC. Risk estimates based on the linear model with cumulative exposure are developed for  
 2 comparison, but the linear model with log cumulative exposure is too steep in the low-exposure  
 3 region (see Figure 4-5) and is not considered further. For more details of the breast cancer  
 4 incidence exposure-response modeling, see Section D.1 of Appendix D.

5  
 6 **Table 4-11. Exposure-response modeling results for breast cancer incidence**  
 7 **in females from the NIOSH cohort for models not presented by [Steenland et](#)**  
 8 **[al. \(2003\)](#)**  
 9

Model <sup>a</sup>	<i>p</i> -value <sup>b</sup>	Coefficient (SE) (per ppm × day)
Full incidence study cohort <sup>c</sup>		
Linear regression of categorical results, excluding the highest exposure quintile	0.33	0.0000264 (0.0000269)
Subcohort with interviews <sup>d</sup>		
2-piece log-linear spline (knot at 5,800 ppm × days)	0.01	low-exposure spline segment: B1 = 0.0000770 (0.0000317)
2-piece linear spline (knot at 5,800 ppm × days)	0.002	low-exposure spline segment: B1 = 0.000119 (0.0000677)
linear	0.003	0.0000304 (0.0000175)
linear with log cumulative exposure	0.01	0.0713 (0.0392)
Linear regression of categorical results, excluding the highest exposure quintile	0.16	0.0000517 (0.0000369)

10  
 11 <sup>a</sup>All with cumulative exposure in ppm × days as the exposure variable and with a 15-yr lag.

12 <sup>b</sup>*p*-value for addition of exposure variables from likelihood ratio test, except for the linear regressions of categorical  
 13 results, where Wald *p*-values are reported.

14 <sup>c</sup>319 breast cancer cases.

15 <sup>d</sup>233 breast cancer cases.

16  
 17 Source: Additional analyses performed by Dr. Steenland (see Section D.2 of Appendix D), except for the linear  
 18 regressions of categorical results, which were performed by EPA using the equations of [Rothman \(1986\)](#) presented  
 19 in Appendix F.

20  
 21  
 22 Risk estimates based on the original linear regression analyses of the categorical results  
 23 are also presented for comparison. For the approach of using a weighted linear regression of the  
 24 results from the Cox regression model with categorical cumulative exposure (and a 15-year lag),  
 25 excluding the highest exposure group, the weights used for the ORs were the inverses of the  
 26 variances, which were calculated from the confidence intervals.<sup>36</sup> Mean and median exposures

---

<sup>36</sup>Equations for this weighted linear regression approach are presented in [Rothman \(1986\)](#) and summarized in  
 Appendix F.

1 for the cumulative exposure groups for the full cohort were kindly provided by Dr. Steenland  
2 (email dated April 21, 2004, from Kyle Steenland, Emory University, to Jennifer Jinot, EPA).<sup>37</sup>  
3 The mean values were used for the weighted regression analysis because the (arithmetic) mean  
4 exposures best represent the model's linear relationship between exposure and cancer response.  
5 Differences between means and medians were not large for the females, especially for the lower  
6 four quintiles. If the median values had been used, a slightly larger regression coefficient would  
7 have been obtained, resulting in slightly larger risk estimates. Although the exposure values are  
8 for risk sets from the full cohort, they should be reasonably close to the values for the subcohort  
9 with interviews. See Table 4-11 for the results from the weighted linear regressions of the  
10 categorical results and Figures 4-4 and 4-5 for a depiction of the resulting linear regression  
11 models.

12 As the subcohort with interviews from the NIOSH incidence study cohort provides the  
13 preferred data set for the derivation of unit risk estimates for breast cancer, a summary of all the  
14 models considered for modeling the breast cancer exposure-response data from the subcohort  
15 and the judgments made about model selection is provided in Table 4-12. See Figure 4-5 for  
16 visual representations of the models. See Tables 4-10 and 4-11 and Section D.1 of Appendix D  
17 for parameter estimates, *p*-values, and other fit statistics. Three of the models presented in  
18 Table 4-12 had a good overall statistical fit, a good visual fit, and a credible low-exposure slope  
19 (the linear and log-linear two-piece spline models and the [continuous exposure] linear RR  
20 model). To better compare these models, they are plotted again in Figure 4-6 this time against  
21 the categorical data in deciles. Earlier categorical results in this assessment were based on the  
22 (log-linear) Cox regression model; however, the deciles in Figure 4-6 are based on a linear RR  
23 categorical model—this model had a lower AIC than the log-linear decile model (1963.94 vs.  
24 1966.91), and it provides a statistically significant fit to the data (*p* = 0.004), so the deciles  
25 should provide a good representation of the data for the purposes of comparing the models (the  
26 decile results from the log-linear and linear RR categorical models and the mean cumulative  
27 exposure estimates for the deciles are presented in Section D.1 of Appendix D). As can be seen  
28 in Figure 4-6, the two-piece linear spline model, in addition to having the lowest AIC (see  
29 Table 4-12), appears to have a better fit to the lower-exposure data, which are of the greatest  
30 interest in estimating low-exposure risk. It also appears from Figure 4-6 that the linear model  
31 has a poorer fit (too shallow) to the lower-exposure data than either of the two-piece spline  
32 models. This is consistent with the analysis presented in Section D.1 of Appendix D  
33

---

<sup>37</sup>Mean exposures for females with a 15-year lag for the exposure categories in Table 3 of [Steenland et al. \(2003\)](#) were 280; 1,241; 3,304; 8,423; and 36,022 ppm × days. Median values were 253; 1,193; 3,241; 7,741; and 26,597 ppm × days. These values are for the risk sets but should provide a good approximation to the full cohort values.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1  
2  
3  
4

**Table 4-12. Models considered for modeling the exposure-response data for breast cancer incidence in females in the subcohort with interviews from the NIOSH incidence study cohort for the derivation of unit risk estimates**

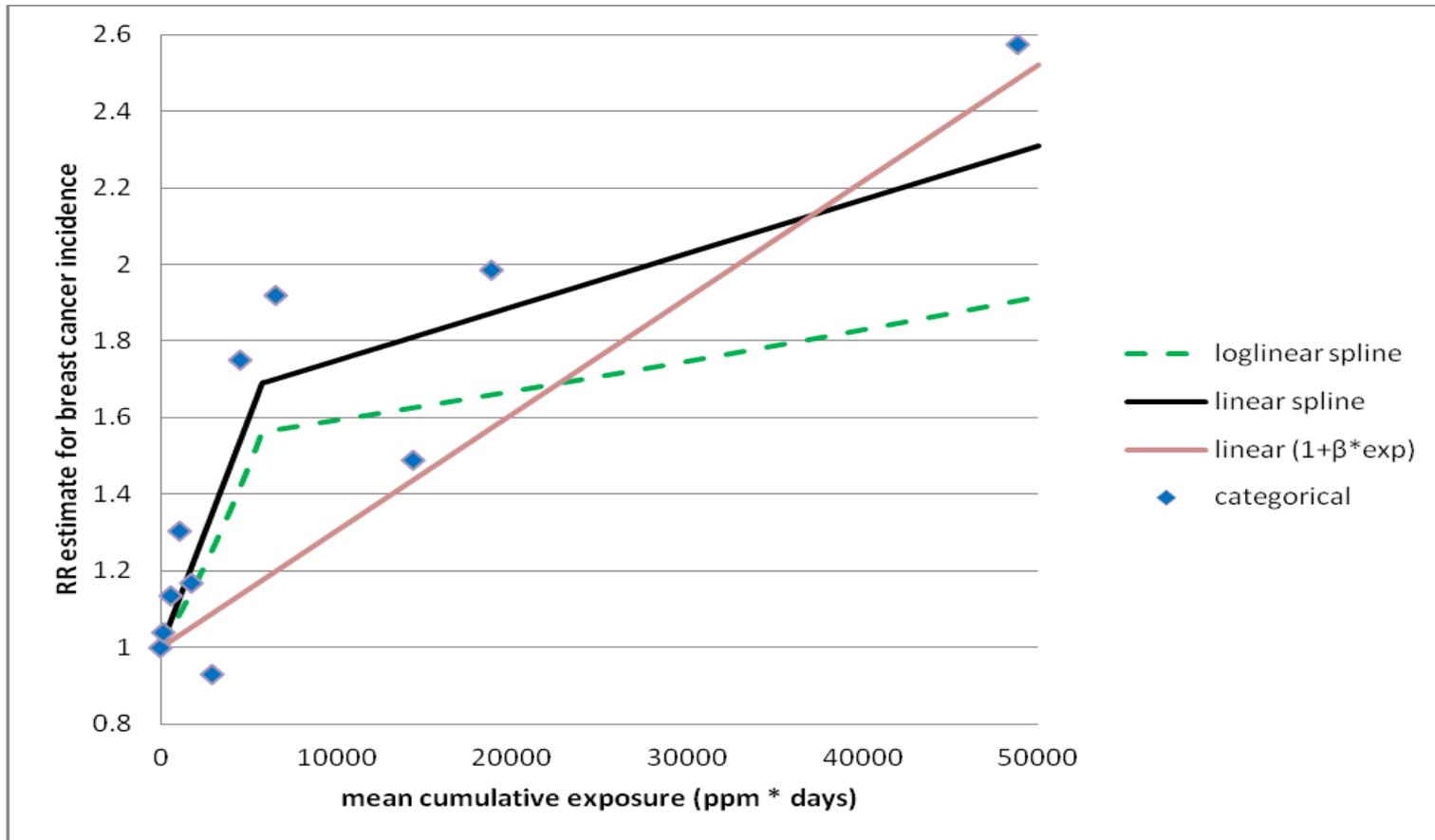
Model <sup>a</sup>	AIC <sup>b</sup>	Comments
Cox regression (log-linear) model	1956.675	Good overall statistical fit but poor visual fit (too shallow in the low-exposure region).
Cox regression model with log cumulative exposure	1956.176	Good overall statistical fit but too steep in the low-exposure region.
Cox regression model with square-root transformation of exposure	1953.028	Good overall statistical fit but still notably supralinear (steep) in the low-exposure region, though less so than with the log transformation; also preference was given to the two-piece spline models as providing a more systematic approach to modeling supralinear data.
Linear regression of categorical results, excluding the highest exposure quintile	-- <sup>c</sup>	Not statistically significant, though that is unsurprising since the approach, which is based on categorical data, has low statistical power; preference given to models that treated exposure as a continuous variable, as recommended by the SAB, and that also provided reasonable representations of the low-exposure region.
2-piece log-linear spline model (knot at 5,800 ppm × days)	1954.485	Good overall statistical fit and good visual fit; preference given to the 2-piece linear spline model because it had a better statistical fit (lower AIC) and better apparent fit to the lower-exposure data.
linear model ( $RR = 1 + \beta \times \text{exposure}$ )	1952.260	Good overall statistical fit and good visual fit; preference given to the 2-piece linear spline model because it had a better statistical fit (lower AIC) and better apparent fit to the lower-exposure data.
linear model with log cumulative exposure	1954.267	Good overall statistical fit but too steep in the low-exposure region.
2-piece linear spline model (knot at 5,800 ppm × days)	1950.935	<b>SELECTED.</b> Good overall statistical fit and good visual fit; lower AIC than 2-piece log-linear spline and linear model and better apparent fit to the lower-exposure data.

5  
6  
7  
8  
9

<sup>a</sup>All with cumulative exposure as the exposure variable, except where noted, and with a 15-yr lag.

<sup>b</sup>AIC = 2p-2LL, where p = # of parameters and LL = ln(likelihood), assuming two exposure parameters for the two-piece spline models.

<sup>c</sup>Not calculated.



**Figure 4-6. RR estimate for breast cancer incidence in subcohort with interviews vs. mean exposure (with 15-year lag, unadjusted for continuous exposure); select models compared to deciles.**

Categorical: linear model ( $RR = 1 + \beta \times \text{exposure}$ ) with categorical exposures; log-linear and linear spline: 2-piece spline models, both with knots at 5,800 ppm × days (see text); linear:  $RR = 1 + \beta \times \text{exposure}$ , with exposure as a continuous variable. (Note that, with the exception of the categorical results and the linear regression of the categorical results, the various models have different implicitly estimated baseline risks; thus, they are not strictly comparable to each other in terms of RR values, i.e., along the y-axis. They are, however, comparable in terms of general shape.)

Source: Steenland analyses in Appendix D.

1 showing the strong influence of the upper tail of cumulative exposures on the results of the  
 2 cumulative exposure Cox regression model. The responses in the upper tail of exposures are  
 3 relatively dampened, such that when the highest 5% of exposures are excluded, the slope of the  
 4 Cox regression model is substantially increased (e.g., at 10,000 ppm × days, the RR estimate  
 5 increases from about 1.1 to almost 1.5; see Figure D-1d in Appendix D). This strong influence  
 6 of the upper tail of exposures would similarly attenuate the slope of the linear model. The  
 7 two-piece spline models, on the other hand, are more flexible, and the influence of the upper tail  
 8 of exposures would be primarily on the upper spline segment; thus, the two-piece models are  
 9 able to provide a better fit to the lower-exposure data.

10 The exposure level ( $EC_x$ ) and the associated 95% lower confidence limit ( $LEC_x$ )  
 11 corresponding to an extra risk of 1% ( $x = 0.01$ ) for breast cancer incidence in females (based on  
 12 invasive + in situ tumors in the subcohort with interviews) for the models discussed above  
 13 (except for the square-root model and the linear model with log cumulative exposure, which  
 14 were not considered further) were estimated using the actuarial program (life-table analysis). As  
 15 noted in Section 4.1.1.2, a 1% extra risk level is a more reasonable response level for defining  
 16 the POD for these epidemiologic data than a 10% level. The results are presented in Table 4-13.

17  
 18 **Table 4-13.  $EC_{01}$ ,  $LEC_{01}$ , and unit risk estimates for breast cancer incidence**  
 19 **in females—invasive and in situ<sup>a</sup>**

20

Model	With interviews			Full cohort		
	$EC_{01}$ (ppm)	$LEC_{01}$ (ppm)	Unit risk (per ppm)	$EC_{01}$ (ppm)	$LEC_{01}$ (ppm)	Unit risk (per ppm)
Cox regression, cumulative exposure, 15-yr lag <sup>b</sup>	0.135	0.0788	-- <sup>c</sup>	0.237	0.115	-- <sup>c</sup>
Cox regression, log cumulative exposure, 15-yr lag <sup>b</sup>	0.0000765	0.0000422	-- <sup>c</sup>	0.000124	0.0000529	-- <sup>c</sup>
Linear regression of categorical results, excluding highest exposure quintile; cumulative exposure, 15-yr lag <sup>b,d</sup>	0.0257	0.0118	0.847	0.0503	0.0188	0.532
Low-exposure log- linear spline, cumulative exposure, 15-yr lag <sup>e</sup>	0.0166	0.00991	1.01 <sup>f</sup>	-- <sup>g</sup>		

21

**Table 4-13. EC<sub>01</sub>, LEC<sub>01</sub>, and unit risk estimates for breast cancer incidence in females—invasive and in situ<sup>a</sup> (continued)**

Linear model with continuous cumulative exposure, 15-yr lag <sup>h</sup>	0.0437	0.0224	0.446 <sup>i</sup>	-- <sup>g</sup>
Low-exposure linear spline, cumulative exposure, 15-yr lag <sup>e</sup>	0.0112	0.00576	<b>1.74</b> <sup>f,j</sup>	-- <sup>g</sup>

<sup>a</sup>All-cause mortality adjusted (to dying of something other than breast cancer or developing breast cancer). Unit risk = 0.01/LEC<sub>01</sub>. Note that the EC<sub>01</sub> and LEC<sub>01</sub> results presented here will not exactly match those presented in Appendix D because, although the regression coefficients reported by Dr. Steenland in Appendix D were used, the life-table analyses using 2004 all-cause mortality and 2002–2006 cause-specific mortality and incidence rates were redone to be more up-to-date; the results presented in Appendix D were based on life-table analyses using 2000 all-cause mortality rates and comparable cause-specific rates.

<sup>b</sup>From Tables 4 and 5 of [Steenland et al. \(2003\)](#), Cox regression models.

<sup>c</sup>Unit risk estimates are not presented for these models because these models were deemed unsuitable for the derivation of risks from (low) environmental exposure levels (see text).

<sup>d</sup>Regression coefficient derived from linear regression of categorical results, as described in Section 4.1.2.3.

<sup>e</sup>From low-exposure segment of two-piece spline analysis; see text and Table D-1c of Appendix D for log-linear model or Table D-1i for linear model; two-piece spline analyses not performed for the full cohort. The EC<sub>01</sub> value is below the value of 0.075 ppm roughly corresponding to the knot of 5,800 ppm × days [(5,800 ppm × days) × (10 m<sup>3</sup>/20 m<sup>3</sup>) × (240 d/365 d) × (365 d/yr)/70 yr = 0.075 ppm] and, thus, appropriately in the range of the low-exposure segment.

<sup>f</sup>For unit risk estimates above 1, convert to risk per ppb (e.g., 1.74 per ppm = 1.74 × 10<sup>-3</sup> per ppb).

<sup>g</sup>Not estimated; two-piece spline and linear RR models not developed for the full cohort.

<sup>h</sup>From linear analyses in Section D.1.b.2 and Table D-1i of Appendix D.

<sup>i</sup>Confidence intervals used in deriving the LEC<sub>01</sub>s were estimated employing the Wald approach. Confidence intervals for linear RR models, however, in contrast to those for the log-linear RR models, may not be symmetrical. EPA also evaluated application of a profile likelihood approach for the linear RR models ([Langholz and Richardson, 2010](#)), which allows for asymmetric CIs, for comparison with the Wald approach. The MLE for the regression coefficient of the linear model is 0.0000304 per ppm × day. Using the profile likelihood method, the (95% one-sided) upper bound on the regression coefficient is 0.0000745 per ppm × day and the (95% one-sided) lower bound on the regression coefficient is 0.00000975 per ppm × day. Based on these profile likelihood estimates, the LEC<sub>01</sub> estimate is 0.0174 ppm, the UEC<sub>01</sub> estimate is 0.133 ppm, and the unit risk estimate for breast cancer incidence from the linear model would have been 0.575 per ppm, slightly higher (29%) than the value of 0.446 per ppm obtained using the Wald approach.

<sup>j</sup>Confidence intervals used in deriving the LEC<sub>01</sub>s were estimated employing the Wald approach. Confidence intervals for linear RR models, however, in contrast to those for the log-linear RR models, may not be symmetrical. EPA also evaluated application of a profile likelihood approach for the linear RR models ([Langholz and Richardson, 2010](#)), which allows for asymmetric CIs, for comparison with the Wald approach. The MLE for the regression coefficient of the first spline segment is 0.000119 per ppm × day. Using the profile likelihood method, the (95% one-sided) upper bound on the regression coefficient is 0.000309 per ppm × day and the (95% one-sided) lower bound on the regression coefficient is 0.000032 per ppm × day. Based on these profile likelihood estimates, the LEC<sub>01</sub> estimate is 0.00430 ppm, the UEC<sub>01</sub> estimate is 0.0415 ppm, and the unit risk estimate for breast cancer incidence from the low-exposure linear spline would have been 2.33 per ppm, slightly higher (34%) than the value of 1.74 per ppm obtained using the Wald approach.

1           Because EtO is DNA-reactive and has direct mutagenic activity (see Section 3.3.3),  
2 which is one of the cases cited by EPA's *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA,](#)  
3 [2005a](#)) for the use of linear low-dose extrapolation, a linear low-exposure extrapolation was  
4 performed. The inhalation unit risk estimates for the different breast cancer incidence models  
5 considered suitable for low-exposure extrapolation are presented in Table 4-13. As discussed  
6 above, the unit risk estimate based on the two-piece linear spline model using cumulative  
7 exposure with a 15-year lag (i.e., 1.74 per ppm, or  $1.74 \times 10^{-3}$  per ppb) is the preferred estimate.  
8 The two-piece log-linear spline model resulted in a unit risk estimate of 1.01 per ppm, while the  
9 linear regression of categorical results yielded a unit risk estimate of 0.847 per ppm and the  
10 continuous exposure linear model produced a unit risk estimate of 0.446 per ppm; these alternate  
11 estimates are about 60%, 50%, and 25%, respectively, of the estimate based on the preferred  
12 two-piece linear spline model.  $EC_{01}$  and  $LEC_{01}$  estimates from the other models examined are  
13 presented for comparison only, to illustrate the differences in model behavior at the low end of  
14 the exposure-response range. Unit risk estimates are not presented for these other models  
15 because, as discussed above, the log cumulative exposure Cox regression model was considered  
16 overly supralinear and the cumulative exposure Cox regression model was considered overly  
17 sublinear for the data in the lower exposure range (e.g., first 4 quintiles of exposure). As one can  
18 see from the results for the subcohort with interviews, the standard Cox regression cumulative  
19 exposure model, with its extreme sublinearity in the lower exposure region, yields a notably  
20 higher  $EC_{01}$  estimate (0.135 ppm) than that from the two-piece linear spline model (0.0112 ppm),  
21 while the log cumulative exposure model, with its extreme supralinearity in the lower exposure  
22 region, yields a substantially lower  $EC_{01}$  estimate (0.0000765 ppm). Converting the units, the  
23 preferred unit risk estimate of 1.74 per ppm corresponds to an estimate of  $9.51 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$   
24 for breast cancer incidence.

25           As discussed above, the primary risk calculations for breast cancer incidence were based  
26 on invasive and in situ tumors in the subcohort of women with interviews, and the primary  
27 model was the two-piece linear spline model. For this assessment, the two-piece spline analyses  
28 were not performed with the full cohort and the life-table analyses were not replicated for the  
29 invasive cancers only. In the 2006 Draft Assessment ([U.S. EPA, 2006a](#)), however, comparison  
30 analyses were done. Using the linear regression of the categorical results, the comparable unit  
31 risk estimate for the full cohort was about 40% lower than the estimate based on the subcohort  
32 with interviews. The corresponding unit risk estimate derived based on the subcohort results but  
33 using invasive breast cancer only for the background incidence rates was about 17% lower than  
34 the estimate based on invasive and in situ tumors, reflecting the difference between incidence  
35 rates for invasive breast cancer only and for combined in situ and invasive breast cancer.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 The unit risk estimate of **1.74 per ppm** ( $1.74 \times 10^{-3}$  per ppb) is the preferred estimate for  
2 female breast cancer risk because it is based on incidence data versus mortality data, it is based  
3 on more cases ( $n = 233$ ) than the mortality estimate ( $n = 103$ ), and information on personal  
4 breast cancer risk factors obtained from the interviews is taken into account. Furthermore, the  
5 two-piece linear spline model, which uses the complete data set with exposure as a continuous  
6 variable, was statistically significant and had the lowest AIC and the best apparent visual fit to  
7 the lower-exposure data of the models considered. Converting the units, 1.74 per ppm  
8 corresponds to a unit risk estimate of  $9.51 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$ .

#### 10 **4.1.3. Total Cancer Risk Estimates**

11 According to EPA's *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)),  
12 cancer risk estimates are intended to reflect total cancer risk, not site-specific cancer risk;  
13 therefore, an additional calculation was made to estimate the combined risk for (incident)  
14 lymphoid and breast cancers, because females would be at risk for both cancer types. Assuming  
15 that the cancer types are independent and that the risk estimates are approximately normally  
16 distributed, one can estimate the 95% UCL (one-sided) on the total risk as the 95% UCL on the  
17 sum of the maximum likelihood estimates (MLEs) of the risk estimates according to the formula

$$18 \qquad \qquad \qquad 19 \qquad \qquad \qquad 20 \qquad \qquad \qquad 95\% \text{ UCL} = \text{MLE} + 1.645(\text{SE}), \qquad \qquad \qquad (4-3)$$

21  
22 where MLE is the MLE of total cancer risk (i.e., the sum of the individual MLEs) and the SE of  
23 the sum of the MLEs is the square root of the sum of the individual variances (i.e., the variance  
24 of the sum is the sum of the variances, and the SE is the square root of the variance). Because  
25 both models are linear in the range around the PODs, the combining-risk calculations can be  
26 done directly from the unit risk and  $0.01/\text{EC}_{01}$  estimates rather than having to do the calculations  
27 at a common exposure level near where the  $\text{EC}_{01}$  and  $\text{LEC}_{01}$  for the combined risk would be.  
28 First, an  $\text{EC}_{01}$  of 0.0078 ppm for the total cancer risk (i.e., lymphoid cancer incidence + breast  
29 cancer incidence) was estimated, as summarized in Table 4-14.

**Table 4-14. Calculation of EC<sub>01</sub> for total cancer risk**

Cancer type	EC <sub>01</sub> (ppm)	0.01/EC <sub>01</sub> (per ppm)	EC <sub>01</sub> for total cancer risk (ppm)
Lymphoid	0.0254	0.394	--
Breast	0.0112	0.893	--
Total <sup>a</sup>	--	1.29	0.00775

<sup>a</sup>The total 0.01/EC<sub>01</sub> value equals the sum of the individual 0.01/EC<sub>01</sub> values; the EC<sub>01</sub> for the total cancer risk then equals 0.01/(0.01/EC<sub>01</sub>).

Then, a unit risk estimate of 2.3 per ppm for the total cancer risk (i.e., lymphoid cancer incidence + breast cancer incidence) was derived, as shown in Table 4-15. An LEC<sub>01</sub> estimate of 0.00441 ppm for the total cancer risk can be calculated as 0.01/(2.27 per ppm).

**Table 4-15. Calculation of total cancer unit risk estimate**

Cancer type	Unit risk estimate (per ppm)	0.01/EC <sub>01</sub> (per ppm)	SE <sup>a</sup> (per ppm)	Variance	Total cancer unit risk estimate (per ppm)	LEC <sub>01</sub> for total cancer risk <sup>d</sup> (ppm)
Lymphoid	0.877	0.394	0.294	0.0864	--	--
Breast	1.74	0.893	0.515	0.265	--	--
Total	--	1.29	(0.593) <sup>b</sup>	0.351	2.27 <sup>c</sup>	0.00441

<sup>a</sup>SE = (unit risk – 0.01/EC<sub>01</sub>)/1.645.

<sup>b</sup>The SE of the total cancer risk is calculated as the square root of the sum of the variances (next column), not as the sum of the SEs.

<sup>c</sup>Total cancer unit risk = 1.29 + 1.645 × 0.593.

<sup>d</sup>The LEC<sub>01</sub> for the total cancer risk equals 0.01/(total cancer unit risk estimate).

Thus, the total cancer unit risk estimate is 2.3 per ppm (or  $2.3 \times 10^{-3}$  per ppb;  $1.2 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$ ). Recall that this is the unit risk estimate derived under the assumption that RR is independent of age (see Section 4.1.1.2). The preferred assumption of increased early-life susceptibility, in accordance with EPA's *Supplemental Guidance* (U.S. EPA, 2005b), is considered in Section 4.4. While there are uncertainties regarding the assumption of a normal distribution of risk estimates, the resulting unit risk estimate is appropriately bounded in the roughly twofold range between estimates based on the sum of the individual MLEs (i.e., 1.29) and the sum of the individual 95% UCLs (i.e., unit risk estimates, 2.6), or more precisely in this

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 case, between the largest individual unit risk estimate (1.74) and the sum of the unit risk  
2 estimates (2.6). Thus, any inaccuracy in the total cancer risk estimate resulting from the  
3 approach used to combine risk estimates across cancer types is relatively minor.  
4

#### 5 **4.1.4. Sources of Uncertainty in the Cancer Risk Estimates**

6 The two major sources of uncertainty in quantitative cancer risk estimates are generally  
7 interspecies extrapolation and high-dose to low-dose extrapolation. The risk estimates derived  
8 from the [Steenland et al. \(2003\)](#), [Steenland et al. \(2004\)](#), and additional Steenland (see  
9 Appendix D) analyses are not subject to interspecies uncertainty because they are based on  
10 human data. Furthermore, the human-based estimates are less affected by high-dose to low-dose  
11 extrapolation than are rodent-based estimates and, thus, uncertainty from that source is reduced  
12 somewhat. For example, the average exposure in the NIOSH cohort was more than 10 times  
13 lower than the lowest exposure level in a rodent bioassay after adjustment to continuous lifetime  
14 exposure. Nonetheless, uncertainty remains in the extrapolation from occupational exposures to  
15 lower environmental exposures. Although the actual exposure-response relationship at low  
16 exposure levels is unknown, the clear evidence of EtO mutagenicity supports the linear  
17 low-exposure extrapolation that was used ([U.S. EPA, 2005a](#)).

18 Because of the existence of endogenous EtO (see Section 3.3.3.1), several members of  
19 the SAB panel that reviewed EPA's external review draft assessment felt that the  
20 exposure-response relationship for cancer at low exposures would be nonlinear and suggested  
21 that it would be consistent with EPA's 2005 *Guidelines for Carcinogen Risk Assessment* ([U.S.](#)  
22 [EPA, 2005a](#)) to present a nonlinear approach for "extrapolation" to lower exposures ([SAB,](#)  
23 [2007](#)). EPA considered this suggestion but judged that the support for a nonlinear approach was  
24 inadequate. In brief, as discussed in Sections 3.1 through 3.3.3, EtO is a DNA-reactive,  
25 mutagenic, multisite carcinogen in humans and experimental species; as such, it has the  
26 hallmarks of a compound for which low-dose linear extrapolation is strongly supported under  
27 EPA's *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)). EPA's *Guidelines for*  
28 *Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)) do provide for presenting alternate approaches  
29 when those alternatives have significant biological support; however, EPA's analysis of the  
30 arguments for using a nonlinear approach presented on page 23 and in Appendix C of the SAB  
31 report ([SAB, 2007](#)) did not find these arguments to be persuasive. The arguments posited by the  
32 SAB panel members who supported using a nonlinear approach were largely that (1) DNA  
33 adducts may show a nonlinear response when identical adducts are formed endogenously and  
34 (2) mutations do not have linear relationships with exposure but exhibit an "inflection point."  
35 However, as discussed in Section 3.3.3.1, recent data from [Marsden et al. \(2009\)](#) support a linear

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 exposure-response relationship for EtO exposure and DNA adducts ( $p < 0.05$ ) and demonstrate  
2 increases of DNA adducts from exogenous EtO exposure above those from endogenous EtO for  
3 very low exposures to exogenous EtO, providing evidence against argument (1). Moreover,  
4 Appendix C of the SAB report ([SAB, 2007](#)) presents two EtO-specific mutation data sets in  
5 support of argument (2); however, EPA's analysis of these data sets finds that they are in fact  
6 consistent with low-dose linearity. See the response to this comment under charge question 2.b  
7 in Appendix H for a more comprehensive discussion of EPA's consideration and rejection of a  
8 nonlinear approach and for the details of EPA's analysis of the two EtO mutation data sets. EPA  
9 also considered more recent (2013) public comments proposing nonlinear modes of action for  
10 EtO carcinogenicity; however, EPA found these hypothetical modes of action to be speculative  
11 at this time (see Appendix L and Section J.3.2 of Appendix J).

12 Other sources of uncertainty emanate from the epidemiologic studies and their analyses  
13 [[Steenland et al. \(2004\)](#);[Steenland et al. \(2003\)](#); Steenland analyses in Appendix D], including  
14 the retrospective estimation of EtO exposures in the cohort, the modeling of the epidemiologic  
15 exposure-response data, the proper dose metric for exposure-response analysis, and potential  
16 confounding or modifying factors. Although these are common areas of uncertainty in  
17 epidemiologic studies, they were generally well addressed in the NIOSH studies.

18 Regarding exposure estimation, the NIOSH investigators conducted a detailed  
19 retrospective exposure assessment to estimate the individual worker exposures. They used  
20 extensive data from 18 facilities, spanning a number of years, to develop a regression model for  
21 estimating EtO exposure levels associated with different jobs (exposure categories), facilities,  
22 and time periods ([Hornung et al., 1994](#); [Greife et al., 1988](#)) [see also Section A.2.8 for more  
23 details about the development and evaluation of the regression model]. The model accounted for  
24 85% of the variation in average EtO exposure levels in an independent set of test data. In  
25 addition, the modeled estimates were not highly biased nor biased in one direction when  
26 compared to the predictions of a panel of 11 industrial hygiene experts familiar with EtO levels  
27 in the sterilization industry. The regression model was used to develop an exposure matrix  
28 stratified by time and exposure category for each facility in the cohort study. Detailed work  
29 history data for the individual workers were collected for the 1987 follow-up and used in  
30 conjunction with the facility-specific exposure matrices to derive cumulative exposure estimates  
31 for the individual workers ([Steenland et al., 1991](#)). For the extended follow-up ([Steenland et al.,](#)  
32 [2004](#); [Steenland et al., 2003](#)), additional information on the date last employed was obtained for  
33 those workers still employed and exposed at the time of the original work history collection for  
34 the plants still using EtO (25% of the cohort). It was then assumed that exposure for these  
35 workers continued until the date of last employment and that their exposure level stayed the

1 same as that in their last job held at the time of the original data collection. Thus, there would be  
2 more exposure misclassification in the extended follow-up. However, when the investigators  
3 compared cumulative exposures estimated with and without the extended work histories, they  
4 found little difference because exposure levels were very low by the mid-1980s and, therefore,  
5 had little impact on cumulative exposure ([Steenland et al., 2004](#); [Steenland et al., 2003](#)). While  
6 the NIOSH regression model performed well in estimating exposures in validation tests  
7 ([Hornung et al., 1994](#)), there is, nonetheless, uncertainty associated with any retrospective  
8 exposure assessment, and this can affect the ability to discriminate among exposure-response  
9 models.

10 With respect to the lymphohematopoietic cancer response, it is not clear exactly which  
11 lymphohematopoietic cancer subtypes are related to EtO exposure, so analyses were done for  
12 both lymphoid cancers and all lymphohematopoietic cancers ([Steenland et al., 2004](#)). The  
13 associations observed for all lymphohematopoietic cancers was largely driven by the lymphoid  
14 cancer responses, and biologically, there is stronger support for an etiologic role for EtO in the  
15 development of the more closely related lymphoid cancers than in the development of the more  
16 diverse cancers in the aggregate all lymphohematopoietic cancer grouping; thus, the lymphoid  
17 cancer analysis is the preferred analysis for the lymphohematopoietic cancers. Nonetheless, the  
18 preferred unit risk estimate for all lymphohematopoietic cancers was similar to (about 50%  
19 greater than) that for the lymphoid cancers.

20 For the lymphoid cancer response ([Steenland et al., 2004](#)), modeling the  
21 exposure-response relationship is limited by the small number of cases ( $n = 53$ ). The Cox  
22 proportional hazards model used by [Steenland et al. \(2004\)](#) is commonly used for this type of  
23 analysis because exposure can be modeled as a continuous variable, competing causes of  
24 mortality can be taken into account, and potential confounding factors can be controlled for in  
25 the regression. Normally, model dependence should be minimized by the practice, under EPA's  
26 2005 *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)), of modeling only in the  
27 observable range and then performing a linear extrapolation from the "POD" (in this case the  
28  $LEC_{01}$ ). However, the log cumulative exposure Cox regression model with 15-year lag, which  
29 provides the best fit to the overall data, is too steep in the low-exposure region and then plateaus  
30 rapidly at higher exposures, making it difficult to derive stable risk estimates (i.e., estimates that  
31 are not highly dependent on the POD). And the alternative cumulative exposure model, though  
32 typically used for epidemiologic data, is too sublinear (shallow) in the low-exposure region for  
33 these data, which exhibit supralinearity. EPA attempted to fit two-piece log-linear and linear  
34 spline models to the individual continuous exposure data to address the supralinearity of the data  
35 while avoiding the extreme low-exposure curvature of the log cumulative exposure model;

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 however, these models resulted in low-exposure slopes that appeared to be implausibly steep  
2 (i.e., they suggested excessively large changes in risk from small changes in exposure). The  
3 steep low-exposure slopes are a manifestation of apparently high risks in workers with relatively  
4 low exposures; however, this elevation is based on small numbers of cancer cases in that  
5 exposure range, and EPA has low confidence in the low-exposure slopes. A two-piece spline  
6 model with the knot at a higher exposure level could have been used, but without model  
7 likelihood as a basis for knot selection, such selection becomes arbitrary, and with the knot at the  
8 higher exposure level which had an apparent local maximum for the log-linear model  
9 (1,600 ppm × days rather than 100 ppm × days), the model was not statistically significant  
10 ( $p = 0.07$ ). Thus, EPA opted for a weighted linear regression model based on the Cox regression  
11 categorical results, excluding the highest exposure group, to reflect the exposure-response  
12 relationship in the exposure region below the “plateau.” The all lymphohematopoietic cancer  
13 data set had more cases ( $n = 74$ ) but was heavily dominated by the lymphoid cancer response and  
14 conveyed the same problems for exposure-response modeling; thus, a linear regression model,  
15 excluding the highest exposure group, was used for this data set as well.

16 The linear model is a parsimonious choice that assumes neither a sublinear nor a  
17 supralinear exposure-response relationship and acknowledges the inherent imprecision in the  
18 epidemiological data. The highest exposure group was excluded because it is less relevant to the  
19 low-exposure risks of interest for low-exposure extrapolation and its inclusion would have overly  
20 influenced the linear regression, resulting in a slope that would have substantially underestimated  
21 the apparent low-exposure risks. Excluding data can appear arbitrary, but EPA aimed to avoid  
22 an arbitrary selection by using the a priori exposure groups presented by [Steenland et al. \(2004\)](#)  
23 and excluding only the highest exposure group, with the exposures least relevant to low  
24 environmental exposure levels. The linear regression has its own limitations (e.g., it is based on  
25 categorical rather than continuous exposure data and the slopes were not statistically significant);  
26 nonetheless, it was judged to be the most reasonable approach for deriving low-exposure risk  
27 estimates from the available lymphohematopoietic cancer data.

28 Although the linear regression model of the categorical results seems to be a reasonable  
29 approach for best reflecting the exposure-response results at the lower end of the exposure range,  
30 clearly there is uncertainty regarding the exposure-response model. The log cumulative  
31 exposure Cox regression model, which was the best-fitting model overall of the models  
32 investigated, yields lower  $EC_{01}$  and  $LEC_{01}$  estimates than the linear regression model of the  
33 categorical results (see Table 4-6), but the estimates based on the linear regression model are  
34 preferred because the linear regression model is more stable. The most suitable alternative  
35 model based on the continuous exposure data is the two-piece log-linear spline model with the

1 knot at 1,600 ppm × days, and thus, unit risk calculations using this model are included in the  
2 assessment for comparison purposes. The unit risk estimate from this alternative model is about  
3 2.0 times the estimate from the linear regression model of the categorical results. If the  
4 alternative model had been selected for lymphoid cancer rather than the linear regression model  
5 of the categorical results, the total cancer unit risk estimate would have been 2.90 per ppm rather  
6 than 2.27 per ppm, that is, about 28% higher than the total cancer risk estimate with the preferred  
7 lymphoid cancer model.<sup>38</sup>

8 Another area of uncertainty related to the exposure-response modeling is the lag period.  
9 The best-fitting models presented by [Steenland et al. \(2004\)](#) for lymphohematopoietic cancer  
10 mortality had a 15-year lag (lag periods of 0, 5, 10, 15, and 20 years were considered). A  
11 15-year lag period means that exposures in the 15 years prior to death or the end of follow-up are  
12 not taken into account. In other words, in the best-fitting models, relevant exposures for the  
13 development of the lymphohematopoietic cancers occurred over 15 years before death. For the  
14 best-fitting continuous exposure model for lymphoid cancer reported by [Steenland et al. \(2004\)](#),  
15 the log cumulative exposure Cox regression model, the actual difference between the regression  
16 coefficients from the 15-year-lagged and the unlagged models was negligible (the regression  
17 coefficient from the unlagged model was about 8% lower than that from the 15-year-lagged  
18 model; however, it should be noted that the unlagged model did not provide a statistically  
19 significant fit to the data ( $p = 0.17$ ) (the results for the unlagged model are presented in  
20 Section D.3.e of Appendix D).

21 In addition, the analyses of the NIOSH investigators indicate that the regression  
22 coefficient for cumulative exposure might have decreased with increasing follow-up, suggesting  
23 that the higher exposure levels encountered by the workers in the more distant past are having  
24 less of an impact on more recent risk. The regression coefficient for lymphoid cancers was  
25  $1.2 \times 10^{-5}$  per ppm × day, for both sexes with a 10-year lag, in the 1987 follow-up ([Stayner et al.,](#)  
26 [1993](#)) versus  $4.7 \times 10^{-6}$  per ppm × day, for both sexes with a 15-year lag, in the 1998 follow-up  
27 (see Steenland reanalyses in Appendix D). A similar decrease was found in the regression  
28 coefficient for cumulative exposure for all lymphohematopoietic cancers. The life-table analysis  
29 used in this dose-response assessment assumes exposure accrues over the full lifetime for the  
30 cumulative exposure metric. If, in fact, exposures in the distant past cease to have a meaningful  
31 impact on the risk of lymphohematopoietic cancers, this approach would tend to overestimate the  
32 unit risk. Thus, a comparison analysis was conducted to evaluate the impact of ignoring  
33 exposures over 55 years in the past in the life-table analysis. The actual value of such a cut

---

<sup>38</sup>The total cancer risk calculations were conducted at 0.003 ppm, an exposure level expected to be near the LEC<sub>01</sub> estimate for the total cancer risk.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 point, if warranted, is unknown. A value less than 55 years might not be appropriate because  
2 exposures for some of the workers began in 1943, so any diminution of potency for past  
3 exposures occurring since 1943 is already reflected in the regression coefficient with follow-up  
4 through 1998, at least for those workers, although it is unknown what proportion of workers had  
5 such early exposures and how long they survived. The comparison analysis for lymphoid cancer  
6 yielded an  $LEC_{01}$  of 0.0156 ppm and a unit risk estimate of 0.64 per ppm, which is about  
7 27% less than the estimate obtained from the unrestricted life-table analysis. Because the  
8 appropriate cut point for excluding past exposures is unknown and the unit risk estimate from the  
9 linear regression model of the categorical results is already substantially less than that obtained  
10 from the best-fitting log cumulative exposure Cox regression model or even that from the  
11 alternative two-piece log-linear spline model with the knot at 1,600 ppm  $\times$  days, the estimate  
12 from the full life-table analysis is preferred. In any event, the preferred estimate is not  
13 appreciably different from the estimate from the analysis which considered only the most recent  
14 55 years of exposure in the life-table analysis.

15 Several dose metrics (cumulative exposure, duration of exposure, maximum [8-hour  
16 TWA] exposure, and average exposure) were analyzed by [Steenland et al. \(2004\)](#), and  
17 cumulative exposure was the best predictor of mortality from lymphohematopoietic cancers.  
18 Cumulative exposure is considered a good measure of total exposure because it integrates  
19 exposure (levels) over time.

20 Also, the important potential modifying/confounding factors of age, sex, race, and  
21 calendar time were taken into account in the analysis, and the plants included in this cohort were  
22 specifically selected for the absence of any known confounding exposures ([Stayner et al., 1993](#)).

23 With respect to the breast cancer mortality response ([Steenland et al., 2004](#)), the  
24 exposure-response modeling was based on 103 deaths. As for the lymphohematopoietic cancer  
25 responses, the exposure-response data for breast cancer mortality are fairly supralinear,  
26 especially for the low-exposure groups. An attempt was again made to fit two-piece log-linear  
27 and linear spline models to the individual continuous exposure data to address the supralinearity  
28 of the data while avoiding the extreme low-exposure curvature of the log cumulative exposure  
29 Cox regression model; however, these models resulted in low-exposure slopes that appeared to  
30 be implausibly steep and the model fits were not statistically significant. Thus, the same linear  
31 regression approach, excluding the highest exposure group, was taken to obtain a regression  
32 coefficient for the life-table analysis. As discussed above, the linear regression has its own  
33 limitations (e.g., it is based on categorical rather than continuous exposure data and the slope is  
34 not statistically significant); nonetheless, it was judged to be the most reasonable approach for  
35 deriving low-exposure risk estimates from the available breast cancer mortality data. The most

1 suitable alternative model based on the continuous exposure data is the two-piece log-linear  
2 spline model with the knot at 13,000 ppm × days (not the knot with the largest likelihood, but a  
3 local maximum), and thus, unit risk calculations using this model are included in the assessment  
4 for comparison purposes. The unit risk estimate from this alternative model is about 34% of the  
5 estimate from the linear regression model of the categorical results.

6 For the lag period, the best-fitting model had a lag of 20 years, which was the longest lag  
7 period investigated. This is a commonly used lag period for solid tumors, which typically have  
8 longer latency periods than lymphohematopoietic cancers. It is unknown whether a lag period  
9 longer than 20 years would have provided a better model fit. The [Steenland et al. \(2004\)](#)  
10 analysis took into account age, race, and calendar time. Other risk factors for breast cancer could  
11 not be included in the mortality analysis, but many of these factors were considered in the breast  
12 cancer incidence study ([Steenland et al., 2003](#)), as discussed below, and the preferred breast  
13 cancer risk estimates are based on the breast cancer incidence data.

14 [Steenland et al. \(2003\)](#) conducted an incidence study for breast cancer; therefore, it was  
15 not necessary to calculate unit risk estimates for breast cancer incidence indirectly from the  
16 mortality data as was done for lymphohematopoietic cancer. Further advantages to using the  
17 results from the incidence study are that more cases were available for the exposure-response  
18 modeling (319 cases) and that the investigators were able to include data on potential  
19 confounders in the modeling for the subcohort with interviews (233 cases). Because the  
20 subcohort with interviews had complete case ascertainment and provided data on potential  
21 confounders, it was the preferred breast cancer incidence data set, although some results based  
22 on the full cohort are presented for comparison. For the full cohort, the continuous exposure Cox  
23 regression model providing the best fit to the data was again the log cumulative exposure model.  
24 With breast cancer incidence, a 15-year lag provided the best model fits. For the subcohort, the  
25 cumulative exposure and log cumulative exposure Cox regression models fit nearly equally well  
26 (e.g., similar AICs). For both groups, the categorical Cox regression results suggest that a linear  
27 model, providing a compromise between the supralinear log cumulative exposure model and the  
28 sublinear cumulative exposure model, would better represent the low-exposure data than either  
29 of the two presented continuous exposure models (see Figures 4-4 and 4-5). Thus, for both  
30 groups, in EPA's original draft analyses based on the published summary data, a linear  
31 regression was fitted to the categorical results, dropping the highest exposure group to provide a  
32 better fit to the lower-exposure data ([U.S. EPA, 2006a](#)). In subsequent analyses by Dr. Steenland  
33 (see Appendix D) of the individual data using exposure as a continuous variable, two-piece  
34 log-linear and linear spline models and other linear RR models were used to model the subcohort

1 data; the two-piece linear spline model was the best-fitting of these models and provided the  
2 preferred breast cancer incidence risk estimates.

3 Confidence intervals were determined using the Wald approach. Confidence intervals for  
4 linear RR models, however, in contrast to those for the log-linear RR models, may not be  
5 symmetrical. EPA also evaluated application of a profile likelihood approach for the linear RR  
6 models ([Langholz and Richardson, 2010](#)), which allows for asymmetric CIs, for comparison with  
7 the Wald approach. Using the profile likelihood method and the two-piece linear spline model,  
8 the resulting unit risk estimate for breast cancer incidence would have been 2.33 per ppm,  
9 slightly higher (34%) than the value of 1.74 per ppm obtained as the unit risk estimate for breast  
10 cancer incidence in this assessment. Thus, if the profile likelihood method had been used for the  
11 linear RR models in this assessment, the total cancer risk estimate, which incorporates the breast  
12 cancer incidence estimate as a component, would be less than 34% higher than the total cancer  
13 risk estimate presented here.

14 With respect to the two-piece spline models, the use of this model form is not intended to  
15 imply that an abrupt change in biological response occurs at the knot but, rather, to allow  
16 description of an exposure-response relationship in which the slope of the relationship differs  
17 notably in the low-exposure versus high-exposure regions. The two-piece model is used here  
18 primarily for its representation of the low-exposure data. The main uncertainty in the two-piece  
19 spline models is in the selection of the knot, and the location of the knot is critical in defining the  
20 low-exposure slope. The model likelihood was used to provide a statistical basis for knot  
21 selection; although, as shown in Appendix D (see Figure D-1a), the likelihood did not generally  
22 change appreciably over a range of possible knots. Thus, because of the importance of knot  
23 selection, a sensitivity analysis was done to examine the impacts of selecting different knots (see  
24 Section D.6 of Appendix D). For the sensitivity analysis, the two-piece log-linear model was run  
25 with knots roughly one increment (1,000 ppm × days) below and one increment above the  
26 selected knot. For breast cancer incidence, this sensitivity analysis yielded EC<sub>01</sub> estimates of  
27 0.0133 ppm and 0.0176 ppm, respectively (i.e., about 14% lower and 14% higher, respectively,  
28 than the EC<sub>01</sub> of 0.0154 ppm obtained with the originally selected knot of 6,000 ppm × days).<sup>39</sup>

29 As can be seen in Table 4-13, there is substantial variation in the EC<sub>01</sub> estimates obtained  
30 from the different models. Although some plateauing is apparent with the highest exposure  
31 group and is evidenced in the subcohort with interviews by the strong influence of the top 5% of  
32 cumulative exposures on dampening the slope of the (cumulative exposure) Cox regression

---

<sup>39</sup> About 12% lower and 17% higher, respectively, than the EC<sub>01</sub> of 0.0151 ppm obtained with the more finely tuned knot of 5,800 ppm × days (see Appendix D). The EC<sub>01</sub> value of 0.0166 presented in this assessment (see Table 4-13) is not directly comparable to the values in the sensitivity analysis because more recent background incidence and mortality rates were used in the lifetable analyses upon which the assessment estimates were based.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 model (see Section D.1 and Figure D-1d of Appendix D), the categorical data for breast cancer  
2 incidence do not display the strong supralinearity (steep rise) in the lower exposure groups seen  
3 in the cases discussed above (i.e., lymphohematopoietic cancers and breast cancer mortality).  
4 Thus, for the subcohort with interviews, the difference between the  $EC_{01}$  estimates from the  
5 standard cumulative exposure Cox regression model and the two-piece spline models or the  
6 linear regression of the categorical results or continuous exposure linear models are not as  
7 dramatic as seen in those cases (the  $EC_{01}$  estimates from the latter four approaches are nearly  
8 within an order of magnitude of that of the cumulative exposure model). For the subcohort with  
9 interviews, the two-piece spline models, the continuous exposure linear model, and the linear  
10 regression of the categorical results gave similar results—the unit risk estimates spanned less  
11 than a fourfold range. This range is bounded by the two best-fitting (based on AIC) continuous  
12 exposure models—the two-piece linear spline model and the linear model. If the continuous  
13 exposure linear model had been selected rather than the two-piece linear spline model, which had  
14 a slightly lower AIC value and a better apparent visual fit of the lower-exposure data, the breast  
15 cancer incidence unit risk estimate would have been 0.446 per ppm rather than 1.74 per ppm, and  
16 the total cancer unit risk estimate would have been 1.15 per ppm rather than 2.27 per ppm. In  
17 other words, of the models investigated, the total cancer unit risk estimate with the best-fitting  
18 alternate breast cancer incidence model (based on AIC) is about 50% lower than that of the  
19 best-fitting model. However, data in the lower-exposure range of greatest relevance for the  
20 derivation of a unit risk estimate support a steeper slope in the lower-exposure range; thus,  
21 although the lower estimate obtained from the continuous exposure linear model is plausible,  
22 unit risk estimates notably lower than that are considered unlikely from the available data.

23 The best-fitting models presented by [Steenland et al. \(2003\)](#) for breast cancer incidence  
24 generally had a 15-year lag (lag periods of 0, 5, 10, 15, and 20 years were considered). A  
25 15-year lag period means that exposures in the 15 years prior to diagnosis or the end of  
26 follow-up are not taken into account. For the various continuous exposure models for breast  
27 cancer incidence in the full cohort and the subcohort with interviews reported by [Steenland et al.](#)  
28 [\(2003\)](#), Tables 4 and 5, none of the unlagged models provided a statistically significant fit to the  
29 data, with the exception of the log cumulative exposure Cox regression model for the subcohort,  
30 where the unlagged model fit marginally better than the 15-year-lagged model. However, as  
31 noted in Section 4.1.2.3, the log cumulative exposure model with no lag was considered less  
32 biologically realistic than the corresponding model with a 15-year lag because some lag period  
33 would be expected for the development of breast cancer; thus, the 15-year-lagged model was  
34 used in this assessment. The regression coefficient from the unlagged log cumulative exposure  
35 Cox regression model was about 90% higher than that from the 15-year-lagged model.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 With respect to dose metrics for breast cancer incidence, models using duration of  
2 exposure provided better model fits than those using cumulative exposure ([Steenland et al.,  
3 2003](#)); however, duration is less useful for estimating unit risks and the cumulative exposure  
4 models also provided statistically significant fits to the data, thus the cumulative exposure metric  
5 was used for the quantitative risk estimates. Models using peak (highest one-time exposure) or  
6 average exposure (cumulative exposure divided by duration) did not fit as well.

7 Regarding potential confounders/modifying factors, analyses for the full cohort were  
8 adjusted for age, race, and calendar time, and exposures to other chemicals in these plants were  
9 reportedly minimal. For the subcohort with interviews, a number of specific breast cancer risk  
10 factors were investigated, including body mass index, breast cancer in a first-degree relative,  
11 parity, age at menopause, age at menarche, socioeconomic status, and diet; however, only parity  
12 and breast cancer in a first-degree relative were determined to be important predictors of breast  
13 cancer and were included in the final models.

14 An area of uncertainty in the life-table analysis for breast cancer incidence pertains to the  
15 rates used for the cause-specific background rate. The regression coefficients presented by  
16 [Steenland et al. \(2003\)](#) represent invasive and in situ cases combined, where 6% of the cases are  
17 in situ, and the preferred unit risk estimates in this assessment are calculated similarly using  
18 background rates for invasive and in situ cases combined. The regression coefficients for  
19 invasive and in situ cases combined should be good approximations for regression coefficients  
20 for invasive cases alone; however, it is uncertain how well they reflect the exposure-response  
21 relationships for in situ cases alone. Diagnosed cases of in situ breast cancer would presumably  
22 be remedied and not progress to invasive breast cancer, so double-counting is unlikely to be a  
23 significant problem. Carcinoma in situ is a risk factor for invasive breast cancer; however, this  
24 observation is most likely explained by the fact that these two types of breast cancer have other  
25 breast cancer risk factors in common, some of which have been considered in the subcohort  
26 analysis. One might hypothesize that EtO exposure could cause a more rapid progression to  
27 invasive tumors; however, there is no specific evidence that this occurs. On the other hand, there  
28 is some indication that in situ cases in the incidence study might have been diagnosed at  
29 relatively low rates in comparison to the invasive cases. [Steenland et al. \(2003\)](#) reported that 6%  
30 of the cases in their study are in situ; according to the National Cancer Institute, however, ductal  
31 carcinoma in situ accounted for about 18% of newly diagnosed cases of breast cancer in 1998  
32 ([NCI, 2004](#)).

33 There are several possible explanations for this difference. One is that it reflects  
34 differences in diagnosis with calendar time because the rate of diagnosis of carcinoma in situ has  
35 increased over time with increased use of mammography. Another is that the difference is

1 partially a reflection of the age distribution in the cohort because the proportion of new cases  
2 diagnosed as carcinoma in situ varies by age. A third possible explanation is that the low  
3 proportion of in situ cases is at least partially a consequence of underascertainment of cases  
4 because in situ cases will not be reported on death certificates, although, even if all 20 in situ  
5 cases were in the subcohort with interviews, that would still be only 8.6% of the cases. In any  
6 event, this is a relatively minor source of uncertainty, and a comparison of the unit risk estimates  
7 using invasive + in situ breast cancer background rates and invasive-only background rates,  
8 using EPA's original analyses in the 2006 Draft Assessment, found that the estimate based on the  
9 invasive + in situ background rates was less than 20% higher than the corresponding estimate  
10 using only invasive breast cancer background rates ([U.S. EPA, 2006a](#)).

11 The results for the subcohort with interviews are used for the primary breast cancer unit  
12 risk calculations because, in addition to including the data on potential confounders, the  
13 subcohort is considered to have full ascertainment of the breast cancer cases, whereas the full  
14 cohort for the incidence study has incomplete case ascertainment, as illustrated by the fact that  
15 death certificates were the only source of case ascertainment for 14% of the cases. Complete  
16 interviews were available for only 68% of the 7,576 women in the full incidence cohort, and  
17 thus, some potential exists for participation selection bias in the subcohort. There is, however,  
18 no basis for considering participation to be associated with breast cancer or EtO exposure, and  
19 the major reason for nonparticipation was a failure to locate (22% of full incidence cohort) and  
20 not lack of response (3% of cohort) or refusal to participate (7% of cohort). Risk estimates based  
21 on the full cohort were calculated for comparison with the subcohort estimates using the original  
22 linear regression analyses of the categorical results. The unit risk estimate based on the  
23 subcohort was about 60% higher than the corresponding estimate from the full cohort ([U.S. EPA,](#)  
24 [2006a](#)).

25 Some additional sources of uncertainty are not so much inherent in the exposure-response  
26 modeling or in the epidemiologic data themselves but, rather, arise in the process of obtaining  
27 more general Agency risk estimates from the epidemiologic results. EPA cancer risk estimates  
28 are typically derived to represent an upper bound on increased risk of cancer incidence for all  
29 sites affected by an agent for the general population. From experimental animal studies, this is  
30 accomplished by using tumor incidence data and summing across all the tumor sites that  
31 demonstrate significantly increased incidences, customarily for the most sensitive sex and  
32 species, to be protective of the general human population. However, in estimating comparable  
33 risks from the NIOSH epidemiologic data, certain limitations are encountered. First, the study  
34 reported by [Steenland et al. \(2004\)](#) is a retrospective mortality study, and cancer incidence data  
35 are not available for lymphohematopoietic cancer [for breast cancer, a separate incidence study

1 ([Steenland et al., 2003](#)) was available]. Second, these occupational epidemiology data represent  
2 a healthy worker cohort. Third, the epidemiologic study may not have sufficient statistical  
3 power and follow-up time to observe associations for all the tumor sites that may be affected by  
4 EtO.

5 The first limitation was addressed quantitatively in the life-table analysis for the  
6 lymphohematopoietic cancer risk estimates. Although assumptions are made in using incidence  
7 rates for the cause-specific background rates, as discussed in Section 4.1.1.3, the resulting  
8 incidence-based estimates are believed to be better estimates of cancer incidence risk than are the  
9 mortality-based estimates. The incidence unit risk estimate is about 120% higher than (i.e.,  
10 2.2 times) the mortality-based estimate, which seems reasonable given the relatively high  
11 survival rates for lymphoid cancers (according to SEER data [[www.seer.cancer.gov](http://www.seer.cancer.gov)], 5-year  
12 survival rates are 65% for NHL; 78% for chronic lymphocytic leukemia, which are the vast  
13 majority of the lymphocytic leukemias in adults; and 40% for multiple myeloma).

14 The healthy-worker effect is often an issue in occupational epidemiology studies, but the  
15 internal exposure-response analyses conducted by these investigators help address this concern,  
16 at least partially. In terms of representing the general population, the NIOSH study cohort was  
17 relatively diverse. It contained both female (55%) and male workers, and the workers were 79%  
18 white, 16% black, and 5% “other.” Furthermore, because of EtO’s mutagenic mode of action,  
19 increased early-life susceptibility is assumed and ADAFs are applied for exposure scenarios  
20 involving early life (see Section 4.4).

21 With respect to other possible tumor sites of concern, the rodent data suggest that  
22 lymphohematopoietic cancers are a major tumor type associated with EtO exposure in female  
23 mice and in male and female rats. Thus, it is reasonable that this might be a cancer type of  
24 concern in humans also. Likewise, the mouse data suggest an increased risk of mammary gland  
25 tumors from EtO exposure, and evidence of that can be seen in the [Steenland et al. \(2004\)](#) and  
26 [Steenland et al. \(2003\)](#) studies. However, the rodent data suggest associations between EtO  
27 exposure and other tumor types as well, and although site concordance across species is not  
28 generally assumed, it is possible that the NIOSH study, despite its relatively large size and long  
29 follow-up (mean length of follow-up was 26.8 years), had insufficient power to observe small  
30 increases in risk in certain other sites. For example, the tumor site with the highest potency  
31 estimate in both male and female mice was the lung. In the NIOSH study, one cannot rule out a  
32 small increase in the risk of lung cancer, which has a high background rate, thus making small  
33 increases difficult to detect.

34 To obtain the risk estimate for total cancer risk (2.3 per ppm, or  $2.3 \times 10^{-3}$  per ppb), the  
35 preferred estimates for lymphoid cancer incidence and breast cancer incidence were combined.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 While there are uncertainties in the approach used to combine the individual estimates, the  
2 resulting unit risk estimate is appropriately bounded in the roughly twofold range between  
3 estimates based on the sum of the individual MLEs of risk and the sum of the individual 95%  
4 UCLs, and thus, any inaccuracy in the total cancer unit risk estimate resulting from the approach  
5 used is relatively minor. Because the breast cancer component of the total cancer risk estimate  
6 applies only to females, the total cancer risk estimate is expected to overestimate the cancer risk  
7 to males somewhat (the preferred unit risk estimate for lymphoid cancer alone was 0.877 per  
8 ppm [or  $8.77 \times 10^{-4}$  per ppb], which is about 40% of the total cancer risk estimate).

9 Despite these uncertainties, the inhalation cancer unit risk estimate of 2.3 per ppm (or  
10  $2.3 \times 10^{-3}$  per ppb) for the total cancer risk from lymphoid cancer incidence and female breast  
11 cancer incidence has the advantages of being based on human data from a large, high-quality  
12 epidemiologic study with individual exposure estimates for each worker. Furthermore, the breast  
13 cancer component of the risk estimate, which contributes approximately 60% of the total cancer  
14 risk, is based on a substantial number of incident cases (233 total, the vast majority of which  
15 were in the exposure range below the knot of 5,800 ppm  $\times$  days [see Table D.1a of  
16 Appendix D]).

17 A further area of uncertainty pertains to the assumption that RR is independent of age,  
18 which is a common assumption in the dose-response modeling of epidemiological data and is an  
19 underlying assumption in the Cox regression model. For the NIOSH worker cohort, the  
20 proportional hazards model assumption of RR being independent of age was tested by checking  
21 the significance of an interaction between age and cumulative exposure, and none of the models  
22 had a significant interaction term. This suggests that, for adults at least, the assumption that RR  
23 is independent of age is valid. However, the worker cohort contains no children and is  
24 uninformative on the issue of early-life susceptibility. In the absence of data on early-life  
25 susceptibility, EPA's *Supplemental Guidance* ([U.S. EPA, 2005b](#)) recommends that increased  
26 early-life susceptibility be assumed for carcinogens with a mutagenic mode of action, and the  
27 conclusion was made in Section 3.4 that the weight of evidence supports a mutagenic mode of  
28 action for EtO. Thus, in accordance with the *Supplemental Guidance*, the alternate assumption  
29 of increased early-life susceptibility is preferred as the basis for risk estimates in this assessment,  
30 and risk estimates derived under this preferred assumption are presented in Section 4.4.

#### 31 32 **4.1.5. Summary**

33 Under the common assumption that RR is independent of age, an inhalation unit risk  
34 estimate for lymphoid cancer incidence of 0.877 per ppm (or  $8.77 \times 10^{-4}$  per ppb;  $4.79 \times 10^{-4}$  per  
35  $\mu\text{g}/\text{m}^3$ ) was calculated using a life-table analysis and a weighted linear regression of the

1 categorical Cox regression results, excluding the highest exposure group, for excess lymphoid  
2 cancer mortality from a high-quality occupational epidemiology study. Similarly, an inhalation  
3 unit risk estimate for female breast cancer incidence of 1.74 per ppm (or  $1.74 \times 10^{-3}$  per ppb;  
4  $9.51 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$ ) was calculated using a life-table analysis and two-piece linear spline  
5 modeling of the continuous exposure data for excess breast cancer incidence from the same  
6 high-quality occupational epidemiology study. The linear regression of the categorical results  
7 with the exclusion of the highest exposure group for the lymphoid cancer results and the  
8 two-piece linear spline analysis for the breast cancer incidence data were different modeling  
9 approaches used to address the supralinearity of the exposure-response data in the two data sets.  
10 Low-dose linear extrapolation was used, as warranted by the clear mutagenicity of EtO. An  
11  $\text{EC}_{01}$  estimate of 0.0078 ppm, a  $\text{LEC}_{01}$  estimate of 0.0044 ppm, and a unit risk estimate of 2.3 per  
12 ppm (or  $2.3 \times 10^{-3}$  per ppb;  $1.2 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$ ) were obtained for the total cancer risk  
13 combined across both cancer types. Despite the uncertainties discussed above, this inhalation  
14 unit risk estimate has the advantages of being based on human data from a high-quality  
15 epidemiologic study with individual exposure estimates for each worker.

16 In the absence of data on early-life susceptibility, EPA's *Supplemental Guidance* ([U.S.](#)  
17 [EPA, 2005a](#)) recommends that increased early-life susceptibility be assumed for carcinogens  
18 with a mutagenic mode of action, and the conclusion was made in Section 3.4 that the weight of  
19 evidence supports a mutagenic mode of action for EtO. Thus, in accordance with the  
20 *Supplemental Guidance*, the alternate assumption of increased early-life susceptibility is  
21 preferred as the basis for risk estimates in this assessment, and risk estimates derived under this  
22 preferred assumption are presented in Section 4.4. Other than the use of the alternate assumption  
23 about early-life susceptibility, the approach used to derive the estimates presented in Section 4.4  
24 is identical to the approach used for the estimates derived here in Section 4.1, and the  
25 comparisons made between various options and the issues and uncertainties discussed here in  
26 Section 4.1 are applicable to the estimates derived in Section 4.4.

27

## 28 **4.2. INHALATION UNIT RISK DERIVED FROM EXPERIMENTAL ANIMAL DATA**

### 29 **4.2.1. Overall Approach**

30 Lifetime animal cancer bioassays of inhaled EtO have been carried out in three  
31 laboratories, as described in Section 3.2. The data from these reports are presented in Tables 3-3  
32 through 3-5. These studies have also been reviewed by the [IARC \(1994b\)](#) and [Health Canada](#)  
33 [\(2001\)](#). Health Canada calculated the  $\text{ED}_{05}$  for each data set using the benchmark dose  
34 methodology. The EOIC report ([EOIC, 2001](#)) tabulated only lymphatic tumors because they  
35 constituted the predominant risk.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1           The overall approach in this derivation is to find a unit risk for each of the  
2 bioassays—keeping data on males and females separate—from data on the incidence of all tumor  
3 types and then to use the maximum of these values as the summary measure of the unit risk from  
4 animal studies (i.e., the unit risk represents the most sensitive species and sex). The unit risk for  
5 the animals in these bioassays is converted to a unit risk in humans by first determining the  
6 continuous exposures in humans that are equivalent to the rodent bioassay exposures and then by  
7 assuming that the lifetime incidence in humans is equivalent to lifetime incidence in rodents, as  
8 is commonly accepted in interspecies risk extrapolations. For cross-species scaling of exposure  
9 levels (see Section 4.2.2 below), an assumption of ppm equivalence is used; thus, no interspecies  
10 conversion is needed for the exposure concentrations. Bioassay exposure levels are adjusted to  
11 equivalent continuous exposures by multiplying by (hours of exposure/24 hours) and by (5/7) for  
12 the number of days exposed per week. The unit risk in humans (risk per unit air concentration)  
13 is then assumed to be numerically equal to that in rodents (after adjustment to continuous  
14 exposures); the calculations from the rodent bioassay data are shown in Tables 3-3 through 3-5.  
15

#### 16 **4.2.2. Cross-Species Scaling**

17           In the absence of chemical-specific information, EPA’s 1994 inhalation dosimetry  
18 methods ([U.S. EPA, 1994](#)) provide standard methods and default scaling factors for  
19 cross-species scaling. Under EPA’s methodology, EtO would be considered a Category 2 gas  
20 because it is reactive and water soluble and has clear systemic distribution and effects.  
21 Dosimetry equations for Category 2 gases are undergoing EPA re-evaluation and are not being  
22 used at this time. For cross-species scaling of extraréspiratory effects, current practice is to treat  
23 Category 2 gases as Category 3 gases. For Category 3 gases, ppm equivalence is assumed (i.e.,  
24 responses across species are equivalent on a ppm exposure basis), unless the air: blood partition  
25 coefficient for the experimental species is less than the coefficient for humans [[U.S. EPA \(1994\)](#),  
26 p. 4–61]. In the case of EtO, measured air: blood partition coefficients are 78 in the mouse  
27 ([Fennell and Brown, 2001](#)), 64 in the rat ([Krishnan et al., 1992](#)), and 61 in the human ([Csanady  
28 et al., 2000](#)); thus, ppm equivalence for cross-species scaling to humans can be assumed for  
29 extraréspiratory effects observed in mice and rats. The assumption of ppm equivalence is further  
30 supported by the PBPK modeling of [Fennell and Brown \(2001\)](#), who reported that simulated  
31 blood AUCs for EtO after 6 hours of exposure to concentrations between 1 ppm and 100 ppm  
32 were similar for mice, rats, and humans and were linearly related to the exposure concentration  
33 (see Section 3.3.1 and Figure 3-2). This modeling was validated against measured blood EtO  
34 concentrations for rodents and humans. For Category 2 gases with respiratory effects, there is no  
35 clear guidance on an interim approach. One suggested approach is to do cross-species scaling

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 using both Category 1 and Category 3 gas equations and then decide which is most appropriate.  
2 In this document, the preferred approach was to assume ppm equivalence was also valid for the  
3 lung tumors in mice because of the clear systemic distribution of EtO (e.g., see Section 3.1).  
4 Treating EtO as a Category 1 gas for cross-species scaling of the lung tumors would presume  
5 that the lung tumors are arising only from the immediate and direct action of EtO as it comes into  
6 first contact with the lung. In fact, some of the EtO dose contributing to lung tumors is likely  
7 attributable to recirculation of systemic EtO through the lung.

8 If one were to treat EtO as a Category 1 gas for the cross-species scaling of the lung  
9 tumor response as a bounding exercise, EPA's 1994 inhalation dosimetry methods present  
10 equations for estimating the  $RGDR_{PU}$ , i.e., the regional gas dose ratio for the pulmonary region,  
11 which acts as an adjustment factor for estimating human equivalent exposure concentrations  
12 from experimental animal exposure concentrations (adjusted for continuous exposure) [[U.S. EPA](#)  
13 [\(1994\)](#), pp. 4–49 to 4–51]. These equations rely on parameters describing mass transport of the  
14 gas (EtO) in the extrathoracic and tracheobronchial regions for both the experimental animal  
15 species (mouse) and humans. Without experimental data for these parameters, it seems  
16 reasonable to estimate  $RGDR_{PU}$  using a simplified equation and the adjusted alveolar ventilation  
17 rates of [Fennell and Brown \(2001\)](#). Fennell and Brown adjusted the alveolar ventilation rates to  
18 reflect limited pulmonary uptake of EtO, a phenomenon commonly observed for highly  
19 water-soluble gases ([Johanson and Filser, 1992](#)). The adjusted ventilation rates were then used  
20 by Fennell and Brown in their PBPK modeling simulations, and good fits to blood concentration  
21 data were reported for both the mouse and human models. In this document, the adjusted  
22 alveolar ventilation rates were used to estimate the  $RGDR_{PU}$  as follows:

$$23 \quad 24 \quad 25 \quad RGDR_{PU} = (RGD_{PU})_m / (RGD_{PU})_h = (Q_{alv} / SA_{PU})_m / (Q_{alv} / SA_{PU})_h, \quad (4-4)$$

26 where:

27  $RGD_{PU}$  = regional gas dose to the pulmonary region,  
28  $Q_{alv}$  = (adjusted) alveolar ventilation rate,  
29  $SA_{PU}$  = surface area of the pulmonary region, and  
30 the subscripts "m" and "h" denote mouse and human values.

31  
32  
33 Then, using adjusted alveolar ventilation rates from [Fennell and Brown \(2001\)](#) and surface area  
34 values from EPA [[U.S. EPA \(1994\)](#), p. 4–26],

$$35 \quad 36 \quad 37 \quad RGDR_{PU} = ((0.78 \text{ L/h}) / (0.05 \text{ m}^2)) / ((255 \text{ L/h}) / (54.0 \text{ m}^2)) = 3.3. \quad (4-5)$$

*This document is a draft for review purposes only and does not constitute Agency policy.*

1  
2  
3 Using this value for the  $RGDR_{PU}$  would increase the human equivalent concentration about  
4 threefold, resulting in a decreased risk for lung tumors of about threefold, as a lower bound. The  
5 true value of the  $RGDR_{PU}$  is expected to be between 1 and 3, and any adjustment to the lung  
6 tumor risks would still be expected to result in unit risk estimates roughly within the range of the  
7 rodent unit risk estimates derived later in Section 4.2 under the assumption of ppm equivalence.  
8

### 9 **4.2.3. Dose-Response Modeling Methods**

10 In this document the following steps were used:

11 1. *Extract the incidence data presented in the original studies.* In order to crudely adjust  
12 for early mortality in the analysis of the [NTP \(1987\)](#) data, the incidence data have been corrected  
13 for a specific tumor type by eliminating the animals that died prior to the occurrence of the first  
14 tumor or prior to 52 weeks, whichever was earlier. It was not possible to make this adjustment  
15 with the other studies where data on individual animals were not available. With these  
16 exceptions, the tumor incidence data in Tables 3-3 through 3-5 match the original data.

17 2. *Fit the multistage model to the dose-response data using the `Tox_Risk` program.*

18 The likelihood-ratio test was used to determine the lowest value of the multistage  
19 polynomial degree that provided the best fit to the data while requiring selection of the most  
20 parsimonious model. In this procedure, if a good fit to the data in the neighborhood of the POD  
21 is not obtained with the multistage model because of a nonmonotonic reduction in risk at the  
22 highest dose tested (as sometimes occurs when there is early mortality from other causes), that  
23 data point is eliminated and the model is fit again to the remaining data. Such a deletion was  
24 found necessary in two cases (mammary tumors in the NTP study and mononuclear cell  
25 leukemia in the Lynch study). The goodness-of-fit measures for the dose-response curves and  
26 the parameters derived from them are shown in Appendix G.

27 In the NTP bioassay, where the individual animal data were available, a time-to-tumor  
28 analysis was undertaken to account for early mortality. The general model used in this analysis  
29 is the multistage Weibull model:

30  
31  
32 
$$P(d,t) = 1 - \exp[-(q_0 + q_1d + q_2d^2 + \dots + q_kd^k) \times (t - t_0)^z], \quad (4-6)$$
  
33  
34

35 where  $P(d,t)$  represents the probability of a tumor by age  $t$  (in bioassay weeks) for dose  $d$  (i.e.,  
36 human equivalent exposure), and the parameter ranges are restricted as follows:  $z \geq 1$ ,  $t_0 \geq 0$ ,  
37 and  $q_i \geq 0$  for  $i = 0, 1, \dots, k$ . The parameter  $t_0$  represents the time between when a potentially

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 fatal tumor becomes observable and when it causes death. The analyses were conducted using  
2 the computer software Tox\_Risk version 3.5, which is based on methods developed by [Krewski](#)  
3 [et al. \(1983\)](#). Parameters are estimated in Tox\_Risk using the method of maximum likelihood.

4 Tumor types can be categorized by tumor context as either fatal or incidental. Incidental  
5 tumors are those tumors thought not to have caused the death of an animal, whereas fatal tumors  
6 are thought to have resulted in animal death. Tumors at all sites were treated as incidental  
7 (although it was recognized that this may not have been the case, the experimental data are not  
8 detailed enough to conclude otherwise). The parameter  $t_0$  was set equal to 0 because there were  
9 insufficient data to reliably estimate it.

10 The likelihood-ratio test was used to determine the lowest value of the multistage  
11 polynomial degree  $k$  that provided the best fit to the data while requiring selection of the most  
12 parsimonious model. The one-stage Weibull (i.e.,  $k = 1$ ) was determined to be the most optimal  
13 value for all the tumor types analyzed.

14 3. *Select the POD and calculate the unit risk for each tumor site.* The effective  
15 concentration that causes a 10% extra risk for tumor incidence,  $EC_{10}$ , and the 95% lower bound  
16 of that concentration,  $LEC_{10}$ , are derived from the dose-response model. The  $LEC_{10}$  is then used  
17 as the POD for a linear low-dose extrapolation, and the unit risk is calculated as  $0.1/LEC_{10}$ . This  
18 is the procedure specified in the EPA's *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA,](#)  
19 [2005a](#)) for agents such as EtO that have direct mutagenic activity. See Section 3.4 for a  
20 discussion of the mode of action for EtO. Tables 3-3 through 3-5 present the unit risk estimates  
21 for the individual tumor sites in each bioassay.

22 4. *Develop a unit risk estimate based on the incidence of all tumors combined.* This  
23 method assumes that occurrences of tumors at multiple sites are independent and, further, that  
24 the risk estimate for each tumor type is normally distributed. Then, at a given exposure level, the  
25 MLEs of extra risk due to each tumor type are added to obtain the MLE of total cancer risk. The  
26 variances corresponding to each tumor type are added to give the variance associated with the  
27 sum of the MLEs. The one-sided 95% UCL of the MLE for the combined risk is then calculated  
28 as:

$$95\% \text{ UCL} = \text{MLE} + 1.645(\text{SE}), \quad (4-7)$$

29  
30  
31  
32  
33  
34 where SE is the standard error and is the square root of the summed variance. (Note that as a  
35 precursor to this step, when Tox\_Risk is used to fit the incidence of a single tumor type, it  
36 provides the MLE and 95% UCL of extra risk at a specific dose. The standard error in the MLE

1 is determined using the above formula). The calculation is repeated for a few exposure levels,  
 2 and the exposure yielding a value of 0.1 for the upper bound on extra risk is determined by  
 3 interpolation. The unit risk is then the slope of the linear extrapolation from this POD. The  
 4 results are given in Table 4-16.

5  
 6 **Table 4-16. Upper-bound unit risks (per  $\mu\text{g}/\text{m}^3$ ) obtained by combining**  
 7 **tumor sites**  
 8

Combination method <sup>a</sup>	<a href="#">NTP (1987)</a> female mouse	<a href="#">Lynch et al. (1984b),</a> <a href="#">Lynch et al. (1984a)</a> male rat	<a href="#">Snellings et al. (1984)<sup>b</sup></a>	
			Male rat	Female rat
UCL on sum of risks <sup>c</sup>	$2.71 \times 10^{-5}$	$4.17 \times 10^{-5}$	$2.19 \times 10^{-5}$	$3.37 \times 10^{-5}$
Sum of unit risks <sup>d</sup>	$4.12 \times 10^{-5}$	$3.66 \times 10^{-5}$	$2.88 \times 10^{-5}$	$3.54 \times 10^{-5}$
Time-to-tumor analysis and u.c.b on sum of risks <sup>c</sup>	$4.55 \times 10^{-5}$	–	–	–

9  
 10 <sup>a</sup>Unit risk in these methods is the slope of the straight line extrapolation from a point of departure at the dose  
 11 corresponding to a value of 0.1 for the 95% upper confidence bound on total extra risk.

12 <sup>b</sup>Includes data on brain tumors from the analysis by [Garman et al. \(1985\)](#). See Table 3-3.

13 <sup>c</sup>UCL = 95% upper confidence bound. At a given dose, the MLE of the combined extra risk was determined by  
 14 summing the MLE of risk due to each tumor type. The variance associated with this value was determined by  
 15 summing over the variances due to each tumor type.

16 <sup>d</sup>Sum of values in last column of Tables 3-1 through 3-3.  
 17  
 18

19 **4.2.4. Description of Experimental Animal Studies**

20 [NTP \(1987\)](#) exposed male and female B6C3F<sub>1</sub> mice to concentrations of 0, 50, and  
 21 100 ppm for 6 hours per day, 5 days per week, for 102 weeks. An elevated incidence of lung  
 22 carcinomas was found in males, and elevated lung carcinomas, malignant lymphomas, uterine  
 23 adenocarcinomas, and mammary carcinomas were found in females. These data are shown in  
 24 Table 3-3.

25 Lynch et al. ([Lynch et al., 1984a](#); [Lynch et al., 1984b](#)) exposed male F344 rats to 0, 50,  
 26 and 100 ppm for 7 hours per day, 5 days per week, for 2 years. They found excess incidence of  
 27 tumors at three sites: mononuclear cell leukemia in the spleen, testicular peritoneal  
 28 mesothelioma, and brain glioma. In this study the survival in the high-dose group (19%) was  
 29 less than that of controls (49%), which reduced the incidence of leukemias. In the animals in the  
 30 high-dose group that survived to the termination of the experiment, the incidence of leukemias  
 31 was statistically significantly higher than for controls ( $p < 0.01$ ). The incidence data are shown  
 32 in Table 3-4, uncorrected for the high-dose-group mortality. If the individual animal data were

1 available to perform the correction, the incidence would be higher. Therefore, using these data  
2 results in an underestimate of risk.

3 [Snellings et al. \(1984\)](#) exposed male and female F344 rats to 0, 10, 33, and 100 ppm for  
4 6 hours per day, 5 days per week, for 2 years and described their results for all sites except the  
5 brain. In two subsequent publications for the same study, ([Garman et al., 1986, 1985](#)) described  
6 the development of brain tumors in a different set of F344 rats. The [Snellings et al. \(1984\)](#)  
7 publication reported an elevated incidence of splenic mononuclear cell leukemia and peritoneal  
8 mesothelioma in males and an elevated incidence of splenic mononuclear cell leukemia in  
9 females. The mortality was higher in the 100-ppm groups than the other three groups for both  
10 males and females. The incidences in the animals killed after 24 months in [Snellings et al.](#)  
11 [\(1984\)](#) are shown in Table 3-5. Table 3-5 also presents the brain tumor incidence data for male  
12 and female rats from the ([Garman et al., 1986, 1985](#)) publications. The brain tumor incidence  
13 was lower than that of the other tumors, particularly the splenic mononuclear cell leukemias.  
14

#### 15 **4.2.5. Results of Data Analysis of Experimental Animal Studies**

16 The unit risks calculated from the individual site-sex-bioassay data sets are presented in  
17 Tables 3-3 through 3-5. The highest unit risk of any individual site is  $3.23 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$ ,  
18 which is for mononuclear cell leukemia in the female rats of the [Snellings et al. \(1984\)](#) study.

19 Table 4-17 presents the results of the time-to-tumor method applied to the individual  
20 animals in the NTP bioassay, compared with the results from the dose group incidence data in  
21 Table 3-3. This comparison was done for each tumor type separately. The time-to-tumor  
22 method of analyzing the individual animals results in generally higher unit risk estimates than  
23 does the analysis of dose group data, as shown in Table 4-17. The ratio is not large (less than  
24 2.2) across the tumor types. (In the case of mammary tumors this ratio is actually less than 1. It  
25 must be noted that the incidence at the highest dose [where the incidence was substantially less  
26 than at the intermediate dose] was deleted from the analysis of grouped data, whereas it was  
27 retained in the time-to-tumor analysis. Therefore, the comparison for the mammary tumors is  
28 not a strictly valid comparison of methods.) The results also show the extent to which a time-to-  
29 tumor analysis of individual animal data increases the risk estimated from data on dose groups.  
30 It is expected that if individual animal data were available for the [Lynch et al. \(1984a\)](#) and [Lynch](#)  
31 [et al., 1984b](#)) and the [Snellings et al. \(1984\)](#) bioassays, then the time-to-tumor analysis would  
32 also result in higher estimates because both those studies also showed early mortality in the  
33 highest dose group.

34 The results of combining tumor types are summarized in Table 4-16. The sums of the  
35 individual unit risks tabulated in Tables 3-3 to 3-5 are given in the second row of Table 4-16.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 **Table 4-17. Unit risk values from multistage Weibull<sup>a</sup> time-to-tumor**  
 2 **modeling of mouse tumor incidence in the [NTP \(1987\)](#) study**  
 3

Tumor type	Unit risk, 0.1/LEC <sub>10</sub> (per µg/m <sup>3</sup> ) from time-to-tumor analysis	Unit risk, 0.1/LEC <sub>10</sub> (per µg/m <sup>3</sup> ) (Table 3-3) <sup>b</sup>	Ratio of unit risks time-to- tumor/grouped data
<b>Males</b>			
Lung: alveolar/bronchiolar adenoma and carcinoma	3.01 × 10 <sup>-5</sup>	2.22 × 10 <sup>-5</sup>	1.4
<b>Females</b>			
Lung: alveolar/bronchiolar adenoma and carcinoma	2.40 × 10 <sup>-5</sup>	1.10 × 10 <sup>-5</sup>	2.2
Malignant lymphoma	1.43 × 10 <sup>-5</sup>	7.18 × 10 <sup>-6</sup>	2.0
Uterine carcinoma	6.69 × 10 <sup>-6</sup>	4.33 × 10 <sup>-6</sup>	1.5
Mammary carcinoma	8.69 × 10 <sup>-6</sup>	1.87 × 10 <sup>-5</sup>	0.5

4  
 5 <sup>a</sup>P(d,t) = 1 - exp[-(q<sub>0</sub> + q<sub>1</sub>d + q<sub>2</sub>d<sup>2</sup> + ... + q<sub>k</sub>d<sup>k</sup>) × (t - t<sub>0</sub>)<sup>2</sup>], where d is inhaled ethylene oxide concentration in ppm, t  
 6 is weeks until death with tumor. In all cases, k = 1 provided the optimal model.

7 <sup>b</sup>Incidence data modeled using multistage model without taking time to tumor into account.  
 8  
 9

10 Note that as expected they are greater than the unit risks computed from the upper bound on the  
 11 sum of risks for all data sets except for the Lynch et al. ([Lynch et al., 1984a](#); [Lynch et al., 1984b](#))  
 12 data. The reason for this exception is not known, but the differences are small. It is likely that  
 13 the problem arises from the methodology used to combine the risks across tumor sites. In an  
 14 attempt to be consistent with the new two-step methodology (i.e., modeling in the observable  
 15 range to a POD and then doing a linear extrapolation to zero extra risk at zero exposure), the  
 16 exposure concentration at which the sum of the independent tumor site risks yielded a 95% upper  
 17 bound on 10% extra risk was estimated and used as the POD. Summing risks in this way results  
 18 in a POD for the combined tumor risk that is different (lower) than the points of departure for  
 19 each individual tumor site risk. Thus, the risk estimate for the sum is not strictly comparable to  
 20 the individual risks that constitute it. These tumor-site-specific risks were based on points of  
 21 departure individually calculated to correspond with a 10% extra risk. In any event, adding the  
 22 upper bound risks of individual tumor sites should overestimate the upper bound of the sum, and  
 23 the latter is the preferred measure of the total cancer risk because it avoids the overestimate.  
 24 However, for the exceptional Lynch et al. ([Lynch et al., 1984a](#); [Lynch et al., 1984b](#)) data, the  
 25 sum of upper bounds, 3.66 × 10<sup>-5</sup> per µg/m<sup>3</sup>, is already an overestimate of the total risk, and this  
 26 value is preferred over the anomalously high value of 4.17 × 10<sup>-5</sup> per µg/m<sup>3</sup> corresponding to the  
 27 upper bound on the sum of risks. The latter value is considered to be an excessive overestimate

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 and is therefore not carried over into the summary Table 4-18. For the [Snellings et al. \(1984\)](#)  
 2 data sets, the upper confidence bound on the sum of risks is used in the summary Table 4-18.  
 3 The results of the sum-of-risks calculations on the NTP bioassay time-to-tumor data are included  
 4 in the third row of Table 4-16. The estimate for the NTP female mice is  $4.55 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$ ,  
 5 which is higher than the other two measures of total tumor risk in that bioassay. This value is  
 6 preferable to the other measures because it utilizes the individual animal data available for that  
 7 bioassay.

8 Summary of results. The summary of unit risks from the five data sets is shown in  
 9 Table 4-18. The data set giving the highest risk ( $4.55 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$ ) is the [NTP \(1987\)](#) data  
 10 on combined tumors in female mice. The other values are within about a factor of 2 of the  
 11 highest value.

12  
 13 **Table 4-18. Summary of unit risk estimates (per  $\mu\text{g}/\text{m}^3$ ) in animal bioassays**  
 14

Assay	Males	Females
<a href="#">NTP (1987)</a> , B6C3F <sub>1</sub> mice	$3.01 \times 10^{-5}$ <sup>a</sup>	$4.55 \times 10^{-5}$ <sup>b</sup>
<a href="#">Lynch et al. (1984b)</a> , <a href="#">Lynch et al. (1984a)</a> , F344 rats	$3.66 \times 10^{-5}$ <sup>c</sup>	–
<a href="#">Snellings et al. (1984)</a> , F344 rats	$2.19 \times 10^{-5}$ <sup>d</sup>	$3.37 \times 10^{-5}$ <sup>d</sup>

15  
 16 <sup>a</sup>From time-to-tumor analysis of lung adenomas and carcinomas, Table 4-17.

17 <sup>b</sup>Upper bound on sum of risks from the time-to-tumor analysis of the NTP data, Table 4-16.

18 <sup>c</sup>Sum of (upper bound) unit risks (see text for explanation), Table 4-16.

19 <sup>d</sup>Upper bound on sum of risks, Table 4-16.  
 20  
 21

22 **4.3. SUMMARY OF INHALATION UNIT RISK ESTIMATES—NOT ACCOUNTING**  
 23 **FOR ASSUMED INCREASED EARLY-LIFE SUSCEPTIBILITY**

24 For both humans and laboratory animals, tumors occur at multiple sites. In humans, there  
 25 was a combination of tumors having lymphohematopoietic, in particular lymphoid, origins in  
 26 both sexes and breast cancer in females, and, in rodents, lymphohematopoietic tumors, mammary  
 27 carcinomas, and tumors of other sites were observed. From human data, an extra cancer unit risk  
 28 estimate of  $4.79 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  ( $8.77 \times 10^{-4}$  per ppb) was calculated for lymphoid cancer  
 29 incidence, and a unit risk estimate of  $9.31 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  ( $1.74 \times 10^{-3}$  per ppb) was calculated  
 30 for breast cancer incidence in females. The total extra cancer unit risk estimate was  $1.2 \times 10^{-3}$   
 31 per  $\mu\text{g}/\text{m}^3$  ( $2.3 \times 10^{-3}$  per ppb) for both cancer types combined ( $\text{EC}_{01} = 0.0078$  ppm;  
 32  $\text{LEC}_{01} = 0.0044$  ppm). Unit risk estimates derived from the three chronic rodent bioassays for

1 EtO ranged from  $2.2 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$  to  $4.6 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$ , over an order of magnitude lower  
2 than the estimates based on human data.

3 Adequate human data, if available, are considered to provide a more appropriate basis  
4 than rodent data for estimating human risks ([U.S. EPA, 2005a](#)), primarily because uncertainties  
5 in extrapolating quantitative risks from rodents to humans are avoided. Although there is a  
6 sizeable difference between the rodent-based and the human-based estimates, the human data are  
7 from a large, high-quality study, with EtO exposure estimates for the individual workers and  
8 little reported exposure to chemicals other than EtO. Therefore, the total extra cancer unit risk  
9 estimate of  $1.2 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$  ( $2.3 \times 10^{-3}$  per ppb) calculated for lymphoid cancers and breast  
10 cancer combined is the preferred estimate of those estimates not taking assumed increased early-  
11 life susceptibility into account (estimates accounting for assumed increased early-life  
12 susceptibility are presented in Section 4.4). The unit risk estimate is intended to be an upper  
13 bound on cancer risk for use with exposures below the POD (i.e., the  $\text{LEC}_{01}$ ). The unit risk  
14 estimate should not generally be used above the POD; however, in the case of this total extra  
15 cancer unit risk, which is based on cancer type-specific unit risk estimates from two linear  
16 models, the estimate should be valid for exposures up to about 0.075 ppm ( $140 \mu\text{g}/\text{m}^3$ ), which is  
17 the minimum of the limits for the lymphoid cancer unit risk estimate (0.090 ppm; see  
18 Section 4.1.1.2) and the breast cancer unit risk estimate (0.075 ppm; see Section 4.1.2.3).

19 Because a mutagenic mode of action for EtO carcinogenicity (see Section 3.3.2) is  
20 “sufficiently supported in (laboratory) animals” and “relevant to humans,” and as there are no  
21 chemical-specific data to evaluate the differences between adults and children, increased  
22 early-life susceptibility should be assumed and, if there is early-life exposure, the age-dependent  
23 adjustment factors (ADAFs) should be applied, as appropriate, in accordance with EPA’s  
24 *Supplemental Guidance* ([U.S. EPA, 2005b](#)) see Section 4.4 below for more details on the  
25 application of ADAFs).

#### 27 **4.4. ADJUSTMENTS FOR POTENTIAL INCREASED EARLY-LIFE** 28 **SUSCEPTIBILITY**

29 There are no chemical-specific data on age-specific susceptibility to EtO-induced  
30 carcinogenesis. However, there is sufficient weight of evidence to conclude that EtO operates  
31 through a mutagenic mode of action (see Section 3.4.1). In such circumstances (i.e., the absence  
32 of chemical-specific data on age-specific susceptibility but sufficient evidence of a mutagenic  
33 mode of action), EPA’s *Supplemental Guidance for Assessing Susceptibility from Early-Life*  
34 *Exposure to Carcinogens* ([U.S. EPA, 2005b](#)) recommends the assumption of increased early-life  
35 susceptibility and the application of default age-dependent adjustment factors (ADAFs) to adjust

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 for this potential increased susceptibility from early-life exposure. See the *Supplemental*  
2 *Guidance* for detailed information on the general application of these adjustment factors. In  
3 brief, the *Supplemental Guidance* establishes ADAFs for three specific age groups. The current  
4 ADAFs and their age groupings are 10 for <2 years, 3 for 2 to <16 years, and 1 for 16 years and  
5 above ([U.S. EPA, 2005b](#)). For risk assessments based on specific exposure assessments, the  
6 10-fold and 3-fold adjustments to the unit risk estimates are to be combined with age-specific  
7 exposure estimates when estimating cancer risks from early-life (<16 years of age) exposure.

8 These ADAFs, however, were formulated based on comparisons of the ratios of cancer  
9 potency estimates from juvenile-only exposures to cancer potency estimates from adult-only  
10 exposures from rodent bioassay data sets with appropriate exposure scenarios, and they are  
11 designed to be applied to cancer potency estimates derived from adult-only exposures. Thus,  
12 alternate life-table analyses were conducted to derive comparable adult-exposure-only unit risk  
13 estimates to which ADAFs would be applied to account for early-life exposure.<sup>40</sup> For these  
14 alternate life-table analyses, it was assumed that RR is independent of age for adults, which  
15 represent the life stage for which the exposure-response data and the Cox regression modeling  
16 results from the NIOSH cohort study specifically pertain, but that there is increased early-life  
17 susceptibility, based on the weight-of-evidence-based conclusion that EtO carcinogenicity has a  
18 mutagenic mode of action (see Section 3.4), which supersedes the assumption that RR is  
19 independent of age for all ages including children.

20 In the alternate analyses, exposure in the life table was taken to start at age 16 years, the  
21 age cut point that was established in EPA's *Supplemental Guidance* ([U.S. EPA, 2005b](#)), to derive  
22 an adult-exposure-only unit risk estimate to which ADAFs would be applied to account for  
23 early-life exposure. Other than the age at which exposure was initiated, the life-table analyses  
24 are identical to those conducted for the results presented in Section 4.1. Adult-exposure-only  
25 unit risk estimates were derived for both cancer incidence and mortality for both lymphoid and  
26 breast cancers. Alternate estimates were not derived for all lymphohematopoietic cancers  
27 because lymphoid cancer was the preferred endpoint (see Section 4.1.1.2). Incidence estimates  
28 are preferred over mortality estimates, but both are calculated here for comparison and because  
29 mortality estimates are sometimes used in addition to incidence estimates in benefit-cost

---

<sup>40</sup> In this assessment, *adult-exposure-only unit risk estimates* refer to estimates derived from the life-table analysis assuming exposure only for ages  $\geq 16$  years. The adult-exposure-only unit risk estimates are merely intermediate values in the calculation of adult-based unit risk estimates and should not be used in any risk calculations. *Adult-based unit risk estimates* refer to estimates derived after rescaling the adult-exposure-only unit risk estimates to a (70-year) lifetime, as described later in Section 4.4. The adult-based unit risk estimates are intended to be used in ADAF calculations (U.S. EPA, 2005b) for the computation of extra risk estimates for specific exposure scenarios. Note that the unit risk estimates in this section, which are derived under an assumption of increased early-life susceptibility, supersede those that were derived in Section 4.1 under the assumption that RR is independent of age.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 analyses. For each cancer endpoint, the same exposure-response model was used as that which  
 2 was selected for the unit risk estimates in Section 4.1 (i.e., linear regression of the categorical  
 3 results, excluding the highest exposure category, for lymphoid cancer and breast cancer mortality  
 4 and two-piece linear spline model for breast cancer incidence). The results are presented in  
 5 Table 4-19 along with the unit risk estimates derived assuming that RR was independent of age  
 6 for all ages (see Section 4.1) for comparison. As can be seen in Table 4-19, the unit risk  
 7 estimates for adult-only exposures range from about 66% to about 72% of the unit risk estimates  
 8 derived under the assumption of age independence across all ages.

9  
 10 **Table 4-19. EC<sub>01</sub>, LEC<sub>01</sub>, and unit risk estimates for adult-only exposures\***

Cancer response	EC <sub>01</sub> (ppm)	LEC <sub>01</sub> (ppm)	Adult-exposure-only unit risk estimate <sup>a</sup> (per ppm)	Lifetime-exposure unit risk estimate under assumption of age independence <sup>b</sup> (per ppm)
Lymphoid cancer mortality (both sexes)	0.0787	0.0352	0.284	0.397
Lymphoid cancer incidence (both sexes)	0.0364	0.0163	0.613	0.877
Breast cancer mortality (females)	0.0590	0.0297	0.337	0.513
Breast cancer incidence (females)	0.0167	0.00863	1.16 <sup>c</sup>	1.74 <sup>c</sup>

12  
 13 \*These are intermediate values. See Table 4-22 below for the final adult-based cancer-type-specific unit risk  
 14 estimates.

15 <sup>a</sup>Unit risk estimate = 0.01/LEC<sub>01</sub>.

16 <sup>b</sup>From Tables 4-5, 4-9, and 4-13 of Section 4.1.

17 <sup>c</sup>For unit risk estimates above 1, convert to risk per ppb (e.g., 1.16 per ppm = 1.16 × 10<sup>-3</sup> per ppb).

18  
 19  
 20 According to EPA's *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#))  
 21 cancer risk estimates are intended to reflect total cancer risk, not site-specific cancer risk;  
 22 therefore, an additional calculation was made to estimate the combined risk for (incident)  
 23 lymphoid and breast cancers from adult-only exposures, because females would be at risk for  
 24 both cancer types. Assuming that the tumor types are independent and that the risk estimates are  
 25 approximately normally distributed, this calculation can be made as described in Section 4.1.3.  
 26 First, an EC<sub>01</sub> of 0.0114 ppm for the total cancer risk (i.e., lymphoid cancer incidence + breast  
 27 cancer incidence) from adult-only exposure was estimated, as summarized in Table 4-20.

28 Then, a unit risk estimate of 1.5 per ppm for the total cancer risk (i.e., lymphoid cancer  
 29 incidence + breast cancer incidence) from adult-only exposure was derived, as shown in

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 Table 4–21. An LEC<sub>01</sub> estimate of 0.00654 ppm for the total cancer risk can be calculated as  
 2 0.01/(1.53 per ppm).

3  
 4 **Table 4-20. Calculation of EC<sub>01</sub> for total cancer (incidence) risk from adult-**  
 5 **only exposure\***  
 6

Cancer type	EC <sub>01</sub> (ppm)	0.01/EC <sub>01</sub> (per ppm)	EC <sub>01</sub> for total risk (ppm)
Lymphoid	0.0364	0.275	--
Breast	0.0167	0.599	--
Total <sup>a</sup>	--	0.874	0.0114

7  
 8 \*These are intermediate values. See Table 4-23 below for the final adult-based cancer-type-  
 9 specific 0.01/EC<sub>01</sub> estimates.

10 <sup>a</sup>The total 0.01/EC<sub>01</sub> value equals the sum of the individual 0.01/EC<sub>01</sub> values; the EC<sub>01</sub> for the  
 11 total cancer risk then equals 0.01/(0.01/EC<sub>01</sub>).

12  
 13  
 14 **Table 4-21. Calculation of total cancer unit risk estimate from adult-only**  
 15 **exposure\***  
 16

Cancer type	Adult-exposure- only unit risk estimate (per ppm)	0.01/EC <sub>01</sub> (per ppm)	SE <sup>a</sup> (per ppm)	Variance	Adult-exposure-only total cancer unit risk estimate (per ppm)
Lymphoid	0.613	0.275	0.205	0.0422	--
Breast	1.16	0.599	0.340	0.115	--
Total	--	0.874	(0.397) <sup>b</sup>	0.158	1.53 <sup>c</sup>

17  
 18 \*These are intermediate values. See Table 4-22 below for the final adult-based cancer-type-specific unit risk  
 19 estimates.

20 <sup>a</sup>SE = (unit risk – 0.01/EC<sub>01</sub>)/1.645.

21 <sup>b</sup>The SE of the total cancer risk is calculated as the square root of the sum of the variances (next column), not as the  
 22 sum of the SEs.

23 <sup>c</sup>Total cancer unit risk = 0.874 + 1.645 × 0.397.

24  
 25  
 26 Thus, the total cancer unit risk estimate from adult-only exposure is 1.53 per ppm (or  
 27  $1.53 \times 10^{-3}$  per ppb;  $8.36 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$ ). While there are uncertainties regarding the  
 28 assumption of a normal distribution of risk estimates, the resulting unit risk estimate is  
 29 appropriately bounded in the roughly twofold range between estimates based on the sum of the  
 30 individual MLEs (i.e., 0.874) and the sum of the individual 95% UCLs (i.e., unit risk estimates,  
 31 1.77), or more precisely in this case, between the largest individual unit risk estimate (1.16) and

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 the sum of the unit risk estimates (1.77), and thus, any inaccuracy in the total cancer risk estimate  
2 resulting from the approach used to combine risk estimates across cancer types is relatively  
3 minor.

4 When EPA derives unit risk estimates from rodent bioassay data, there is a blurring of the  
5 distinction between lifetime and adult-only exposures because the relative amount of time that a  
6 rodent spends as a juvenile is negligible (<8%) compared to its lifespan. [According to EPA's  
7 *Supplemental Guidance*, puberty begins around 5–7 weeks of age in rats and around 4–6 weeks  
8 in mice ([U.S. EPA, 2005b](#))]. Thus, when exposure in a rodent is initiated at 5–8 weeks, as in the  
9 typical rodent bioassay, and the bioassay is terminated after 104 weeks of exposure, the unit risk  
10 estimate derived from the resulting cancer incidence data is considered a unit risk estimate from  
11 lifetime exposure, except when the ADAFs were formulated and are applied, in which case the  
12 same estimate is considered to apply to adult-only exposure. Yet, when adult exposures are  
13 considered in the application of ADAFs, the adult-exposure-only unit risk estimate is pro-rated  
14 over the full default (average) human lifespan of 70 years, presumably because that is how adult  
15 exposures are treated when a unit risk estimate calculated in the same manner from the same  
16 bioassay exposure paradigm is taken as a lifetime unit risk estimate.

17 However, in humans, a greater proportion of time is spent in childhood (e.g., 16 of  
18 70 years = 23%), and the distinction between lifetime exposure and adult-only exposure cannot  
19 be ignored when human data are used as the basis for the unit risk estimates. Thus, as described  
20 above, adult-exposure-only unit risk estimates were calculated distinct from the lifetime  
21 estimates that were derived in Section 4.1 under the assumption of age independence for all ages.  
22 In addition, the adult-exposure-only unit risk estimates need to be rescaled to a 70-year lifespan  
23 in order to be used in the ADAF calculations and risk estimate calculations involving less-than-  
24 lifetime exposure scenarios in the standard manner, which includes prorating even adult-based  
25 unit risk estimates over 70 years. Thus, the adult-exposure-only unit risk estimates are  
26 multiplied by 70/54 to rescale the 54-year adult period of the 70-year default lifespan to 70 years.  
27 Then, for example, if a risk estimate were calculated for a less-than-lifetime exposure scenario  
28 involving exposure only for the full adult period of 54 years, the rescaled unit risk estimate  
29 would be multiplied by 54/70 in the standard calculation and the adult-exposure-only unit risk  
30 estimate would be appropriately reproduced. Without rescaling the adult-exposure-only unit risk  
31 estimates, the example calculation just described for exposure only for the full adult period of  
32 54 years would result in a risk estimate 77% (i.e., 54/70) of that obtained directly from the  
33 adult-exposure-only unit risk estimates, which would be illogical. The rescaled adult-based unit  
34 risk estimates for use in ADAF calculations and risk estimate calculations involving less-than-

lifetime exposure scenarios are presented in Table 4-22. Rescaled LEC<sub>01</sub> and EC<sub>01</sub> estimates for adult-based total cancer risk are  $5.0 \times 10^{-3}$  ppm ( $9.2 \mu\text{g}/\text{m}^3$ ) and  $8.8 \times 10^{-3}$  ppm ( $16 \mu\text{g}/\text{m}^3$ ).

**Table 4-22. Adult-based unit risk estimates for use in ADAF calculations and risk estimate calculations involving less-than-lifetime exposure scenarios**

Cancer response	Adult-based unit risk estimate (per ppm)	Adult-based unit risk estimate (per $\mu\text{g}/\text{m}^3$ )
<b>Preferred Models<sup>a</sup></b>		
Lymphoid cancer mortality	0.368	$2.01 \times 10^{-4}$
Lymphoid cancer incidence	0.795	$4.35 \times 10^{-4}$
Breast cancer mortality	0.436	$2.39 \times 10^{-4}$
Breast cancer incidence	1.50 <sup>b</sup>	$8.21 \times 10^{-4}$
Total cancer incidence	<b>1.98<sup>b</sup></b>	<b><math>1.08 \times 10^{-3}</math></b>
<b>Alternative Models</b>		
Breast cancer incidence <sup>c</sup>	0.386	$2.11 \times 10^{-4}$
Total cancer incidence <sup>d</sup>	1.03 <sup>b</sup>	$5.64 \times 10^{-4}$
Lymphoid cancer incidence <sup>e</sup>	1.56 <sup>b</sup>	$8.53 \times 10^{-4}$
Total cancer incidence <sup>f</sup>	2.56 <sup>b</sup>	$1.40 \times 10^{-3}$

<sup>a</sup>Linear regression of categorical results for lymphoid cancer and breast cancer mortality; two-piece linear spline model for breast cancer incidence.

<sup>b</sup>For unit risk estimates above 1, convert to risk per ppb (e.g., 1.16 per ppm =  $1.16 \times 10^{-3}$  per ppb).

<sup>c</sup>Linear RR model for breast cancer incidence, representing the other (low) end of the range of reasonable alternatives to the two-piece linear spline model, which is at the high end of the range.

<sup>d</sup>With preferred lymphoid cancer incidence model but alternative linear RR model for breast cancer incidence, representing the low end of the range of reasonable estimates for total cancer risk.

<sup>e</sup>Two-piece log-linear spline model with knot at 1,600 ppm  $\times$  days, an alternative model for lymphoid cancer based on continuous exposure data.

<sup>f</sup>With preferred breast cancer incidence model but alternative two-piece log-linear spline model with knot at 1,600 ppm  $\times$  days for lymphoid cancer incidence.

An example calculation illustrating the application of the ADAFs to the human-data-derived adult-based (rescaled as discussed above) unit risk estimate for EtO for a lifetime exposure scenario is presented below. For inhalation exposures, assuming ppm equivalence across age groups, i.e., equivalent risk from equivalent exposure levels, independent of body size, the ADAF calculation is fairly straightforward. Thus, the ADAF-adjusted lifetime total-cancer unit risk estimate is calculated as follows:

*This document is a draft for review purposes only and does not constitute Agency policy.*

total cancer risk from exposure to constant EtO exposure level of  $1 \mu\text{g}/\text{m}^3$  from ages 0–70 years:

<u>Age group</u>	<u>ADAF</u>	<u>unit risk (per <math>\mu\text{g}/\text{m}^3</math>)</u>	<u>exposure conc (<math>\mu\text{g}/\text{m}^3</math>)</u>	<u>duration adjustment</u>	<u>partial risk</u>
0 to <2 years	10	$1.08 \times 10^{-3}$	1	2 years/70 years	$3.09 \times 10^{-4}$
2 to <16 years	3	$1.08 \times 10^{-3}$	1	14 years/70 years	$6.48 \times 10^{-4}$
$\geq 16$ years	1	$1.08 \times 10^{-3}$	1	54 years/70 years	<u><math>8.33 \times 10^{-4}</math></u>
				<i>total lifetime risk =</i>	$1.80 \times 10^{-3}$

The partial risk for each age group is the product of the values in columns 2–5 [e.g.,  $10 \times (1.08 \times 10^{-3}) \times 1 \times 2/70 = 3.09 \times 10^{-4}$ ], and the total risk is the sum of the partial risks.

This 70-year risk estimate for a constant exposure of  $1 \mu\text{g}/\text{m}^3$  is equivalent to a **lifetime unit risk estimate of  $1.8 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$**  (3.3 per ppm, or  $3.3 \times 10^{-3}$  per ppb), adjusted for potential increased early-life susceptibility, assuming a 70-year lifetime and constant exposure across age groups. Note that because of the use of the rescaled adult-based unit risk estimate, the partial risk for the  $\geq 16$  years age group is the same as would be obtained for a  $1 \mu\text{g}/\text{m}^3$  constant exposure directly from the total cancer adult-exposure-only unit risk estimate of  $8.36 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  that was presented above, as it should be (the small difference in the second decimal place is due to round-off error).

In addition to the uncertainties discussed above for the inhalation unit risk estimate, there are uncertainties in the application of ADAFs to adjust for potential increased early-life susceptibility. The ADAFs reflect an expectation of increased risk from early-life exposure to carcinogens with a mutagenic mode of action ([U.S. EPA, 2005b](#)), but they are general adjustment factors and are not specific to EtO. With respect to the breast cancer estimates, for example, evidence suggests that puberty/early adulthood is a particularly susceptible life stage for breast cancer induction ([U.S. EPA, 2005b](#); [Russo and Russo, 1999](#)); however, EPA has not, at this time, developed alternate ADAFs to reflect such a pattern of increased early-life susceptibility, and there is currently no EPA guidance on an alternate approach for adjusting for early-life susceptibility to potential breast carcinogens.

#### 4.5. INHALATION UNIT RISK ESTIMATES—CONCLUSIONS

For both humans and laboratory animals, tumors occur at multiple sites. In humans, there was a combination of tumors having lymphohematopoietic, in particular lymphoid, origins in both sexes and breast cancer in females, and, in rodents, lymphohematopoietic tumors, mammary carcinomas, and tumors of other sites were observed. From human data, an extra cancer unit risk

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 estimate of  $4.79 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  ( $8.77 \times 10^{-4}$  per ppb) was calculated for lymphoid cancer  
2 incidence, and a unit risk estimate of  $9.49 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  ( $1.74 \times 10^{-3}$  per ppb) was calculated  
3 for breast cancer incidence in females, under the assumption that RR is independent of age for all  
4 ages (see Section 4.1). The total extra cancer unit risk estimate was  $1.24 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$   
5 ( $2.27 \times 10^{-3}$  per ppb) for both cancer types combined ( $\text{EC}_{01} = 0.00775$  ppm;  $\text{LEC}_{01} =$   
6  $0.00441$  ppm). Unit risk estimates derived from the three chronic rodent bioassays for EtO  
7 ranged from  $2.2 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$  to  $4.6 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$ , over an order of magnitude lower than  
8 the estimates based on human data.

9 Because a mutagenic mode of action for EtO carcinogenicity (see Section 3.4.1) is  
10 “sufficiently supported in (laboratory) animals” and “relevant to humans,” and as there are no  
11 chemical-specific data to evaluate the differences between adults and children, increased  
12 early-life susceptibility should be assumed, in accordance with EPA’s *Supplemental Guidance*  
13 ([U.S. EPA, 2005b](#)). This assumption of increased early-life susceptibility supersedes the  
14 assumption of age independence under which the human-data-based estimates presented in the  
15 previous paragraph were derived. Thus, as described in Section 4.4, adult-exposure-only (i.e.,  
16 ages  $\geq 16$  years) unit risk estimates were calculated from the human data under an alternate  
17 assumption that RR is independent of age for adults, which represent the life stage for which the  
18 data upon which the exposure-response modeling was conducted pertain. These adult-exposure-  
19 only unit risk estimates were then rescaled to a 70-year basis to derive adult-based unit risk  
20 estimates for use in the standard ADAF calculations and risk estimate calculations involving  
21 less-than-lifetime exposure scenarios. The resulting adult-based unit risk estimates were  $4.35 \times$   
22  $10^{-4}$  per  $\mu\text{g}/\text{m}^3$  ( $7.95 \times 10^{-4}$  per ppb) for lymphoid cancer incidence and  $8.21 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$   
23 ( $1.50 \times 10^{-3}$  per ppb) for breast cancer incidence in females. The adult-based total extra cancer  
24 unit risk estimate for use in ADAF calculations and risk estimate calculations involving  
25 less-than-lifetime exposure scenarios was  $1.08 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$  ( $1.98 \times 10^{-3}$  per ppb) for both  
26 cancer types combined.

27 For exposure scenarios involving early-life exposure, the ADAFs should be applied, in  
28 accordance with EPA’s *Supplemental Guidance* ([U.S. EPA, 2005b](#)). Applying the ADAFs to  
29 obtain a full lifetime unit risk estimate yields

$$\begin{aligned} & 1.98/\text{ppm} \times ((10 \times 2 \text{ years}/70 \text{ years}) + (3 \times 14/70) + (1 \times 54/70)) && (4-8) \\ & = 3.29/\text{ppm} = 1.80 \times 10^{-3}/(\mu\text{g}/\text{m}^3). \end{aligned}$$

1 Applying the ADAFs to the unit risk estimates derived from the three chronic rodent bioassays  
2 for EtO yields estimates ranging from  $3.7 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$  to  $7.6 \times 10^{-5}$  per  $\mu\text{g}/\text{m}^3$ , still over an  
3 order of magnitude lower than the estimate based on human data.

4 Adequate human data, if available, are considered to provide a more appropriate basis  
5 than rodent data for estimating human risks ([U.S. EPA, 2005a](#)), primarily because uncertainties  
6 in extrapolating quantitative risks from rodents to humans are avoided. Although there is a  
7 sizeable difference between the rodent-based and the human-based estimates, the human data are  
8 from a large, high-quality study, with EtO exposure estimates for the individual workers and  
9 little reported exposure to chemicals other than EtO. Therefore, the human-based **full lifetime**  
10 **total extra cancer unit risk estimate of  $1.8 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$  ( $3.3 \times 10^{-3}$  per ppb)** calculated  
11 for lymphoid cancers and breast cancer combined and applying the ADAFs is the preferred  
12 lifetime unit risk estimate.<sup>41</sup> For less-than-lifetime exposure scenarios, the human-data-derived  
13 (rescaled) adult-based unit risk estimate of  $1.1 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$  ( $2.0 \times 10^{-3}$  per ppb) should be  
14 used, in conjunction with the ADAFs if early-life exposures occur.

15 Although there are uncertainties in this unit risk estimate, the major ones related to  
16 exposure misclassification, model uncertainty, and low-dose extrapolation, as discussed in  
17 Section 4.1.4, confidence in the unit risk estimate is relatively high. First, there is strong  
18 confidence in the hazard characterization of EtO as “carcinogenic to humans,” which is based on  
19 strong epidemiological evidence supplemented by other lines of evidence, such as genotoxicity  
20 in both rodents and humans (see Section 3.5.1). Second, the unit risk estimate is based on human  
21 data from a large, high-quality epidemiology study with individual worker exposures estimated  
22 using a high-quality regression model (see Section 4.1 and Section A.2.8 of Appendix A).  
23 Finally, the use of low-exposure linear extrapolation is strongly supported by the conclusion that  
24 EtO carcinogenicity has a mutagenic mode of action (see Section 3.4.1).

25 Confidence in the unit risk estimate is particularly high for the breast cancer component,  
26 the largest contributor to the total cancer unit risk estimate, which is based on over 200 incident  
27 cases for which the investigators had information on other potential breast cancer risk factors  
28 (see Section 4.1.2.3). The selected model for the breast cancer incidence data was the  
29 best-fitting model of the models investigated as well as the model that provided the best  
30 representation of the categorical results, particularly in the lower exposure range of greatest

---

<sup>41</sup>Technically, this unit risk estimate reflects the total (upper bound) cancer risk to females and not to the general population because the breast cancer risk applies only to females. As a practical matter for regulatory purposes, however, females comprise roughly half the general population, and this unit risk estimate enables risk managers to evaluate the individual risk for this substantial population group. For the purposes of estimating numbers of cancer cases attributable to specific exposure levels, e.g., for benefits analyses, it would be more appropriate to use the cancer-specific unit risk estimates (or central tendency estimates), taking sex into account.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 relevance for the derivation of a unit risk estimate. Alternate estimates calculated from other  
2 reasonable models suggest that a unit risk estimate for breast cancer incidence that is up to  
3 fourfold lower (corresponding to a total cancer unit risk estimate up to twofold lower) is  
4 plausible; however, unit risk estimates notably lower than that are considered unlikely from the  
5 available data.

6         There is lower confidence in the lymphoid cancer component of the unit risk estimate  
7 because it is based on fewer events (40 lymphoid cancer deaths); incidence risk was estimated  
8 from mortality data; and the exposure-response relationship is exceedingly supralinear, such that  
9 continuous exposure models yield apparently implausibly steep low-exposure slopes (see  
10 Figure 4-1). Although these continuous exposure models provided statistically significant slope  
11 coefficients, there was low confidence in such steep slopes, which, particularly for the two-piece  
12 spline model with the maximum likelihood, are highly dependent on a small number of cases in  
13 the low-exposure range. Thus, a linear regression model of the categorical results for the lowest  
14 three quartiles was used to derive the unit risk estimate for lymphoid cancer, and there was  
15 greater confidence in the more moderate slope resulting from that model, although it was not  
16 statistically significant, because it was based on more data and provided a good representation of  
17 the categorical results across this larger data range in the lower-exposure region (see  
18 Section 4.1.1.2). So, while there is lower confidence in the lymphoid cancer unit risk estimate  
19 than in the breast cancer unit risk estimate, the lymphoid cancer estimate is considered a  
20 reasonable estimate from the available data, and overall, there is relatively high confidence in the  
21 total cancer unit risk estimate. An alternative lymphoid cancer model (two-piece log-linear  
22 spline model with the knot defined by a local maximum likelihood rather than the largest  
23 likelihood), judged to be the most suitable model based on the continuous exposure data, was  
24 also considered for comparison. This alternative model yielded a unit risk estimate for lymphoid  
25 cancer incidence two times that from the linear regression of the categorical results, resulting in a  
26 total cancer unit risk estimate about 28% higher than the total cancer risk estimate with the  
27 preferred lymphoid cancer model.

28         The unit risk estimate is intended to be an upper bound on cancer risk for use with  
29 exposures below the POD (i.e., the  $LEC_{01}$ ). The unit risk estimate should not generally be used  
30 above the POD; however, in the case of this total extra cancer unit risk, which is based on cancer  
31 type-specific unit risk estimates from two linear models, the estimate should be valid for  
32 exposures up to about 0.075 ppm ( $140 \mu\text{g}/\text{m}^3$ ), which is the minimum of the limits for the  
33 lymphoid cancer unit risk estimate (0.090 ppm: see Section 4.1.1.2) and the breast cancer unit  
34 risk estimate (0.075 ppm; see Section 4.1.2.3). (See Section 4.7 for risk estimates based on  
35 occupational exposure scenarios.)

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 Using the above full lifetime unit risk estimate of  $3.3 \times 10^{-3}$  per ppb ( $1.8 \times 10^{-3}$  per  
2  $\mu\text{g}/\text{m}^3$ ), the (lower bound) lifetime chronic exposure level of EtO corresponding to an increased  
3 cancer risk of  $10^{-6}$  can be estimated as follows:

$$(10^{-6})/(3.3/\text{ppm}) = 3.0 \times 10^{-7} \text{ ppm} = 0.00030 \text{ ppb} = 0.0006 \mu\text{g}/\text{m}^3. \quad (4-9)$$

4  
5  
6  
7  
8  
9 The inhalation unit risk estimate presented above, which is calculated based on a linear  
10 extrapolation from the POD ( $\text{LEC}_{01}$ ), is expected to provide an upper bound on the risk of cancer  
11 incidence. For some applications, however, estimates of “central tendency” for the risk below  
12 the POD are desired. Thus, adult-based extra risk estimates per ppm for some of the cancer  
13 responses based on linear extrapolation from the adult-exposure-only  $\text{EC}_{01}$  (i.e.,  $0.01/\text{EC}_{01}$ ), and  
14 rescaled to a 70-year basis for use in ADAF calculations and risk estimate calculations involving  
15 less-than-lifetime exposure scenarios (see Section 4.4), are reported in Table 4-23. The  
16 adult-exposure-only  $\text{EC}_{01}$ s were from the linear regression models of the categorical results for  
17 lymphoid cancers and breast cancer mortality and from the two-piece linear spline model  
18 (low-dose segment) for breast cancer incidence. (Note that, for each of these models, the  
19 low-exposure extrapolated estimates are a straight linear continuation of the linear models used  
20 above the PODs, and thus, the statistical properties of the models are preserved.) These estimates  
21 are dependent on the suitability of the  $\text{EC}_{01}$  estimates as well as on the applicability of the linear  
22 low-dose extrapolation. The assumption of low-dose linearity is supported by the mutagenicity  
23 of EtO (see Section 3.4). If these estimates are to be used, ADAFs should be applied if early-life  
24 exposure occurs, in accordance with EPA’s *Supplemental Guidance* ([U.S. EPA, 2005b](#)).

25 As can be seen by comparing the adult-based  $0.01/\text{EC}_{01}$  estimates in Table 4-23 with the  
26 adult-based unit risk estimates (i.e.,  $0.01/\text{LEC}_{01}$  estimates) in Table 4-22, the  $0.01/\text{EC}_{01}$  estimates  
27 are about 45% of the unit risk estimates for the lymphoid cancer responses and about 50% of the  
28 unit risk estimates for the breast cancer responses.

1 **Table 4-23. Adult-based extra risk estimates per ppm based on**  
 2 **adult-exposure-only EC<sub>01</sub>s (0.01/EC<sub>01</sub> estimates)<sup>a</sup>**  
 3

Cancer response	Adult-exposure-only EC <sub>01</sub> (ppm)	Adult-based 0.01/EC <sub>01</sub> (per ppm) <sup>b,c</sup>
Lymphoid cancer mortality (both sexes)	0.0787	0.165
Lymphoid cancer incidence (both sexes)	0.0364	0.356
Breast cancer mortality (females)	0.0590	0.219
Breast cancer incidence (females)	0.0167	0.776
Total cancer incidence	0.0114	1.14 <sup>d</sup>

4  
 5 <sup>a</sup>ADAFs should be applied to the adult-based 0.01/EC<sub>01</sub> estimates if early-life exposure occurs, in  
 6 accordance with EPA’s *Supplemental Guidance*.

7 <sup>b</sup>These estimates are calculated as 0.01/EC<sub>01</sub> for the adult-exposure-only extra risk estimate per  
 8 ppm rescaled to a 70-yr basis by multiplying by 70/54 (see Section 4.4).

9 <sup>c</sup>For conversion to per µg/m<sup>3</sup>, divide by 1830.

10 <sup>d</sup>For unit risk estimates above 1, convert to risk per ppb (e.g., 1.14 per ppm = 1.14 × 10<sup>-3</sup> per ppb).  
 11  
 12

13 Finally, it should be noted that some investigators have posited that the high and variable  
 14 background levels of endogenous EtO-induced DNA damage in the body (see Section 3.3.3.1)  
 15 may overwhelm any contribution from low levels of exogenous EtO exposure ([Marsden et al.,](#)  
 16 [2009](#); [SAB, 2007](#)). It is true that the existence of these high and variable background levels may  
 17 make it hard to observe statistically significant increases in risk from low levels of exogenous  
 18 exposure. However, there is clear evidence of carcinogenic hazard from the rodent bioassays  
 19 and strong evidence from human studies (see Section 3.5), and the genotoxicity/mutagenicity of  
 20 EtO (see Section 3.4) supports low-dose linear extrapolation of risk estimates from those studies  
 21 ([U.S. EPA, 2005a](#)). In fact, as discussed in Section 3.3.3.1, [Marsden et al. \(2009\)](#), using  
 22 sensitive detection techniques and an approach designed to separately quantify endogenous  
 23 N7-HEG adducts and “exogenous” N7-HEG adducts induced by EtO treatment in rats, reported  
 24 increases in exogenous adducts in DNA of spleen and liver at the lowest dose administered  
 25 (0.0001 mg/kg injected i.p. daily for 3 days, which is a very low dose compared to the LOAELs  
 26 in the carcinogenicity bioassays; see Section C.7 of Appendix C). [Marsden et al. \(2009\)](#) also  
 27 reported statistically significant linear dose-response relationships ( $p < 0.05$ ) for exogenous  
 28 adducts in all three tissues examined (spleen, liver, and stomach), although they caution that  
 29 some of the adduct levels induced at low EtO concentrations are below the limit of accurate  
 30 quantitation. Furthermore, while the contributions to DNA damage from low exogenous EtO  
 31 exposures may be relatively small compared to those from endogenous EtO exposure, low levels  
 32 of exogenous EtO may nonetheless be responsible for additional risk (above background risk).

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 This is not inconsistent with the much higher levels of background cancer risk, to which  
2 endogenous EtO may contribute, for the two cancer types observed in the human  
3 studies—lymphoid cancers have a background lifetime incidence risk on the order of 3%, while  
4 the background lifetime incidence risk for breast cancer is on the order of 15%.<sup>42</sup>

5 Also related to the issue of endogenous EtO, [Starr and Swenberg \(2013\)](#) have proposed  
6 an approach for bounding the cancer risk from low levels of exogenous exposure to chemicals  
7 that also exist endogenously. In brief, [Starr and Swenberg \(2013\)](#) assume that all background  
8 cancer risk ( $P_0$ ) for a specific cancer type is attributable to background levels ( $C_0$ ) of some  
9 endogenous adduct (as a marker of exposure) of the chemical of interest [[Starr and Swenberg](#)  
10 [\(2013\)](#) use formaldehyde as an example] in that tissue; they then use the ratio  $P_0/C_0$  (actually the  
11 lower bound on  $C_0$ ) to estimate a linear slope for risk as a function of endogenous adduct level  
12 down to zero adducts, which they claim is a conservative (upper) bound on cancer risk from low  
13 levels of exogenous exposure (similarly expressed in terms of adduct levels). EPA disagrees that  
14 this approach necessarily yields a conservative bound on risk, however, because, *even if* the  
15 adduct level is an appropriate dose metric for comparing cancer risks, the approach relies on the  
16 assumption that the dose-response relationship over the dose range of endogenous adducts is  
17 linear down to zero adducts. In contrast to this assumption, EPA considers it highly plausible  
18 that the dose-response relationship over the endogenous range is sublinear (e.g., that the baseline  
19 levels of DNA repair enzymes and other protective systems evolved to deal with endogenous  
20 DNA damage would work more effectively for lower levels of endogenous adducts), that is, that  
21 the slope of the dose-response relationship for risk per adduct would increase as the level of  
22 endogenous adducts increases. If the dose-response relationship over the endogenous range is  
23 sublinear, rather than linear as assumed by [Starr and Swenberg \(2013\)](#), then the approach  
24 proposed by [Starr and Swenberg \(2013\)](#) does not necessarily produce a conservative bound on  
25 risk.

26 See Table 4-24 for a summary of key unit risk estimates derived in this assessment. See  
27 Section 4.7 for risk estimates based on occupational exposure scenarios.

---

<sup>42</sup>These background lifetime incidence values were obtained from the lifetable analysis, based on SEER rates, as discussed in Sections 4.1.1.3 and 4.1.2.3. For lymphoid cancer, for example, see the value of  $R_0$  at the bottom of the lifetable analysis in Appendix E.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1  
2  
3

**Table 4-24. Summary of key unit risk estimates from this assessment (see Section 4.7 for risk estimates based on occupational exposure scenarios)**

Basis	Inhalation unit risk estimate <sup>a</sup> (per $\mu\text{g}/\text{m}^3$ ) <sup>b</sup>
<b>Full lifetime unit risk estimate<sup>c</sup></b>	
Total cancer risk based on human data (NIOSH cohort of sterilizer workers)—lymphoid cancer incidence (linear regression of categorical results) and breast cancer incidence in females (2-piece linear spline model)	$1.8 \times 10^{-3}$
<b>Adult-based unit risk estimates<sup>d</sup></b>	
Total cancer risk based on human data (NIOSH cohort)—lymphoid cancer incidence (linear regression of categorical results) and breast cancer incidence in females (2-piece linear spline model)	$1.1 \times 10^{-3}$
Lymphoid cancer incidence based on human data (NIOSH cohort)—preferred model: linear regression of categorical results	$4.3 \times 10^{-4}$
Breast cancer incidence in females based on human data (NIOSH cohort)—preferred model: 2-piece linear spline model	$8.2 \times 10^{-4}$
Lymphoid cancer incidence based on human data (NIOSH cohort)—range based on 2 models: the preferred model (linear regression of categorical results) and an alternative continuous exposure model (2-piece log-linear spline model)	$4.3 \times 10^{-4}$ to $8.5 \times 10^{-4}$
Breast cancer incidence in females based on human data (NIOSH cohort)—range based on three reasonable statistically significant continuous exposure models: 2-piece linear spline model, 2-piece log-linear spline model, and linear model	$2.1 \times 10^{-4}$ to $8.2 \times 10^{-4}$
Total cancer risk based on human data (NIOSH cohort)—range based on total estimates from range of lymphoid cancer incidence estimates (linear regression of categorical results and 2-piece log-linear spline model) and range of female breast cancer incidence estimates (2-piece linear spline model, 2-piece log-linear spline model, and linear model)	$5.6 \times 10^{-4}$ to $1.4 \times 10^{-3}$
Lymphoid cancer mortality based on human data (NIOSH cohort)—linear regression of categorical results	$2.0 \times 10^{-4}$
Breast cancer mortality in females based on human data (NIOSH cohort)—linear regression of categorical results	$2.4 \times 10^{-4}$
Preferred total cancer incidence risk estimate from rodent data (female mouse)	$4.6 \times 10^{-5}$
Range of total cancer incidence risk estimates from rodent data (mouse and rat)	$2.2 \times 10^{-5}$ to $4.6 \times 10^{-5}$

*This document is a draft for review purposes only and does not constitute Agency policy.*

**Table 4–24. Summary of key unit risk estimates from this assessment (see Section 4.7 for risk estimates based on occupational exposure scenarios) (continued)**

Basis	Inhalation unit risk estimate <sup>a</sup> (per $\mu\text{g}/\text{m}^3$ ) <sup>b</sup>
<b>0.01/EC<sub>01</sub> estimates<sup>c</sup></b>	
Lymphoid cancer incidence based on human data (NIOSH cohort)—linear regression of categorical results	$1.9 \times 10^{-4}$
Breast cancer incidence in females based on human data (NIOSH cohort)—estimate based on best-fitting model: the 2-piece linear spline model	$4.2 \times 10^{-4}$
Lymphoid cancer mortality based on human data (NIOSH cohort)—linear regression of categorical results	$9.0 \times 10^{-5}$
Breast cancer mortality in females based on human data (NIOSH cohort)—linear regression of categorical results	$1.2 \times 10^{-4}$
Total cancer incidence based on human data (NIOSH cohort)	$6.2 \times 10^{-4}$

<sup>a</sup>Technically, the values listed in this table are not all unit risk estimates as defined by EPA, but they are all potency estimates that, when multiplied by an exposure value, give an estimate of extra cancer risk. These potency estimates are not intended for use with continuous lifetime exposure levels above  $140 \mu\text{g}/\text{m}^3$ . See Section 4.7 for risk estimates based on occupational exposure scenarios. Preferred estimates are in bold.

<sup>b</sup>To convert unit risk estimates to  $(\text{ppm})^{-1}$ , multiply the  $(\mu\text{g}/\text{m}^3)^{-1}$  estimates by 1,830  $(\mu\text{g}/\text{m}^3)/\text{ppm}$ .

<sup>c</sup>Because the weight of evidence supports a mutagenic mode of action for EtO carcinogenicity, and because of the lack of chemical-specific data, EPA assumes increased early-life susceptibility and recommends the application of ADAFs, in accordance with EPA’s *Supplemental Guidance* ([U.S. EPA, 2005b](#)), for exposure scenarios that include early-life exposures. For the full lifetime (upper-bound) unit risk estimate presented here, ADAFs have been applied, as described in Section 4.4.

<sup>d</sup>These (upper-bound) unit risk estimates are intended for use in ADAF calculations and less-than-lifetime adult exposure scenarios ([U.S. EPA, 2005b](#)). Note that these are not the same as the unit risk estimates derived directly from the human data in Section 4.1 under the assumption that RRs are independent of age. Under that assumption, the key unit risk estimates were  $4.8 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  for lymphoid cancer incidence,  $9.5 \times 10^{-4}$  per  $\mu\text{g}/\text{m}^3$  for breast cancer incidence, and  $1.2 \times 10^{-3}$  per  $\mu\text{g}/\text{m}^3$  for the combined cancer incidence risk from those two cancers. See Section 4.4 for the derivation of the adult-based unit risk estimates.

<sup>e</sup>These are not upper-bound risk estimates but, rather, estimates based on linear extrapolation from the EC<sub>01</sub>. ADAFs should be applied if early-life exposure occurs, in accordance with EPA’s *Supplemental Guidance* ([U.S. EPA, 2005b](#)).

## 4.6. COMPARISON WITH OTHER PUBLISHED RISK ESTIMATES

The unit risk values derived in this document are compared with other recent risk estimates presented in the published literature (see Table 4-25).

### 4.6.1. Unit Risk Estimates Based on Human Studies

[Kirman et al. \(2004\)](#) used leukemia data only and pooled data from both the [Stayner et al. \(1993\)](#) and the UCC studies ([Teta et al., 1999](#); [Teta et al., 1993](#)). Based on the assumption that

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 leukemias are due to chromosome translocations, requiring two independent events  
 2 (chromosome breaks), [Kirman et al. \(2004\)](#) proposed that two independent EtO-induced events  
 3 are required for EtO-induced leukemias and used a dose-squared model, yielding a unit risk  
 4 value of  $4.5 \times 10^{-8} (\mu\text{g}/\text{m}^3)^{-1}$  as their preferred estimate.

5  
6  
7 **Table 4-25. Comparison of unit risk estimates<sup>a</sup>**  
8

Assessments	Data source	Inhalation unit risk estimate <sup>b</sup> (per $\mu\text{g}/\text{m}^3$ )
<b>Based on human data</b>		
EPA (this document) <sup>c</sup>	Lymphoid cancer incidence in sterilizer workers (NIOSH cohort) <sup>d</sup>	$7.2 \times 10^{-4}$
	Breast cancer incidence in female sterilizer workers (NIOSH cohort) <sup>e</sup>	$1.4 \times 10^{-3}$
	Total cancer risk based on the NIOSH data	$1.8 \times 10^{-3}$
<a href="#">Kirman et al. (2004)</a>	Leukemia mortality in combined NIOSH and UCC cohorts (earlier follow-ups)	$4.5 \times 10^{-8}$ Range of $1.4 \times 10^{-8}$ to $1.4 \times 10^{-7}$ <sup>f</sup>
<a href="#">Valdez-Flores et al. (2010)</a>	multiple individual cancer endpoints, including all lymphohematopoietic, lymphoid, and breast cancers, in combined updated NIOSH and updated UCC cohorts	$5.5 \times 10^{-7}$ to $1.6 \times 10^{-6}$ <sup>g</sup>
<b>Based on rodent data</b>		
EPA (this document) <sup>c</sup>	Female mouse tumors	$7.6 \times 10^{-5}$
<a href="#">Kirman et al. (2004)</a>	Mononuclear cell leukemia in rats and lymphomas in mice	$2.6 \times 10^{-8}$ to $1.5 \times 10^{-5}$ <sup>h</sup>

9  
10 <sup>a</sup>Upper-bound estimates except where footnoted to indicate that estimates are based on EC values (i.e., estimates  
11 with footnotes f and g).

12 <sup>b</sup>Because the weight of evidence supports a mutagenic mode of action for EtO carcinogenicity, and in the absence of  
13 chemical-specific data, EPA assumes increased early-life susceptibility, in accordance with EPA's *Supplemental*  
14 *Guidance* ([U.S. EPA, 2005b](#)), and for the EPA lifetime unit risk estimates presented in this table, ADAFs have been  
15 applied, as described in Section 4.4. The corresponding adult-based unit risk estimates are  $4.4 \times 10^{-4} (\mu\text{g}/\text{m}^3)^{-1}$  for  
16 human-based lymphoid cancer incidence,  $8.2 \times 10^{-4} (\mu\text{g}/\text{m}^3)^{-1}$  for human-based breast cancer incidence,  
17  $1.1 \times 10^{-3} (\mu\text{g}/\text{m}^3)^{-1}$  for human-based total cancer incidence, and  $4.6 \times 10^{-5} (\mu\text{g}/\text{m}^3)^{-1}$  for rodent-based total cancer  
18 incidence. The non-EPA estimates in the table are shown as reported and do not account for potential increased  
19 early-life susceptibility for lifetime exposures that include childhood, with the exception of the [Valdez-Flores et al.](#)  
20 [\(2010\)](#) estimates, which are purported to include the ADAFs, but the ADAFs were in fact misapplied and have  
21 essentially no impact (see Appendix A.2.20).

22 <sup>c</sup>See Table 4-24 in Section 4.5 for a more complete summary of estimates from this assessment. See Section 4.7 for  
23 risk estimates for occupational exposure scenarios.

24 <sup>d</sup>For lymphoid cancer mortality, the ADAF-adjusted lifetime unit risk estimate is  $3.3 \times 10^{-4} (\mu\text{g}/\text{m}^3)^{-1}$  and the adult-  
25 based unit risk estimate is  $2.0 \times 10^{-4} (\mu\text{g}/\text{m}^3)^{-1}$ .

1 **Table 4-25. Comparison of unit risk estimates<sup>a</sup> (continued)**

2  
3  
4 <sup>e</sup>For breast cancer mortality, the ADAF-adjusted lifetime unit risk estimate is  $4.0 \times 10^{-4} (\mu\text{g}/\text{m}^3)^{-1}$  and the adult-  
5 based unit risk estimate is  $2.4 \times 10^{-4} (\mu\text{g}/\text{m}^3)^{-1}$ .

6 <sup>f</sup>Estimates based on linear extrapolation from EC0001–EC000001 obtained from the quadratic model.

7 <sup>g</sup>Estimates based on range of EC(1/million)s of 0.001–0.003 ppm obtained from the model  $\text{RR} = e^{(\beta \times \text{exposure})}$  for  
8 relevant cancer endpoints.

9 <sup>h</sup>Estimates based on quadratic extrapolation model below the observable range of the data (i.e., below the  $\text{LEC}_{10}$  or  
10  $\text{LEC}_{01}$  obtained using multistage model) with various points of departure ( $\text{LEC}_{01}$ – $\text{LEC}_{000001}$ ) for final linear  
11 extrapolation (see Section 4.4.2).  
12  
13

14 The [Kirman et al. \(2004\)](#) values are different from those in the current document because  
15 of the different assumptions inherent in the [Kirman et al. \(2004\)](#) approach and because the study  
16 used unpublished data from earlier follow-ups of the two cohorts. A key difference is that EPA  
17 uses a linear model or a two-piece linear spline model rather than a quadratic (dose-squared)  
18 model in the range of observation. Then, EPA uses a higher extra risk level (1%) for  
19 establishing the POD, whereas [Kirman et al. \(2004\)](#) used a risk level of  $10^{-5}$  for their best  
20 estimate and a risk range of  $10^{-4}$  to  $10^{-6}$  for their range of values. The extra risk level and the  
21 corresponding POD are not critical with the linear model; however, with the quadratic model  
22 used by [Kirman et al. \(2004\)](#), the lower the risk level (and hence the POD), the greater the  
23 impact of the quadratic model and the lower the resulting unit risk estimates.

24 In addition, EPA (1) uses data for lymphoid cancers (and female breast cancers) rather  
25 than leukemias, (2) includes ages up to 85 years in the life-table analysis rather than stopping at  
26 70 years, (3) calculates unit risk estimates for cancer incidence as well as mortality, (4) uses a  
27 lower bound as the POD rather than the maximum likelihood estimate, (5) uses the results of  
28 lagged analyses rather than unlagged analyses, and (6) uses adult-based unit risk estimates in  
29 conjunction with ADAFs (see Section 4.4) to derive the lifetime unit risk estimates.

30 Another key difference is that [Kirman et al. \(2004\)](#) relied on earlier NIOSH results  
31 ([Stayner et al., 1993](#)), whereas EPA uses the results of NIOSH's more recent follow-up of the  
32 cohort ([Steenland et al., 2004](#)). [Kirman et al. \(2004\)](#) claim that a quadratic dose-response model  
33 provided the best fit to the data in the observable range and that this provides support for their  
34 assumed mode of action. However, the 2004 NIOSH data for lymphohematopoietic cancers  
35 suggest a supralinear exposure-response relationship (see Section 4.1.1.2 and Figures 4-1 and  
36 4-2), which is inconsistent with a dose-squared model. Furthermore, EPA's review of the mode  
37 of action evidence does not support the mode of action assumed by [Kirman et al. \(2004\)](#) (see  
38 Section 3.4).

1 The [Valdez-Flores et al. \(2010\)](#) unit risk estimates (see Table 4–25) are similarly much  
2 lower than those in the current document because of the different assumptions used. A key  
3 difference is that EPA uses a linear model or a two-piece linear spline model in the range of  
4 observation rather than an exponential model ( $RR = e^{\beta \times \text{exposure}}$ ), which was used by [Valdez-](#)  
5 [Flores et al. \(2010\)](#) despite its lack of fit. Then, EPA uses a 1% extra risk level for establishing  
6 the POD for linear extrapolation, whereas [Valdez-Flores et al. \(2010\)](#) used a risk level of  $10^{-6}$ . In  
7 addition, EPA (1) includes ages up to 85 years in the life-table analysis rather than stopping at  
8 70 years, (2) calculates unit risk estimates for cancer incidence as well as mortality, (3) uses a  
9 lower bound as the POD rather than the maximum likelihood estimate, and (4) uses the results of  
10 lagged analyses rather than unlagged analyses. See Appendix A.2.20 for a more detailed  
11 discussion of the differences between the EPA and [Valdez-Flores et al. \(2010\)](#) analyses.  
12

#### 13 **4.6.2. Unit Risk Estimates Based on Laboratory Animal Studies**

14 [Kirman et al. \(2004\)](#) also used linear and dose-squared extrapolation models to derive  
15 unit risk estimates based on the rat mononuclear cell leukemia data and the mouse lymphoma  
16 data. First, they used the multistage model to calculate the  $LEC_{10}$  ( $LEC_{01}$  for the male mouse  
17 lymphoma data) for the POD from the observable range. Then, using these PODs for linear  
18 extrapolation, [Kirman et al. \(2004\)](#) obtained a unit risk range of  $3.9 \times 10^{-6} (\mu\text{g}/\text{m}^3)^{-1}$  to  
19  $1.5 \times 10^{-5} (\mu\text{g}/\text{m}^3)^{-1}$ . Alternatively, [Kirman et al. \(2004\)](#) used a quadratic extrapolation model  
20 below the observable range to estimate secondary points of departure ( $LEC_{01}$ – $LEC_{000001}$ ;  
21  $LEC_{001}$ – $LEC_{000001}$  for the male mouse) for final linear low-dose extrapolation, yielding unit risks  
22 ranging from  $2.6 \times 10^{-8} (\mu\text{g}/\text{m}^3)^{-1}$  to  $4.9 \times 10^{-6} (\mu\text{g}/\text{m}^3)^{-1}$ . These values are all smaller than the  
23 unit risks derived from the rodent data in this document.  
24

#### 25 **4.7. RISK ESTIMATES FOR SOME OCCUPATIONAL EXPOSURE SCENARIOS**

26 The unit risk estimates derived in the preceding sections were developed for  
27 environmental exposure levels, where maximum modeled levels are on the order of  $1\text{--}2 \mu\text{g}/\text{m}^3$   
28 (email dated October 3, 2005, from Mark Morris, EPA, to Jennifer Jinot, EPA), i.e., roughly  
29  $0.5\text{--}1$  ppb, and are not applicable to higher exposures, including some occupational exposure  
30 levels. However, occupational exposure levels of EtO are of concern to EPA when EtO is used  
31 as a pesticide (e.g., sterilizing agent or fumigant). The occupational exposure scenarios of  
32 interest to EPA include some cumulative exposures corresponding to exposure levels in the  
33 nonlinear range of some of the models (i.e., above the maximum exposure level at which the  
34 low-dose-linear unit risk estimates apply). Therefore, extra risk estimates were calculated for a  
35 number of occupational exposure scenarios of possible concern. Extra risk estimates are

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 estimates of the extra cancer risk above background and are the same type of estimate that one  
2 gets from multiplying a unit risk estimate by an exposure level. In this case, the exposure level is  
3 used directly in the exposure-response model, thus accounting for any nonlinearities in the model  
4 above the range of exposure levels for which the linear unit risk estimate is applicable. For these  
5 occupational exposure scenarios, exposure-response models based on data from the NIOSH  
6 cohort were used in conjunction with the life-table program, as previously discussed in  
7 Section 4.1. A 35-year exposure occurring between ages 20 and 55 years was assumed, and  
8 exposure levels ranging from 0.1 to 1 ppm 8-hour TWA were examined (i.e., ranging from about  
9 1,300 to 13,000 ppm × days). (Note that the current Occupational Safety and Health  
10 Administration Permissible Exposure Limit is 1 ppm [8-hour TWA].)

11 For *lymphoid cancer* mortality in both sexes, the best-fitting (natural) log cumulative  
12 exposure Cox regression model (see Steenland reanalyses in Appendix D; see also  
13 Section 4.1.1.2), lagged 15 years, was used. The log cumulative exposure Cox regression model  
14 was the best-fitting model for lymphoid cancer in males in the [Steenland et al. \(2004\)](#) study, and  
15 the same model form is used here but with the data from both sexes. Although this model was  
16 deemed too steep in the low-exposure region to be useful for the derivation of unit risk estimates  
17 for lower (environmental) exposures, the model is well suited for the occupational exposure  
18 scenarios of interest in this assessment because the corresponding cumulative exposures are well  
19 within the range of the cumulative exposures in the NIOSH cohort. The model was statistically  
20 significant ( $p = 0.02$ ) and provided a better fit, based on AIC, than the two-piece spline models  
21 with maximum likelihoods that were considered in Section 4.1.1.2 (the AICs were 460.426,  
22 461.847, and 461.48 for the log cumulative exposure Cox regression, log-linear two-piece spline  
23 and linear two-piece spline models, respectively [as reported in Section D.3 of Appendix D]; a  
24 lower AIC indicates a better fit), as well as a more plausible, smoothly curved exposure-response  
25 relationship than the two-piece spline models, both of which, with knots at 100 ppm × days, had  
26 a very steep rise and then a very sharp change in slope at the knot. In addition, the log  
27 cumulative exposure Cox regression model had a slightly lower AIC (460.426 versus 460.54)  
28 than the log cumulative exposure linear model (see Section D.3.c of Appendix D) and has the  
29 advantage of being a standard epidemiological model for continuous exposure data (the Cox  
30 regression model, albeit with log cumulative exposure to accommodate the supralinearity of the  
31 exposure-response data). The log cumulative exposure linear model yields slightly higher RR  
32 estimates than the log cumulative exposure Cox regression model, as can be seen by comparing  
33 the log cumulative exposure models in Figures D-3b and D-3c in Appendix D, and would thus  
34 result in slightly higher extra risk estimates than the log cumulative exposure Cox regression  
35 model. For example, the MLEs of extra risk from the log cumulative exposure linear model

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 would range from about 23% higher for the 0.1 ppm 8-hour TWA to 6% higher for the 1 ppm  
2 8-hour TWA. (The two-piece log-linear spline model with the knot at 1,600 ppm × days [for  
3 which there was a local maximum likelihood], which was considered as an alternative model for  
4 the derivation of a unit risk estimate for lymphoid cancer, was not considered for the  
5 occupational exposure extra risk estimates because there are better fitting models, discussed  
6 above, that are suitable for the range of occupational exposures of interest.)

7 The extra risk results for lymphoid cancer mortality and incidence in both sexes for the  
8 log cumulative exposure Cox regression model are presented in Table 4-26. For lymphoid  
9 cancer incidence, the exposure-response relationship was assumed to be the same as for mortality  
10 (see Section 4.1.1.3). As can be seen in Table 4-26, the extra risks for these occupational  
11 exposure levels are in the “plateau” region of the exposure-response relationships and increase  
12 less than proportionately with exposure. For occupational exposures less than about 1,000 ppm  
13 × days, or about 0.08 ppm 8-hour TWA for 35 years, risk estimates are no longer in the plateau  
14 region (see Figure 4-1) but rather in the steep low-exposure region, which is a region of greater  
15 uncertainty for the log cumulative exposure model, and one might want to use the linear  
16 regression of the categorical results that was used for lower exposures (see Section 4.1.1.2;  
17 Appendix D). Furthermore, if one is using the linear regression model in this range and also  
18 estimating risks for exposure levels in the range between about 0.08 and 0.6 ppm 8-hour TWA  
19 (near where the linear regression and log cumulative exposure Cox regression models meet),  
20 then one might want to use the linear regression model for the entire range up to 0.6 ppm 8-hour  
21 TWA to avoid a discontinuity between the two models; thus, results for the linear regression  
22 model for exposure levels up to 0.6 ppm 8-hour TWA are also presented in Table 4-26. While  
23 the best-fitting continuous exposure model, the log cumulative exposure Cox regression model,  
24 would generally be preferred in the exposure range between 0.08 and 0.6 ppm 8-hour TWA,  
25 there is model uncertainty, so the use of either model could be justified. For exposures higher  
26 than where the linear regression and log cumulative exposure Cox regression models meet, the  
27 log cumulative exposure model exclusively is recommended. The models used to derive the  
28 extra risk estimates presented in Table 4-26 for lymphoid cancer for the occupational exposure  
29 scenarios are displayed in Figure 4-7 over the range of occupational cumulative exposures of  
30 interest; the categorical results are included for comparison.

31 For *breast cancer*, incidence data were available from the NIOSH incidence study; thus,  
32 only incidence estimates were calculated. In addition to being the preferred type of cancer risk  
33 estimate, the breast cancer incidence risk estimates are based on more cases than were available  
34 in the mortality study and the incidence data (for the subcohort with interviews) are adjusted for  
35 a number of breast cancer risk factors (see Section 4.1.2.3). In terms of the incidence data, the

1 subcohort data are preferred to the full cohort data because the subcohort data are adjusted for  
2 these potential confounders and also because the full cohort data have incomplete ascertainment  
3 of breast cancer cases.

4 For breast cancer incidence in the subcohort with interviews, a number of Cox regression  
5 exposure-response models from the [Steenland et al. \(2003\)](#) breast cancer incidence study fit  
6 almost equally well (see Section 4.1.2.3). These include a log cumulative exposure model and a  
7 cumulative exposure model, both with a 15-year lag, and a log cumulative exposure model with  
8 no lag. The latter model was omitted from the calculations because the inclusion of a 15-year lag  
9 for the development of breast cancer was considered more biologically realistic than not  
10 including a lag. [Steenland et al. \(2003\)](#) also provide a duration-of-exposure Cox regression  
11 model with a marginally better fit; however, models using duration of exposure are less useful  
12 for estimating exposure-related risks, and duration of exposure and cumulative exposure are  
13 correlated. Thus, only the lagged cumulative exposure models are considered here.

14

**Table 4-26. Extra risk estimates for lymphoid cancer in both sexes for various occupational exposure levels<sup>a</sup>**

8-hr TWA (ppm)	Lymphoid cancer mortality				Lymphoid cancer incidence <sup>b</sup>			
	Log cumulative exposure Cox regression model <sup>c</sup>		Linear regression model of categorical results <sup>d</sup>		Log cumulative exposure Cox regression model <sup>c</sup>		Linear regression model of categorical results <sup>d</sup>	
	MLE	95% UCL	MLE	95% UCL	MLE	95% UCL	MLE	95% UCL
0.1	0.014	0.032	0.003	0.007	<b>0.031</b>	<b>0.071</b>	<b>0.007</b>	<b>0.016</b>
0.2	0.016	0.038	0.007	0.014	<b>0.035</b>	<b>0.084</b>	<b>0.014</b>	<b>0.031</b>
0.3	0.017	0.042	0.010	0.022	<b>0.038</b>	<b>0.093</b>	<b>0.021</b>	<b>0.047</b>
0.4	0.018	0.045	0.013	0.029	<b>0.040</b>	<b>0.099</b>	<b>0.028</b>	<b>0.062</b>
0.5	0.018	0.047	0.016	0.036	<b>0.042</b>	<b>0.10</b>	<b>0.035</b>	<b>0.076</b>
0.6	0.019	0.049	0.019	0.042	<b>0.043</b>	<b>0.11</b>	<b>0.042</b>	<b>0.090</b>
0.7	0.019	0.051	--	0.049	<b>0.044</b>	<b>0.11</b>	--	--
0.8	0.020	0.052	--	--	<b>0.045</b>	<b>0.12</b>	--	--
0.9	0.020	0.054	--	--	<b>0.046</b>	<b>0.12</b>	--	--
1.0	0.021	0.055	--	--	<b>0.047</b>	<b>0.12</b>	--	--

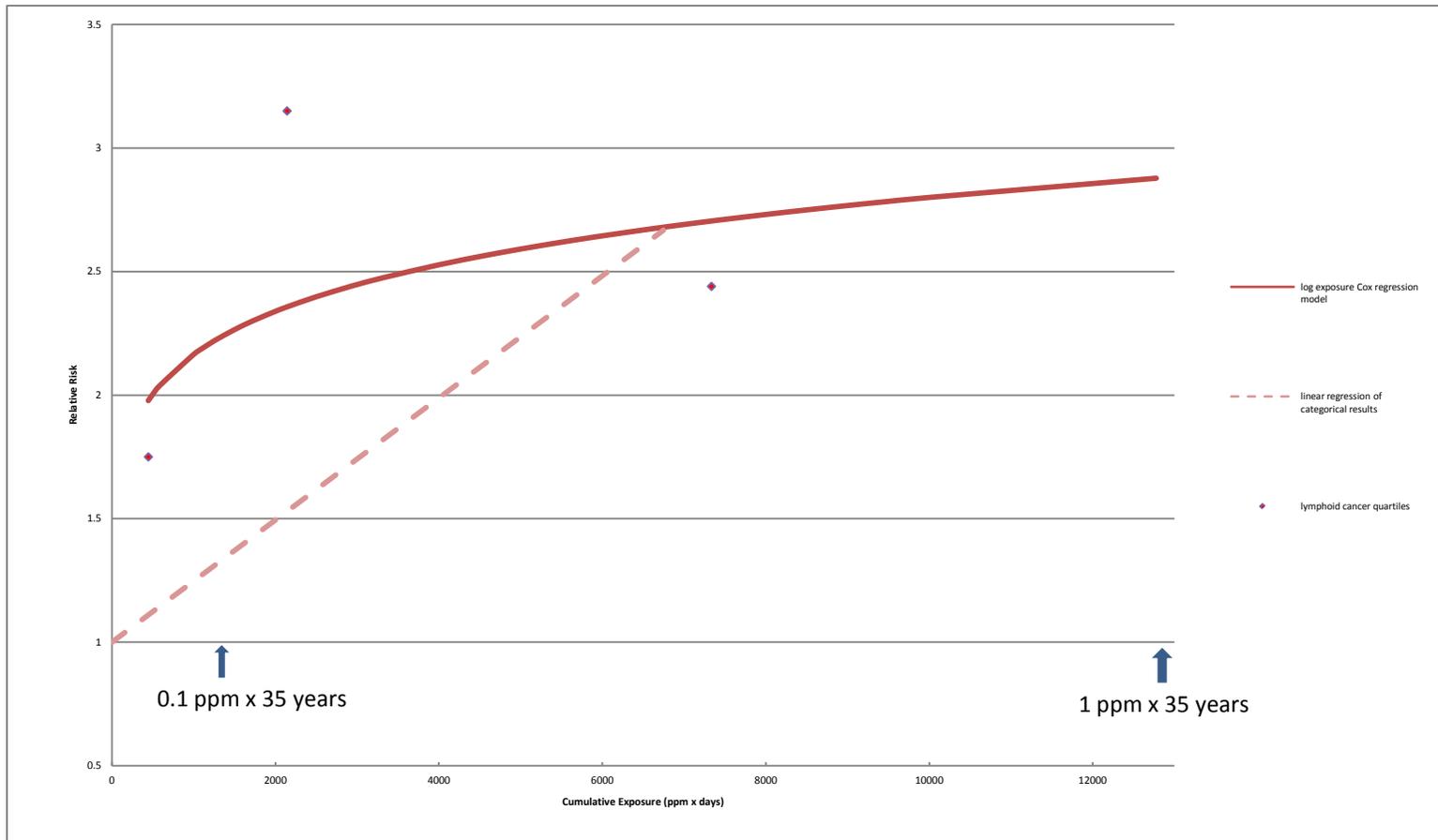
<sup>a</sup>Assuming a 35-yr exposure between ages 20 and 55 years (see Section 4.7).

<sup>b</sup>Assumes same exposure-response relationship as for lymphoid cancer mortality.

<sup>c</sup>From the best-fitting log cumulative exposure Cox regression model for lymphoid cancer mortality in both sexes; 15-yr lag (see Appendix D; see also Section 4.1.1.2).

<sup>d</sup>Linear regression of categorical results for both sexes (see Appendix D; 15-yr lag), excluding the highest exposure group (see Section 4.1.1.2); extra risk estimates from the linear model are provided only up to the exposure level where the linear model meets the log cumulative Cox regression model.

MLE: maximum likelihood estimate; UCL: (one-sided) upper confidence limit estimate.



**Figure 4-7. RR estimates for lymphoid cancer from occupational EtO exposures (with 15-year lag).**

Lymphoid cancer models (see Section 4.1.1.2): log cumulative exposure Cox regression model; categorical results from Cox regression model; linear regression of categorical results, excluding highest exposure group. (Note that, with the exception of the categorical results and the linear regression of the categorical results, the various models have different implicitly estimated baseline risks; thus, they are not strictly comparable to each other in terms of RR values, i.e., along the y-axis. They are, however, comparable in terms of general shape.)

1           The extra risk estimates for breast cancer incidence in females from the lagged  
2 cumulative exposure and log cumulative exposure Cox regression models listed above are  
3 presented in Table 4-27. As can be seen in Table 4-27, the extra risk estimates for the lagged log  
4 cumulative exposure and cumulative exposure models differ substantially. Furthermore, the  
5 categorical Cox regression results for breast cancer incidence in the subcohort with interviews  
6 suggest a more linear low-exposure exposure-response relationship than that obtained with either  
7 the continuous variable log cumulative exposure (supralinear) or cumulative exposure (sublinear)  
8 Cox regression models (see Figure 4-5). (The lowest four exposure quintiles represent individual  
9 worker exposures ranging from 0 to about 15,000 ppm × days, which covers the range of  
10 cumulative exposures for the occupational exposure scenarios of interest in this assessment, the  
11 maximum of which is 12,775 ppm × days.) Therefore, the two-piece linear spline model (with a  
12 15-year lag) (see Section 4.1.2.3) was also used to calculate the extra risk estimates. The  
13 two-piece linear spline model provides a better fit to the data than the log cumulative exposure or  
14 cumulative exposure Cox regression models, as indicated by a lower AIC value (1,950.9 for  
15 two-piece linear spline model vs. 1,956.2 for the log cumulative exposure Cox regression model  
16 and 1,956.8 for the cumulative exposure Cox regression model; Table 4-12 and Appendix D). In  
17 fact, the two-piece linear spline model provided the best fit to the breast cancer incidence data of  
18 all the models investigated in Section 4.1.2.3, and it provides the best representation of the  
19 categorical RR results, particularly for the range of cumulative exposures for the occupational  
20 exposure scenarios of interest (see Figures 4-5, 4-6, and 4-8). The extra risk estimates calculated  
21 using the two-piece linear spline model are also presented in Table 4-27 and are the preferred  
22 estimates because they are derived from the best-fitting model.

23           In addition, extra risk estimates for breast cancer incidence in females from the  
24 continuous exposure linear model (with a 15-year lag) (see Section 4.1.2.3) are presented in  
25 Table 4-27 for comparison. This model, with an AIC of 1,952.3 (see Table 4-12), was the  
26 second-best-fitting model and also provided an adequate visual fit to the categorical data (see  
27 Figure 4-6). Moreover, the two best-fitting models (i.e., the continuous exposure linear model  
28 and two-piece linear spline model) span the range of RR estimates from the three best-fitting  
29 models investigated in Section 4.1.2.3 (the third being the two-piece log-linear spline model)  
30 over the range of cumulative exposures for the occupational exposure scenarios of interest in this  
31 assessment (see Figure 4-6). Comparing the results of the two best-fitting models shows that the  
32 extra risk estimates differ by just under fourfold at the lowest exposure level (0.1 ppm 8-hour

**Table 4-27. Extra risk estimates for breast cancer incidence in females for various occupational exposure levels<sup>a,b</sup>**

8-hr TWA (ppm)	Log cumulative exposure Cox regression model <sup>c</sup>		Cumulative exposure Cox regression model <sup>c</sup>		Continuous linear model <sup>d</sup>		Two-piece linear spline model <sup>e</sup>	
	MLE	95% UCL	MLE	95% UCL	MLE	95% UCL <sup>f</sup>	MLE	95% UCL <sup>g</sup>
0.1	0.055	0.11	0.0013	0.0023	0.0042	0.0081	<b>0.016</b>	<b>0.031</b>
0.2	0.061	0.12	0.0026	0.0046	0.0084	0.016	<b>0.032</b>	<b>0.061</b>
0.3	0.065	0.13	0.0040	0.0069	0.012	0.024	<b>0.048</b>	<b>0.090</b>
0.4	0.068	0.14	0.0053	0.0092	0.017	0.032	<b>0.063</b>	<b>0.118</b>
0.5	0.070	0.14	0.0067	0.012	0.021	0.040	<b>0.075</b>	<b>0.139</b>
0.6	0.072	0.14	0.0081	0.014	0.025	0.048	<b>0.081</b>	<b>0.150</b>
0.7	0.073	0.15	0.0095	0.017	0.029	0.055	<b>0.086</b>	<b>0.157</b>
0.8	0.074	0.15	0.011	0.019	0.033	0.063	<b>0.089</b>	<b>0.162</b>
0.9	0.076	0.15	0.012	0.022	0.037	0.070	<b>0.093</b>	<b>0.167</b>
1.0	0.077	0.16	0.014	0.024	0.041	0.078	<b>0.095</b>	<b>0.171</b>

<sup>a</sup>Assuming a 35-yr exposure between ages 20 and 55 years.

<sup>b</sup>From incidence data for subcohort with interviews; invasive and in situ tumors ([Steenland et al., 2003](#)).

<sup>c</sup>Cox regression models from Table 5 of [Steenland et al. \(2003\)](#), with 15-yr lag.

<sup>d</sup>Linear model with cumulative exposure as a continuous variable (see Section 4.1.2.3 and Section D.2 of Appendix D).

<sup>e</sup>Two-piece linear spline model results for occupational exposures use both spline segments (see Section D.2 of Appendix D), knot at 5,800 ppm × days; with 15-yr lag. For the 95% UCL, for exposures below the knot,  $RR = 1 + (\beta_1 + 1.645 \times SE_1) \times \text{exposure}$ ; for exposures above the knot,  $RR = 1 + (\beta_1 \times \text{exp} + \beta_2 \times (\text{exp} - \text{knot}) + 1.645 \times \sqrt{\text{exp}^2 \times \text{var}_1 + (\text{exp} - \text{knot})^2 \times \text{var}_2 + 2 \times \text{exp} \times (\text{exp} - \text{knot}) \times \text{covar}}$ ), where exp = cumulative exposure, var = variance, covar = covariance (see Section D.2 of Appendix D for the parameter values).

<sup>f</sup>Confidence intervals used in deriving the 95% UCLs were estimated employing the Wald approach. Confidence intervals for linear RR models, however, in contrast to those for the log-linear RR models, may not be symmetrical. EPA also evaluated application of a profile likelihood approach for the linear RR models ([Langholz and Richardson, 2010](#)), which allows for asymmetric CIs, for comparison with the Wald approach. Using the profile likelihood method, the resulting extra risk estimates for breast cancer incidence for the linear model would have been about 29% higher than those obtained using the Wald approach.

**Table 4-27. Extra risk estimates for breast cancer incidence in females for various occupational exposure levels<sup>a,b</sup> (continued)**

<sup>a</sup>Confidence intervals used in deriving the 95% UCLs were estimated employing the Wald approach. Confidence intervals for linear RR models, however, in contrast to those for the log-linear RR models, may not be symmetrical. EPA also evaluated application of a profile likelihood approach for the linear RR models ([Langholz and Richardson, 2010](#)), which allows for asymmetric CIs, for comparison with the Wald approach. Using the profile likelihood method, the resulting extra risk estimates for breast cancer incidence for the low-exposure linear spline segment (i.e., below 0.4 ppm 8-hr TWA) would have been about 34% higher than those obtained using the Wald approach. Calculating the profile likelihood CIs in the region of the second spline segment is computationally difficult and was not pursued here.

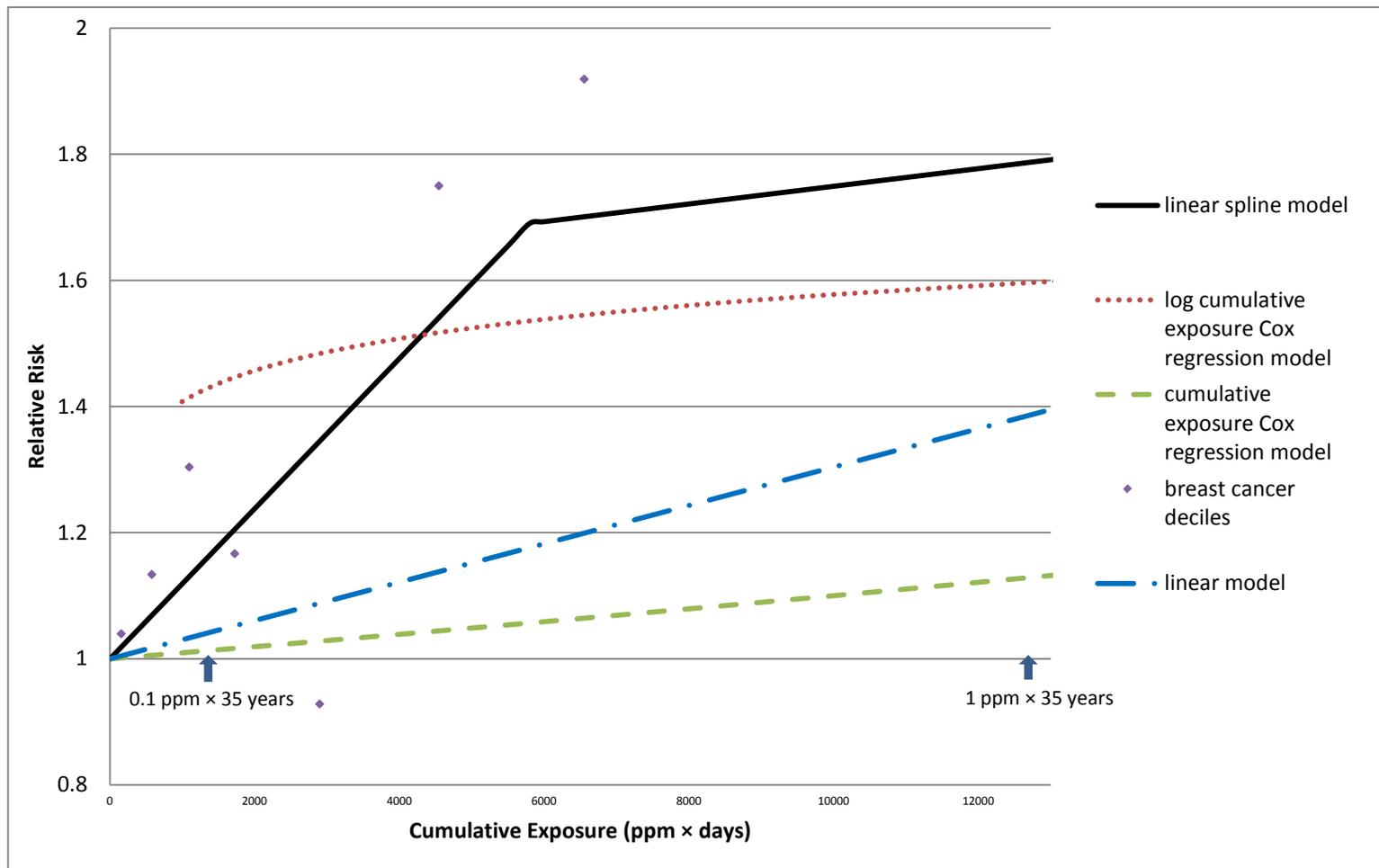
MLE: maximum likelihood estimates; UCL: (one-sided) upper confidence limit estimate.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 TWA) and the difference tapers to just over twofold at the highest exposure level (1 ppm 8-hour  
2 TWA), with the estimates of the best-fitting model, the two-piece linear spline model, yielding  
3 the higher extra risk estimates across the range.

4 Finally, also for comparison, maximum likelihood estimates (MLEs) of extra risk for the  
5 log cumulative exposure Cox regression model with a 10-year lag were calculated. The model  
6 with a 10-year lag also provided a statistically significant fit to the data ( $p = 0.03$ ), whereas, the  
7 models with 5- and 20-year lags did not. These estimates ranged from 0.067 for 0.1 ppm  
8 exposure to 0.094 for 1.0 ppm exposure. Thus, the MLEs of extra risk with a 10-year lag were  
9 about 20% higher than those with the 15-year lag.

10 The continuous exposure models (with a 15-year lag) considered for deriving the extra  
11 risk estimates for breast cancer incidence in females for the occupational exposure scenarios are  
12 displayed in Figure 4-8 over the range of occupational cumulative exposures of interest.  
13 Categorical results are also presented for comparison (deciles from the categorical linear model  
14 are presented because it had a better fit than the log-linear categorical model, as indicated by the  
15 AICs, which were 1,963.9 and 1,966.9, respectively; Appendix D). The recommended model is  
16 the two-piece linear spline model; this was the best-fitting continuous exposure model of those  
17 evaluated in this assessment, and it provides the best visual fit in comparison to the categorical  
18 results in the range of the occupational exposure scenarios of interest. As shown in Figure 4-8,  
19 the log cumulative exposure Cox regression model is too flat across the range of exposures of  
20 interest. It also appears from Figure 4-8 that the slopes of the cumulative exposure Cox  
21 regression model and the (continuous exposure) linear model are both insufficiently steep across  
22 the range of exposures of interest. This is consistent with the analysis presented in Section D.1  
23 of Appendix D showing the strong influence of the upper tail of cumulative exposures on the  
24 results of the cumulative exposure Cox regression model. The responses in the upper tail of  
25 exposures are relatively dampened, such that when the highest 5% of exposures (exposures >  
26 27,500 ppm × days, which are well in excess of the exposures corresponding to the occupational  
27 exposure scenarios considered here) are excluded, the slope of the Cox regression model is  
28 substantially increased (e.g., at 10,000 ppm × days, the RR estimate increases from about 1.1 to  
29 almost 1.5; see Figure D-1d in Appendix D). This strong influence of the upper tail of exposures  
30 would similarly attenuate the slope of the linear model, resulting in underestimation of the  
31 lower-exposure risks. The two-piece linear spline model, on the other hand, is more flexible, and  
32 the influence of the upper tail of exposures would be primarily on the upper spline segment; thus,  
33 the two-piece model is able to provide a better fit to the lower-exposure data.



**Figure 4-8. RR estimates for breast cancer incidence from occupational EtO exposures (with 15-year lag).**

Breast cancer models (see Section 4.1.2.3): linear 2-piece spline model, with knot at 5,800 ppm × days; log cumulative exposure Cox regression model; (cumulative exposure) Cox regression model; (continuous exposure) linear model; categorical results (deciles) from continuous exposure linear model. (Note that the various models have different implicitly estimated baseline risks; thus, they are not strictly comparable to each other in terms of RR values, i.e., along the y-axis. They are, however, comparable in terms of general shape.)

1 For the *total cancer risk* combined across the two cancer types, the MLE can be obtained  
2 directly by summing the MLEs for the individual cancer types. An upper bound can be  
3 approximated by summing the 95% UCL estimates for the individual cancer types; however, this  
4 will overestimate the corresponding 95% UCL on total cancer risk.<sup>43</sup>

5 Although there is model uncertainty, as discussed above, there is less overall uncertainty  
6 associated with the extra risk estimates for occupational exposure scenarios than with the unit  
7 risk estimates for environmental exposures. The extra risk estimates are derived for occupational  
8 exposure scenarios that yield cumulative exposures well within the range of the exposures in the  
9 NIOSH study. Moreover, the NIOSH study is a study of sterilizer workers who used EtO for the  
10 sterilization of medical supplies or spices ([Steenland et al., 1991](#)); thus, the results are directly  
11 applicable to workers in these occupations, and these are among the occupations of primary  
12 concern to EPA.

13  
14 *Calculation of Extra Risk Estimates for Other Occupational Exposure Scenarios:*

15  
16 Some detailed guidance is provided here for calculating extra risk estimates outside of the  
17 range of occupational scenarios considered above. Note that for 35-year exposures to exposure  
18 levels between the exposure levels presented in Tables 4-26 and 4-27, e.g., 0.15 ppm, one could  
19 interpolate between the extra risk estimates presented for the closest exposure levels on either  
20 side.

21  
22 For occupational exposures with durations other than 35 years:

23  
24 Extra risk estimates for a 45-year exposure to the same exposure levels were nearly  
25 identical to those from the 35-year exposure for both lymphoid cancer in both sexes and breast  
26 cancer in females (results not shown). With the 15-year lag, the assumption of an additional  
27 10 years of exposure only negligibly affects the risks above age 70 and has little impact on  
28 lifetime risk. For exposure scenarios of 35–45 years but with 8-hour TWAs falling between  
29 those presented in the tables, one can estimate the extra risk by interpolation. For exposure

---

<sup>43</sup>Technically, these sums would reflect the total cancer risk to females and not a mixed-sex workforce because the breast cancer risk applies only to females. As a practical matter for regulatory purposes, females typically comprise a substantial proportion of the sterilizer workforce and summing the extra risk estimates enables risk managers to evaluate the individual risk for this substantial workforce group. In a situation in which the workforce of concern is comprised predominantly of males, it might be appropriate to use a sex-weighted sum of the extra risks from the two cancer types. For the purposes of estimating numbers of cancer cases attributable to specific exposure levels, e.g., for benefits analyses, it would be most suitable to use the cancer-specific unit risk estimates (or central tendency estimates), taking sex into account.

*This document is a draft for review purposes only and does not constitute Agency policy.*

1 scenarios with durations of exposure less than 30–35 years, one could roughly estimate extra  
2 risks by calculating the cumulative exposure and finding the extra risks for a similar cumulative  
3 exposure in Tables 4-26 and 4-27. For a more precise estimation, or for exposure scenarios of  
4 much shorter duration or for specific age groups, one should do the calculations using a life-table  
5 analysis, as presented in Appendix E but modified for the specific exposure scenarios.

6  
7 For occupational exposures below 0.1 ppm:

8  
9 *For lymphoid cancer*, use of the log cumulative exposure Cox regression model is not  
10 advised below 0.1 ppm ( $\times$  35 years). Instead, the low-exposure continuation of the linear  
11 regression model presented in Table 4-26 of the assessment is recommended. For 35-year  
12 exposures, the following formulae would apply:

13  
14  
15 
$$95\% \text{ UCL on extra risk for lymphoid cancer incidence} \approx (8\text{-h TWA occ exp [in ppm]}) \times$$
  
16 
$$(0.016/0.1 \text{ ppm}) = (8\text{-h TWA occ exp [in ppm]}) \times (0.16/\text{ppm})$$

17  
18 
$$\text{MLE of extra risk for lymphoid cancer incidence} = (8\text{-h TWA occ exp [in ppm]}) \times$$
  
19 
$$(0.007/0.1 \text{ ppm}) = (8\text{-h TWA occ exp [in ppm]}) \times (0.07/\text{ppm})$$

20  
21  
22 If one is considering occupational exposure scenarios using a range of 8-h TWA  
23 exposure levels on both sides of 0.1 ppm, one might want to use the linear regression model for  
24 all the exposure levels up to about 0.6 ppm 8-h TWA (approximately where the linear regression  
25 model intersects the log cumulative exposure Cox regression model) to avoid the discontinuity  
26 between the two models below where they intersect. Note that the extra risk estimates from the  
27 different models differ by at most about 4.5-fold (at 0.1 ppm) and that there is model uncertainty  
28 in this range, so the use of either model could be justified. Above where the models intersect,  
29 only the log cumulative exposure Cox regression model should be used.

30  
31 *For breast cancer*, the low-exposure continuation of the two-piece linear spline model  
32 presented in Table 4-27 of the assessment is recommended. For 35-year exposures, the  
33 following formulae would apply:

34  
35  
36 
$$95\% \text{ UCL on extra risk for breast cancer incidence} \approx (8\text{-h TWA occ exp [in ppm]}) \times$$
  
37 
$$(0.031/0.1 \text{ ppm}) = (8\text{-h TWA occ exp [in ppm]}) \times (0.31/\text{ppm})$$

*This document is a draft for review purposes only and does not constitute Agency policy.*

1  
2 MLE of extra risk for breast cancer incidence = (8-h TWA occ exp [in ppm]) ×  
3 (0.016/0.1ppm) = (8-h TWA occ exp [in ppm]) × (0.16/ppm)

4  
5  
6 *Alternatively*, for exposures below 0.1 ppm, one could use the formulae presented below,  
7 which are based on the unit risk estimates presented in Tables 4-22 (for 95% UCLs) and  
8 Table 4-23 (for MLEs), with conversions for adjusting occupational to environmental exposures.  
9 Note, however, that the extra risk results for 35 years of exposure based on these unit risk values  
10 do not exactly match the values in Tables 4-26 and 4-27 for the linear models (the formulae  
11 below yield extra risk estimates that are 15–20% lower than the values in Tables 4-26 and 4-27  
12 for the low end of the exposure range [e.g., 0.1–0.4 ppm] where the comparison with the  
13 unit-risk-based estimates is appropriate). This is because the results in Tables 4-26 and 4-27 are  
14 based on life-table analyses, which take into account age-specific background rates of the  
15 cancers and ages of exposure (assumed to be from 20 to 55 years of age in these occupational  
16 exposure scenarios), whereas the formulae below are approximations that do not take  
17 age-specific considerations into account. The advantage of the formulae based on the unit risk  
18 values is that they can incorporate durations other than ~ 35 years.

19  
20  
21 
$$8\text{-h TWA occ exp [in ppm]} \times (10 \text{ m}^3/\text{day}/20 \text{ m}^3/\text{day}) \times (240 \text{ days}/\text{year}/365 \text{ days}/\text{year}) \times$$
  
22 
$$(35 \text{ years}/70 \text{ years}) = (\text{continuous lifetime}) \text{ env exp [in ppm]}$$

23 (Note that for exposure durations other than 35 years, replace 35 years with the  
24 alternative duration in the formula above.)

25  
26 
$$95\% \text{ UCL on extra risk for lymphoid cancer incidence} \approx 0.795/\text{ppm} \times \text{env exp [in ppm]}$$

27  
28 
$$\text{MLE of extra risk for lymphoid cancer incidence} = 0.356/\text{ppm} \times \text{env exp [in ppm]}$$

29  
30 
$$95\% \text{ UCL on extra risk for breast cancer incidence (in females)} = 1.50/\text{ppm} \times \text{env exp [in}$$
  
31 
$$\text{ppm]}$$

32  
33 
$$\text{MLE of extra risk for breast cancer incidence (in females)} = 0.776/\text{ppm} \times \text{env exp [in}$$
  
34 
$$\text{ppm]}$$

## REFERENCES

This reference list includes all the references cited in the document except for Appendix B, which is a reference list pertaining to Figure 3–3. References added after the 2007 external peer review are also listed separately in Appendix I. References identified in a May 2013 literature search but appearing after the 30 June 2010 cutoff date for literature inclusion into this carcinogenicity assessment are cited and discussed in Appendix J.

- [Adám, B; Bárdos, H; Adány, R.](#) (2005). Increased genotoxic susceptibility of breast epithelial cells to ethylene oxide. *Mutat Res* 585: 120-126.  
<http://dx.doi.org/10.1016/j.mrgentox.2005.04.009>
- [Arias, E.](#) (2007). United States life tables, 2004. Atlanta, GA: Centers of Disease Control and Prevention; National Center for Health Statistics.  
[http://www.cdc.gov/nchs/data/nvsr/nvsr56/nvsr56\\_09.pdf](http://www.cdc.gov/nchs/data/nvsr/nvsr56/nvsr56_09.pdf)
- [Bastlová, T; Andersson, B; Lambert, B; Kolman, A.](#) (1993). Molecular analysis of ethylene oxide-induced mutations at the HPRT locus in human diploid fibroblasts. *Mutat Res* 287: 283-292.
- [BEIR](#) (Committee on the Biological Effects of Ionizing Radiation). (1988). Health risks of radon and other internally deposited alpha-emitters. Washington, DC: National Academy Press.
- [Benson, LO; Teta, MJ.](#) (1993). Mortality due to pancreatic and lymphopietic cancers in chlorohydrin production workers. *Br J Ind Med* 50: 710-716.  
<http://dx.doi.org/10.1136/oem.50.8.710>
- [Beranek, DT.](#) (1990). Distribution of methyl and ethyl adducts following alkylation with monofunctional alkylating agents [Review]. *Mutat Res* 231: 11-30.
- [Bisanti, L; Maggini, M; Raschetti, R; Alegiani, SS; Ippolito, FM; Caffari, B; Segnan, N; Ponti, A.](#) (1993). Cancer mortality in ethylene oxide workers. *Br J Ind Med* 50: 317-324.
- [Boffetta, P; van der Hel, O; Norppa, H; Fabianova, E; Fucic, A; Gundy, S; Lazutka, J; Cebulka-Wasilewska, A; Puskaierova, D; Znaor, A; Kelecsenyi, Z; Kurtinaitis, J; Rachtan, J; Forni, A; Vermeulen, R; Bonassi, S.](#) (2007). Chromosomal aberrations and cancer risk: results of a cohort study from Central Europe. *Am J Epidemiol* 165: 36-43.  
<http://dx.doi.org/10.1093/aje/kwj367>
- [Bolt, HM.](#) (1996). Quantification of endogenous carcinogens. The ethylene oxide paradox [Review]. *Biochem Pharmacol* 52: 1-5.
- [Bonassi, S; Znaor, A; Ceppi, M; Lando, C; Chang, WP; Holland, N; Kirsch-Volders, M; Zeiger, E; Ban, S; Barale, R; Bigatti, MP; Bolognesi, C; Cebulka-Wasilewska, A; Fabianova, E; Fucic, A; Hagmar, L; Joksic, G; Martelli, A; Migliore, L; Mirkova, E; Scarfi, MR; Zijno, A; Norppa, H; Fenech, M.](#) (2007). An increased micronucleus frequency in peripheral blood lymphocytes predicts the risk of cancer in humans. *Carcinogenesis* 28: 625-631.  
<http://dx.doi.org/10.1093/carcin/bgl177>
- [Boogaard, PJ.](#) (2002). Use of haemoglobin adducts in exposure monitoring and risk assessment [Review]. *J Chromatogr B Analyt Technol Biomed Life Sci* 778: 309-322.  
[http://dx.doi.org/10.1016/S0378-4347\(01\)00445-5](http://dx.doi.org/10.1016/S0378-4347(01)00445-5)
- [Brown, CD; Bahman, A; Turner, MJ; Fennell, TR.](#) (1998). Ethylene oxide dosimetry in the mouse. *Toxicol Appl Pharmacol* 148: 215-222. <http://dx.doi.org/10.1006/taap.1997.8349>

*This document is a draft for review purposes only and does not constitute Agency policy.*

- [Brown, CD; Wong, BA; Fennell, TR.](#) (1996). In vivo and in vitro kinetics of ethylene oxide metabolism in rats and mice. *Toxicol Appl Pharmacol* 136: 8-19.  
<http://dx.doi.org/10.1006/taap.1996.0002>
- [Chaganti, SR; Chen, W; Parsa, N; Offit, K; Louie, DC; Dalla-Favera, R; Chaganti, RS.](#) (1998). Involvement of BCL6 in chromosomal aberrations affecting band 3q27 in B-cell non-Hodgkin lymphoma. *Genes Chromosomes Cancer* 23: 323-327.
- [Christiansen, DH; Andersen, MK; Desta, F; Pedersen-Bjergaard, J.](#) (2005). Mutations of genes in the receptor tyrosine kinase (RTK)/RAS-BRAF signal transduction pathway in therapy-related myelodysplasia and acute myeloid leukemia. *Leukemia* 19: 2232-2240.  
<http://dx.doi.org/10.1038/sj.leu.2404009>
- [Christiansen, DH; Andersen, MK; Pedersen-Bjergaard, J.](#) (2001). Mutations with loss of heterozygosity of p53 are common in therapy-related myelodysplasia and acute myeloid leukemia after exposure to alkylating agents and significantly associated with deletion or loss of 5q, a complex karyotype, and a poor prognosis. *J Clin Oncol* 19: 1405-1413.
- [Clare, MG; Dean, BJ; De Jong, G; Van Sittert, NJ.](#) (1985). Chromosome analysis of lymphocytes from workers at an ethylene oxide plant. *DNA Repair* 156: 109-116.
- [Clewell, HJ; Teeguarden, J; McDonald, T; Sarangapani, R; Lawrence, G; Covington, T; Gentry, R; Shipp, A.](#) (2002). Review and evaluation of the potential impact of age- and gender-specific pharmacokinetic differences on tissue dosimetry [Review]. *Crit Rev Toxicol* 32: 329-389. <http://dx.doi.org/10.1080/20024091064264>
- [Coggon, D; Harris, EC; Poole, J; Palmer, KT.](#) (2004). Mortality of workers exposed to ethylene oxide: extended follow up of a British cohort. *Occup Environ Med* 61: 358-362.
- [Csanady, GA; Denk, B; Putz, C; Kreuzer, PE; Kessler, W; Baur, C; Gargas, ML; Filser, JG.](#) (2000). A physiological toxicokinetic model for exogenous and endogenous ethylene and ethylene oxide in rat, mouse, and human: formation of 2-hydroxyethyl adducts with hemoglobin and DNA. *Toxicol Appl Pharmacol* 165: 1-26.  
<http://dx.doi.org/10.1006/taap.2000.8918>
- [Dellarco, VL; Generoso, WM; Sega, GA; Fowle, JR, III; Jacobson-Kram, D.](#) (1990). Review of the mutagenicity of ethylene oxide [Review]. *Environ Mol Mutagen* 16: 85-103.
- [Donner, EM; Wong, BA; James, RA; Preston, RJ.](#) (2010). Reciprocal translocations in somatic and germ cells of mice chronically exposed by inhalation to ethylene oxide: implications for risk assessment. *Mutagenesis* 25: 49-55. <http://dx.doi.org/10.1093/mutage/kep042>
- [Dunkelberg, H.](#) (1982). Carcinogenicity of ethylene oxide and 1,2-propylene oxide upon intragastric administration to rats. *Br J Cancer* 46: 924-933.
- [Ehrenberg, L; Hussain, S.](#) (1981). Genetic toxicity of some important epoxides [Review]. *DNA Repair* 86: 1-113.
- [EOIC](#) (Ethylene Oxide Industry Council). (2001). Toxicological review of ethylene oxide in support of summary information on the integrated risk information system. Arlington, VA.
- [Fennell, TR; Brown, CD.](#) (2001). A physiologically based pharmacokinetic model for ethylene oxide in mouse, rat, and human. *Toxicol Appl Pharmacol* 173: 161-175.  
<http://dx.doi.org/10.1006/taap.2001.9184>
- [Filser, JG; Denk, B; Tornqvist, M; Kessler, W; Ehrenberg, L.](#) (1992). Pharmacokinetics of ethylene in man: body burden with ethylene oxide and hydroxyethylation of hemoglobin due to endogenous and environmental ethylene. *Arch Toxicol* 66: 157-163.

*This document is a draft for review purposes only and does not constitute Agency policy.*

- [Föst, U; Hallier, E; Ottenwälder, H; Bolt, HM; Peter, H.](#) (1991). Distribution of ethylene oxide in human blood and its implications for biomonitoring. *Hum Exp Toxicol* 10: 25-31.
- [Galloway, SM; Berry, PK; Nichols, WW; Wolman, SR; Soper, KA; Stolley, PD; Archer, P.](#) (1986). Chromosome aberrations in individuals occupationally exposed to ethylene oxide, and in a large control population. *Mutat Res Genet Toxicol* 170: 55-74.  
[http://dx.doi.org/10.1016/0165-1218\(86\)90082-0](http://dx.doi.org/10.1016/0165-1218(86)90082-0)
- [Gardner, MJ; Coggon, D; Pannett, B; Harris, EC.](#) (1989). Workers exposed to ethylene oxide: a follow up study. *Occup Environ Med* 46: 860-665.
- [Garman, RH; Snellings, WM; Maronpot, RR.](#) (1985). Brain tumors in F344 rats associated with chronic inhalation exposure to ethylene oxide. *Neurotoxicology* 6: 117-137.
- [Garman, RH; Snellings, WM; Maronpot, RR.](#) (1986). Frequency, size and location of brain tumours in F-344 rats chronically exposed to ethylene oxide. *Food Chem Toxicol* 24: 145-153. [http://dx.doi.org/10.1016/0278-6915\(86\)90349-2](http://dx.doi.org/10.1016/0278-6915(86)90349-2)
- [Garry, VF; Hozier, J; Jacobs, D; Wade, RL; Gray, DG.](#) (1979). Ethylene oxide: evidence of human chromosomal effects. *Environ Mutagen* 1: 375-382.  
<http://dx.doi.org/10.1002/em.2860010410>
- [Gelehrter, TD; Collins, FS; Ginsburg, D.](#) (1990). Principles of medical genetics. Baltimore, MD: Williams and Wilkins.
- [Generoso, WM; Cain, KT; Cornett, CV; Cacheiro, NLA; Hughes, LA.](#) (1990). Concentration-response curves for ethylene-oxide-induced heritable translocations and dominant lethal mutations. *Environ Mol Mutagen* 16: 126-131.
- [Godderis, L; Aka, P; Matecuca, R; Kirsch-Volders, M; Lison, D; Veulemans, H.](#) (2006). Dose-dependent influence of genetic polymorphisms on DNA damage induced by styrene oxide, ethylene oxide and gamma-radiation. *Toxicology* 219: 220-229.  
<http://dx.doi.org/10.1016/j.tox.2005.11.021>
- [Golberg, L.](#) (1986). Chemical and physical properties. In Hazard assessment of ethylene oxide. Boca Raton, FL: CRC Press.
- [Greenberg, HL; Ott, MG; Shore, RE.](#) (1990). Men assigned to ethylene oxide production or other ethylene oxide related chemical manufacturing: A mortality study. *Br J Ind Med* 47: 221-230. <http://dx.doi.org/10.1136/oem.47.4.221>
- [Greife, AL; Hornung, RW; Stayner, LG; Steenland, KN.](#) (1988). Development of a model for use in estimating exposure to ethylene oxide in a retrospective cohort mortality study. *Scand J Work Environ Health* 1: 29-30.
- [Hagmar, L; Mikoczy, Z; Welinder, H.](#) (1995). Cancer incidence in Swedish sterilant workers exposed to ethylene oxide. *Occup Environ Med* 52: 154-156.  
<http://dx.doi.org/10.1136/oem.52.3.154>
- [Hagmar, L; Stromberg, U; Bonassi, S; Hansteen, IL; Knudsen, LE; Lindholm, C; Norppa, H.](#) (2004). Impact of types of lymphocyte chromosomal aberrations on human cancer risk: results from Nordic and Italian cohorts. *Cancer Res* 64: 2258-2263.  
<http://dx.doi.org/10.1158/0008-5472.CAN-03-3360>
- [Hagmar, L; Welinder, H; Lindén, K; Attewell, R; Osterman-Golkar, S; Törnqvist, M.](#) (1991). An epidemiological study of cancer risk among workers exposed to ethylene oxide using hemoglobin adducts to validate environmental exposure assessments. *Int Arch Occup Environ Health* 63: 271-277.

- [Hansen, JP; Allen, J; Brock, K; Falconer, J; Helms, MJ; Shaver, GC; Strohm, B.](#) (1984). Normal sister chromatid exchange levels in hospital sterilization employees exposed to ethylene oxide. *J Occup Med* 26: 29-32.
- [Harada, H; Harada, Y; Tanaka, H; Kimura, A; Inaba, T.](#) (2003). Implications of somatic mutations in the AML1 gene in radiation-associated and therapy-related myelodysplastic syndrome/acute myeloid leukemia. *Blood* 101: 673-680. <http://dx.doi.org/10.1182/blood-2002-04-1010>
- [Harris, NL; Jaffe, ES; Diebold, J; Flandrin, G; Muller-Hermelink, HK; Vardiman, J; Lister, TA; Bloomfield, CD.](#) (1999). World Health Organization classification of neoplastic diseases of the hematopoietic and lymphoid tissues: report of the Clinical Advisory Committee meeting--Airlie House, Virginia, November 1997. *J Clin Oncol* 17: 3835-3849.
- [Haufroid, V; Merz, B; Hofmann, A; Tschopp, A; Lison, D; Hotz, P.](#) (2007). Exposure to ethylene oxide in hospitals: biological monitoring and influence of glutathione S-transferase and epoxide hydrolase polymorphisms. *Cancer Epidemiol Biomarkers Prev* 16: 796-802. <http://dx.doi.org/10.1158/1055-9965.EPI-06-0915>
- [Health Canada.](#) (2001). Priority substances list assessment report: Ethylene oxide. (Cat. no. En40-215/51E). Ottawa, Ontario. [http://www.hc-sc.gc.ca/ewh-semt/pubs/contaminants/psl2-lsp2/ethylene\\_oxide/index-eng.php](http://www.hc-sc.gc.ca/ewh-semt/pubs/contaminants/psl2-lsp2/ethylene_oxide/index-eng.php)
- [Hill, AB.](#) (1965). The environment and disease: Association or causation? *Proc R Soc Med* 58: 295-300.
- [Högstedt, B; Gullberg, B; Hedner, K; Kolnig, AM; Mitelman, F; Skerfving, S; Widegren, B.](#) (1983). Chromosome aberrations and micronuclei in bone marrow cells and peripheral blood lymphocytes in humans exposed to ethylene oxide. *Hereditas* 98: 105-113.
- [Hogstedt, C.](#) (1988). Methods for detecting DNA damaging agents in humans: Applications in cancer epidemiology and prevention Epidemiologic studies on ethylene oxide and cancer: An updating. In IARC Sci Publ. Lyon, France: International Agency for Research on Cancer.
- [Hogstedt, C; Aringer, L; Gustavsson, A.](#) (1986). Epidemiologic support for ethylene oxide as a cancer-causing agent. *JAMA* 255: 1575-1578.
- [Hong, HH; Houle, CD; Ton, TV; Sills, RC.](#) (2007). K-ras mutations in lung tumors and tumors from other organs are consistent with a common mechanism of ethylene oxide tumorigenesis in the B6C3F1 mouse. *Toxicol Pathol* 35: 81-85. <http://dx.doi.org/10.1080/01926230601063839>
- [Horner, MJ; Ries, LAG; Krapcho, M; Neyman, N; Aminou, R; Howlader, N; Altekruse, SF; Feuer, EJ; Huang, L; Mariotto, A; Miller, BA; Lewis, DR; Eisner, MP; Stinchcomb, DG; Edwards, BK.](#) (2009). SEER cancer statistics review, 1975-2006. Bethesda, MD: National Cancer Institute. [http://seer.cancer.gov/csr/1975\\_2006/](http://seer.cancer.gov/csr/1975_2006/)
- [Hornung, RW; Greife, AL; Stayner, LT; Steenland, NK; Herrick, RF; Elliott, LJ; Ringenburg, VL; Morawetz, J.](#) (1994). Statistical model for prediction of retrospective exposure to ethylene oxide in an occupational mortality study. *Am J Ind Med* 25: 825-836.
- [Houle, CD; Ton, TV; Clayton, N; Huff, J; Hong, HH; Sills, RC.](#) (2006). Frequent p53 and H-ras mutations in benzene- and ethylene oxide-induced mammary gland carcinomas from B6C3F1 mice. *Toxicol Pathol* 34: 752-762. <http://dx.doi.org/10.1080/01926230600935912>

*This document is a draft for review purposes only and does not constitute Agency policy.*

- [IARC](#) (International Agency for Research on Cancer). (1994a). Ethylene. In IARC monographs on the evaluation of carcinogenic risks to humans Some industrial chemicals (pp. 45-71). Lyon, France. <http://monographs.iarc.fr/ENG/Monographs/vol60/volume60.pdf>
- [IARC](#) (International Agency for Research on Cancer). (1994b). Ethylene oxide. In IARC Monographs on the Evaluation of Carcinogenic Risks to Humans (pp. 73-159). Lyon, France. <http://monographs.iarc.fr/ENG/Monographs/vol60/volume60.pdf>
- [IARC](#) (International Agency for Research on Cancer). (2008). 1,3-butadiene, ethylene oxide and vinyl halides (vinyl fluoride, vinyl chloride and vinyl bromide). In IARC monographs on the evaluation of carcinogenic risks to humans (pp. 3-471). Lyon, France. <http://monographs.iarc.fr/ENG/Monographs/vol97/mono97.pdf>
- [Ingvarsson, S.](#) (1999). Molecular genetics of breast cancer progression [Review]. *Semin Cancer Biol* 9: 277-288. <http://dx.doi.org/10.1006/scbi.1999.0124>
- [Johanson, G; Filser, JG.](#) (1992). Experimental data from closed chamber gas uptake studies in rodents suggest lower uptake rate of chemical than calculated from literature values on alveolar ventilation. *Arch Toxicol* 66: 291-295.
- [Kardos, L; Széles, G; Gombkőto, G; Szeremi, M; Tompa, A; Adány, R.](#) (2003). Cancer deaths among hospital staff potentially exposed to ethylene oxide: an epidemiological analysis. *Environ Mol Mutagen* 42: 59-60. <http://dx.doi.org/10.1002/em.10167>
- [Kiesselbach, N; Ulm, K; Lange, HJ; Korallus, U.](#) (1990). A multicentre mortality study of workers exposed to ethylene oxide. *Br J Ind Med* 47: 182-188.
- [Kirman, CR; Sweeney, LM; Teta, MJ; Sielken, RL; Valdez-Flores, C; Albertini, RJ; Gargas, ML.](#) (2004). Addressing nonlinearity in the exposure-response relationship for a genotoxic carcinogen: cancer potency estimates for ethylene oxide. *Risk Anal* 24: 1165-1183. <http://dx.doi.org/10.1111/j.0272-4332.2004.00517.x>
- [Kolman, A; Chovanec, M.](#) (2000). Combined effects of gamma-radiation and ethylene oxide in human diploid fibroblasts. *Mutagenesis* 15: 99-104. <http://dx.doi.org/10.1093/mutage/15.2.99>
- [Kolman, A; Chovanec, M; Osterman-Golkar, S.](#) (2002). Genotoxic effects of ethylene oxide, propylene oxide and epichlorohydrin in humans: update review (1990-2001) [Review]. *DNA Repair* 512: 173-194.
- [Krewski, D; Crump, KS; Farmer, J; Gaylor, DW; Howe, R; Portier, C; Salsburg, D; Sielken, RL; Van Ryzin, J.](#) (1983). A comparison of statistical methods for low dose extrapolation utilizing time-to-tumour data. *Fundam Appl Toxicol* 3: 140-160. [http://dx.doi.org/10.1016/S0272-0590\(83\)80075-X](http://dx.doi.org/10.1016/S0272-0590(83)80075-X)
- [Krishnan, K; Gargas, ML; Fennell, TR; Andersen, ME.](#) (1992). A physiologically based description of ethylene oxide dosimetry in the rat. *Toxicol Ind Health* 8: 121-140.
- [Lambert, B; Andersson, B; Bastlova, T; Hou, SM; Hellgren, D; Kolman, A.](#) (1994). Mutations induced in the hypoxanthine phosphoribosyl transferase gene by three urban air pollutants: Acetaldehyde, benzo[a]pyrene diolepoxide, and ethylene oxide. *Environ Health Perspect Suppl* 102: 135-138.
- [Langholz, B; Richardson, DB.](#) (2010). Fitting general relative risk models for survival time and matched case-control analysis. *Am J Epidemiol* 171: 377-383. <http://dx.doi.org/10.1093/aje/kwp403>
- [Laurent, C; Frederic, J; Léonard, AY.](#) (1984). Sister chromatid exchange frequency in workers exposed to high levels of ethylene oxide, in a hospital sterilization service. *Int Arch Occup Environ Health* 54: 33-43.

*This document is a draft for review purposes only and does not constitute Agency policy.*

- [Lerda, D; Rizzi, R.](#) (1992). Cytogenetic study of persons occupationally exposed to ethylene oxide. *Mutat Res* 281: 31-37.
- [Lewis, SE; Barnett, LB; Felton, C; Johnson, FM; Skow, LC; Cacheiro, N; Shelby, MD.](#) (1986). Dominant visible and electrophoretically expressed mutations induced in male mice exposed to ethylene oxide by inhalation. *Environ Mol Mutagen* 8: 867-872.
- [Liou, SH; Lung, JC; Chen, YH; Yang, T; Hsieh, LL; Chen, CJ; Wu, TN.](#) (1999). Increased chromosome-type chromosome aberration frequencies as biomarkers of cancer risk in a blackfoot endemic area. *Cancer Res* 59: 1481-1484.
- [Lorenti Garcia, C; Darroudi, F; Tates, AD; Natarajan, AT.](#) (2001). Induction and persistence of micronuclei, sister-chromatid exchanges and chromosomal aberrations in splenocytes and bone-marrow cells of rats exposed to ethylene oxide. *Mutat Res Genet Toxicol Environ Mutagen* 492: 59-67. [http://dx.doi.org/10.1016/S1383-5718\(01\)00149-8](http://dx.doi.org/10.1016/S1383-5718(01)00149-8)
- [Lossos, IS; Levy, R.](#) (2000). Mutation analysis of the 5' noncoding regulatory region of the BCL-6 gene in non-Hodgkin lymphoma: evidence for recurrent mutations and intraclonal heterogeneity. *Blood* 95: 1400-1405.
- [Lynch, DW; Lewis, TR; Moorman, WJ; Burg, JR; Groth, DH; Khan, A; Ackerman, LJ; Cockrell, BY.](#) (1984a). Carcinogenic and toxicologic effects of inhaled ethylene oxide and propylene oxide in F344 rats. *Toxicol Appl Pharmacol* 76: 69-84.
- [Lynch, DW; Lewis, TR; Moorman, WJ; Burg, JR; Lal, JB; Setzer, JV; Groth, DH; Gulati, DK; Zavos, PM; Sabharwal, PS; Ackerman, LJ; Cockrell, BY; Sprinz, H.](#) (1984b). Effects on monkeys and rats of long-term inhalation exposure to ethylene oxide: Major findings of the NIOSH study. In *Inhospital ethylene oxide sterilization: Current issues in ethylene oxide toxicity and occupational exposure* (pp. 7-10). (AAMI Technology Assessment Report No. 8-84). Arlington, VA: Association for the Advancement of Medical Instrumentation.
- [Major, J; Jakab, MG; Tompa, A.](#) (1996). Genotoxicological investigation of hospital nurses occupationally exposed to ethylene-oxide: I Chromosome aberrations, sister-chromatid exchanges, cell cycle kinetics, and UV-induced DNA synthesis in peripheral blood lymphocytes. *Environ Mol Mutagen* 27: 84-92. [http://dx.doi.org/10.1002/\(SICI\)1098-2280\(1996\)27:2<84::AID-EM2>3.0.CO;2-E](http://dx.doi.org/10.1002/(SICI)1098-2280(1996)27:2<84::AID-EM2>3.0.CO;2-E)
- [Major, J; Jakab, MG; Tompa, A.](#) (2001). Genotoxicological investigation of hospital nurses occupationally exposed to ethylene oxide. II. HPRT mutation frequencies. *Central Eur J Occup Env Med* 7: 195-208.
- [Marsden, DA; Jones, DJ; Britton, RG; Ognibene, T; Ubick, E; Johnson, GE; Farmer, PB; Brown, K.](#) (2009). Dose-response relationships for N7-(2-hydroxyethyl)guanine induced by low-dose [<sup>14</sup>C]ethylene oxide: evidence for a novel mechanism of endogenous adduct formation. *Cancer Res* 69: 3052-3059. <http://dx.doi.org/10.1158/0008-5472.CAN-08-4233>
- [Mayer, J; Warburton, D; Jeffrey, AM; Pero, R; Walles, S; Andrews, L; Toor, M; Latriano, L; Wazneh, L; Tang, D; Tsai, WY; Kuroda, M; Perera, F.](#) (1991). Biologic markers in ethylene oxide exposed workers and controls. *Mutat Res* 248: 163-176. [http://dx.doi.org/10.1016/0027-5107\(91\)90098-9](http://dx.doi.org/10.1016/0027-5107(91)90098-9)
- [Memisoglu, A; Samson, L.](#) (2000). Base excision repair in yeast and mammals [Review]. *Mutat Res* 451: 39-51.

- Mertes, I; Fleischmann, R; Glatt, H; Oesch, F. (1985). Interindividual variations in the activities of cytosolic and microsomal epoxide hydrolase in human liver. *Carcinogenesis* 6: 219-223.
- Miniño, AM; Arias, E; Kochanek, KD; Murphy, SL; Smith, BL. (2002). Deaths: Final Data for 2000. In *National Vital Statistics Reports*. Hyattsville, MD: National Center for Health Statistics. [http://www.cdc.gov/nchs/data/nvsr/nvsr50/nvsr50\\_15.pdf](http://www.cdc.gov/nchs/data/nvsr/nvsr50/nvsr50_15.pdf)
- Natarajan, AT; Preston, RJ; Dellarco, V; Ehrenberg, L; Generoso, W; Lewis, S; Tates, AD. (1995). Ethylene oxide: evaluation of genotoxicity data and an exploratory assessment of genetic risk [Review]. *DNA Repair* 330: 55-70.
- NCI (National Cancer Institute). (2004). Breast cancer screening (PDQ®): Health professional version. Washington, DC: U.S. Department of Health, Education, and Welfare, National Institutes of Health. <http://www.cancer.gov/cancertopics/pdq/screening/breast/HealthProfessional/>
- Norman, SA; Berlin, JA; Soper, KA; Middendorf, BF; Stolley, PD. (1995). Cancer incidence in a group of workers potentially exposed to ethylene oxide. *Int J Epidemiol* 24: 276-284. <http://dx.doi.org/10.1093/ije/24.2.276>
- NRC (National Research Council). (1983). Risk assessment in the federal government: Managing the process. Washington, DC: National Academies Press. [http://www.nap.edu/openbook.php?record\\_id=366&page=R1](http://www.nap.edu/openbook.php?record_id=366&page=R1)
- NRC (National Research Council). (2011). Review of the Environmental Protection Agency's draft IRIS assessment of formaldehyde. Washington, DC: National Academies Press. <http://www.nap.edu/catalog/13142.html>
- NTP (National Toxicology Program). (1987). Toxicology and carcinogenesis studies of ethylene oxide (CAS no 75-21-8) in B6C3F1 mice (inhalation studies).
- Olsen, GW; Lacy, SE; Bodner, KM; Chau, M; Arceneaux, TG; Cartmill, JB; Ramlow, JM; Boswell, JM. (1997). Mortality from pancreatic and lymphopoietic cancer among workers in ethylene and propylene chlorohydrin production. *Occup Environ Med* 54: 592-598. <http://dx.doi.org/10.1136/oem.54.8.592>
- Pauwels, W; Veulemans, H. (1998). Comparison of ethylene, propylene and styrene 7,8-oxide in vitro adduct formation on N-terminal valine in human haemoglobin and on N-7-guanine in human DNA. *Mutat Res* 418: 21-33.
- Paz-y-Miño, C; Pérez, JC; Fiallo, BF; Leone, PE. (2002). A polymorphism in the hMSH2 gene (gIVS12-6T&gt;C) associated with non-Hodgkin lymphomas. *Cancer Genet Cytogenet* 133: 29-33.
- Pedersen-Bjergaard, J; Christiansen, DH; Desta, F; Andersen, MK. (2006). Alternative genetic pathways and cooperating genetic abnormalities in the pathogenesis of therapy-related myelodysplasia and acute myeloid leukemia [Review]. *Leukemia* 20: 1943-1949. <http://dx.doi.org/10.1038/sj.leu.2404381>
- Pfeiffer, P; Goedecke, W; Obe, G. (2000). Mechanisms of DNA double-strand break repair and their potential to induce chromosomal aberrations [Review]. *Mutagenesis* 15: 289-302.
- Popp, W; Vahrenholz, C; Przygoda, H; Brauksiepe, A; Goch, S; Müller, G; Schell, C; Norporth, K. (1994). DNA-protein cross-links and sister chromatid exchange frequencies in lymphocytes and hydroxyethyl mercapturic acid in urine of ethylene oxide-exposed hospital workers. *Int Arch Occup Environ Health* 66: 325-332.

*This document is a draft for review purposes only and does not constitute Agency policy.*

- Preston, RJ. (1999). Cytogenetic effects of ethylene oxide, with an emphasis on population monitoring [Review]. *Crit Rev Toxicol* 29: 263-282.  
<http://dx.doi.org/10.1080/10408449991349212>
- Preston, RJ; Fennell, TR; Leber, AP; Sielken, RL; Swenberg, JA. (1995). Reconsideration of the genetic risk assessment for ethylene oxide exposures. *Environ Mol Mutagen* 26: 189-202.  
<http://dx.doi.org/10.1002/em.2850260303>
- Preudhomme, C; Warot-Loze, D; Roumier, C; Gardel-Duflos, N; Garand, R; Lai, JL; Dastugue, N; Macintyre, E; Denis, C; Bauters, F; Kerckaert, JP; Cosson, A; Fenaux, P. (2000). High incidence of biallelic point mutations in the Runt domain of the AML1/PEBP2 alpha B gene in Mo acute myeloid leukemia and in myeloid malignancies with acquired trisomy 21. *Blood* 96: 2862-2869.
- Rapoport, IA. (1948). The effect of ethylene oxide, glycide and glycol on genetic mutations. *Dokl Biochem Biophys* 60: 469-472.
- Recio, L; Donner, M; Abernethy, D; Pluta, L; Steen, AM; Wong, BA; James, A; Preston, RJ. (2004). In vivo mutagenicity and mutation spectrum in the bone marrow and testes of B6C3F1 lacI transgenic mice following inhalation exposure to ethylene oxide. *Mutagenesis* 19: 215-222.
- Ribeiro, LR; Salvadori, DM; Rios, AC; Costa, SL; Tates, AD; Törnqvist, M; Natarajan, AT. (1994). Biological monitoring of workers occupationally exposed to ethylene oxide. *Mutat Res* 313: 81-87.
- Richmond, GW; Abrahams, RH; Nemenzo, JH; Hine, CH. (1985). An evaluation of possible effects on health following exposure to ethylene oxide. *Arch Environ Occup Health* 40: 20-25.
- Ries, LAG; Eisner, MP; Kosary, CL; Hankey, BF; Miller, BA; Clegg, L; Mariotto, A; Feuer, EF; Edwards, BK. (2004). SEER (Surveillance Epidemiology and End Results) cancer statistics review, 1975-2001. Bethesda, MD: National Cancer Institute, U.S. Department of Health, Education, and Welfare, National Institutes of Health.  
[http://seer.cancer.gov/csr/1975\\_2001](http://seer.cancer.gov/csr/1975_2001)
- Ries, LAG; Melbert, D; Krapcho, M; Mariotto, A; Miller, BA; Feuer, EJ; Clegg, L; Horner, MJ; Howlander, N; Eisner, MP. (2007). SEER (Surveillance Epidemiology and End Results) cancer statistics review, 1975–2004. Bethesda, MD: National Cancer Institute, U.S. Department of Health, Education, and Welfare, National Institutes of Health.  
[http://seer.cancer.gov/csr/1975\\_2004](http://seer.cancer.gov/csr/1975_2004)
- Rossner, P; Boffetta, P; Ceppi, M; Bonassi, S; Smerhovsky, Z; Landa, K; Juzova, D; Sram, RJ. (2005). Chromosomal aberrations in lymphocytes of healthy subjects and risk of cancer. *Environ Health Perspect* 113: 517-520. <http://dx.doi.org/10.1289/ehp.6925>
- Rothman, KJ. (1986). *Modern epidemiology*. Boston, MA: Little Brown & Co.
- Russo, J; Russo, IH. (1999). Cellular basis of breast cancer susceptibility [Review]. *Oncol Res* 11: 169-178.
- Rusyn, I; Asakura, S; Li, Y; Kosyk, O; Koc, H; Nakamura, J; Upton, PB; Swenberg, JA. (2005). Effects of ethylene oxide and ethylene inhalation on DNA adducts, apurinic/aprimidinic sites and expression of base excision DNA repair genes in rat brain, spleen, and liver. *DNA Repair* 4: 1099-1110. <http://dx.doi.org/10.1016/j.dnarep.2005.05.009>
- SAB (Science Advisory Board). (2007). Review of Office of Research and Development (ORD) draft assessment entitled "Evaluation of the carcinogenicity of ethylene oxide". Washington, DC: Science Advisory Board, U.S. Environmental Protection Agency.

*This document is a draft for review purposes only and does not constitute Agency policy.*

- [http://yosemite.epa.gov/sab/sabproduct.nsf/368203f97a15308a852574ba005bbd01/5D661BC118B527A3852573B80068C97B/\\$File/EPA-SAB-08-004-unsigned.pdf](http://yosemite.epa.gov/sab/sabproduct.nsf/368203f97a15308a852574ba005bbd01/5D661BC118B527A3852573B80068C97B/$File/EPA-SAB-08-004-unsigned.pdf)
- [Sarto, F; Clonfero, E; Bartolucci, GB; Franceschi, C; Chiricolo, M; Levis, AG.](#) (1987). Sister chromatid exchanges and DNA repair capability in sanitary workers exposed to ethylene oxide: Evaluation of the dose-effect relationship. *Am J Ind Med* 12: 625-637.  
<http://dx.doi.org/10.1002/ajim.4700120515>
- [Sarto, F; Cominato, I; Pinton, AM; Brovedani, PG; Faccioli, CM; Bianchi, V; Levis, AG.](#) (1984). Cytogenetic damage in workers exposed to ethylene oxide. *Mutat Res* 138: 185-195.
- [Sarto, F; Tomanin, R; Giacomelli, L; Iannini, G; Cupiraggi, AR.](#) (1990). The micronucleus assay in human exfoliated cells of the nose and mouth: application to occupational exposures to chromic acid and ethylene oxide. *Mutat Res* 244: 345-351.
- [Sarto, F; Törnqvist, MA; Tomanin, R; Bartolucci, GB; Osterman-Golkar, SM; Ehrenberg, L.](#) (1991). Studies of biological and chemical monitoring of low-level exposure to ethylene oxide. *Scand J Work Environ Health* 17: 60-64.
- [Schulte, PA; Boeniger, M; Walker, JT; Schober, SE; Pereira, MA; Gulati, DK; Wojciechowski, JP; Garza, A; Froelich, R; Strauss, G.](#) (1992). Biologic markers in hospital workers exposed to low levels of ethylene oxide. *Mutat Res* 278: 237-251.
- [Segerbäck, D.](#) (1990). Reaction products in hemoglobin and DNA after in vitro treatment with ethylene oxide and N-(2-hydroxyethyl)-N-nitrosourea. *Carcinogenesis* 11: 307-312.
- [Sielken, RL; Valdez-Flores, C.](#) (2009). Life-table calculations of excess risk for incidence versus mortality: ethylene oxide case study. *Regul Toxicol Pharmacol* 55: 82-89.  
<http://dx.doi.org/10.1016/j.yrtph.2009.06.003>
- [Sisk, SC; Pluta, LJ; Meyer, KG; Wong, BC; Recio, L.](#) (1997). Assessment of the in vivo mutagenicity of ethylene oxide in the tissues of B6C3F1 lacI transgenic mice following inhalation exposure. *Mutat Res* 391: 153-164.
- [Snellings, WM; Weil, CS; Maronpot, RR.](#) (1984). A two-year inhalation study of the carcinogenic potential of ethylene oxide in Fischer 344 rats. *Toxicol Appl Pharmacol* 75: 105-117.
- [Starr, TB; Swenberg, JA.](#) (2013). A novel bottom-up approach to bounding low-dose human cancer risks from chemical exposures. *Regul Toxicol Pharmacol* 65: 311-315.  
<http://dx.doi.org/10.1016/j.yrtph.2013.01.004>
- [Stayner, L; Steenland, K; Dosemeci, M; Hertz-Picciotto, I.](#) (2003). Attenuation of exposure-response curves in occupational cohort studies at high exposure levels. *Scand J Work Environ Health* 29: 317-324.
- [Stayner, L; Steenland, K; Greife, A; Hornung, R; Hayes, RB; Nowlin, S; Morawetz, J; Ringenburg, V; Elliot, L; Halperin, W.](#) (1993). Exposure-response analysis of cancer mortality in a cohort of workers exposed to ethylene oxide. *Am J Epidemiol* 138: 787-798.
- [Steenland, K; Deddens, J; Piacitelli, L.](#) (2001). Risk assessment for 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) based on an epidemiologic study. *Am J Epidemiol* 154: 451-458.
- [Steenland, K; Deddens, JA.](#) (2004). A practical guide to dose-response analyses and risk assessment in occupational epidemiology [Review]. *Epidemiology* 15: 63-70.  
<http://dx.doi.org/10.1097/01.ede.0000100287.45004.e7>
- [Steenland, K; Stayner, L; Deddens, J.](#) (2004). Mortality analyses in a cohort of 18 235 ethylene oxide exposed workers: follow up extended from 1987 to 1998. *Occup Environ Med* 61: 2-7.

*This document is a draft for review purposes only and does not constitute Agency policy.*

- [Steenland, K; Stayner, L; Greife, A; Halperin, W; Hayes, R; Hornung, R; Nowlin, S.](#) (1991). Mortality among workers exposed to ethylene oxide. *N Engl J Med* 324: 1402-1407. <http://dx.doi.org/10.1056/NEJM199105163242004>
- [Steenland, K; Whelan, E; Deddens, J; Stayner, L; Ward, E.](#) (2003). Ethylene oxide and breast cancer incidence in a cohort study of 7576 women (United States). *Cancer Causes Control* 14: 531-539.
- [Stolley, PD; Soper, KA; Galloway, SM; Nichols, WW; Norman, SA; Wolman, SR.](#) (1984). Sister-chromatid exchanges in association with occupational exposure to ethylene oxide. *Mutat Res* 129: 89-102. [http://dx.doi.org/10.1016/0027-5107\(84\)90127-1](http://dx.doi.org/10.1016/0027-5107(84)90127-1)
- [Swaen, GM; Burns, C; Teta, JM; Bodner, K; Keenan, D; Bodnar, CM.](#) (2009). Mortality study update of ethylene oxide workers in chemical manufacturing: a 15 year update. *J Occup Environ Med* 51: 714-723. <http://dx.doi.org/10.1097/JOM.0b013e3181a2ca20>
- [Swaen, GMH; Slangen, JMM; Ott, MG; Kusters, E; Van Den Langenbergh, G; Arends, JW; Zober, A.](#) (1996). Investigation of a cluster of ten cases of Hodgkin's disease in an occupational setting. *Int Arch Occup Environ Health* 68: 224-228. <http://dx.doi.org/10.1007/BF00381432>
- [Swenberg, JA; Fedtke, N; Fennell, TR; Walker, VE.](#) (1990). Relationships between carcinogen exposure, DNA adducts and carcinogenesis. In DB Clayson; IC Munro; P Shubik; JA Swenberg (Eds.), (pp. 161-184). New York, NY: Elsevier.
- [Tates, AD; Boogaard, PJ; Darroudi, F; Natarajan, AT; Caubo, ME; Van Sittert, NJ.](#) (1995). Biological effect monitoring in industrial workers following incidental exposure to high concentrations of ethylene oxide. *DNA Repair* 329: 63-77.
- [Tates, AD; Grummt, T; Törnqvist, M; Farmer, PB; van Dam, FJ; van Mossel, H; Schoemaker, HM; Osterman-Golkar, S; Uebel, C; Tang, YS; Zwinderman, AH; Natarajan, AT; Ehrenberg, L.](#) (1991). Biological and chemical monitoring of occupational exposure to ethylene oxide. *Mutat Res* 250: 483-497. [http://dx.doi.org/10.1016/0027-5107\(91\)90205-3](http://dx.doi.org/10.1016/0027-5107(91)90205-3)
- [Tates, AD; Van Dam, FJ; Natarajan, AT; Van Teylingen, CMM; De Zwart, FA; Zwinderman, AH; Van Sittert, NJ; Nilsen, A; Nilsen, OG; Zahlsen, K; Magnusson, AL; Tornqvist, M.](#) (1999). Measurement of HPRT mutations in splenic lymphocytes and haemoglobin adducts in erythrocytes of Lewis rats exposed to ethylene oxide. *DNA Repair* 431: 397-415.
- [Teta, MJ; Benson, LO; Vitale, JN.](#) (1993). Mortality study of ethylene oxide workers in chemical manufacturing: a 10 year update. *Br J Ind Med* 50: 704-709.
- [Teta, MJ; Sielken, RL; Valdez-Flores, C.](#) (1999). Ethylene oxide cancer risk assessment based on epidemiological data: application of revised regulatory guidelines. *Risk Anal* 19: 1135-1155.
- [Thier, R; Bolt, HM.](#) (2000). Carcinogenicity and genotoxicity of ethylene oxide: new aspects and recent advances [Review]. *Crit Rev Toxicol* 30: 595-608. <http://dx.doi.org/10.1080/10408440008951121>
- [Tompa, A; Major, J; Jakab, MG.](#) (1999). Is breast cancer cluster influenced by environmental and occupational factors among hospital nurses in Hungary. *Pathol Oncol Res* 5: 117-121.
- [Tompkins, EM; McLuckie, KI; Jones, DJ; Farmer, PB; Brown, K.](#) (2009). Mutagenicity of DNA adducts derived from ethylene oxide exposure in the pSP189 shuttle vector replicated in

*This document is a draft for review purposes only and does not constitute Agency policy.*

- human Ad293 cells. *Mutat Res* 678: 129-137.  
<http://dx.doi.org/10.1016/j.mrgentox.2009.05.011>
- Törnqvist, M. (1996). Ethylene oxide as a biological reactive intermediate of endogenous origin [Review]. *Adv Exp Med Biol* 387: 275-283.
- U.S. EPA (U.S. Environmental Protection Agency). (1986). Guidelines for carcinogen risk assessment. *Fed Reg* 51: 33993-34003.
- U.S. EPA (U.S. Environmental Protection Agency). (1994). Methods for derivation of inhalation reference concentrations and application of inhalation dosimetry. (EPA/600/8-90/066F). Research Triangle Park, NC: U.S. Environmental Protection Agency, Environmental Criteria and Assessment Office.  
<http://cfpub.epa.gov/ncea/cfm/recordisplay.cfm?deid=71993>
- U.S. EPA (U.S. Environmental Protection Agency). (1997). Chemical and radiation leukemogenesis in humans and rodents and the value of rodent models for assessing risks of lymphohematopoietic cancers [EPA Report]. (EPA/600/R-97/090). Washington, DC: National Center for Environmental Assessment, Office of Research and Development.  
<http://cfpub.epa.gov/ncea/cfm/recordisplay.cfm?deid=12181>
- U.S. EPA (U.S. Environmental Protection Agency). (2000). Science policy council handbook: risk characterization. (EPA/100/B-00/002). Washington, D.C.: U.S. Environmental Protection Agency, Office of Science Policy.  
<http://www.epa.gov/osa/spc/pdfs/rhandbk.pdf>
- U.S. EPA (U.S. Environmental Protection Agency). (2005a). Guidelines for carcinogen risk assessment. (EPA/630/P-03/001F). Washington, DC: U.S. Environmental Protection Agency, Risk Assessment Forum. <http://www.epa.gov/cancerguidelines/>
- U.S. EPA (U.S. Environmental Protection Agency). (2005b). Supplemental guidance for assessing susceptibility from early-life exposure to carcinogens. (EPA/630/R-03/003F). Washington, DC: U.S. Environmental Protection Agency, Risk Assessment Forum.  
[http://www.epa.gov/raf/publications/pdfs/childrens\\_supplement\\_final.pdf](http://www.epa.gov/raf/publications/pdfs/childrens_supplement_final.pdf)
- U.S. EPA (U.S. Environmental Protection Agency). (2006a). Evaluation of the carcinogenicity of ethylene oxide: external review draft [EPA Report]. (EPA/635/R-06/003). Washington, DC: National Center for Environmental Assessment, Office of Research and Development. <http://cfpub.epa.gov/ncea/cfm/recordisplay.cfm?deid=157664>
- U.S. EPA (U.S. Environmental Protection Agency). (2006b). Science policy council peer review handbook 3rd edition. (EPA/100/B-06/002). U.S. Environmental Protection Agency, Science Policy Council. <http://cfpub.epa.gov/ncea/cfm/recordisplay.cfm?deid=157664>
- U.S. EPA (U.S. Environmental Protection Agency). (2012). Benchmark dose technical guidance. (EPA/100/R-12/001). Washington, DC: Risk Assessment Forum.  
[http://www.epa.gov/raf/publications/pdfs/benchmark\\_dose\\_guidance.pdf](http://www.epa.gov/raf/publications/pdfs/benchmark_dose_guidance.pdf)
- Valdez-Flores, C; Sielken, RL; Teta, MJ. (2010). Quantitative cancer risk assessment based on NIOSH and UCC epidemiological data for workers exposed to ethylene oxide. *Regul Toxicol Pharmacol* 56: 312-320. <http://dx.doi.org/10.1016/j.yrtph.2009.10.001>
- van Delft, JH; van Winden, MJ; Luiten-Schuite, A; Ribeiro, LR; Baan, RA. (1994). Comparison of various immunochemical assays for the detection of ethylene oxide-DNA adducts with monoclonal antibodies against imidazole ring-opened N7-(2-hydroxyethyl)guanosine: application in a biological monitoring study. *Carcinogenesis* 15: 1867-1873.

*This document is a draft for review purposes only and does not constitute Agency policy.*

- [van Sittert, NJ; Beulink, GD; van Vliet, EW; van der Waal, H.](#) (1993). Monitoring occupational exposure to ethylene oxide by the determination of hemoglobin adducts. *Environ Health Perspect* 99: 217-220.
- [van Sittert, NJ; Boogaard, PJ; Natarajan, AT; Tate, AD; Ehrenberg, LG; Törnqvist, MA.](#) (2000). Formation of DNA adducts and induction of mutagenic effects in rats following 4 weeks inhalation exposure to ethylene oxide as a basis for cancer risk assessment. *Mutat Res-Fundam Mol Mech Mutagen* 447: 27-48. [http://dx.doi.org/10.1016/S0027-5107\(99\)00208-0](http://dx.doi.org/10.1016/S0027-5107(99)00208-0)
- [Van Sittert, NJ; de Jong, G.](#) (1985). Biomonitoring of exposure to potential mutagens and carcinogens in industrial populations. *Food Chem Toxicol* 23: 23-31.
- [van Wijngaarden, E; Hertz-Picciotto, I.](#) (2004). A simple approach to performing quantitative cancer risk assessment using published results from occupational epidemiology studies. *Sci Total Environ* 332: 81-87. <http://dx.doi.org/10.1016/j.scitotenv.2004.04.005>
- [Walker, VE; Fennell, TR; Boucheron, JA; Fedtke, N; Ciroussel, F; Swenberg, JA.](#) (1990). Macromolecular adducts of ethylene oxide: a literature review and a time-course study on the formation of 7-(2-hydroxyethyl)guanine following exposures of rats by inhalation [Review]. *DNA Repair* 233: 151-164.
- [Walker, VE; Fennell, TR; Upton, PB; MacNeela, JP; Swenberg, JA.](#) (1993). Molecular dosimetry of DNA and hemoglobin adducts in mice and rats exposed to ethylene oxide. *Environ Health Perspect* 99: 11-17.
- [Walker, VE; Fennell, TR; Upton, PB; Skopek, TR; Prevost, V; Shuker, DEG; Swenberg, JA.](#) (1992a). Molecular dosimetry of ethylene oxide: formation and persistence of 7-(2-hydroxyethyl)guanine in DNA following repeated exposures of rats and mice. *Cancer Res* 52: 4328-4334.
- [Walker, VE; MacNeela, JP; Swenberg, JA; Turner, MJ, Jr; Fennell, TR.](#) (1992b). Molecular dosimetry of ethylene oxide: formation and persistence of N-(2-hydroxyethyl)valine in hemoglobin following repeated exposures of rats and mice. *Cancer Res* 52: 4320-4327.
- [Walker, VE; Sisk, SC; Upton, PB; Wong, BA; Recio, L.](#) (1997). In vivo mutagenicity of ethylene oxide at the hprt locus in T-lymphocytes of B6C3F1 lacI transgenic mice following inhalation exposure. *Mutat Res* 392: 211-222.
- [Walker, VE; Skopek, TR.](#) (1993). A mouse model for the study of in vivo mutational spectra: sequence specificity of ethylene oxide at the hprt locus. *Mutat Res* 288: 151-162. [http://dx.doi.org/10.1016/0027-5107\(93\)90216-3](http://dx.doi.org/10.1016/0027-5107(93)90216-3)
- [Warwick, GP.](#) (1963). The mechanism of action of alkylating agents [Review]. *Cancer Res* 23: 1315-1333.
- [WHO](#) (World Health Organization). (2003). Concise International Chemical Assessment Document: Ethylene oxide. Geneva: International Programme on Chemical Safety (IPCS). <http://www.inchem.org/documents/cicads/cicad54.htm>
- [Wong, O; Trent, LS.](#) (1993). An epidemiological study of workers potentially exposed to ethylene oxide. *Occup Environ Med* 50: 308-316.
- [Yager, JW; Hines, CJ; Spear, RC.](#) (1983). Exposure to ethylene oxide at work increases sister chromatid exchanges in human peripheral lymphocytes. *Science* 219: 1221-1223. <http://dx.doi.org/10.1126/science.6828851>
- [Yong, LC; Schulte, PA; Kao, CY; Giese, RW; Boeniger, MF; Strauss, GH; Petersen, MR; Wiencke, JK.](#) (2007). DNA adducts in granulocytes of hospital workers exposed to ethylene oxide. *Am J Ind Med* 50: 293-302. <http://dx.doi.org/10.1002/ajim.20443>

*This document is a draft for review purposes only and does not constitute Agency policy.*

[Yong, LC; Schulte, PA; Wiencke, JK; Boeniger, MF; Connally, LB; Walker, JT; Whelan, EA; Ward, EM.](#) (2001). Hemoglobin adducts and sister chromatid exchanges in hospital workers exposed to ethylene oxide: effects of glutathione S-transferase T1 and M1 genotypes. *Cancer Epidemiol Biomarkers Prev* 10: 539-550.

[Zharlyganova, D; Harada, H; Harada, Y; Shinkarev, S; Zhumadilov, Z; Zhunusova, A; Tchaizhunosova, NJ; Apsalikov, KN; Kemaikin, V; Zhumadilov, K; Kawano, N; Kimura, A; Hoshi, M.](#) (2008). High frequency of AML1/RUNX1 point mutations in radiation-associated myelodysplastic syndrome around Semipalatinsk nuclear test site. *J Radiat Res (Tokyo)* 49: 549-555.