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IL-10 reduces grain dust-induced airway inflammation and airway hyperreactivity

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Quinn, Timothy J., Somer Taylor, Christine L. Wohlford-Lenane, and David A. Schwartz. IL-10 reduces grain dust-induced airway inflammation and airway hyperreactivity. J. Appl. Physiol. 88: 173–179, 2000.—To determine whether interleukin-10 (IL-10) could alter the development of grain dust-induced airway disease, we pretreated mice with either saline or IL-10 intravenously, exposed the mice to an inhalation challenge with corn dust extract (CDE), and measured inflammation and the development of airway hyperreactivity. Pretreatment with IL-10, in comparison to saline, reduced the concentration and percentage of polymorphonuclear cells in the lavage fluid 30 min after the inhalation challenge with CDE (P < 0.05). In comparison to salinetreated mice, IL-10 did not significantly alter the degree of airway hyperreactivity 30 min after the exposure to CDE. IL-10-treated mice lavaged 18 h after challenge with CDE also exhibited a lower percentage of polymorphonuclear cells in the lavage fluid (P < 0.05) and had significantly less airway hyperreactivity than did mice pretreated with the saline placebo (P < 0.05). These findings indicate that exogenous IL-10 is effective in reducing airway inflammation and airway hyperreactivity due to the inhalation of CDE.

interleukin-10; neutrophils; asthma; lipopolysaccharide

OCCUPATIONAL AND ENVIRONMENTAL exposure to grain dust can cause a spectrum of clinical syndromes, including asthma, acute and chronic changes in airway reactivity, bronchitis, and progressive irreversible airflow obstruction (5). Among grain handlers, the prevalence of work shift changes in forced expiratory volume in 1 s (≥10% decline) varies between 3.9 and 11% (5). Grain dust exposure causes seasonal decrements in airflow (26) and an accelerated longitudinal decline in forced expired volume in 1 s (23). Recent studies in Barcelona, Spain, suggest that environmental exposure to grain dust significantly affects the health of the exposed population, resulting in a higher incidence of asthmatic attacks (1). Chronic exposure to grain dust can cause irreversible and progressive airway disease. Epidemiological studies performed in North America (40), the United Kingdom (3), and South Africa (47) demonstrate that workers chronically exposed to grain dust are at increased risk of developing chronic cough, wheeze, and dyspnea, irrespective of smoking habits. Long-term follow-up studies have shown that grain workers (6), as well as other workers exposed to organic dusts (9, 38), have accelerated development of airflow obstruction.

Several lines of evidence indicate that endotoxin is one of the primary agents in grain dust and other organic dusts that cause airway inflammation and airflow obstruction. First, the concentration of inhaled endotoxin in the bioaerosol is strongly associated with the development of acute decrements in airflow among cotton workers (29), swine-confinement workers (13), and poultry workers (44). The concentration of endotoxin in the bioaerosol is the most important occupational exposure associated with the development (40) and progression (38) of airway disease in agricultural workers. Second, physiologically, inhaled endotoxin (34), grain dust (10), or cotton dust (4) can cause airflow obstruction in naïve or previously unexposed subjects. Naïve, healthy study subjects challenged with dust from animal confinement buildings develop airflow obstruction and an increase in the serum concentration of neutrophils and interleukin (IL)-6, all of which are most strongly associated with the concentration of endotoxin (not dust) in the bioaerosol (48). Finally, previous exposure-response studies from our laboratory have shown that inhaled grain dust and endotoxin produce similar physiological and biological effects in humans (10) and mice (39); the concentration of endotoxin in grain dust plays an important role in the acute biological response to grain dust in humans (25) and mice (39); a competitive antagonist for lipopolysaccharide (LPS; Rhodobacter spheroides diphosphoryl lipid A) reduces the inflammatory response to inhaled grain dust in mice (24); and genetic or acquired hyporesponsiveness to endotoxin substantially reduces the biological response to grain dust in mice (39). Taken together, these studies indicate that endotoxin is an important cause of grain dust-induced airway disease.

IL-10, which was originally called "cytokine inhibitory factor," is a 35-kDa homodimer produced by Th2 cells, B cells, monocytes, and keratinocytes (17). The major effect of IL-10 is to attenuate the release of proinflammatory cytokines. Although IL-10 has direct effects on neutrophils (28), T cells (46), and B cells (37), its principal target is the macrophage (46). One of the primary effects of IL-10 is the inhibition of macrophage IL-12 production; by inhibiting the release of IL-12,
IL-10 decreases the subsequent secretion of interferon-γ (IFN-γ), and this, in turn, decreases the release of cytokine effect on molecules such as tumor necrosis factor-α (TNF-α) (28, 37, 46). The ability of IL-10 to antagonize IFN-γ is macrophage and monocyte dependent, and IL-10 is not an effective inhibitor of IFN-γ in systems in which exogenous IL-12 is present (46). Thus IL-10 targets non-IL-12-secreting (nonactivated) macrophages and inhibits activation and IL-12 secretion (37). Importantly, after intravenous exposure to IL-10, whole blood stimulated by LPS ex vivo released significantly less TNF-α and IL-1β without affecting the release of IL-1-receptor antagonist (IL-1ra) or soluble TNF-α receptor (7). IL-10 substantially reduces the inflammatory response to intravenous endotoxin that appears to be mediated by downregulating the release of TNF-α, IL-8, and IL-12 (45). We have found that serum concentrations of IL-10 can be significantly increased by treating mice with oligonucleotides containing unmethylated cytosine-guanine dinucleotide motifs (41). These studies, which indicate that IL-10 can suppress LPS-induced inflammation, suggest that IL-10 may be effective in preventing the inflammation and physiological effects of grain dust, an airway disease that is primarily mediated by endotoxin.

Given the anti-inflammatory properties of IL-10 and its specific effects on endotoxin-mediated inflammation, we hypothesized that pretreatment with IL-10 would substantially reduce the airflow obstruction and airway inflammation after inhalation of grain dust, an inflammatory response that appears to be primarily mediated by endotoxin.

Methods

Experimental protocol. This investigation was designed to determine whether IL-10 was effective in reducing the physiological or inflammatory response to inhaled grain dust extract. Thus mice were treated with either IL-10 (50 µg/kg) or saline intravenously 2 h before a 4-h challenge with corn dust extract (CDE). A previous study has shown that IL-10 is effective in protecting mice against staphylococcal enterotoxin B-induced lethal shock if it is given up to 3 h before the challenge (2).

Lung physiology. Lung physiology and whole lung lavage were performed at 30 min and 18 h after the inhalation challenge and 18 h after the challenge to assess the effects of grain dust, an airway disease that is primarily mediated by endotoxin.

Lung lavage. Lung lavage was performed 30 min after the inhalation challenge and 18 h after the challenge to assess whether administration of IL-10 altered the degree of lower airway inflammation after inhalation of CDE. Mice were euthanized and underwent in situ lavage by using PE-90 tubing inserted into the exposed trachea. The lungs were lavaged with 6.0 ml of sterile pyrogen-free saline. After lavage, the lungs were isolated and frozen in liquid nitrogen and stored at −70°C.

Treatment of bronchoalveolar lavage fluid. Lavage samples were centrifuged for 5 min at 200 g, and the supernatant was decanted and frozen at −70°C for subsequent use. Cell concentrations were determined by resuspending the residual pellet in Hanks' balanced salt solution (without Ca2+ or Mg2+) and counting on a hemocytometer. Cells were placed on a glass slide by using a cytocentrifuge (Cytospin-2, Shandon Southern, Sewickley, PA) and stained by using a Diff Quik Stain (Dade, Miami, FL) for review by a blinded reviewer. Cells were then counted on a hemocytometer.

Chemicals. Mice were pretreated with commercially available recombinant murine IL-10 (catalog no. 417-ML-025, R&D Systems, Minneapolis, MN). Corn dust used in this study was obtained from the air filtration system at an eastern Iowa grain-handling facility. CDE was prepared by mixing 3.0 g of dust in 30 ml of 0.1% solution of Hanks' balanced salt solution purchased from the University of Iowa Tissue Culture Facility. This mixture was vortexed for 2 min and shaken for 1 h at 4°C. Next, the mixture was centrifuged at 800 g for 20 min, and the supernatant solution was collected, resulting in CDE. This solution was filter sterilized through a 0.22-µm filter (Durapore polyvinylidifluoroide filter unit, Millipore, Bedford, MA) and then stored at −70°C before use.

Endotoxin assay. The endotoxin concentrations of the CDE aerosols were assayed by using the chromogenic Limulus amebocyte lysate assay (QCL-1000, Whittaker Bioproducts, Walkersville, MD) with sterile, pyrogen-free labware and a temperature-controlled microplate block and microplate reader (405 nm). The solution was serially diluted in pyrogen-free water (pfw) and assayed. The airborne concentration of endotoxin was assessed by sampling 0.30 m³ of air drawn from the exposure chamber through 47-mm binder-free glass microfiber filters (EMI-2000, Whatman International, Maidstone, UK) held within a 47-mm stainless steel in-line air-sampling filter holder (Gelman Sciences, Ann Arbor, MI). Air-sampling filters were extracted with 30 ml of pfw at room temperature with gentle shaking for 1 h. The solutions were then serially diluted with pfw and assayed for endotoxin. Four air samples were assayed for each exposure at evenly spaced time intervals during each 4-h exposure.

Exposure protocol and monitoring equipment. CDE aerosols were generated into a glass 20-liter exposure chamber by using a Collison nebulizer (BGI, Waltham, MA). High-efficiency particulate-filtered air was supplied to the nebulizer at a flow rate of 17 1/min, and the chamber atmosphere was exchanged at 1 change/min. Endotoxin concentrations in the aerosol were determined by sampling the total chamber outflow. The concentration of endotoxin was 4.2 µg/m³ in the CDE aerosol used for the mice tested 30 min after exposure and 5.3 µg/m³ in the CDE aerosols for the mice tested 18 h after exposure.

Lung lavage. Lung lavage was performed 30 min after the inhalation challenge and 18 h after the challenge to assess whether administration of IL-10 altered the degree of lower airway inflammation after inhalation of CDE. Mice were euthanized and underwent in situ lavage by using PE-90 tubing inserted into the exposed trachea. The lungs were lavaged with 6.0 ml of sterile pyrogen-free saline. After lavage, the lungs were isolated and frozen in liquid nitrogen and stored at −70°C.

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compression (due to forced expirations). Airway resistance is expressed as enhanced pause (Penh). Penh = (expiratory time/40% of relaxation time) × peak expiratory flow/peak inspiratory flow × 0.67. The validity of Penh as a measure of bronchoconstriction has been examined (42). Penh was measured at baseline and after stimulation with inhaled methacholine (12.5 and 25 mg/ml) according to a standard protocol (42).

Cytokine analysis of lavage fluid. Lavage fluid was assayed for TNF-α, macrophage inflammatory protein-2 (MIP-2), and IL-1β. Polyclonal antibodies specific for each recombinant murine cytokine were used as a capture reagent in a standard commercially available sandwich ELISA (R&D Systems). The expression of these specific cytokines and chemokines has been shown to be associated with the acute inflammatory response to inhaled grain dust (12).

Preparation of RNA and multiprobe Rnase protection assay. Total RNA was extracted from lung specimens by using a single-step method, lysing flash-frozen lung in RNA STAT-60 (Tel-Test; Friendswood, TX). The composition of RNA STAT-60 includes phenol and guanidinium thiocyanate in a monophase solution. The lung parenchyma was homogenized in the RNA STAT-60 by using a polytron homogenizer. Chloroform was then added, and the total RNA was precipitated from the aqueous phase by addition of isopropanol. The total RNA was then washed with ethanol and solubilized in water. After the pellet was dried in a vacuum desiccator, the yield and purity of RNA were quantitated by measuring the ratio of absorbances at 260 and 280 nm. Minigel electrophoresis was used to confirm the integrity of the 28S and 18S rRNA bands. Equivalent amounts of RNA were examined, as judged by the amount of L32, which encodes a ubiquitously expressed ribosome subunit protein (928), in each sample. Commercially available probes were used to detect TNF-α, IL-1β, and MIP-2 (Pharmingen, San Diego, CA). Densitometry was performed to control for the concentration of RNA in each lane by using Sigma gel (Jandel Scientific Software, San Rafael, CA).

Data analysis. The purpose of this experiment was to compare the inflammatory and physiological response to inhaled CDE between mice pretreated with IL-10 and those pretreated with saline. Inflammation was assessed by using lavage cellularity, lavage fluid cytokine concentrations, and the relative concentrations of mRNA for specific cytokines in the lung parenchyma. The physiological response to inhaled CDE was evaluated by measuring the airway response to inhaled methacholine. Given the number of mice for each comparison (6 per group), the Mann-Whitney U nonparametric statistical test was used for all comparisons (36). A P value of <0.05 was considered statistically significant. Six mice for each condition were used on the basis of a sample-size calculation that included a desired power of 80% and anticipated effect of IL-10 of 50% reduction in lung inflammation and that used estimates of lung inflammation from previous work from our laboratory (12).

Results

In a preliminary dosing study, 10 µg/kg IL-10 administered intravenously were ineffective in reducing inflammation due to inhaled LPS. However, 50 µg/kg intravenous IL-10 resulted in a reduced concentration of cells and neutrophils in the lavage fluid of mice challenged with inhaled LPS. On the basis of these
findings, subsequent experiments with CDE aerosols were performed by using 50 µg/kg. Pretreatment with exogenous IL-10 reduced the recruitment of inflammatory cells to the lung after inhalation of CDE. Thirty minutes after the exposure to CDE, mice pretreated with IL-10 had lower concentrations of total cells (P = 0.07) and neutrophils (P = 0.02) and percent neutrophils (P = 0.03) in the lung lavage fluid compared with mice pretreated with saline (Fig. 1). This trend was also present 18 h after inhalation of CDE (Fig. 1), although the percent neutrophils was the only statistically significant difference (P = 0.02). No differences were observed in the concentration of macrophages, lymphocytes, or eosinophils between mice treated with IL-10 and saline for either of the time points. The CDE inhalation challenge involved two separate exposures, one for those killed 30 min after the exposure and one for those killed 18 h after the exposure. Thus the absolute increase in total lavage cells observed at 18 h may have been caused by minor differences in the exposure conditions.

Pretreatment with IL-10 did not significantly alter the concentration of TNF-α, IL-1β, or MIP-2 in the lavage fluid after challenge with CDE (Fig. 2). Thirty minutes after inhalation of CDE, mice pretreated with IL-10 had lower concentrations of TNF-α and MIP-2 in the lavage fluid than did mice pretreated with saline. However, these differences did not prove to be statistically significant. IL-1β does not appear to be affected by pretreatment with IL-10. The concentration of cytokines in the lavage fluid returned to negligible baseline values 18 h after the exposure, and no differences were observed between IL-10- and saline-treated mice (data not presented). IL-10 did not affect the induction of mRNA for TNF-α, IL-1β, or MIP-2 in the lung after inhalation of CDE, and 18 h after inhalation of CDE, mRNA for these cytokines was not detectable (Fig. 3).

Pretreatment with IL-10 resulted in less airway hyperreactivity after inhalation of CDE. Figure 4 presents the results of the lung physiology after inhalation of CDE. The Penh was expressed as the percent change from preexposure values (performed 24 h before the inhalation challenge with CDE) at 30 min or 18 h after the inhalation challenge with CDE. Thirty minutes after the inhalation challenge with CDE, mice pretreated with IL-10 had less (albeit not statistically significant) airway hyperreactivity than did mice pretreated with saline. However, consistent and significant differences in airway hyperreactivity were observed 18 h after inhalation challenge with CDE, with IL-10-treated mice demonstrating significantly lower levels of airway hyperreactivity than saline-treated mice. These findings suggest that IL-10 may reduce the degree of airflow obstruction observed after inhalation of grain dust.
CD14 receptor. Because the CD14 receptor is the lipid A moiety of LPS and prevents stimulation of the permeability-increasing protein (16) that binds the use of LPS antagonists (8) and a bactericidal and response to endotoxin. Such interventions include theinhaled grain dust could focus on the initial biological reduced the severity of the response to intravenousinfusion of IL-1ra to human volunteers significantlycan block LPS-mediated release of IgE in vitro (43), andas homeostatic mechanisms. Administration of IL-1raaggregate, these findings suggest that control of theacute inflammatory response with agents such as IL-10may prevent the long-term complications of chronicinhalaion of grain dust (3, 6, 40, 47).

Our findings indicate that IL-10 can decrease thephysiological and inflammatory response to inhaledcorn dust. Although the specific relationship betweenacute airway inflammation and the development ofchronic airway disease is not fully understood, thepresent evidence in asthmatic children (31) indicates thatrecurrent episodes of airway inflammation (notabnormal lung function in the first year of life) result inpersistent airway hyperresponsiveness and chronicairway disease. There is increasing evidence amongasthmatic individuals that control of the acute inflam-matory response substantially improves airflow andchronic airway inflammation (11, 21). Prolonged treat-ment of newly diagnosed mildly asthmatic (19), chroni-cally stable asthmatic (27), and severely asthmaticindividuals (15) with inhaled corticosteroids resulted insignificant improvement in airflow. Patients with se-vere intrinsic asthma treated for 10 yr with inhaledsteroids demonstrated reduced airway inflammatorycells and restoration of ciliated epithelia (30). In theaggregate, these findings suggest that control of theacute inflammatory response with agents such as IL-10may prevent the long-term complications of chronicinhalaion of grain dust (3, 6, 40, 47).

Discussion

Our results indicate that pretreatment with exog-enous IL-10 reduces airway inflammation and airwayhyperreactivity after inhalation of CDE. Our resultsuggest that IL-10 or other anti-inflammatory cyto-kines may prove to be beneficial in treating grain-dustinduced airway disease. Moreover, prevention of theacute biological and physiological response to inhaledgrain dust with agents such as IL-10 may reduce therisk of developing chronic grain dust-induced airwaydisease.

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Given the importance of endotoxin in mediatingorganic dust-induced airway disease, one could specu-late that several other agents, in addition to IL-10, could effectively prevent the acute response to inhaledgrain dust. IL-1ra is an 18- to 22-kDa protein isolatedfrom monocytes (20) and alveolar macrophages (35)that appears to specifically antagonize the IL-1 recep-tor and has no agonist effects. The production of IL-1rais triggered by stimuli that differ from those that lead toproduction of IL-1. For instance, in human volunteerschallenged with grain dust, we have found that bothprotein and mRNA for IL-1β and IL-1ra are upregulatedin the bronchoalveolar space and inflammatorycells (10), suggesting that these cytokines are inducedas homeostatic mechanisms. Administration of IL-1ra canblock LPS-mediated release of IgE in vitro (43), andinfusion of IL-1ra to human volunteers significantlyreduced the severity of the response to intravenousLPS (18). Other strategies to control the response toinhaled grain dust could focus on the initial biologicalresponse to endotoxin. Such interventions include theuse of LPS antagonists (8) and a bactericidal andpermeability-increasing protein (16) that binds thelipid A moiety of LPS and prevents stimulation of theCD14 receptor. Because the CD14 receptor is theprimary receptor for endotoxin, interfering with theinteraction between the CD14 receptor and endotoxinshould significantly reduce the chronic airway response toinhaled grain dust.

Grain dust-induced airway disease serves as anexcellent model to investigate the biological features of reversible airway inflammation that are fundamental to the development of chronic asthma. In fact, recentreports have indicated that the concentration of endo-toxin in the domestic setting is related to the clinicalseverity of asthma (33). Moreover, asthmatic individu-als develop airflow obstruction at lower concentrationsof inhaled endotoxin (32), and inhalation of allergensincreases the lung’s biological responsiveness to endo-toxin (14). Interestingly, inhaled allergens appear to increase the concentration of LPS-binding protein, which allows the lung inflammatory cells to respond tovery low concentrations of endotoxin that are com-monly present in the airways of uninfected lungs (14).These findings may explain why antigen preparationscontaminated with very low levels of endotoxin causeaneutrophilic response in asthmatic subjects (22). Thusagents that prove to be effective in preventing graindust-induced airway disease may also prove to beeffective in asthma. In fact, IL-10 inhibits eosinophilrecruitment to the airway in mice sensitized andchallenged with ovalbumin (49).

In conclusion, our findings indicate that IL-10 ap-pears to reduce the inflammatory and physiologicaleffects of inhaled CDE. These findings suggest thatcytokine regulation is important in controlling theinflammatory response to inhaled grain dust. In addi-tion, our results suggest that IL-10, as well as otheranti-inflammatory cytokines, may provide a novel ap-proach to the treatment of grain-dust induced airwaydisease.

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IL-10 AND GRAIN DUST-INDUCED AIRWAY DISEASE


