



# Computational Modeling and Simulation of Developmental Toxicity

*What can we learn from a virtual embryo?*

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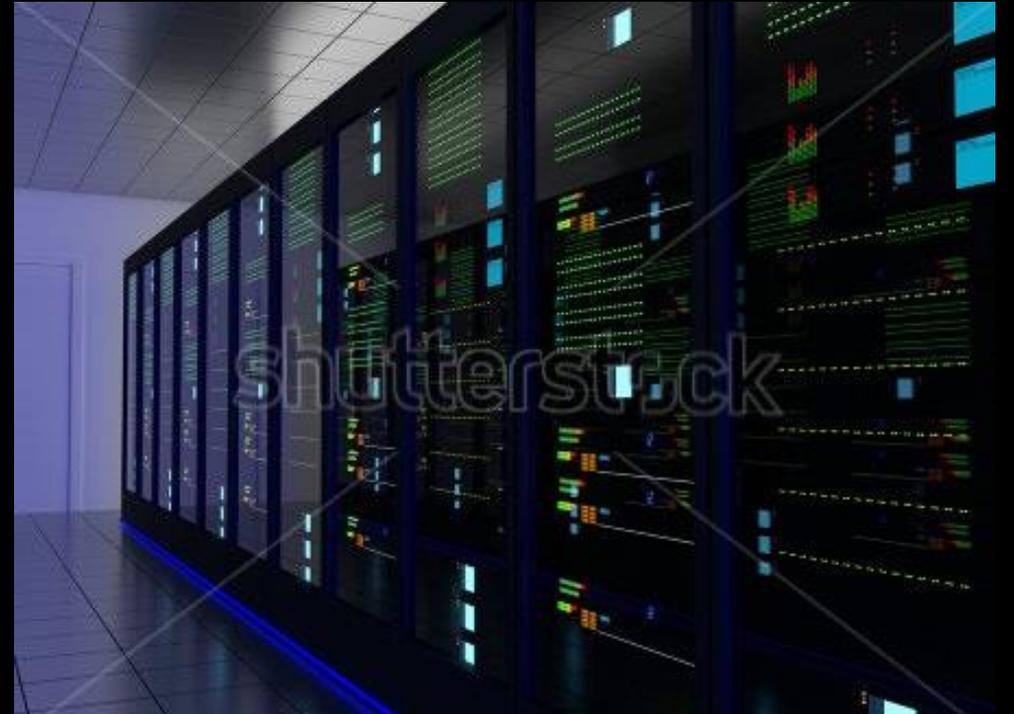
Chemical Safety for Sustainability Research Program



Virtual Tissue Models (VTM) project

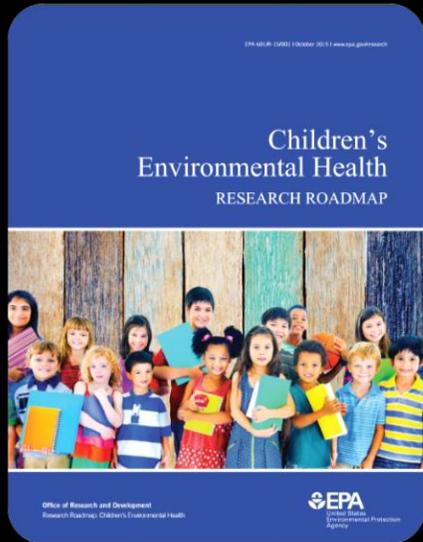
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US FDA, December 2, 2016



**DISCLAIMER:** *The views expressed in this presentation are those of the presenter and do not necessarily reflect the views or policies of the US EPA*

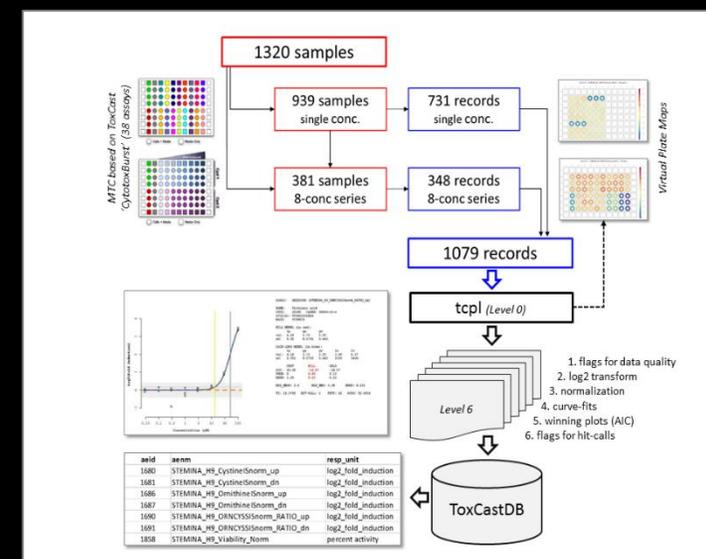
# Developmental health and disease



- ▶ Unique vulnerability of fetuses, infants, and children has a critical role in setting health and environmental policy (Executive Order 13045).
- ▶ Frank R. Lautenberg Chemical Safety for the 21st Century Act of 2016 explicitly requires protection of children and pregnant women.
- ▶ EPA's Office of Children's Health Protection (OCHP) ensures that all EPA actions and programs address the unique vulnerabilities of children.
- ▶ Children's Environmental Health (CEH) is a cross-cutting research goal implemented in EPA/ORD's CEH research roadmap for 2016-2019.

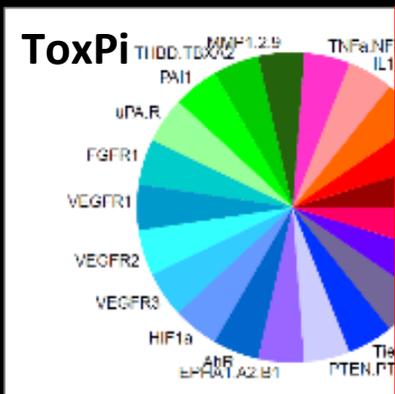
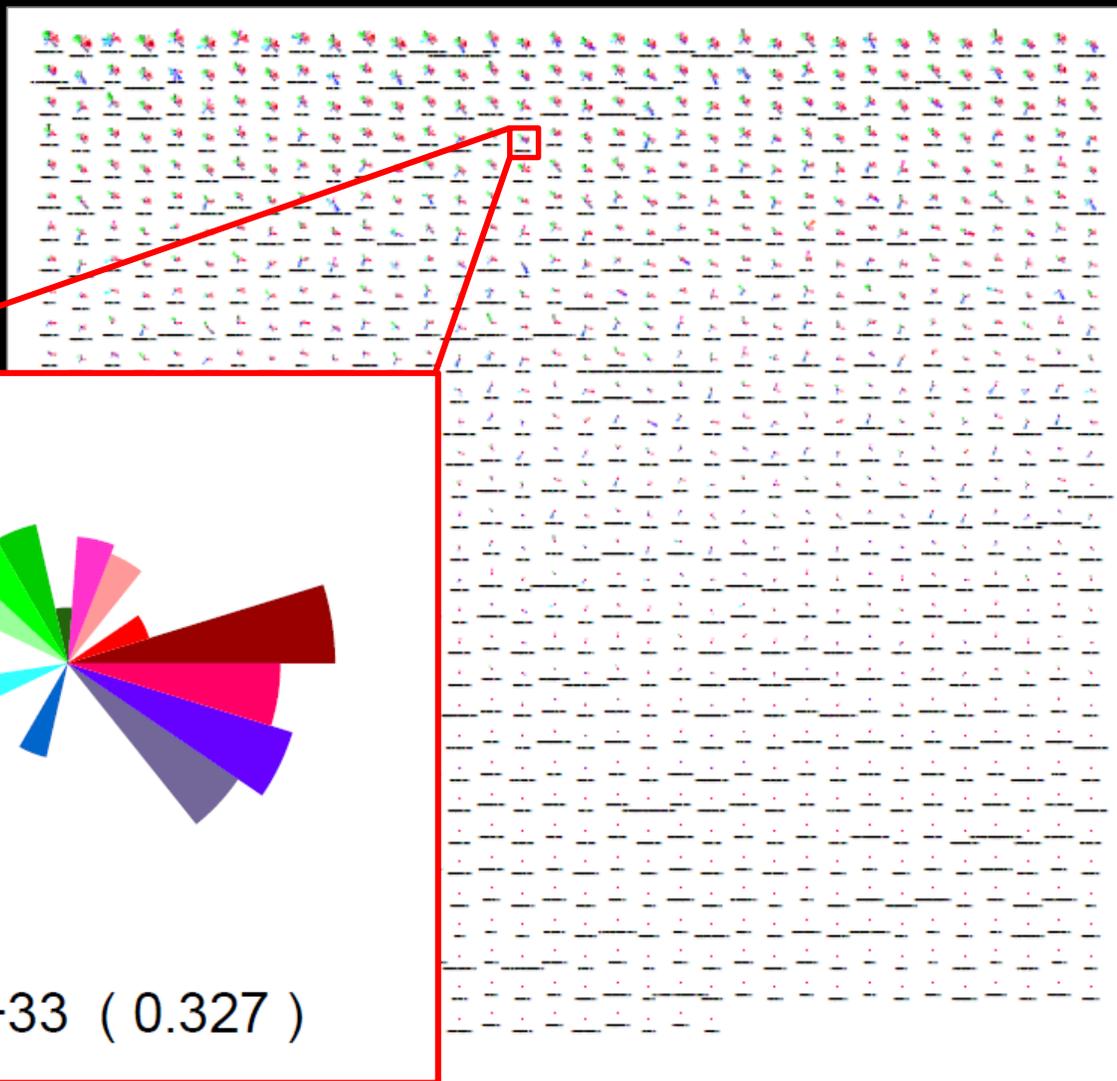
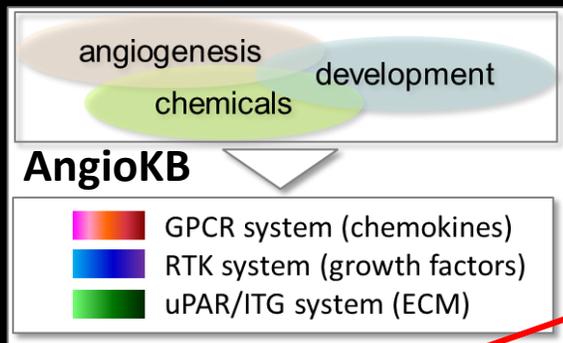
# Predicting developmental toxicity

- ▶ Automated high-throughput assays are providing vast *in vitro* data streams for profiling the bioactivity of large chemical inventories in ToxCast/Tox21.
- ▶ One use of the high-throughput screening (HTS) data is to prioritize chemicals, based on their cellular and molecular bioactivity profiles, for potential developmental toxicity.
- ▶ 1066 ToxCast compounds were tested in a human stem cell-based assay (Stemina devTOX<sup>qP</sup>).
- ▶ STM platform identified 190 positives with a 90% BA model (0.80 sensitivity, 1.00 specificity; n=33 reference compounds).



Source: Knudsen et al. (in preparation)

# Angiogenesis: chemicals sorted by predicted potential to disrupt angiogenesis (pVDCs)

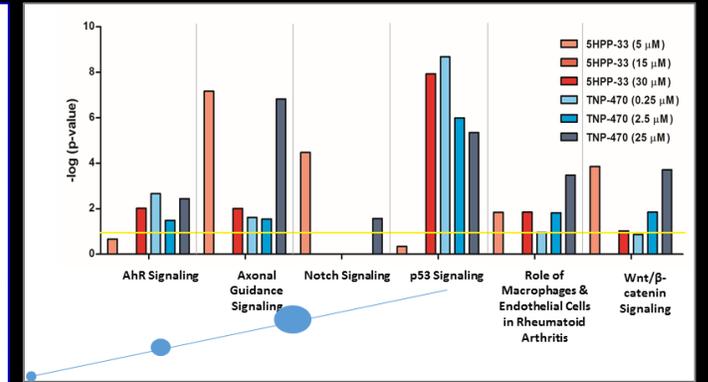
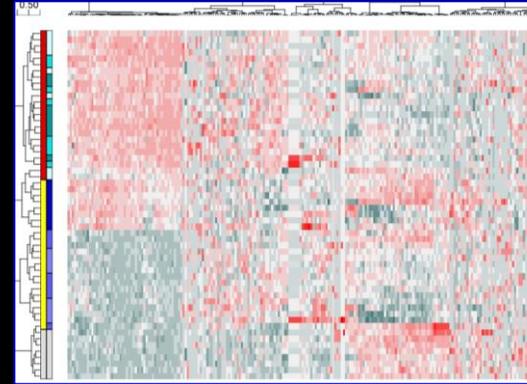
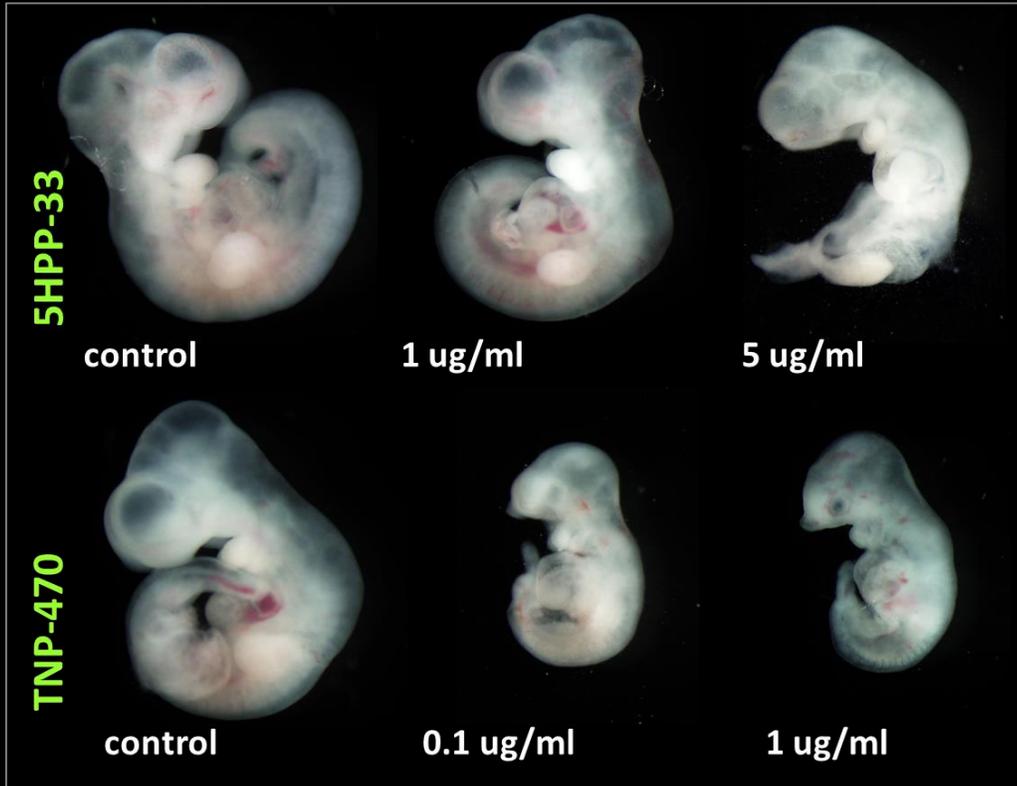


<http://epa.gov/ncc>  
<http://comptox.unc.edu>





# Rat WEC: GD10 embryos exposed for 48h



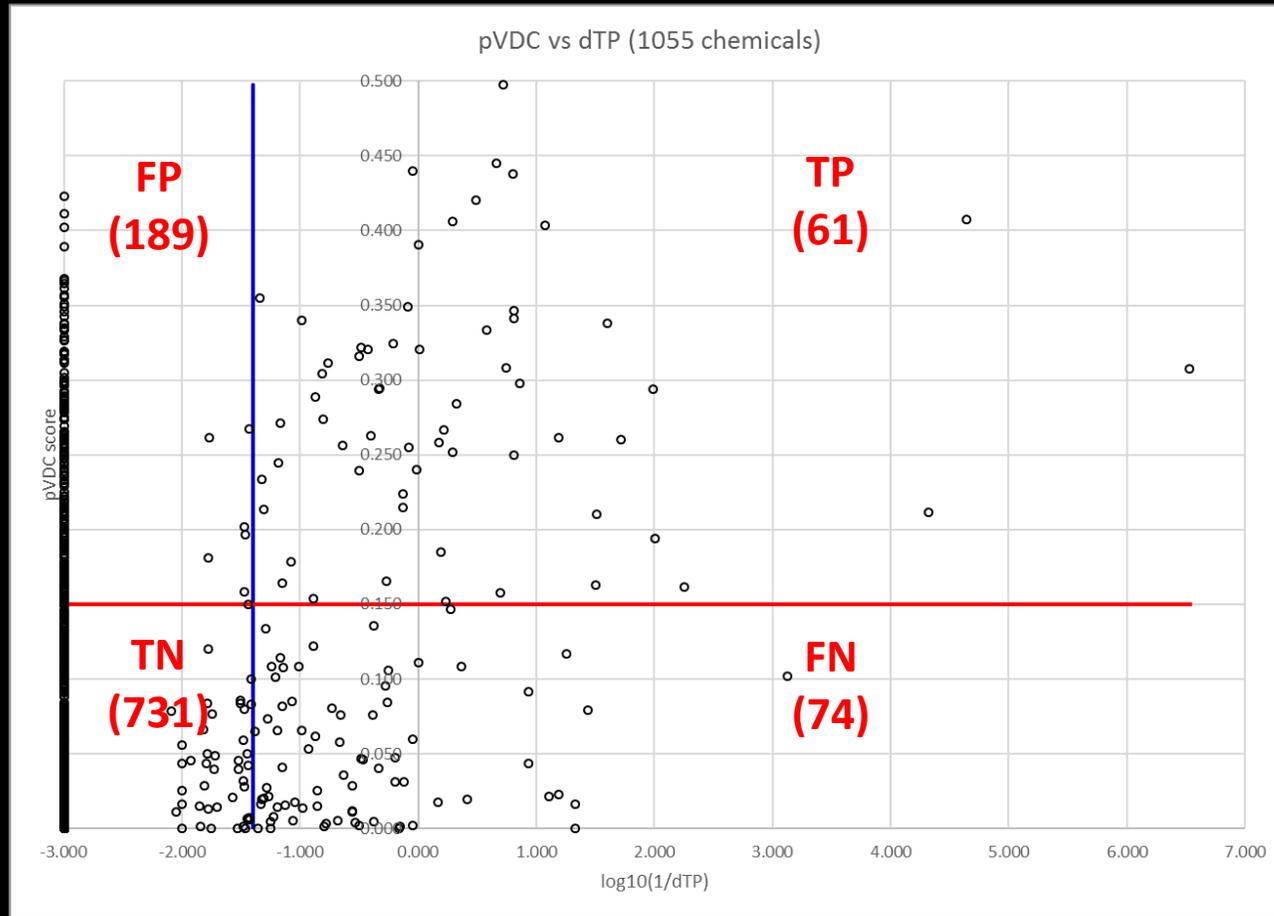
**RNA-seq analysis: p53** was most significantly altered pathway in both cases (5HPP-33, TNP-470); alterations in Notch and Wnt expression unique to 5HPP-33.

*SOURCE: Franzosa et al. (in preparation)*

**5HPP-33:** embryolethal  $\geq 15$   $\mu$ M, AC50 = 21.2  $\mu$ M; STM predicts human teratogenicity at  $\geq 4.37$   $\mu$ M

**TNP-470:** dysmorphogenic  $\geq 0.01$   $\mu$ M, AC50 = 0.038  $\mu$ M; STM predicts human teratogenicity at  $\geq 0.01$   $\mu$ M

# DevTox: how well does the pVDC score match-up with DevTox potential in a human system?



AOP-based **pVDC** score vs **DevTox** potential from the STM hES cell platform

Balanced Accuracy = 75.1%  
(modeled on the 38-test set)

24.4% pVDC(+) also STM(+)  
90.8% pVDC(-) also STM(-)

# Virtual reconstruction of developmental toxicity



- ▶ The question of how tissues and organs are shaped during development is crucial for understanding (and predicting) human birth defects.
- ▶ While ToxCast HTS data may predict developmental toxicity with reasonable accuracy, mechanistic models are still necessary to capture the relevant biology.
- ▶ Subtle microscopic changes induced chemically may amplify to an adverse outcome but coarse changes may override lesion propagation in any complex adaptive system.
- ▶ Modeling system dynamics in a developing tissue is a multiscale problem that challenges our ability to predict toxicity from *in vitro* profiling data (ToxCast/Tox21).

# Anatomical homeostasis in a self-regulating Virtual Embryo

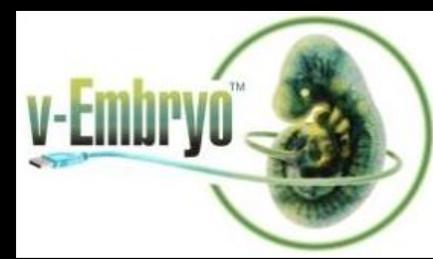


**Mouse Morula**  
*SOURCE: Science Photo Library*



*SOURCE: Andersen, Newman and Otter  
(2006) Am. Assoc. Artif. Intel.*

# Breathing life into a 'Virtual Embryo'



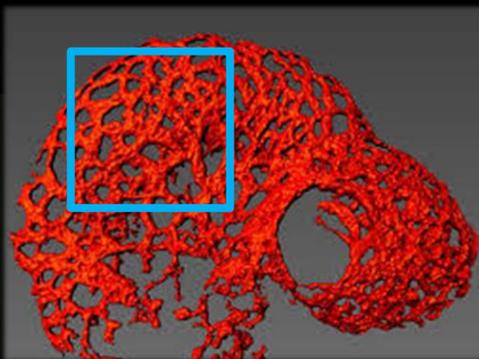
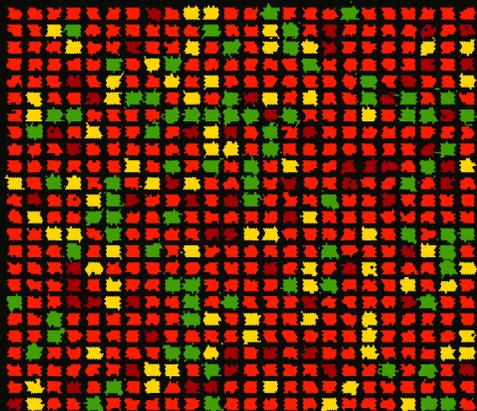
- ▶ **Hypothesis:** computer models that recapitulate a morphogenetic series of events can be used analytically (to understand) and theoretically (to predict) developmental toxicity.
- ▶ **Agent-Based Modeling and Simulation (ABMS):** a heuristic approach to reconstruct tissue dynamics from the bottom-up, cell-by-cell and interaction-by-interaction.
- ▶ **CompuCell3D:** open source modeling environment
  - engineered at Indiana University by James Glazier and colleagues;
  - steppables for distinct cell behaviors (growth, proliferation, apoptosis, differentiation, polarization, motility, ECM, signal secretion, ...);
  - rules coded in Python for cell-autonomous 'agents' that interact in shared microenvironment and self-organize into emergent phenotypes.

# Angiogenesis

- Endothelial Stalk
- Endothelial Tip
- Mural Cell
- Inflammatory Cell

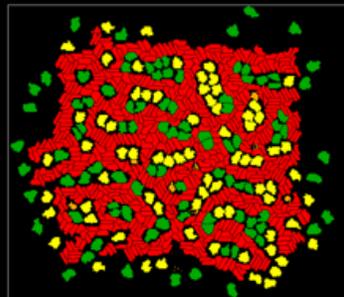
ABMS

VEGF165  
MMPs  
VEGF121  
sFlit1  
TIE2  
CXCL10  
CCL2

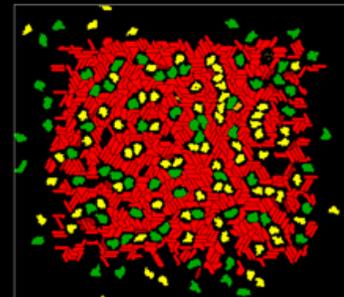


Exposure to 5HPP-33, a synthetic thalidomide analog

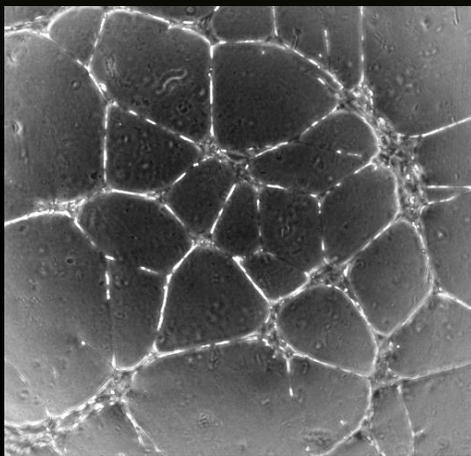
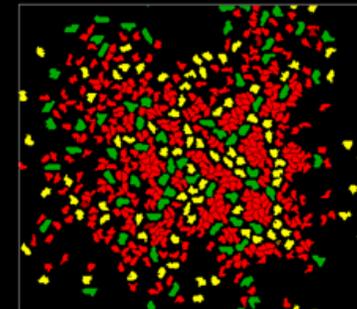
control



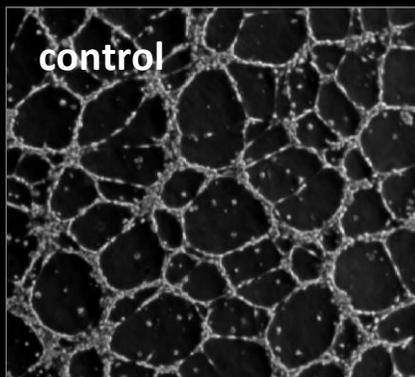
4  $\mu$ M



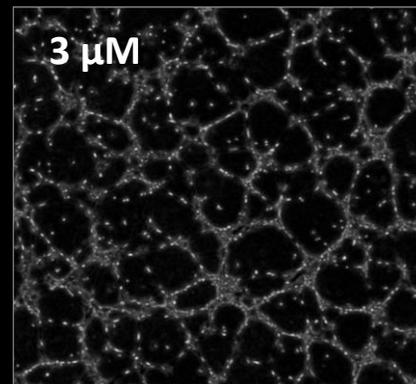
40  $\mu$ M



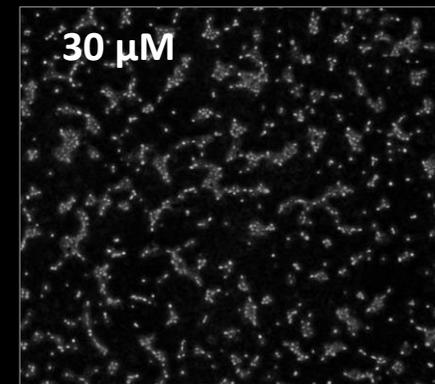
control



3  $\mu$ M



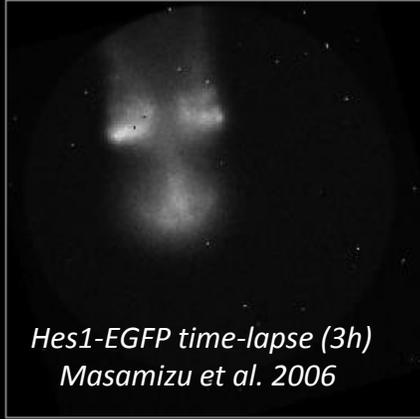
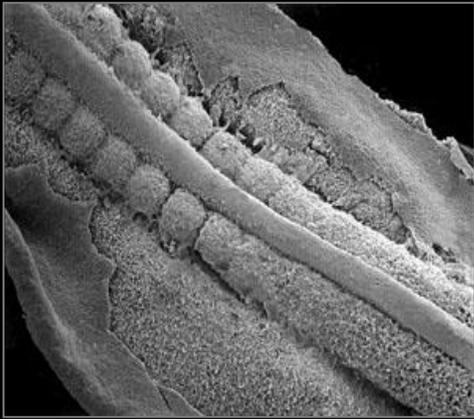
30  $\mu$ M



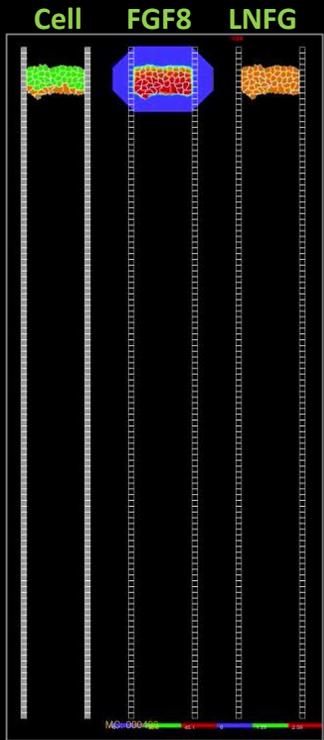
SOURCE: J Glazier, Indiana University

SOURCE: Kleinstreuer et al. 2013, PLoS Comp Biol

# Somite formation

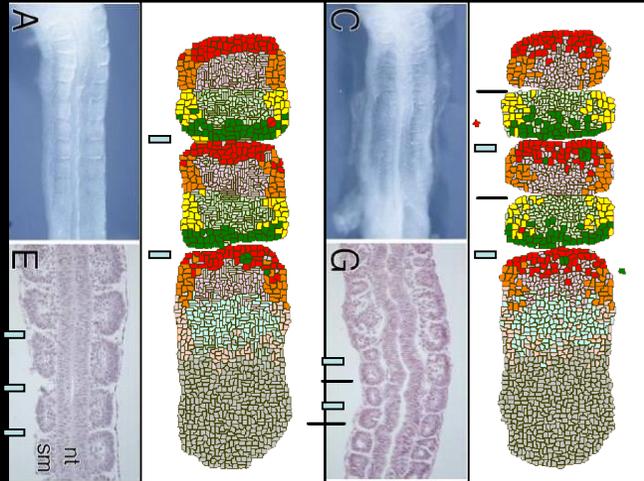


*Hes1-EGFP time-lapse (3h)*  
Masamizu et al. 2006

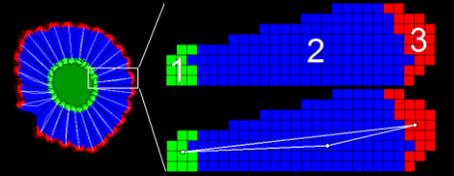
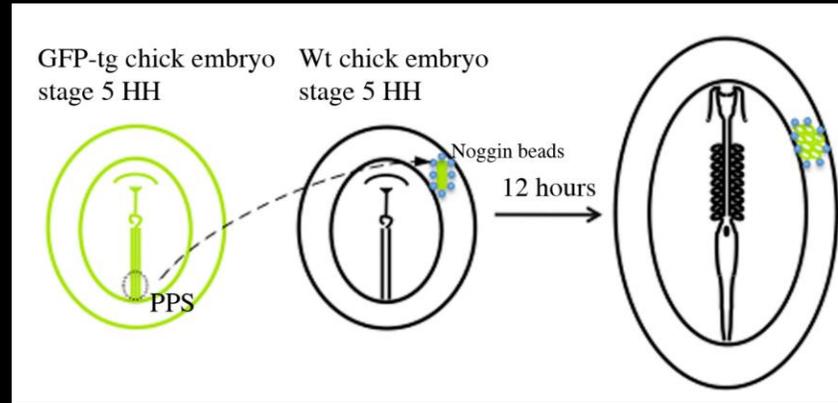


**Clock and Wavefront model:**

- signal gradients along AP axis (eg, FGF8, RA)
- oscillating gene expression (eg, LNFG, Hes1)
- cell adhesion (eg, ND, ephrin system)

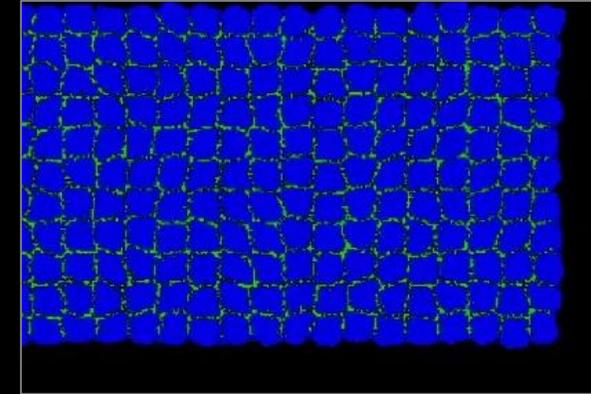


SOURCE: Hester et al. (2011) PLoS Comp Biol



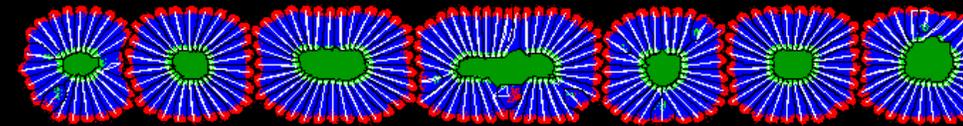
**Epithelialization model:**

- clock genes do not oscillate
- somites form simultaneously



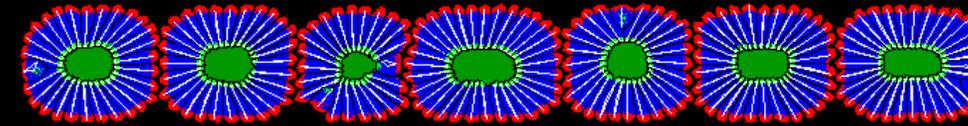
**Adding the Wavefront:**

- restores sequentiality



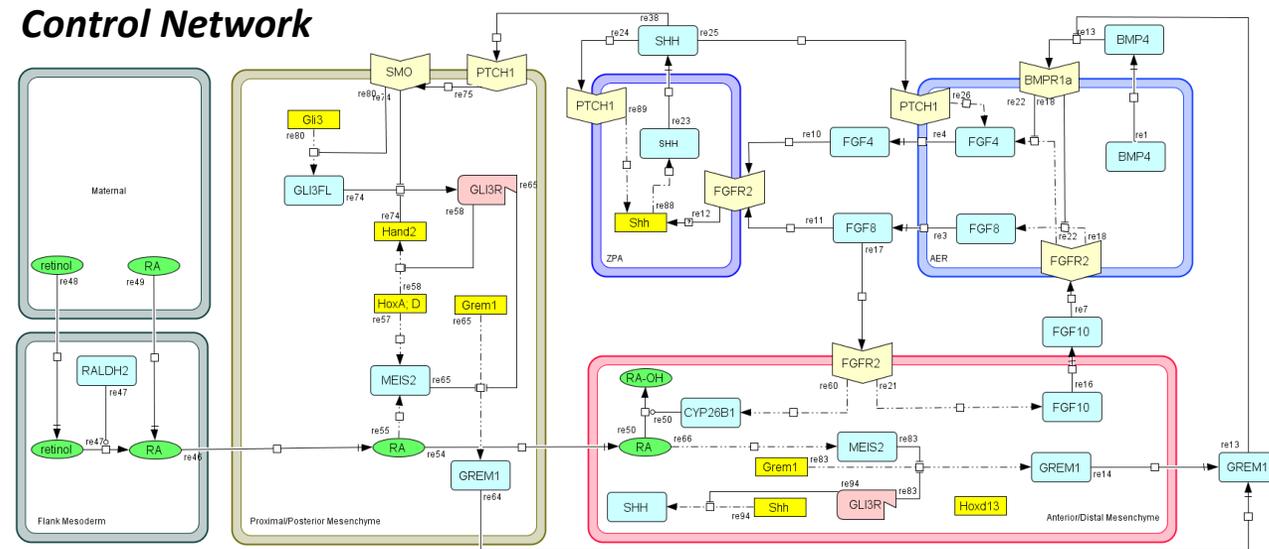
**And adding the Clock:**

- improves regularity

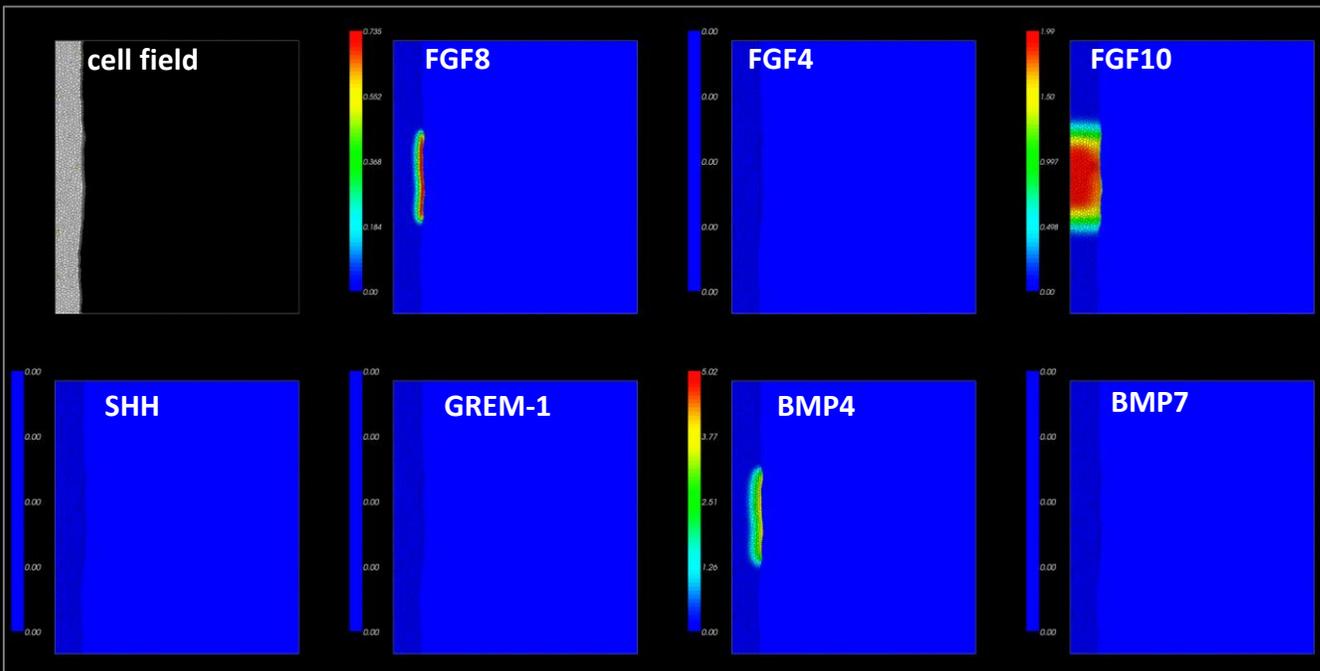
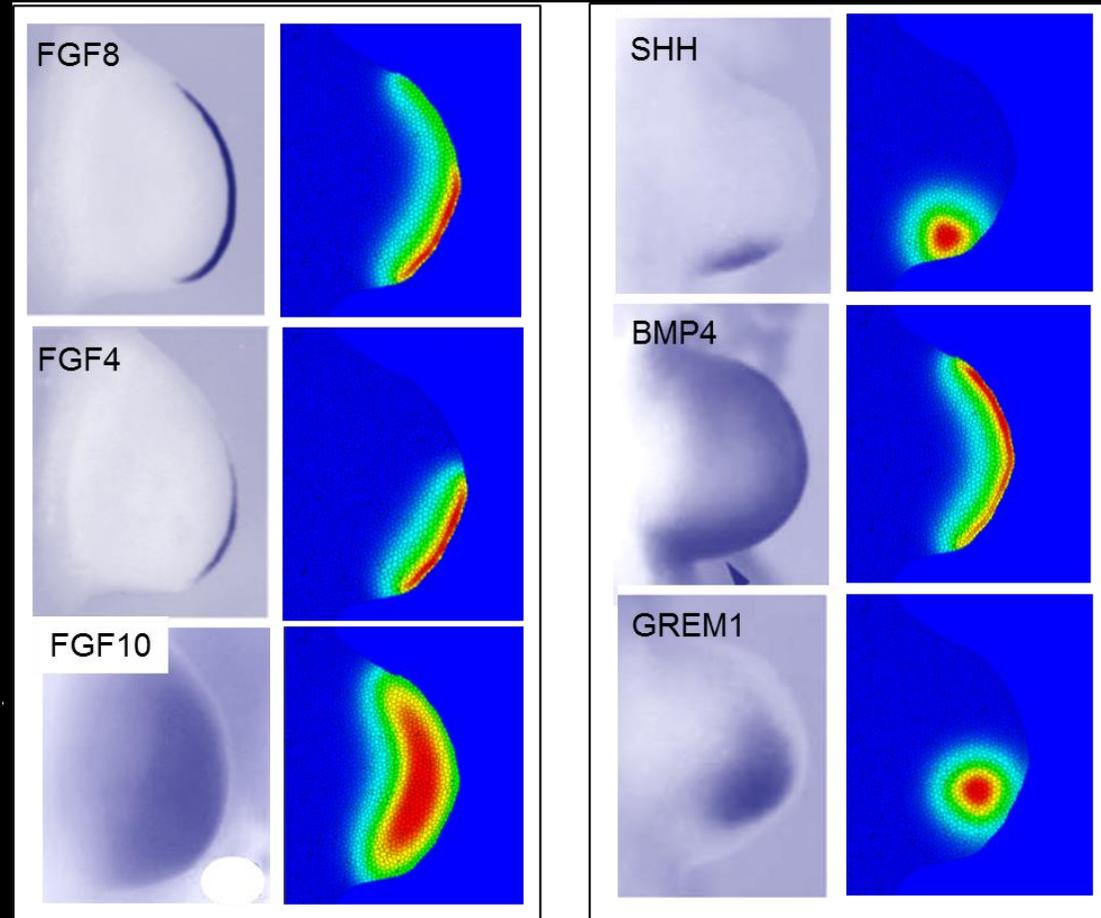


SOURCE: Dias et al. (2014) Science

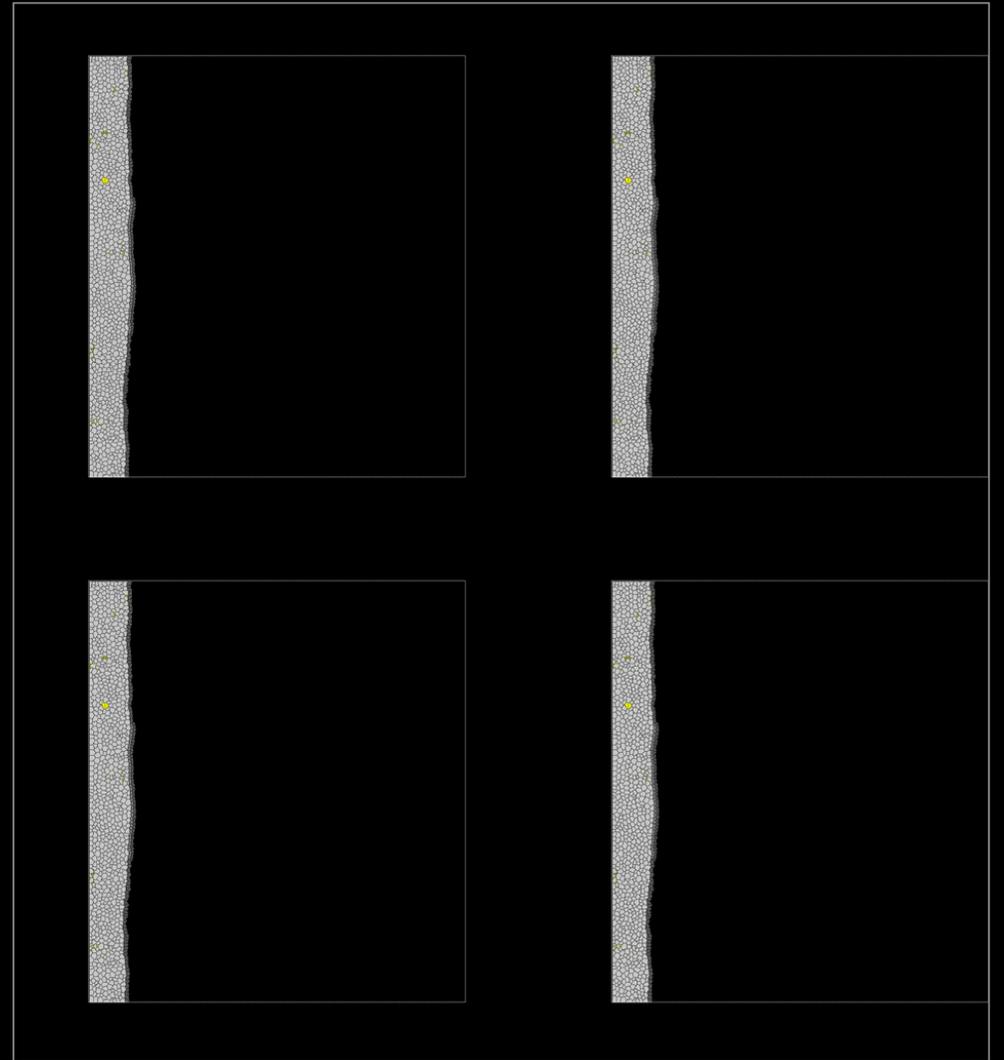
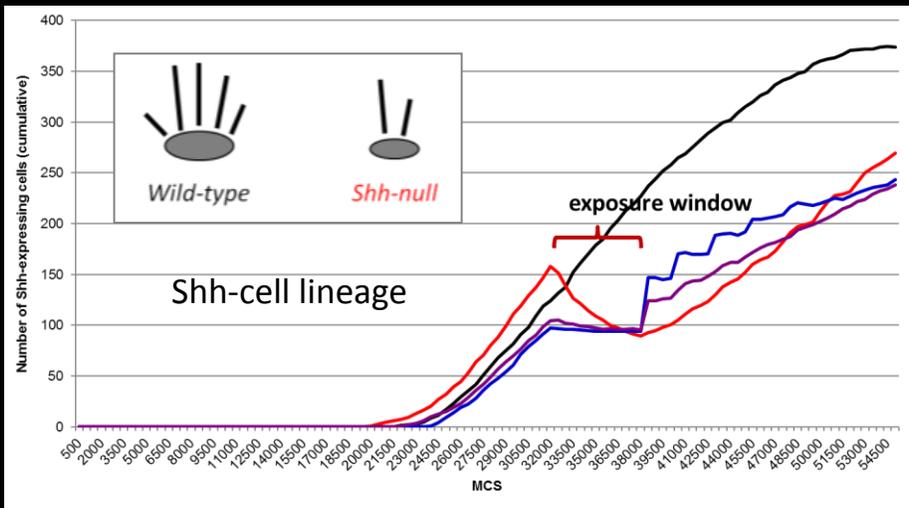
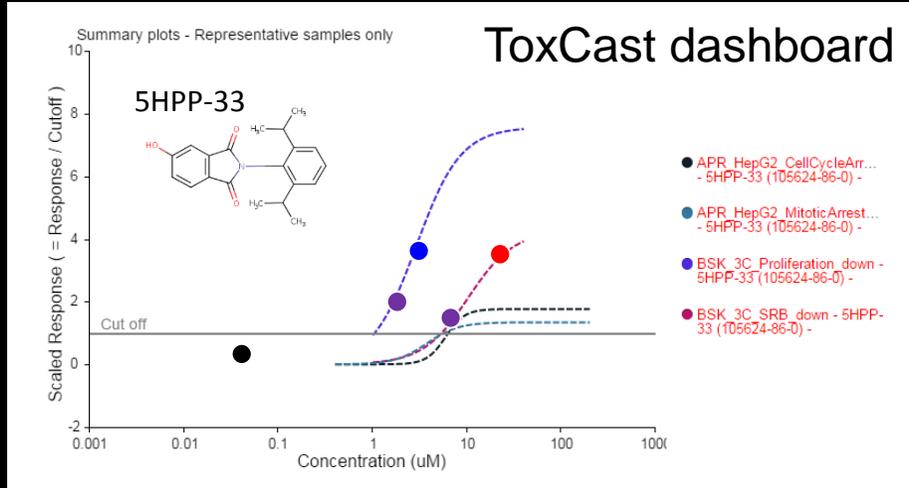
# Control Network



# Limb-bud outgrowth

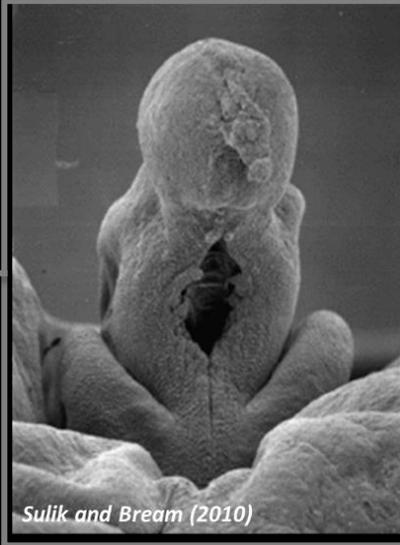


# Teratogenesis *in silico*

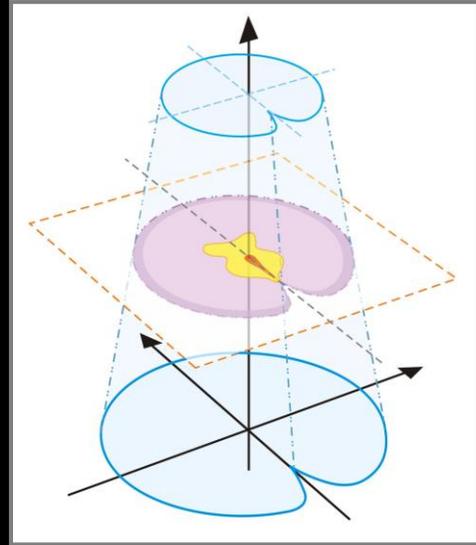


# Genital Tubercle (GT) differentiation

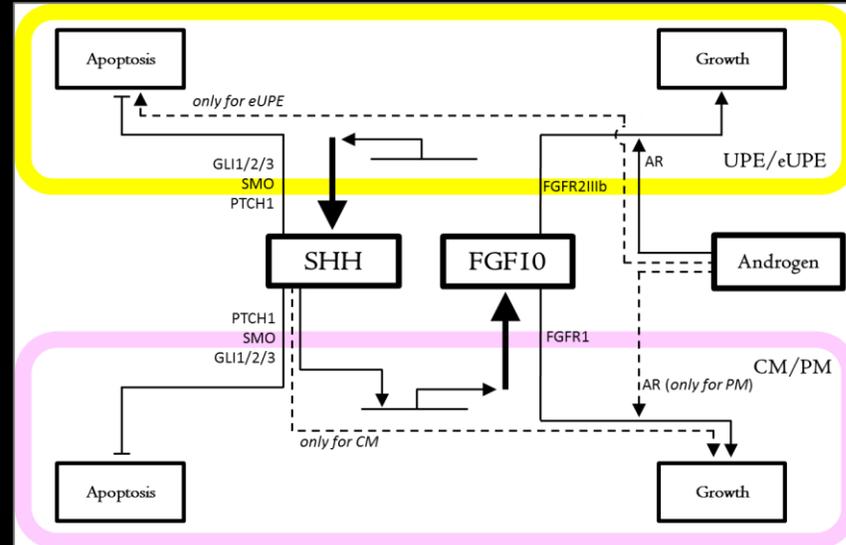
Embryonic GT



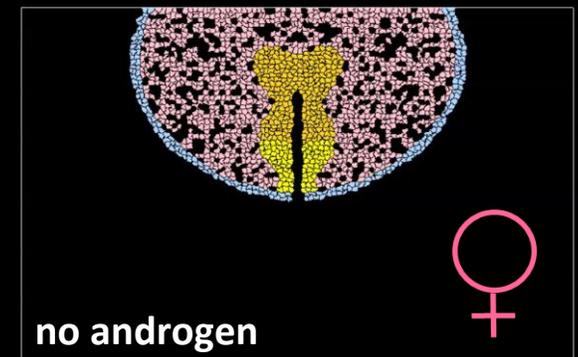
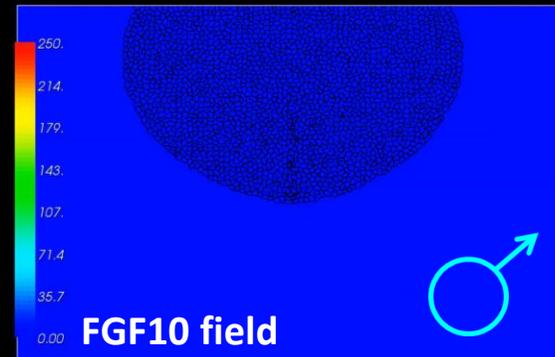
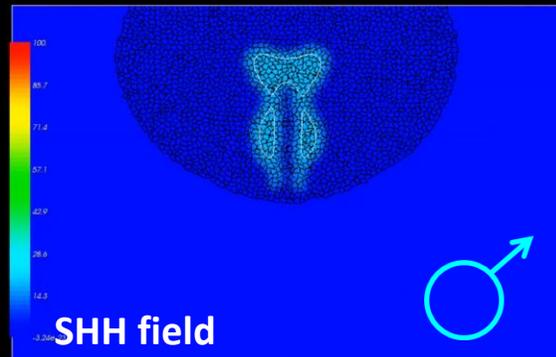
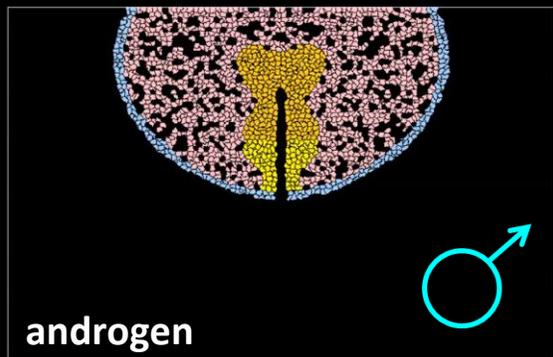
Abstracted GT



Control Network (mouse)

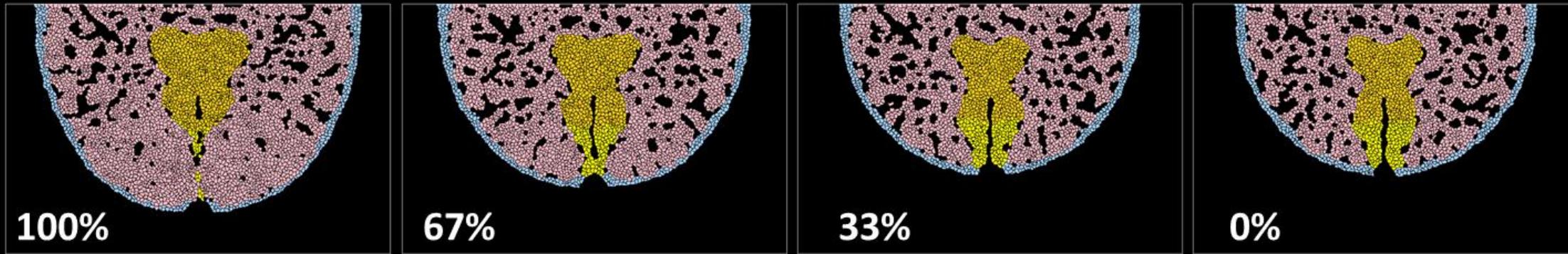


ABM simulation for sexual dimorphism (mouse GD13.5 – 17.5)

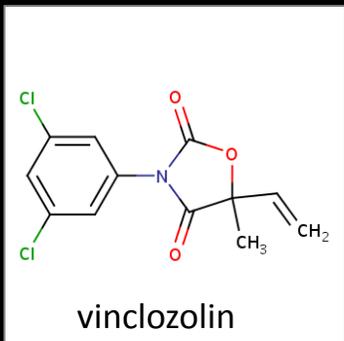


# Urethral Closure: complex process disrupted in 'hypospadias'

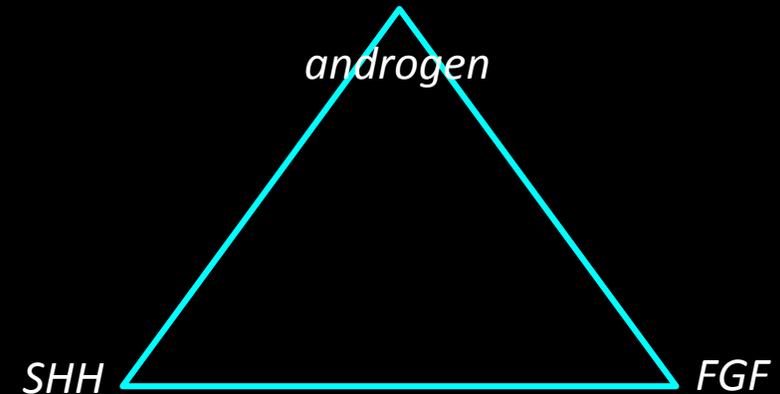
Driven by urethral endoderm (contact, fusion apoptosis) and androgen-dependent effects on preputial mesenchyme (proliferation, condensation, migration) via FGFR2-IIIb.



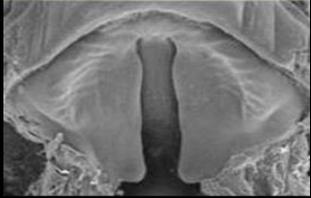
Leung et al. (2016) *Reproductive Toxicology*



Androgenization (n = 10 sims)		<u>Closure Index</u>
100%		0.80
67%		0.57
33%		0.13
0%		0.07

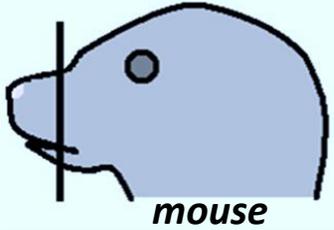


# Cleft Palate

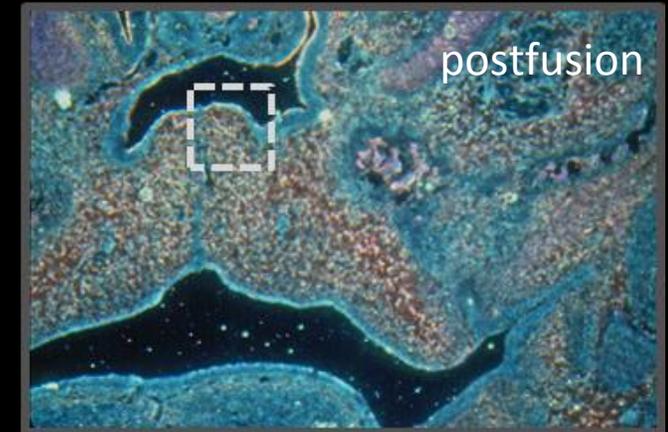
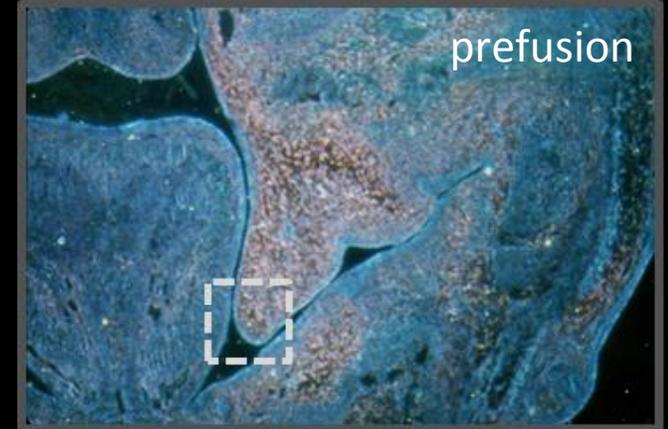
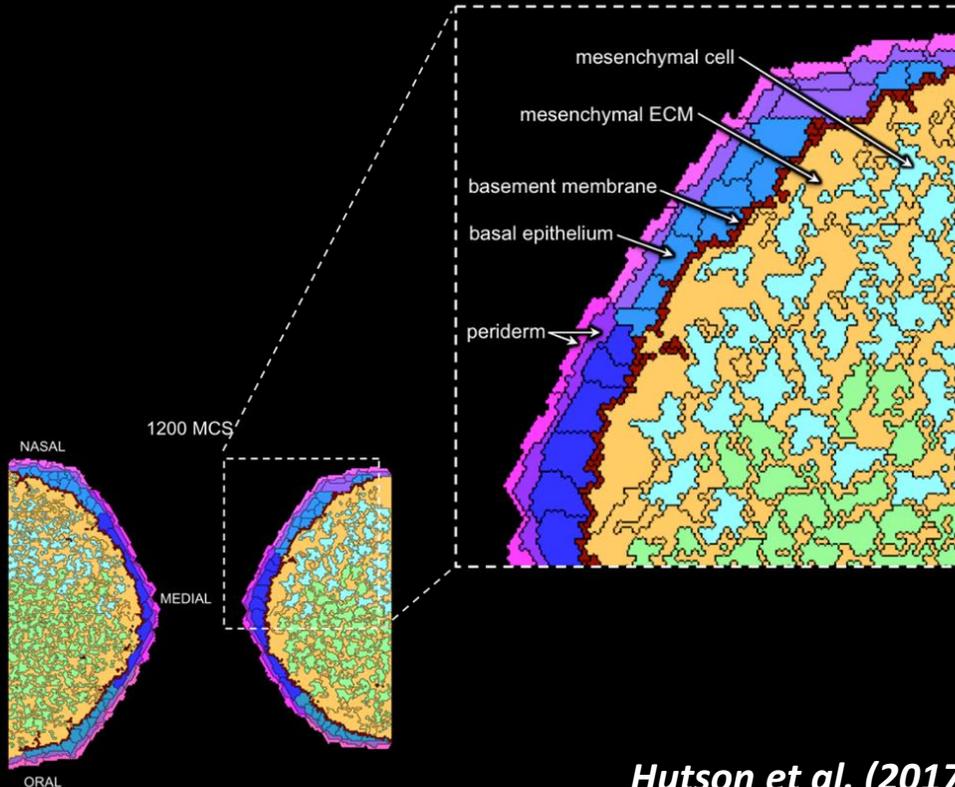


- ▶ Most prevalent craniofacial birth defect (annually ~7000 newborns in the USA and ~200,000 worldwide).
- ▶ Etiology is multifactorial: interaction of genetic, environmental, and lifestyle factors.
- ▶ Vulnerable period encompasses outgrowth of right-left palatal process in the oral cavity of the 1<sup>st</sup> trimester embryo.
- ▶ Local disruption of epithelial-mesenchymal interaction impairs outgrowth and fusion of the palatal processes.

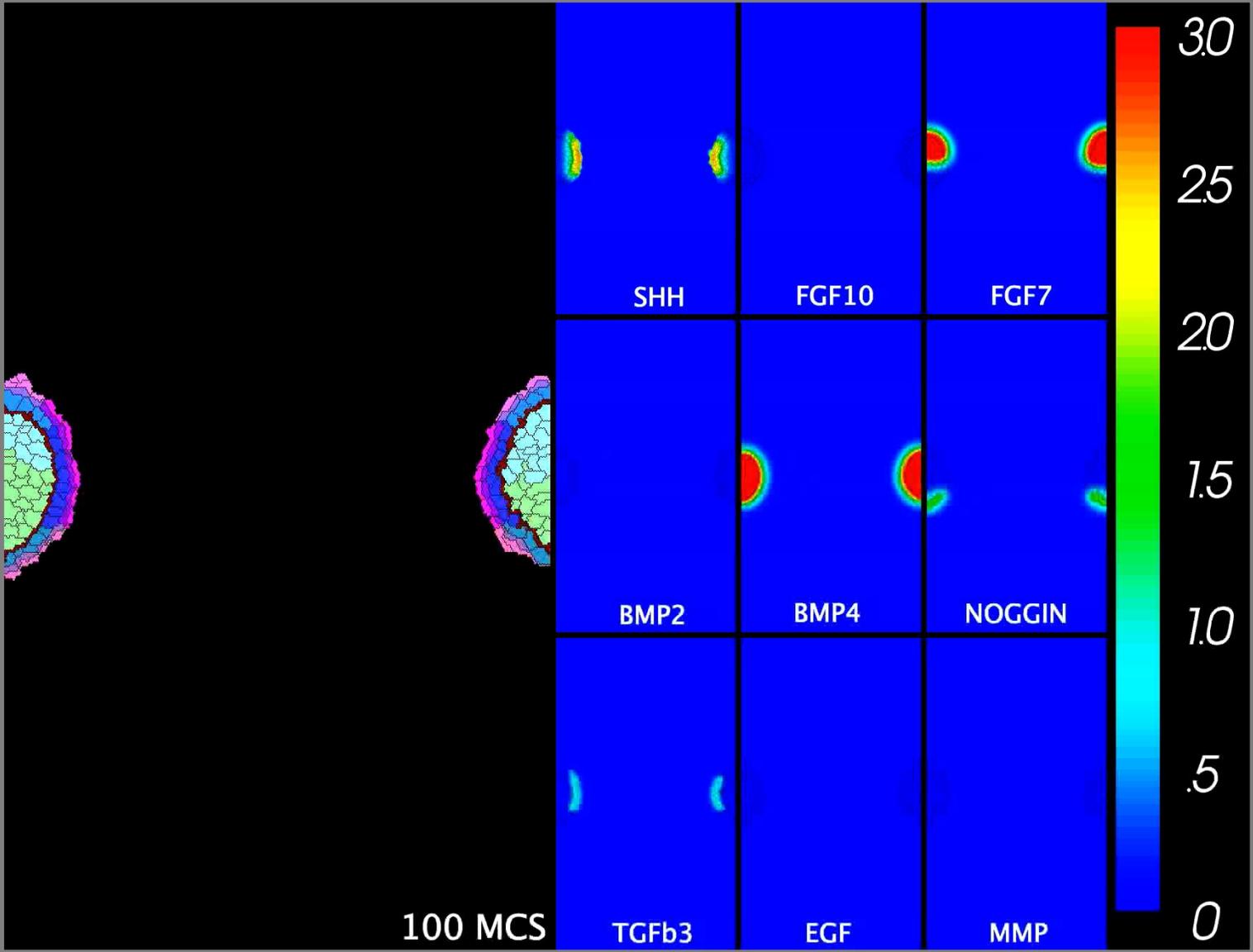
# Modeling Palatal Development



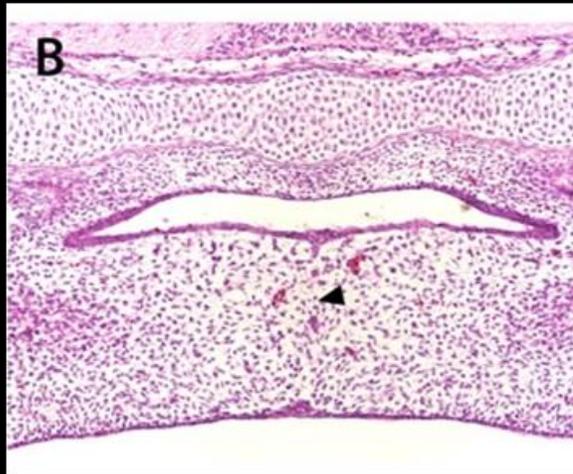
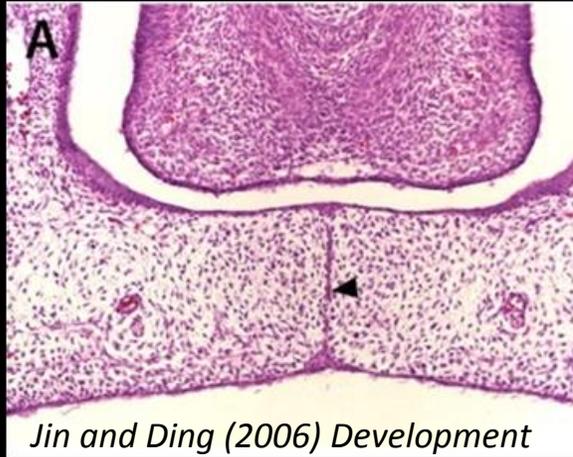
- E12.5 initial outgrowth of palatal shelves
- E13.5 expansion alongside the tongue
- E14.5 elevate, meet, and adhere at medial edge
- E15.5 fusion complete, mesenchymal confluence
- E16.5 osteogenic differentiation



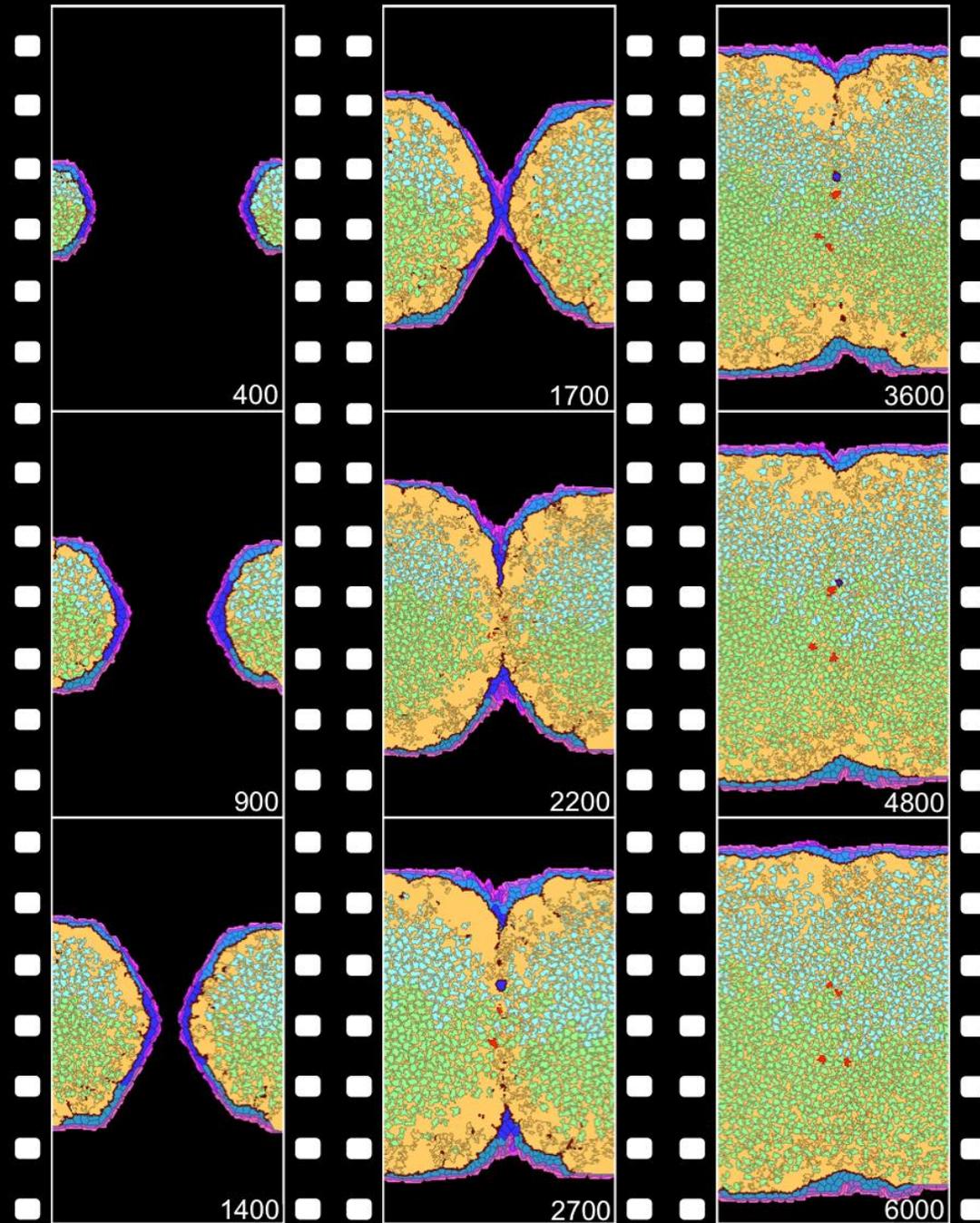
# Spatially-dynamic ABMS for palate development



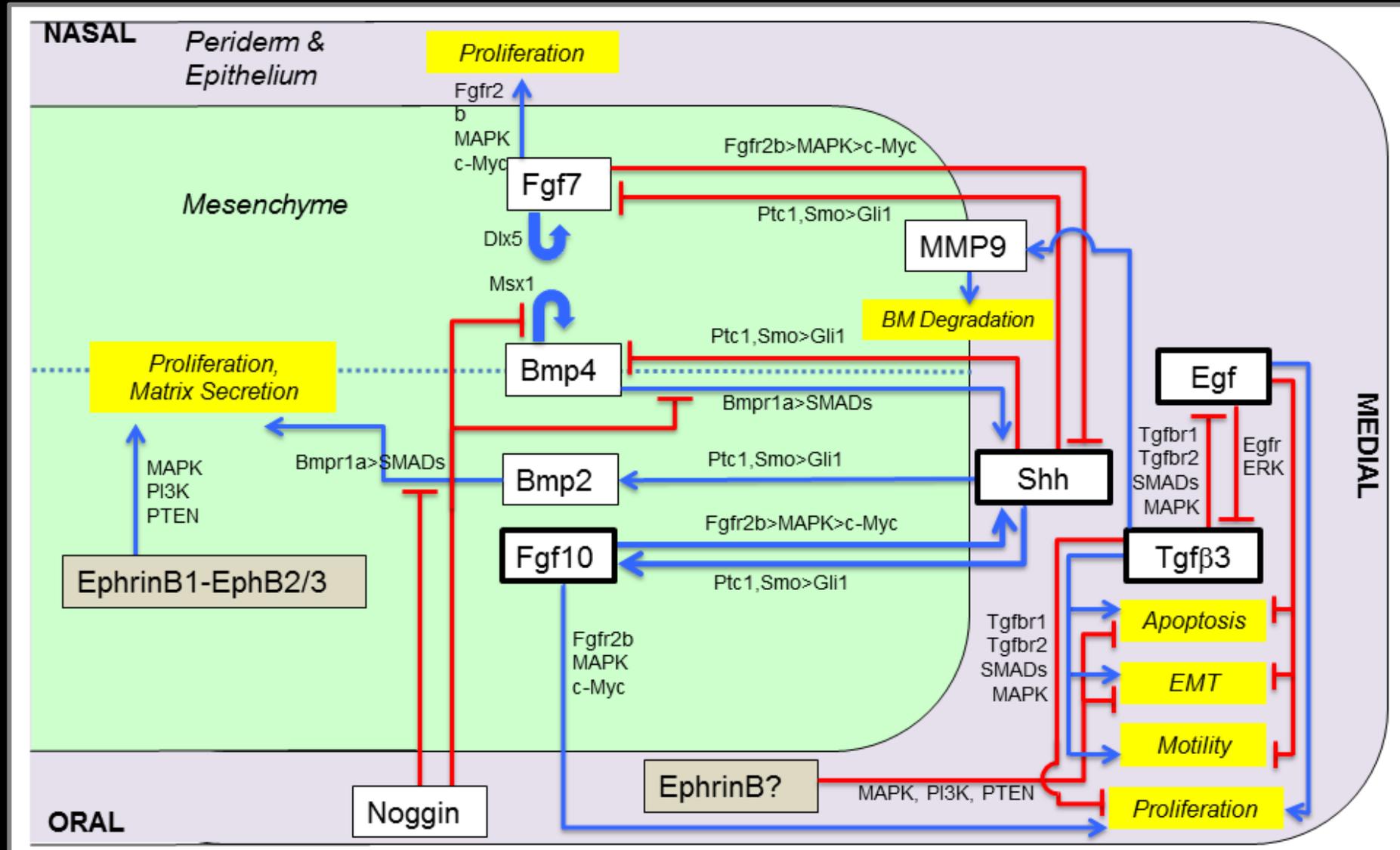
# Morphogenetic fusion



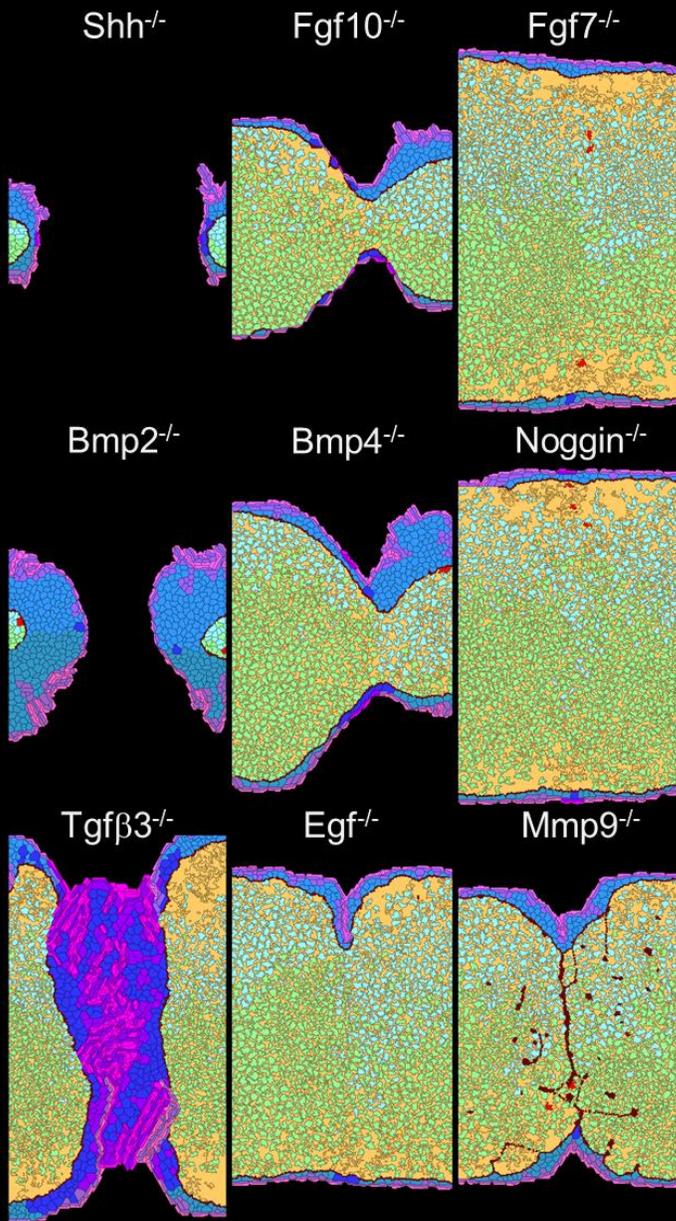
*MES breakdown is programmed genetically to coincide with MEE apposition*



# Control network



# Hacking the Control Network: *in silico* knockouts → ‘Cybermorphs’



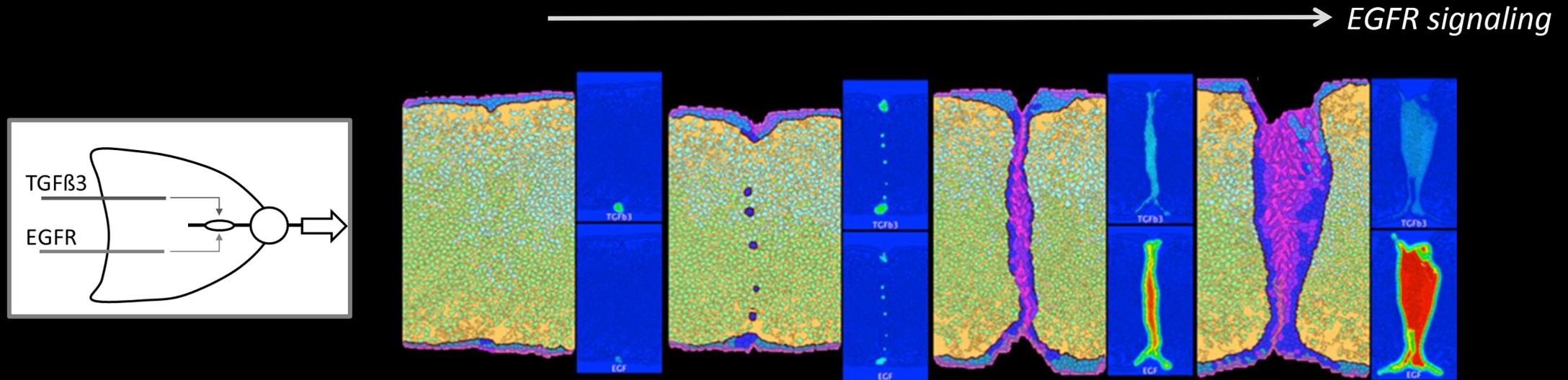
## Signals driving outgrowth to apposition and MEE contact (MCS 200-2000)

- SHH from the MEE drives mesenchymal proliferation and ECM production via FGFs/BMPs.
- Positive and negative feedback loops modulate epithelial-mesenchymal signaling cell-by-cell and interaction-by-interaction.

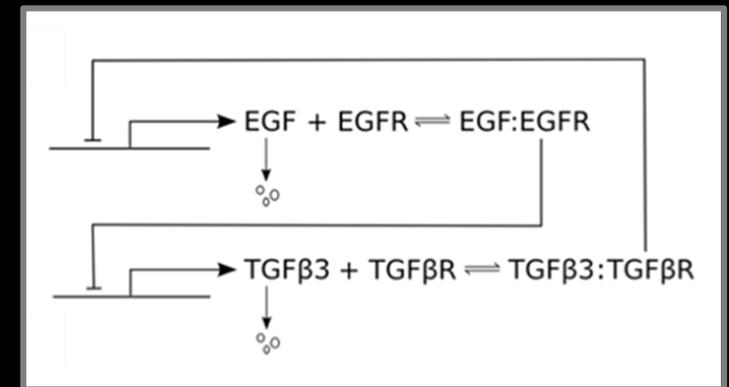
## Signals driving MES breakdown (MCS 2000-3000)

- TGF $\beta$ 3 triggers MEE cells to undergo apoptosis (PCD), epithelial-mesenchymal transition (EMT), and migration (retraction).
- EGF has the opposite effect, maintaining MEE cell growth, proliferation, and survival.

# Messin' with the switch: system fragility and fault tolerance

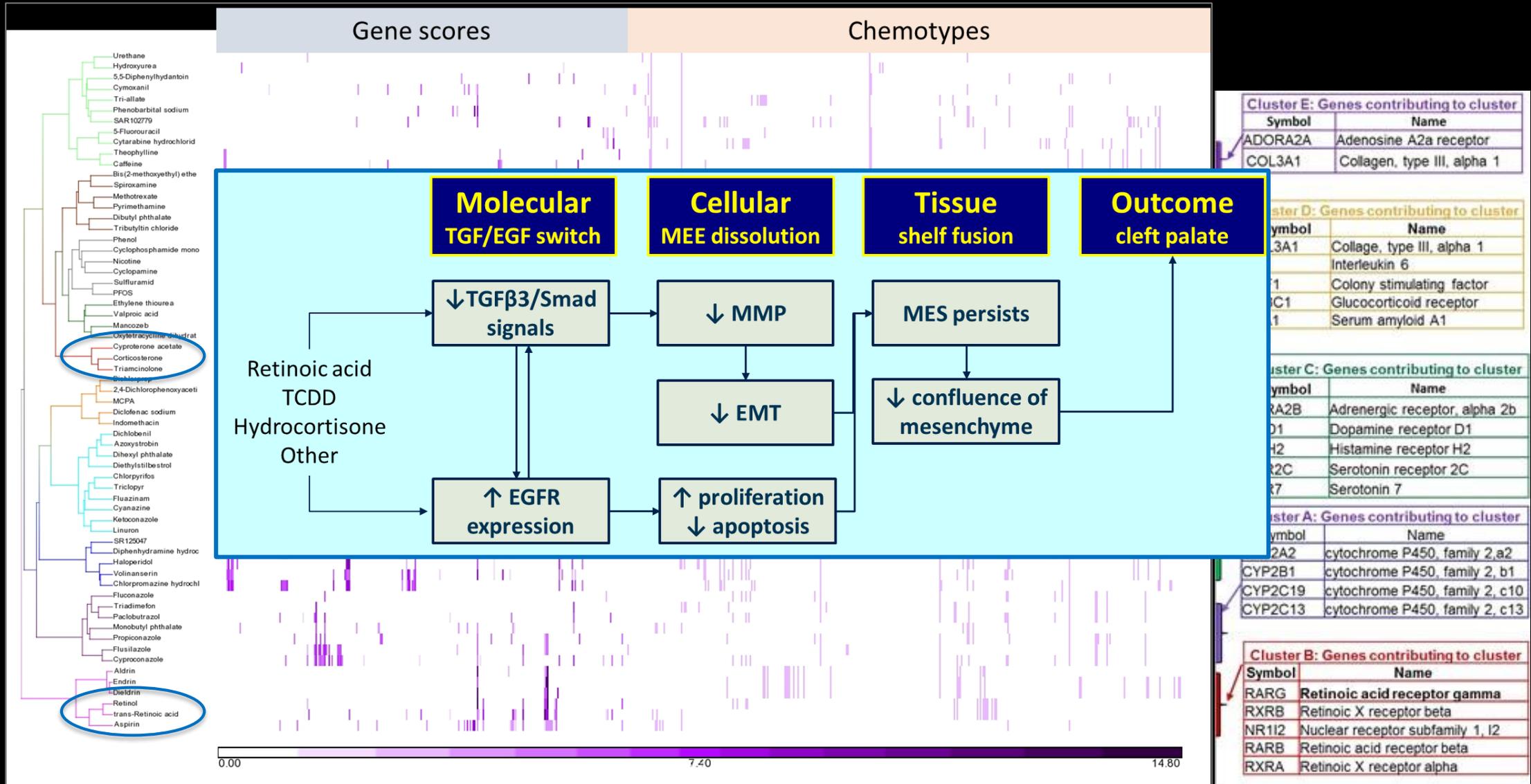


- A mutual inhibitory gene regulatory circuit exhibits switch-like behavior in the MEE.
- $EGFR$  expression normally wanes several hours prior to MEE apposition to flip the switch to the  $TGF\beta 3$  state.
- Several cleft palate teratogens are known to maintain  $EGFR$  expression (Retinoic acid, Hydrocortisone, TCDD) [Abbott 2010]).



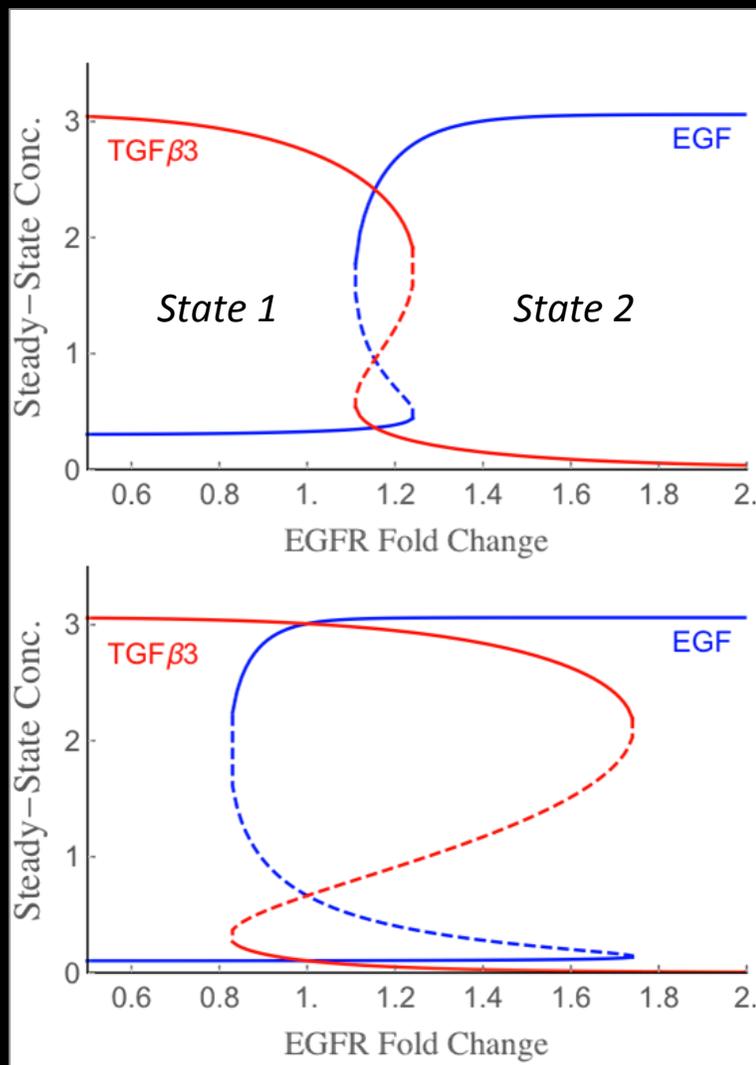
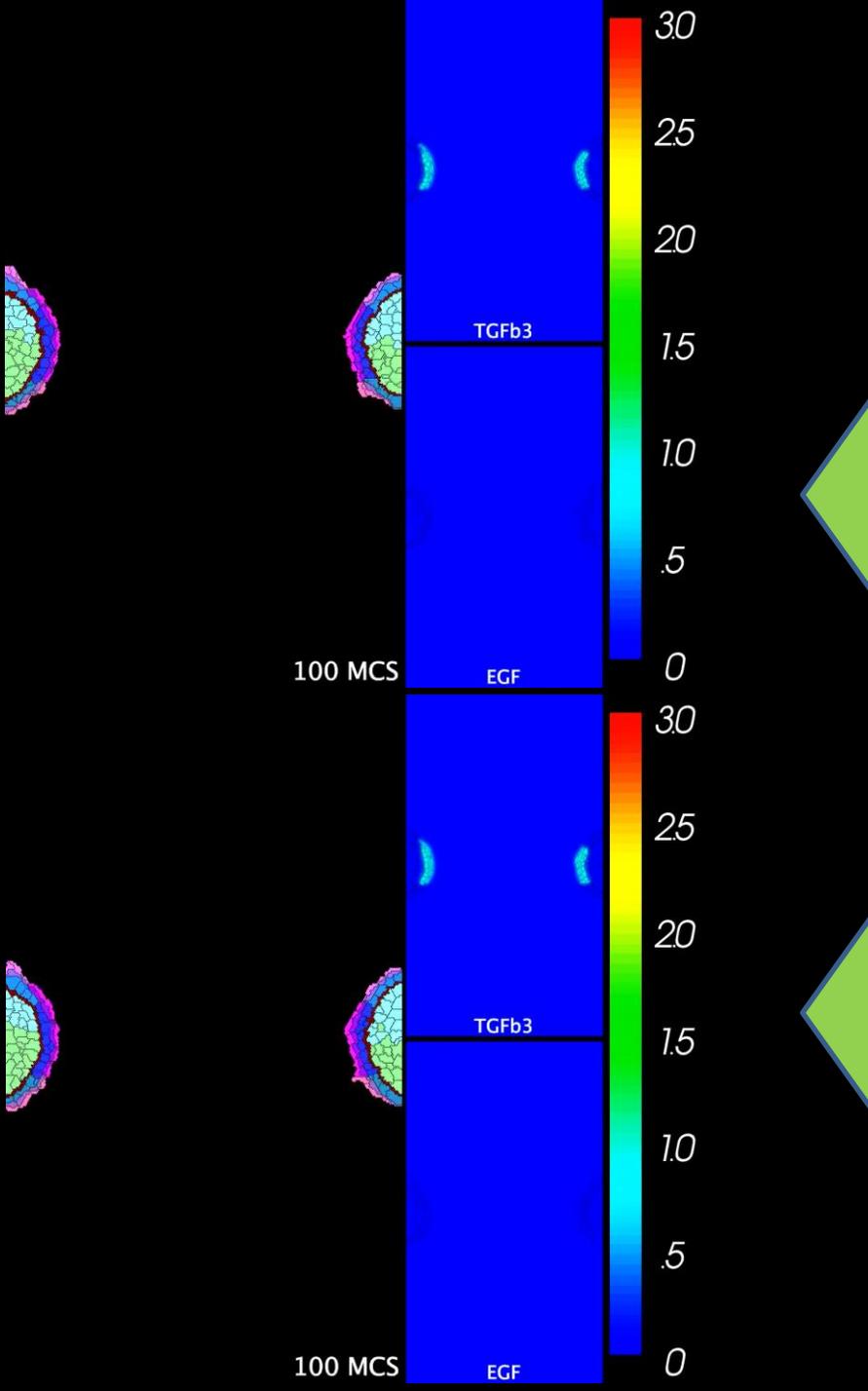
# 63 cleft palate (animal) teratogens in ToxCast

Baker et al. (in preparation)

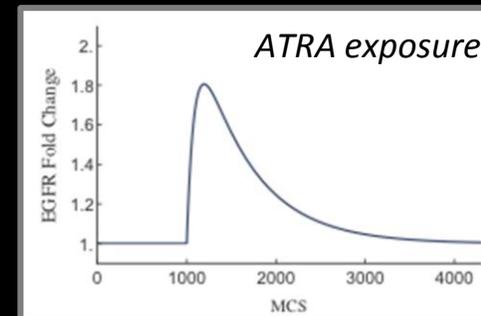


# Dissecting circuit dynamics *in silico*:

two scenarios for differential teratogenicity



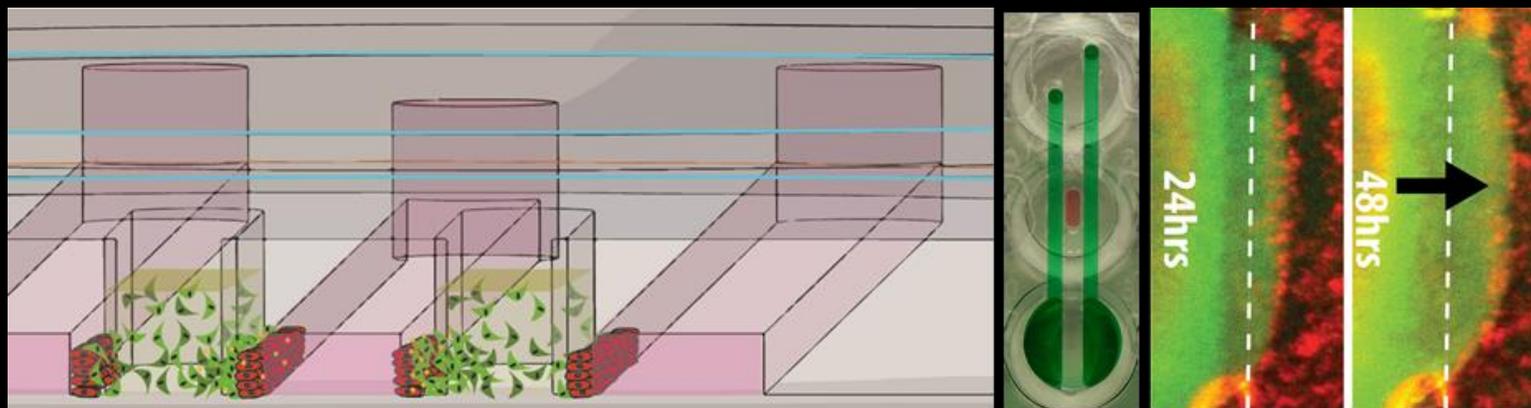
*tipping point >1.8x (n=24)  
(reversible)*



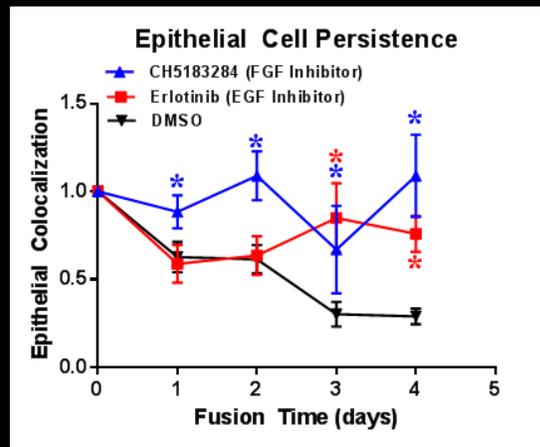
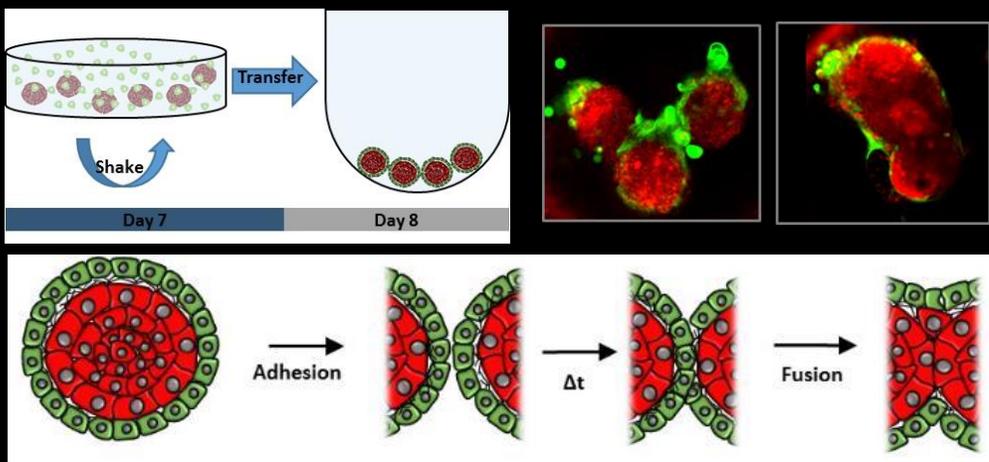
*tipping point ~1.5x (n=16)  
(not reversible)*

# Microscale systems: engineered 3D OCMs that use human cells for predictive toxicology

Beebe lab – U Wisconsin, HMAPS [Johnson et al.] – with permission



Abbott lab - NHEERL/TAD [Belair et al.] – with permission



## VTM WORKFLOW

bioactivity profiles



biological circuits



cellular dynamics



ABMS 'cybermorphs'

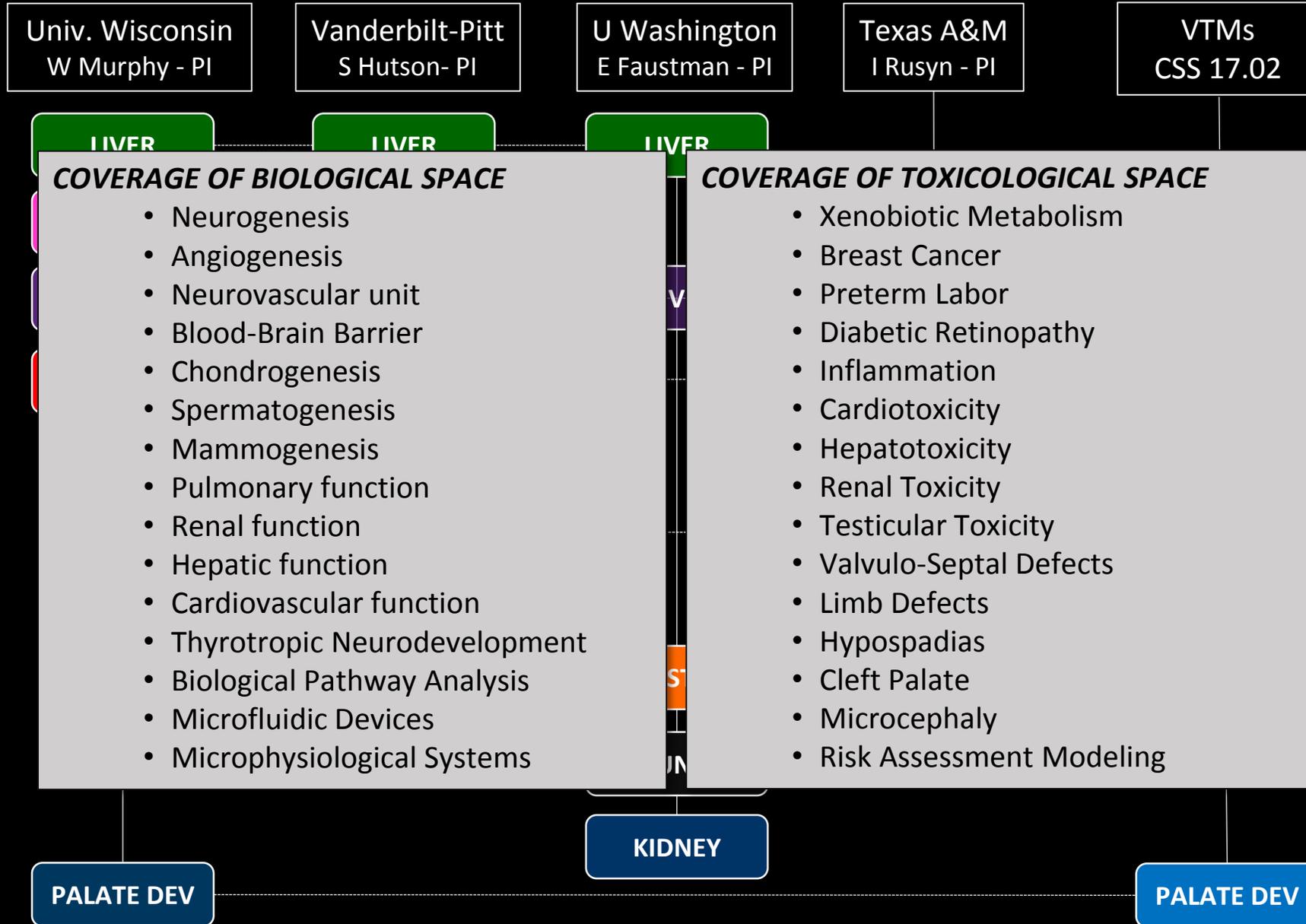


organotypic cultures



functional analysis

# Integration with OCM-PT STAR Centers



# Computational and Organotypic Modeling of Microcephaly

Knudsen TB<sup>1</sup>, Baker NC<sup>2</sup>, Faustman EM<sup>3</sup>, Murphy WL<sup>4</sup> and Daly W<sup>4</sup>.

<sup>1</sup>USEPA, National Center for Computational Toxicology; and <sup>2</sup>Lockheed-Martin, Research Triangle Park NC

<sup>3</sup>University of Washington, UW-PT Center, Seattle WA; and <sup>4</sup>University of Wisconsin, HMAPs Center, Madison WI

## Microcephaly and Maternal Zika Infection

- Cluster of birth defects in N Brazil linked to mosquito-borne Zika virus by the Brazilian Health Ministry (November) and validated by CDC (December) [1].



"On the basis of this review, we conclude that a causal relationship exists between prenatal Zika virus infection and microcephaly and other serious brain anomalies."

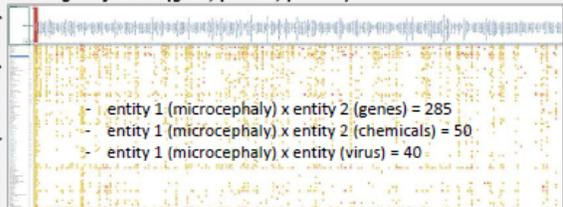
- Reduction in brain volume, ventricular dilations, brain calcifications, retinal defects, and placental insufficiency are all part of the congenital Zika story.
- A broader scientific need exists for Adverse Outcome Pathway (AOP) models of microcephaly because it has many possible causes [2]:
  - intrauterine infections (e.g., Rubella, CMV, ZIKV)
  - inborn errors of metabolism (e.g., urea cycle, mitochondriopathies)
  - maternal smoking, drug and alcohol abuse
  - environmental chemicals (e.g., methylmercury)
  - genetic factors (autosomal recessive traits; microdeletions, duplications)
  - prenatal malnutrition, socioeconomic factors, ...
- OBJECTIVE: capture information on 'microcephaly' into an AOP framework.

## Microcephaly Information Retrieval

- MGI Mammalian Phenotype Browser: 'microcephaly' (MP:0000433) returns 85 gene associations including candidate genes for microcephaly in humans.
- ToxRefDB returns ~75 chemicals invoking dilated ventricles/hydrocephaly (39), and/or reduced brain size/cellular mass (40).
- MicrocephalyConnections tool sweeps literature (PubMed) to produce a multidimensional database of MeSH co-annotations (350,651 records).

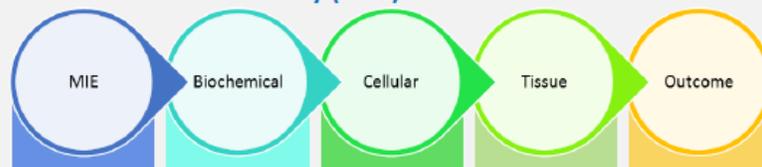
### Biological feature (gene, protein, process)

Stressor (chemical, virus)



- ToxCast high-throughput screening (HTS) data for bioactivity profiling (<https://actor.epa.gov/dashboard/>), in progress.

## Adverse Outcome Pathway (AOP)



**KEY EVENT 4:** reduced neurogenic capacity in the ventricular zone of the brain, leading to cortical thinning and reduced brain size at the 2<sup>nd</sup> trimester.

- Human brain size is determined by the number of neurogenic cells available to form neocortex and is a function of the precursor pool size of neuroprogenitor cells (NPCs) [3].
- NPCs self-replicate in the ventricular zone of the brain during 1<sup>st</sup> trimester; this growth period is followed by differentiation to form neocortex by the 2<sup>nd</sup> trimester [4].

**KEY EVENT 3:** altered neuroprogenitor growth kinetics, leading to hypoplasia of the neurogenic niche in the 1<sup>st</sup> trimester.

- Zika virions infect hNPCs (but not hES cells or neurons) *in vitro*, and the resulting consequences on cell growth/apoptosis has a demonstrable effect on hNPC-derived neurosphere size [5,6,7].
- Chemical injury (alcohol, methylmercury) during rodent neurodevelopment alters NPC pool sizes via adverse effects on the growth kinetics (cell growth/migration/apoptosis) [8].

**KEY EVENT 2:** misorientation of hNPC mitotic division, leading to premature loss of neuroprogenitors from the proliferative cycle.

- hNPCs divide symmetrically before switching to asymmetrical (neurogenic) divisions; premature switching (or apoptosis) results in loss of NPCs from the proliferative cycle [3,4].
- Alignment of the mitotic spindle determines the polarity of mitotic divisions to self-replicate hNPCs (equal division) or spinoff a daughter cell that enters the neurogenic lineage (unequal division) [9].

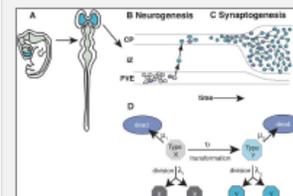
**KEY EVENT 1:** dysregulation of the centrosome cycle, leading to misalignment of the centrioles and the microtubule organizing center of the cell.

- Many candidate genes for human 'primary microcephaly' function in the structural organization and regulation of the centrosome, containing two centrioles at right angles to each other [10].
- Transcriptomic analysis showed hNPCs express five genes for human primary microcephaly (MCPH1, ASPM, CENJ, STIL, CDSRAP2) as an indication of a centrosomal cycle for further investigation of this hypothesis.

**MOLECULAR INITIATING EVENT:** ToxCast bioactivity profiles are a resource for building predictive signature(s) for microcephaly.

- 30 ToxCast chemicals that invoke decreases in brain developmental parameters had bioactivity profiles mapping to one or more targets in a ToxCast assay (top gene scores = p53, NRF2, PXR, AhR, HSF1, VDR, ...).
- Bioactivity profiles are being used to build predictive signatures for microcephaly that can be used in modeling the system, both *in silico* (computational) and *in vitro* (human brain mimics).

## Computational Model of Neurodevelopment



LEFT: model for the neurogenic switch to assess criticality of NPC loss on neurodevelopment [8].  
 BELOW: parameters to simulate NPC dynamics mathematically following chemical (or viral) exposure [9]. This model can be applied to simulate Key Events 3-4 in the AOP.

Parameterization and equations of the dynamic model for neurodevelopment

Kolmogorov forward differential equation for transition probabilities

Equation  $\frac{dP(x, y, t)}{dt} = (x - 1)\lambda_1(t)P(x-1, y, t) + (x+1)\lambda_1(t)P(x+1, y, t) + (y - 1)\lambda_2(t)P(x, y-1, t) + (y+1)\lambda_2(t)P(x, y+1, t) + (x+1)\mu_1(t)P(x+1, y-1, t) - [x\lambda_1(t) + y\lambda_2(t) + x\mu_1(t)]P(x, y, t)$

Parameters  $P(x, y, t) = P(X(t)=x, Y(t)=y|X(t_0)=x_0, Y(t_0)=y_0)$ .  $X(t)$  and  $Y(t)$  denote the numbers of X and Y cells at time  $t$ .  $x_0$  is the number of X cells at initial time  $t_0$ .  $\lambda_1(t)$  and  $\lambda_2(t)$  are the reproduction and death rates for X cells,  $\lambda_2(t)$  and  $\mu_1(t)$  are the reproduction and death rates for Y cells,  $\nu(t)$  is the rate of transformation of X cells to Y cells.

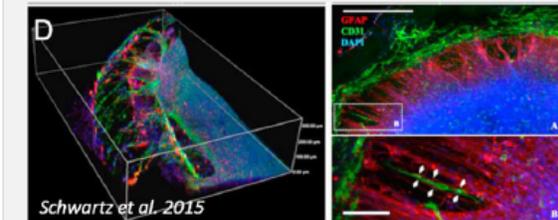
Matrix to approximate solution of the above equation

	$x_1(t)$	0	0	0	0
	$\nu(t)$	$x_2(t)$	0	0	0
Matrix	$\lambda_1(t) + \mu_1(t) + \nu(t)$	0	$2x_1(t)$	0	0
	$\nu(t)$	$\lambda_2(t) + \mu_2(t)$	0	$2x_2(t)$	$\nu(t)$
	$\nu(t)$	0	$\nu(t)$	0	$x_1(t) + x_2(t)$

Parameters  $x_1(t) = \lambda_1(t) - \mu_1(t) - \nu(t)$  and  $x_2(t) = \lambda_2(t) - \mu_2(t)$

## Human Brain Mimics

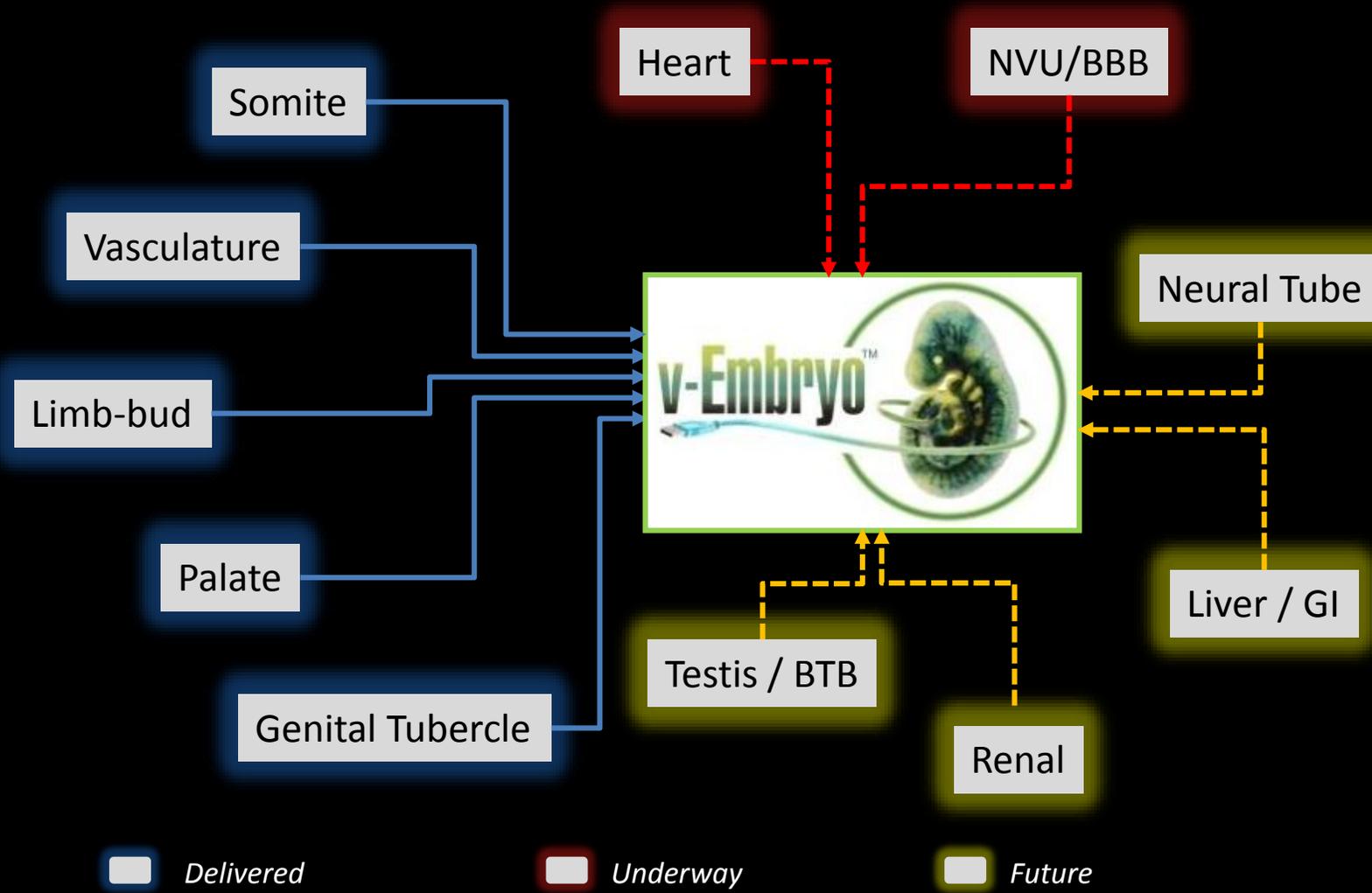
Dynamics of hNPC growth, migration, and apoptosis for the computational model can be assessed in miniorganoids developed from hNPCs + iPSC-derived endothelial and microglia [11]. Studies are planned to develop human brain mimics from microcephalic patient-derived iPSCs and to provide evidence for Key Events 1-3 in the AOP for chemicals and Zika.



## REFERENCES

- [1] Rasmussen et al. (2016) N Engl J Med, April 14
- [2] Von Der Hagen et al. (2014) Dev Med Child Neurol 56
- [3] Otani et al. (2016) Cell Stem Cell 18
- [4] Tyler and Haydar (2010) Nat Neurosci 13
- [5] Tang et al. (2016) Cell Stem Cell 18
- [6] Garcez et al. (2016) Science
- [7] Cugola et al. (2016) Nature
- [8] Gohlke et al. (2005) Toxicol Sci 86
- [9] Faustman et al. (2005) Envi Toxicol Pharmacol 19
- [10] Barbelanne and Tsang (2014) Biomed Res International
- [11] Schwartz et al. (2015)

# Toward a 'Virtual Embryo'



- Hester et al. (2011) PLoS Comp Bio; Dias et al (2014) Science
- Kleinstreuer et al. (2013) PLoS Comp Bio.
- Ahir et al. (MS in preparation).
- Hutson et al. (2017) Chem Res Toxicol (revision).

- Leung et al. (2016) Reprod Toxicol.
- Zurlinden/Saili et al. (FY17 product).
- Hunter et al. (FY18 product).
- Your name here.

# Virtual Tissue Laboratory System (VTLS)

The screenshot shows a web browser window with the URL `iris2.rtpnc.epa.gov` and the page title `VT-LS(beta)`. The main header features the EPA logo and the text "Virtual Tissues Laboratory System (VT-LS): A computational framework for developmental toxicity".

On the left side, there is a navigation menu with two main sections:

- Information**
  - × Login
  - Logout
  - Help
  - Contact Us
- Jobs**
  - × Submit
  - × Status

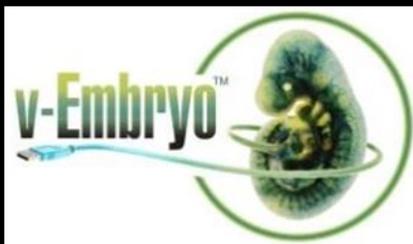
The main content area displays "Welcome to vEmbryo" with a logo for "v-Embryo" showing a green embryo. Below the logo are three input fields: "User Name:", "Email:", and "Affiliation:", each followed by a small downward arrow icon.

A "Login" modal dialog box is open in the center, containing:

- Username:
- Password:
- A "Login" button

# Special Thanks

- Sid Hunter – NHEERL / ISTD
- Max Leung – NCCT (now U Pittsburgh)
- Bhavesh Ahir – NCCT (now U Illinois – Chicago)
- Nicole Kleinstreuer - NCCT (now NIH/NICEATM)
- Nisha Sipes – NCCT (now NTP)
- Richard Spencer – Leidos / EMVL
- Nancy Baker – Leidos / NCCT
- Ed Carney† – Dow Chemical Company
- Rob Ellis-Hutchings – Dow Chemical Company
- Tuula Heinonen – U Tampere / FICAM
- Jessica Palmer – Stemina Biomarker Discovery
- Parth Kothiya – NCCT (now Indiana U)
- James Glazier – Indiana U (TIVS)
- Shane Hutson – Vanderbilt U (VPROMPT)
- Kate Sali – NCCT
- Todd Zurlinden – NCCT
- BeiBei Cai – Vala Sciences
- Dan Rines – Vala Sciences
- Jill Franzosa – NCCT (now CSS)
- Brian Johnson – U Wisconsin (HMAPS)
- Eric Nguyen – U Wisconsin (HMAPS)
- William Murphy – U Wisconsin (HMAPS)
- William Daly – U Wisconsin (HMAPS)
- Tamara Tal – NHEERL/ISTD
- David Belair – NHEERL/ISTD
- Barbara Abbott – NHEERL/ISTD
- Imran Shah - NCCT
- Alex Tseutaki – HS intern



[http://www2.epa.gov/sites/production/files/2015-08/documents/virtual\\_tissue\\_models\\_fact\\_sheet\\_final.pdf](http://www2.epa.gov/sites/production/files/2015-08/documents/virtual_tissue_models_fact_sheet_final.pdf)