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Toxicological Review of *tert*-Butyl Alcohol (*tert*-Butanol)

(CAS No. 75-65-0)

In Support of Summary Information on the Integrated Risk Information System (IRIS)

September 2014

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ABBREVIATIONS

α 2u-g	alpha2u-globulin	LOAEL	lowest-observed-adverse-effect level
ACGIH	American Conference of Governmental Industrial Hygienists	MN	micronuclei
AIC	Akaike's information criterion	MNPCE	micronucleated polychromatic erythrocyte
ALD	approximate lethal dosage	MTD	maximum tolerated dose
ALT	alanine aminotransferase	NAG	N-acetyl- β -D-glucosaminidase
AST	aspartate aminotransferase	NCEA	National Center for Environmental Assessment
atm	atmosphere	NCI	National Cancer Institute
ATSDR	Agency for Toxic Substances and Disease Registry	NOAEL	no-observed-adverse-effect level
BMD	benchmark dose	NTP	National Toxicology Program
BMDL	benchmark dose lower confidence limit	NZW	New Zealand White (rabbit breed)
BMDS	Benchmark Dose Software	OCT	ornithine carbamoyl transferase
BMR	benchmark response	ORD	Office of Research and Development
BUN	blood urea nitrogen	PBPK	physiologically based pharmacokinetic
BW	body weight	PCNA	proliferating cell nuclear antigen
CA	chromosomal aberration	POD	point of departure
CASRN	Chemical Abstracts Service Registry Number	POD _[ADJ]	duration-adjusted POD
CBI	covalent binding index	QSAR	quantitative structure-activity relationship
CHO	Chinese hamster ovary (cell line cells)	RDS	replicative DNA synthesis
CL	confidence limit	RfC	inhalation reference concentration
CNS	central nervous system	RfD	oral reference dose
CPN	chronic progressive nephropathy	RGDR	regional gas dose ratio
CYP450	cytochrome P450	RNA	ribonucleic acid
DAF	dosimetric adjustment factor	SAR	structure activity relationship
DEN	diethylnitrosamine	SCE	sister chromatid exchange
DMSO	dimethylsulfoxide	SD	standard deviation
DNA	deoxyribonucleic acid	SDH	sorbitol dehydrogenase
EPA	Environmental Protection Agency	SE	standard error
FDA	Food and Drug Administration	SGOT	glutamic oxaloacetic transaminase, also known as AST
FEV ₁	forced expiratory volume of 1 second	SGPT	glutamic pyruvic transaminase, also known as ALT
GD	gestation day	SSD	systemic scleroderma
GDH	glutamate dehydrogenase	TCA	trichloroacetic acid
GGT	γ -glutamyl transferase	TCE	trichloroethylene
GSH	glutathione	TWA	time-weighted average
GST	glutathione-S-transferase	UF	uncertainty factor
Hb/g-A	animal blood:gas partition coefficient	UF _A	animal-to-human uncertainty factor
Hb/g-H	human blood:gas partition coefficient	UF _H	human variation uncertainty factor
HEC	human equivalent concentration	UF _L	LOAEL-to-NOAEL uncertain factor
HED	human equivalent dose	UF _S	subchronic-to-chronic uncertainty factor
i.p.	intraperitoneal	UF _D	database deficiencies uncertainty factor
IRIS	Integrated Risk Information System	U.S.	United States of America
IVF	in vitro fertilization		
LC ₅₀	median lethal concentration		
LD ₅₀	median lethal dose		

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PREFACE

This Toxicological Review critically reviews the publicly available studies on *tert*-butyl alcohol (*tert*-butanol) in order to identify its adverse health effects and to characterize exposure-response relationships. It was prepared under the auspices of EPA's Integrated Risk Information System (IRIS) Program. The assessment covers an oral RfD, an inhalation RfC, and a cancer weight of evidence descriptor.

The Toxicological Reviews for ethyl tert-butyl ether (ETBE) and *tert*-butanol are being developed simultaneously because they have a number of overlapping scientific issues:

- *tert*-Butanol is a metabolite of ETBE, so some of the toxicological effects of ETBE may be due to *tert*-butanol. Therefore, data on *tert*-butanol may inform hazard identification and dose-response assessment of ETBE, and vice versa.
- The scientific literature for chemicals include data on α_{2u} -globulin-related nephropathy. Therefore, a common approach was employed to evaluate those data as they relate to the mode of action for kidney effects.
- A combined PBPK model for ETBE and *tert*-butanol in rats was developed to support the dose-response assessments for these chemicals.

This assessment was conducted in accordance with EPA guidance, which is cited and summarized in the Preamble to IRIS Toxicological Reviews. This is the first IRIS assessment for this compound. The findings of this assessment and related documents produced during its development are available on the IRIS Web site (<http://www.epa.gov/iris>).

A public meeting was held in December 2013 to obtain input on preliminary materials for *tert*-butanol, including draft literature searches and associated search strategies, evidence tables, and exposure-response arrays prior to the development of the IRIS assessment. All public comments provided were taken into consideration in developing the draft assessment. The complete set of public comments are available on the docket at <http://www.regulations.gov> (Docket ID No. EPA-HQ-ORD-2013-0111).

In April 2011, the National Research Council (NRC) released its *Review of the Environmental Protection Agency's Draft IRIS Assessment of Formaldehyde*. In addition to offering comments specifically about EPA's draft formaldehyde assessment, the NRC made several recommendations to EPA for improving the development of IRIS assessments. EPA agreed with the recommendations and is implementing them consistent with the Panel's "Roadmap for Revision," which viewed the full implementation of their recommendations by the IRIS Program as a multi-year process.

In response to the NRC's 2011 recommendations, the IRIS Program has made changes to streamline the assessment development process, improve transparency, and create efficiencies in

1 the Program. The NRC has acknowledged EPA's successes in this area. In May 2014, the NRC
2 released their report *Review of EPA's Integrated Risk Information System Process* reviewing the IRIS
3 assessment development process and found that EPA has made substantial improvements to the
4 IRIS Program in a short amount of time.

5 The draft *tert*-butanol assessment represents a significant advancement in implementing
6 the NRC recommendations. This assessment is streamlined, and uses tables, figures, and
7 appendices to increase transparency and clarity. It is structured to have distinct sections for the
8 literature search and screening strategy, study selection and evaluation, hazard identification, and
9 dose-response assessment. The assessment includes a comprehensive, systematic, and
10 documented literature search and screening approach, provides the database search strategy in a
11 table (databases, keywords), visually represents the inclusion and exclusion of studies in a flow
12 diagram, and all of the references are integrated within the Health and Environmental Research
13 Online (HERO) database. A study evaluation section provides a systematic review of
14 methodological aspects of epidemiology and experimental animal studies, including study design,
15 conduct, and reporting, that was subsequently taken into consideration in the evaluation and
16 synthesis of data from these studies. The evidence is presented in standardized evidence tables,
17 and exposure-response arrays. The hazard identification and dose-response sections include
18 subsections based on organ/system-specific effects in which the evidence is synthesized within and
19 integrated across all evidence for each target organ/systems.

20 In the draft *tert*-butanol assessment, the IRIS Program has attempted to transparently and
21 uniformly identify strengths and limitations that would affect interpretation of results. All animal
22 studies of *tert*-butanol that were considered to be of acceptable quality, whether yielding positive,
23 negative, or null results, were considered in assessing the evidence for health effects associated
24 with chronic exposure to *tert*-butanol. These studies were evaluated for aspects of design, conduct,
25 and reporting that could affect the interpretation of results and the overall contribution to the
26 synthesis of evidence for determination of human hazard potential using the study quality
27 considerations outlined in the Preamble. A brief summary of the evaluation is included in the
28 section on methods for study selection and evaluation. Information on study features related to this
29 evaluation is reported in evidence tables and documented in the synthesis of evidence. Discussion
30 of study strengths and limitations (that ultimately supported preferences for the studies and data
31 relied upon) were included in the text where relevant.

32 In this assessment, the IRIS Program is using existing guidelines to systematically approach
33 the integration of noncancer human, animal, and mechanistic evidence. In conducting this analysis
34 and developing the synthesis, the IRIS Program evaluates the data for the: strength of the
35 relationship between the exposure and response and the presence of a dose-response relationship;
36 specificity of the response to chemical exposure and whether the exposure precedes the effect;
37 consistency of the association between the chemical exposure and response; and biological
38 plausibility of the response or effect and its relevance to humans. The IRIS Program uses this

1 weight-of-evidence approach to identify the potential human hazards associated with chemical
2 exposure.

3 The IRIS *tert*-butanol assessment provides a streamlined presentation of information,
4 integrated hazard identification of all toxic effects, and derivation of organ/system-specific
5 reference values. Additionally, consistent with the goal that assessments should provide a
6 scientifically sound and transparent evaluation of the relevant scientific literature and presentation
7 of the analyses performed, this assessment contains an expanded discussion of study selection and
8 evaluation, as well as increased documentation of key assessment decisions.

9 For additional information about this assessment or for general questions regarding IRIS,
10 please contact EPA's IRIS Hotline at 202-566-1676 (phone), 202-566-1749 (fax), or
11 hotline.iris@epa.gov.

12 **Chemical Properties**

13 *tert*-Butanol is a white crystalline solid or colorless liquid (above 77°F) with a camphor-like
14 odor that is highly flammable ([NIOSH, 2005](#); [IPCS, 1987a](#)). *tert*-Butanol contains a hydroxyl
15 chemical functional group and is miscible with alcohol, ether, and other organic solvents and
16 soluble in water ([IPCS, 1987a](#)). Selected chemical and physical properties of *tert*-butanol are
17 presented in Table P-1.

18

1 **Table P-1. Physicochemical properties and chemical identity of *tert*-butanol**

Characteristic	Information	Reference
Chemical name	<i>tert</i> -Butanol	HSDB (2007)
Synonyms/Trade Names	<i>t</i> -butyl alcohol; <i>tert</i> -Butanol; <i>tert</i> -butyl alcohol; <i>t</i> -Butyl hydroxide; 1,1-Dimethylethanol; NCI-C55367; 2-Methyl-2-propanol; <i>tertiary</i> butanol; Trimethyl carbinol; Trimethyl methanol, <i>t</i> -butyl alcohol, TBA	HSDB (2007) IPCS (1987b)
Chemical Formula	C ₄ H ₁₀ O	HSDB (2007)
CASRN	75-65-0	HSDB (2007)
Molecular weight	74.12	HSDB (2007)
Melting point	25.7°C	HSDB (2007)
Boiling point	82.41°C	HSDB (2007)
Vapor pressure	40.7 mm Hg @ 25°C	HSDB (2007)
Density/Specific Gravity	0.78581	HSDB (2007)
Flashpoint	11°C (closed cup)	HSDB (2007)
Water solubility at 25°C	1 x 10 ⁶ mg/L	HSDB (2007)
Octanol/Water Partition Coefficient (Log K _{OW})	0.35	HSDB (2007)
Henry's Law Constant	9.05 x 10 ⁻⁶ atm·m ³ /mole	HSDB (2007)
Odor threshold	219 mg/m ³	HSDB (2007)
Conversion factors	1 ppm = 3.031 mg/m ³ 1 mg/m ³ = 0.324 ppm	IPCS (1987b)
Chemical structure	$ \begin{array}{c} \text{CH}_3 \\ \\ \text{H}_3\text{C} - \text{C} - \text{OH} \\ \\ \text{CH}_3 \end{array} $	HSDB (2007)

2

3 **Uses**

4 *tert*-Butanol is primarily an anthropogenic substance that is produced in large quantities

5 ([HSDB, 2007](#)) from a number of precursors, including 1-butene, isobutylene, acetyl chloride and

6 dimethylzinc, and *tert*-butyl hydroperoxide. The domestic production volume of *tert*-butanol,

7 including imports, was approximately four billion pounds in 2012 ([U.S. EPA, 2014](#)).

8 *tert*-Butanol has been used as a fuel oxygenate, an octane booster in unleaded gasoline, and

9 a denaturant for ethanol. From 1997 to 2005, the annual *tert*-butanol volume found in gasoline

10 ranged from approximately 4 million to 6 million gallons. During that time, larger quantities were

11 used to make methyl *tert*-butyl ether (MTBE) and ETBE. MTBE and ETBE are fuel oxygenates that

1 were used in the U.S. prior to 2007 at levels of more than 2 billion gallons annually. Current use
2 levels of MTBE and ETBE in the U.S. are much lower, but use in Europe and Asia remains strong¹.
3 *tert*-Butanol has been used for a variety of other purposes including as a dehydrating agent
4 and solvent. As such, it is added to lacquers, paint removers, and nail enamels and polishes.
5 *tert*-Butanol is also used to manufacture methyl methacrylate plastics and flotation devices.
6 Cosmetic and food-related uses include the manufacture of flavors and, because of its camphor-like
7 aroma, it is also used to create artificial musk, fruit essences, and perfume (HSDB, 2007). It is also
8 used in coatings on metal and paperboard food containers (Cal/EPA, 1999), industrial cleaning
9 compounds, and can be used for chemical extractions in pharmaceutical application (HSDB, 2007).

10 **Fate and Transport**

11 ***Soil***

12 The mobility of *tert*-butanol in soil is expected to be high due its low affinity for soil organic
13 matter. Rainwater or other water percolating through soil is expected to dissolve and transport
14 most *tert*-butanol present in soil, potentially leading to groundwater contamination. Based on
15 *tert*-butanol's vapor pressure, volatilization from soil surfaces is expected to be an important
16 dissipation process (HSDB, 2007). *tert*-Butanol is a tertiary alcohol and this class of alcohols
17 generally degrades more slowly in the environment compared to primary (e.g., ethanol) or
18 secondary (e.g., isopropanol) alcohols. In anoxic soil conditions, the half-life of *tert*-butanol is
19 estimated to be on the order of months (approximately 200 days). Microbial degradation rates are
20 increased in soils supplemented with nitrate and sulfate nutrients (HSDB, 2007).

21 ***Water***

22 *tert*-Butanol is expected to volatilize from water surfaces within 2 to 29 days and does not
23 readily adsorb to suspended solids and sediments in water (HSDB, 2007). Biodegradation in
24 aerobic water is on the magnitude of weeks to months and in anaerobic aquatic conditions, the
25 biodegradation rate decreases. Bioconcentration of *tert*-butanol in aquatic organisms is low
26 (HSDB, 2007).

27 ***Air***

28 *tert*-Butanol exists primarily as a vapor in the ambient atmosphere. Vapor-phase *tert*-
29 butanol is degraded in the atmosphere by reacting with photochemically-produced hydroxyl
30 radicals with a half-life of 14 days (HSDB, 2007).

31 **Occurrence in the Environment**

32 The Toxics Release Inventory (TRI) Program National Analysis Report estimated that over
33 one million pounds of *tert*-butanol has been released into the soil from landfills, land treatment,

² <http://www.ihs.com/products/chemical/planning/ceh/gasoline-octane-improvers.aspx>

1 underground injection, surface impoundments, and other land disposal sources. The TRI program
2 also estimated that 476,266 pounds of *tert*-butanol was released into the atmosphere from fugitive
3 emissions and point sources (U.S. EPA, 2012c). In California, air emissions of *tert*-butanol from
4 stationary sources are estimated to be at least 27,000 pounds per year, based on data reported by
5 the state's Air Toxics Program (Scorecard, 2014). The TRI National Analysis Report estimated
6 7,469 pounds of *tert*-butanol was released into surface waters from point and nonpoint sources in
7 2011 (U.S. EPA, 2012c).

8 *tert*-Butanol has been identified in drinking wells throughout the United States (HSDB,
9 2007). California's Geotracker Database² lists 3496 detections of *tert*-butanol in groundwater
10 associated with contaminated sites in that state since 2011. *tert*-Butanol has also been detected in
11 drinking water wells in the vicinity of landfills (U.S. EPA, 2012c). Additionally, *tert*-Butanol leaking
12 from underground storage tanks may be a product of MTBE and ETBE, which can degrade to form
13 *tert*-butanol in soils (HSDB, 2007). The industrial chemical *tert*-butyl acetate also can degrade to
14 form *tert*-butanol in animals and in the environment.

15 Ambient outdoor air concentrations of *tert*-butanol vary, seemingly according to proximity
16 to urban areas (HSDB, 2007).

17 **General Population Exposure**

18 *tert*-Butanol exposure can occur in many different settings. Contamination resulting from
19 leaking underground storage tanks could potentially result in exposure to a large number of people
20 who get their drinking water from wells. Due to its high environmental mobility and resistance to
21 biodegradation, *tert*-butanol has the potential to contaminate and persist in groundwater and soil;
22 therefore, exposure through ingestion of contaminated drinking water is likely occurring (HSDB,
23 2007).

24 Contaminated food can also contribute to *tert*-butanol ingestion through its use as a coating
25 in metallic and paperboard food containers (Cal/EPA, 1999). *tert*-Butanol has been detected in
26 food, namely beer and chickpeas, and identified in mother's milk (HSDB, 2007). Indirect exposure
27 to *tert*-butanol may also occur as a result of ingestion of MTBE or ETBE, as *tert*-butanol is a
28 metabolite of these compounds (NSF International, 2003).

29 Alternate human exposure pathways of *tert*-butanol include inhalation and, to a lesser
30 extent, dermal contact. *tert*-Butanol inhalation exposure can occur due to the chemical's volatility
31 and release from industrial processes, consumer products and contaminated sites (HSDB, 2007).
32 Dermal contact is a viable route of exposure through handling consumer products containing
33 *tert*-butanol (NSF International, 2003).

² <http://geotracker.waterboards.ca.gov/>

1 **Assessments by Other Federal, State and International Health Agencies**

2 Toxicity information on *tert*-butanol has been evaluated by the American Conference of
3 Governmental Industrial Hygienists ([ACGIH, 2012](#)), the National Institute for Occupational Safety
4 and Health ([NIOSH, 2007](#)), the Occupational Safety and Health Administration ([OSHA, 2006](#)), and
5 Food and Drug Administration. The results of these assessments are presented in Appendix A of
6 the Supplemental Information. The California EPA carried out an expedited risk assessment for
7 *tert*-butanol in drinking water and calculated a cancer slope factor based on rat kidney tumors
8 observed in the NTP bioassays. It is important to recognize that these earlier assessments were
9 prepared for different purposes using different methods and could consider only the studies that
10 were available at the time.

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12

DRAFT

PREAMBLE TO IRIS TOXICOLOGICAL REVIEWS

1. Scope of the IRIS Program

3 Soon after the EPA was established in
4 1970, it was at the forefront of developing
5 risk assessment as a science and applying it in
6 decisions to protect human health and the
7 environment. The Clean Air Act, for example,
8 mandates that the EPA provide “an ample
9 margin of safety to protect public health”; the
10 Safe Drinking Water Act, that “no adverse
11 effects on the health of persons may
12 reasonably be anticipated to occur, allowing
13 an adequate margin of safety.” Accordingly,
14 the EPA uses information on the adverse
15 effects of chemicals and on exposure levels
16 below which these effects are not anticipated
17 to occur.

18 IRIS assessments critically review the
19 publicly available studies to identify adverse
20 health effects from exposure to chemicals and
21 to characterize exposure-response
22 relationships. In terms set forth by the
23 National Research Council ([NRC, 1983](#)), IRIS
24 assessments cover the hazard identification
25 and dose-response assessment steps of risk
26 assessment, not the exposure assessment or
27 risk characterization steps that are conducted
28 by the EPA’s program and regional offices and
29 by other federal, state, and local health
30 agencies that evaluate risk in specific
31 populations and exposure scenarios. IRIS
32 assessments are distinct from and do not
33 address political, economic, and technical
34 considerations that influence the design and
35 selection of risk management alternatives.

36 An IRIS assessment may cover a single
37 chemical, a group of structurally or
38 toxicologically related chemicals, or a
39 complex mixture. These agents may be found
40 in air, water, soil, or sediment. Exceptions are
41 chemicals currently used exclusively as
42 pesticides, ionizing and non-ionizing
43 radiation, and criteria air pollutants listed
44 under Section 108 of the Clean Air Act

45 (carbon monoxide, lead, nitrogen oxides,
46 ozone, particulate matter, and sulfur oxides).

47 Periodically, the IRIS Program asks other
48 EPA programs and regions, other federal
49 agencies, state health agencies, and the
50 general public to nominate chemicals and
51 mixtures for future assessment or
52 reassessment. Agents may be considered for
53 reassessment as significant new studies are
54 published. Selection is based on program and
55 regional office priorities and on availability of
56 adequate information to evaluate the
57 potential for adverse effects. Other agents
58 may also be assessed in response to an urgent
59 public health need.

2. Process for developing and peer-reviewing IRIS assessments

60 The process for developing IRIS
61 assessments (revised in May 2009 and
62 enhanced in July 2013) involves critical
63 analysis of the pertinent studies,
64 opportunities for public input, and multiple
65 levels of scientific review. The EPA revises
66 draft assessments after each review, and
67 external drafts and comments become part of
68 the public record ([U.S. EPA, 2009](#)).

69 Before beginning an assessment, the IRIS
70 Program discusses the scope with other EPA
71 programs and regions to ensure that the
72 assessment will meet their needs. Then a
73 public meeting on problem formulation
74 invites discussion of the key issues and the
75 studies and analytical approaches that might
76 contribute to their resolution.

77 **Step 1. Development of a draft**
78 **Toxicological Review.** The draft
79 assessment considers all pertinent
80 publicly available studies and applies
81 consistent criteria to evaluate study
82 quality, identify health effects, identify
83 mechanistic events and pathways,

1 integrate the evidence of causation for
2 each effect, and derive toxicity values. A
3 public meeting prior to the integration of
4 evidence and derivation of toxicity values
5 promotes public discussion of the
6 literature search, evidence, and key
7 issues.

8 **Step 2. Internal review by scientists in**
9 **EPA programs and regions.** The draft
10 assessment is revised to address the
11 comments from within the EPA.

12 **Step 3. Interagency science consultation**
13 **with other federal agencies and the**
14 **Executive Offices of the President.** The
15 draft assessment is revised to address the
16 interagency comments. The science
17 consultation draft, interagency comments,
18 and the EPA's response to major
19 comments become part of the public
20 record.

21 **Step 4. Public review and comment,**
22 **followed by external peer review.** The
23 EPA releases the draft assessment for
24 public review and comment. A public
25 meeting provides an opportunity to
26 discuss the assessment prior to peer
27 review. Then the EPA releases a draft for
28 external peer review. The peer review
29 meeting is open to the public and includes
30 time for oral public comments. The peer
31 reviewers assess whether the evidence
32 has been assembled and evaluated
33 according to guidelines and whether the
34 conclusions are justified by the evidence.
35 The peer review draft, written public
36 comments, and peer review report
37 become part of the public record.

38 **Step 5. Revision of draft Toxicological**
39 **Review and development of draft IRIS**
40 **summary.** The draft assessment is
41 revised to reflect the peer review
42 comments, public comments, and newly
43 published studies that are critical to the
44 conclusions of the assessment. The
45 disposition of peer review comments and
46 public comments becomes part of the
47 public record.

48 **Step 6. Final EPA review and interagency**
49 **science discussion with other federal**
50 **agencies and the Executive Offices of**
51 **the President** The draft assessment and
52 summary are revised to address the EPA
53 and interagency comments. The science
54 discussion draft, written interagency
55 comments, and EPA's response to major
56 comments become part of the public
57 record.

58 **Step 7. Completion and posting.** The
59 Toxicological Review and IRIS summary
60 are posted on the IRIS website
61 (<http://www.epa.gov/iris/>).

62 The remainder of this Preamble addresses
63 step 1, the development of a draft
64 Toxicological Review. IRIS assessments
65 follow standard practices of evidence
66 evaluation and peer review, many of
67 which are discussed in EPA guidelines
68 ([U.S. EPA, 2005a, b, 2000b, 1998b, 1996,](#)
69 [1991b, 1986a, b](#)) and other methods ([U.S.](#)
70 [EPA, 2012a, b, 2011, 2006a, b, 2002,](#)
71 [1994](#)). Transparent application of
72 scientific judgment is of paramount
73 importance. To provide a harmonized
74 approach across IRIS assessments, this
75 Preamble summarizes concepts from
76 these guidelines and emphasizes
77 principles of general applicability.

3. Identifying and selecting pertinent studies

78 3.1. Identifying studies

79 Before beginning an assessment, the EPA
80 conducts a comprehensive search of the
81 primary scientific literature. The literature
82 search follows standard practices and
83 includes the PubMed and ToxNet databases of
84 the National Library of Medicine, Web of
85 Science, and other databases listed in the
86 EPA's HERO system (Health and
87 Environmental Research Online,
88 <http://hero.epa.gov/>). Searches for
89 information on mechanisms of toxicity are
90 inherently specialized and may include

1 studies on other agents that act through
2 related mechanisms.

3 Each assessment specifies the search
4 strategies, keywords, and cut-off dates of its
5 literature searches. The EPA posts the results
6 of the literature search on the IRIS web site
7 and requests information from the public on
8 additional studies and ongoing research.

9 The EPA also considers studies received
10 through the IRIS Submission Desk and studies
11 (typically unpublished) submitted under the
12 Toxic Substances Control Act or the Federal
13 Insecticide, Fungicide, and Rodenticide Act.
14 Material submitted as Confidential Business
15 Information is considered only if it includes
16 health and safety data that can be publicly
17 released. If a study that may be critical to the
18 conclusions of the assessment has not been
19 peer-reviewed, the EPA will have it peer-
20 reviewed.

21 The EPA also examines the toxicokinetics
22 of the agent to identify other chemicals (for
23 example, major metabolites of the agent) to
24 include in the assessment if adequate
25 information is available, in order to more
26 fully explain the toxicity of the agent and to
27 suggest dose metrics for subsequent
28 modeling.

29 In assessments of [chemical mixtures](#),
30 mixture studies are preferred for their ability
31 to reflect interactions among components.

32 The literature search seeks, in decreasing
33 order of preference ([U.S. EPA, 2000b, §2.2](#);
34 [1986b, §2.1](#)):

- 35 - Studies of the mixture being assessed.
- 36 - Studies of a sufficiently similar
37 mixture. In evaluating similarity, the
38 assessment considers the alteration of
39 mixtures in the environment through
40 partitioning and transformation.
- 41 - Studies of individual chemical
42 components of the mixture, if there
43 are not adequate studies of
44 sufficiently similar mixtures.

45 **3.2. Selecting pertinent epidemiologic** 46 **studies**

47 Study design is the key consideration for
48 selecting pertinent epidemiologic studies
49 from the results of the literature search.

- 50 - Cohort studies, case-control studies,
51 and some population-based surveys
52 (for example, NHANES) provide the
53 strongest epidemiologic evidence,
54 especially if they collect information
55 about individual exposures and
56 effects.
- 57 - Ecological studies (geographic
58 correlation studies) relate exposures
59 and effects by geographic area. They
60 can provide strong evidence if there
61 are large exposure contrasts between
62 geographic areas, relatively little
63 exposure variation within study
64 areas, and population migration is
65 limited.
- 66 - Case reports of high or accidental
67 exposure lack definition of the
68 population at risk and the expected
69 number of cases. They can provide
70 information about a rare effect or
71 about the relevance of analogous
72 results in animals.

73 The assessment briefly reviews ecological
74 studies and case reports but reports details
75 only if they suggest effects not identified by
76 other studies.

77 **3.3. Selecting pertinent experimental** 78 **studies**

79 Exposure route is a key design
80 consideration for selecting pertinent
81 experimental animal studies or human
82 clinical studies.

- 83 - Studies of oral, inhalation, or dermal
84 exposure involve passage through an
85 absorption barrier and are considered
86 most pertinent to human
87 environmental exposure.
- 88 - Injection or implantation studies are
89 often considered less pertinent but may

1 provide valuable toxicokinetic or
2 mechanistic information. They also may
3 be useful for identifying effects in animals
4 if deposition or absorption is problematic
5 (for example, for particles and fibers).

6 Exposure duration is also a key design
7 consideration for selecting pertinent
8 experimental animal studies.

9 - Studies of effects from chronic
10 exposure are most pertinent to
11 lifetime human exposure.

12 - Studies of effects from less-than-
13 chronic exposure are pertinent but
14 less preferred for identifying effects
15 from lifetime human exposure. Such
16 studies may be indicative of effects
17 from less-than-lifetime human
18 exposure.

19 Short-duration studies involving animals
20 or humans may provide toxicokinetic or
21 mechanistic information.

22 For developmental toxicity and
23 reproductive toxicity, irreversible effects may
24 result from a brief exposure during a critical
25 period of development. Accordingly,
26 specialized study designs are used for these
27 effects ([U.S. EPA, 2006b](#), [1998b](#), [1996](#),
28 [1991b](#)).

4. Evaluating the quality of individual studies

29 After the subsets of pertinent
30 epidemiologic and experimental studies have
31 been selected from the literature searches,
32 the assessment evaluates the quality of each
33 individual study. This evaluation considers
34 the design, methods, conduct, and
35 documentation of each study, but not
36 whether the results are positive, negative, or
37 null. The objective is to identify the stronger,
38 more informative studies based on a uniform
39 evaluation of quality characteristics across
40 studies of similar design.

4.1. Evaluating the quality of epidemiologic studies

41 The assessment evaluates design and
42 methodological aspects that can increase or
43 decrease the weight given to each
44 epidemiologic study in the overall evaluation
45 ([U.S. EPA, 2005a, 1998b, 1996, 1994, 1991b](#)):

46 - Documentation of study design,
47 methods, population characteristics,
48 and results.

49 - Definition and selection of the study
50 group and comparison group.

51 - Ascertainment of exposure to the
52 chemical or mixture.

53 - Ascertainment of disease or health
54 effect.

55 - Duration of exposure and follow-up
56 and adequacy for assessing the
57 occurrence of effects.

58 - Characterization of exposure during
59 critical periods.

60 - Sample size and statistical power to
61 detect anticipated effects.

62 - Participation rates and potential for
63 selection bias as a result of the
64 achieved participation rates.

65 - Measurement error (can lead to
66 misclassification of exposure, health
67 outcomes, and other factors) and
68 other types of information bias.

69 - Potential confounding and other
70 sources of bias addressed in the study
71 design or in the analysis of results.
72 The basis for consideration of
73 confounding is a reasonable
74 expectation that the confounder is
75 related to both exposure and outcome
76 and is sufficiently prevalent to result
77 in bias.

78 For developmental toxicity, reproductive
79 toxicity, neurotoxicity, and cancer there is
80 further guidance on the nuances of evaluating
81 epidemiologic studies of these effects ([U.S.](#)
82 [EPA, 2005a, 1998b, 1996, 1991b](#)).

1 **4.2. Evaluating the quality of**
2 **experimental studies**

3 The assessment evaluates design and
4 methodological aspects that can increase or
5 decrease the weight given to each
6 experimental animal study, in-vitro study, or
7 human clinical study ([U.S. EPA, 2005a, 1998b,](#)
8 [1996, 1991b](#)). Research involving human
9 subjects is considered only if conducted
10 according to ethical principles.

- 11 - Documentation of study design,
12 animals or study population, methods,
13 basic data, and results.
- 14 - Nature of the assay and validity for its
15 intended purpose.
- 16 - Characterization of the nature and
17 extent of impurities and contaminants
18 of the administered chemical or
19 mixture.
- 20 - Characterization of dose and dosing
21 regimen (including age at exposure)
22 and their adequacy to elicit adverse
23 effects, including latent effects.
- 24 - Sample sizes and statistical power to
25 detect dose-related differences or
26 trends.
- 27 - Ascertainment of survival, vital signs,
28 disease or effects, and cause of death.
- 29 - Control of other variables that could
30 influence the occurrence of effects.

31 The assessment uses statistical tests to
32 evaluate whether the observations may be
33 due to chance. The standard for determining
34 statistical significance of a response is a trend
35 test or comparison of outcomes in the
36 exposed groups against those of concurrent
37 controls. In some situations, examination of
38 historical control data from the same
39 laboratory within a few years of the study
40 may improve the analysis. For an uncommon
41 effect that is not statistically significant
42 compared with concurrent controls, historical
43 controls may show that the effect is unlikely
44 to be due to chance. For a response that
45 appears significant against a concurrent

46 control response that is unusual, historical
47 controls may offer a different interpretation
48 ([U.S. EPA, 2005a, §2.2.2.1.3](#)).

49 For developmental toxicity, reproductive
50 toxicity, neurotoxicity, and cancer there is
51 further guidance on the nuances of evaluating
52 experimental studies of these effects ([U.S.](#)
53 [EPA, 2005a, 1998b, 1996, 1991b](#)). In multi-
54 generation studies, agents that produce
55 developmental effects at doses that are not
56 toxic to the maternal animal are of special
57 concern. Effects that occur at doses
58 associated with mild maternal toxicity are not
59 assumed to result only from maternal
60 toxicity. Moreover, maternal effects may be
61 reversible, while effects on the offspring may
62 be permanent ([U.S. EPA, 1998b, §3.1.2.4.5.4;](#)
63 [1991b, §3.1.1.4](#)),.

64 **4.3. Reporting study results**

65 The assessment uses evidence tables to
66 present the design and key results of
67 pertinent studies. There may be separate
68 tables for each site of toxicity or type of study.

69 If a large number of studies observe the
70 same effect, the assessment considers the
71 study quality characteristics in this section to
72 identify the strongest studies or types of
73 study. The tables present details from these
74 studies, and the assessment explains the
75 reasons for not reporting details of other
76 studies or groups of studies that do not add
77 new information. Supplemental information
78 provides references to all studies considered,
79 including those not summarized in the tables.

80 The assessment discusses strengths and
81 limitations that affect the interpretation of
82 each study. If the interpretation of a study in
83 the assessment differs from that of the study
84 authors, the assessment discusses the basis
85 for the difference.

86 As a check on the selection and evaluation
87 of pertinent studies, the EPA asks peer
88 reviewers to identify studies that were not
89 adequately considered.

5. Evaluating the overall evidence of each effect

5.1. Concepts of causal inference

For each health effect, the assessment evaluates the evidence as a whole to determine whether it is reasonable to infer a causal association between exposure to the agent and the occurrence of the effect. This inference is based on information from pertinent human studies, animal studies, and mechanistic studies of adequate quality. Positive, negative, and null results are given weight according to study quality.

Causal inference involves scientific judgment, and the considerations are nuanced and complex. Several health agencies have developed frameworks for causal inference, among them the U.S. Surgeon General (CDC, 2004; HEW, 1964), the International Agency for Research on Cancer (IARC, 2006), the Institute of Medicine (IOM, 2008), and the EPA (2010, §1.6; 2005a, §2.5). Although developed for different purposes, the frameworks are similar in nature and provide an established structure and language for causal inference. Each considers aspects of an association that suggest causation, discussed by Hill (Hill, 1965) and elaborated by Rothman and Greenland (Rothman and Greenland, 1998), and U.S. EPA (2005a, §2.2.1.7; 1994, Appendix C).

Strength of association: The finding of a large relative risk with narrow confidence intervals strongly suggests that an association is not due to chance, bias, or other factors. Modest relative risks, however, may reflect a small range of exposures, an agent of low potency, an increase in an effect that is common, exposure misclassification, or other sources of bias.

Consistency of association: An inference of causation is strengthened if elevated risks are observed in independent studies of different populations and exposure scenarios. Reproducibility of findings constitutes one of the strongest

arguments for causation. Discordant results sometimes reflect differences in study design, exposure, or confounding factors.

Specificity of association: As originally intended, this refers to one cause associated with one effect. Current understanding that many agents cause multiple effects and many effects have multiple causes make this a less informative aspect of causation, unless the effect is rare or unlikely to have multiple causes.

Temporal relationship: A causal interpretation requires that exposure precede development of the effect.

Biologic gradient (exposure-response relationship): Exposure-response relationships strongly suggest causation. A monotonic increase is not the only pattern consistent with causation. The presence of an exposure-response gradient also weighs against bias and confounding as the source of an association.

Biologic plausibility: An inference of causation is strengthened by data demonstrating plausible biologic mechanisms, if available. Plausibility may reflect subjective prior beliefs if there is insufficient understanding of the biologic process involved.

Coherence: An inference of causation is strengthened by supportive results from animal experiments, toxicokinetic studies, and short-term tests. Coherence may also be found in other lines of evidence, such as changing disease patterns in the population.

“Natural experiments”: A change in exposure that brings about a change in disease frequency provides strong evidence, as it tests the hypothesis of causation. An example would be an intervention to reduce exposure in the workplace or environment that is

1 followed by a reduction of an adverse
2 effect.

3 **Analogy:** Information on structural
4 analogues or on chemicals that induce
5 similar mechanistic events can provide
6 insight into causation.

7 These considerations are consistent with
8 guidelines for systematic reviews that
9 evaluate the quality and weight of evidence.
10 Confidence is increased if the magnitude of
11 effect is large, if there is evidence of an
12 exposure-response relationship, or if an
13 association was observed and the plausible
14 biases would tend to decrease the magnitude
15 of the reported effect. Confidence is
16 decreased for study limitations, inconsistency
17 of results, indirectness of evidence,
18 imprecision, or reporting bias ([Guyatt et al.,
19 2008b](#); [Guyatt et al., 2008a](#)).

20 5.2. Evaluating evidence in humans

21 For each effect, the assessment evaluates
22 the evidence from the epidemiologic studies
23 as a whole. The objective is to determine
24 whether a credible association has been
25 observed and, if so, whether that association
26 is consistent with causation. In doing this, the
27 assessment explores alternative explanations
28 (such as chance, bias, and confounding) and
29 draws a conclusion about whether these
30 alternatives can satisfactorily explain any
31 observed association.

32 To make clear how much the
33 epidemiologic evidence contributes to the
34 overall weight of the evidence, the
35 assessment may select a standard descriptor
36 to characterize the epidemiologic evidence of
37 association between exposure to the agent
38 and occurrence of a health effect.

39 **Sufficient epidemiologic evidence of an 40 association consistent with causation:**

41 The evidence establishes a causal
42 association for which alternative
43 explanations such as chance, bias, and
44 confounding can be ruled out with
45 reasonable confidence.

46 **Suggestive epidemiologic evidence of an 47 association consistent with causation:**

48 The evidence suggests a causal
49 association but chance, bias, or
50 confounding cannot be ruled out as
51 explaining the association.

52 **Inadequate epidemiologic evidence to infer 53 a causal association:** The available 54 studies do not permit a conclusion 55 regarding the presence or absence of an 56 association.

57 **Epidemiologic evidence consistent with no
58 causal association:** Several adequate
59 studies covering the full range of human
60 exposures and considering susceptible
61 populations, and for which alternative
62 explanations such as bias and
63 confounding can be ruled out, are
64 mutually consistent in not finding an
65 association.

66 5.3. Evaluating evidence in animals

67 For each effect, the assessment evaluates
68 the evidence from the animal experiments as
69 a whole to determine the extent to which they
70 indicate a potential for effects in humans.
71 Consistent results across various species and
72 strains increase confidence that similar
73 results would occur in humans. Several
74 concepts discussed by Hill ([Hill, 1965](#)) are
75 pertinent to the weight of experimental
76 results: consistency of response, dose-
77 response relationships, strength of response,
78 biologic plausibility, and coherence ([U.S. EPA,
79 2005a, §2.2.1.7](#); [1994, Appendix C](#)).

80 In weighing evidence from multiple
81 experiments, U.S. EPA ([2005a, §2.5](#))
82 distinguishes:

83 **Conflicting evidence** (that is, mixed positive
84 and negative results in the same sex and
85 strain using a similar study protocol)
86 from

87 **Differing results** (that is, positive results and
88 negative results are in different sexes or
89 strains or use different study protocols).

1 Negative or null results do not invalidate
2 positive results in a different experimental
3 system. The EPA regards all as valid
4 observations and looks to explain differing
5 results using mechanistic information (for
6 example, physiologic or metabolic differences
7 across test systems) or methodological
8 differences (for example, relative sensitivity
9 of the tests, differences in dose levels,
10 insufficient sample size, or timing of dosing or
11 data collection).

12 It is well established that there are critical
13 periods for some developmental and
14 reproductive effects ([U.S. EPA, 2006b, 2005a,](#)
15 [b, 1998b, 1996, 1991b](#)). Accordingly, the
16 assessment determines whether critical
17 periods have been adequately investigated.
18 Similarly, the assessment determines
19 whether the database is adequate to evaluate
20 other critical sites and effects.

21 In evaluating evidence of genetic toxicity:

- 22 - Demonstration of gene mutations,
23 chromosome aberrations, or
24 aneuploidy in humans or
25 experimental mammals (*in vivo*)
26 provides the strongest evidence.
- 27 - This is followed by positive results in
28 lower organisms or in cultured cells
29 (*in vitro*) or for other genetic events.
- 30 - Negative results carry less weight,
31 partly because they cannot exclude
32 the possibility of effects in other
33 tissues ([IARC, 2006](#)).

34 For germ-cell mutagenicity, The EPA has
35 defined categories of evidence, ranging from
36 positive results of human germ-cell
37 mutagenicity to negative results for all effects
38 of concern ([U.S. EPA, 1986a, §2.3](#)).

39 **5.4. Evaluating mechanistic data**

40 Mechanistic data can be useful in
41 answering several questions.

- 42 - The biologic plausibility of a causal
43 interpretation of human studies.
- 44 - The generalizability of animal studies
45 to humans.

- 46 - The susceptibility of particular
47 populations or lifestages.

48 The focus of the analysis is to describe, if
49 possible, mechanistic pathways that lead to a
50 health effect. These pathways encompass:

- 51 - *Toxicokinetic processes* of absorption,
52 distribution, metabolism, and
53 elimination that lead to the formation
54 of an active agent and its presence at
55 the site of initial biologic interaction.
- 56 - *Toxicodynamic processes* that lead to a
57 health effect at this or another site
58 (also known as a *mode of action*).

59 For each effect, the assessment discusses
60 the available information on its *modes of*
61 *action* and associated *key events* (*key events*
62 being empirically observable, necessary
63 precursor steps or biologic markers of such
64 steps; *mode of action* being a series of key
65 events involving interaction with cells,
66 operational and anatomic changes, and
67 resulting in disease). Pertinent information
68 may also come from studies of metabolites or
69 of compounds that are structurally similar or
70 that act through similar mechanisms.
71 Information on mode of action is not required
72 for a conclusion that the agent is causally
73 related to an effect ([U.S. EPA, 2005a, §2.5](#)).

74 The assessment addresses several
75 questions about each hypothesized mode of
76 action ([U.S. EPA, 2005a, §2.4.3.4](#)).

77 **1) Is the hypothesized mode of action** 78 **sufficiently supported in test animals?**

79 Strong support for a key event being
80 necessary to a mode of action can come
81 from experimental challenge to the
82 hypothesized mode of action, in which
83 studies that suppress a key event observe
84 suppression of the effect. Support for a
85 mode of action is meaningfully
86 strengthened by consistent results in
87 different experimental models, much
88 more so than by replicate experiments in
89 the same model. The assessment may
90 consider various aspects of causation in
91 addressing this question.

1 2) **Is the hypothesized mode of action**
 2 **relevant to humans?** The assessment
 3 reviews the key events to identify critical
 4 similarities and differences between the
 5 test animals and humans. Site
 6 concordance is not assumed between
 7 animals and humans, though it may hold
 8 for certain effects or modes of action.
 9 Information suggesting quantitative
 10 differences in doses where effects would
 11 occur in animals or humans is considered
 12 in the dose-response analysis. Current
 13 levels of human exposure are not used to
 14 rule out human relevance, as IRIS
 15 assessments may be used in evaluating
 16 new or unforeseen circumstances that
 17 may entail higher exposures.

18 3) **Which populations or lifestyles can be**
 19 **particularly susceptible to the**
 20 **hypothesized mode of action?** The
 21 assessment reviews the key events to
 22 identify populations and lifestyles that
 23 might be susceptible to their occurrence.
 24 Quantitative differences may result in
 25 separate toxicity values for susceptible
 26 populations or lifestyles.

27 The assessment discusses the likelihood
 28 that an agent operates through multiple
 29 modes of action. An uneven level of support
 30 for different modes of action can reflect
 31 disproportionate resources spent
 32 investigating them ([U.S. EPA, 2005a, §2.4.3.3](#)).
 33 It should be noted that in clinical reviews, the
 34 credibility of a series of studies is reduced if
 35 evidence is limited to studies funded by one
 36 interested sector ([Guyatt et al., 2008a](#)).

37 For cancer, the assessment evaluates
 38 evidence of a mutagenic mode of action to
 39 guide extrapolation to lower doses and
 40 consideration of susceptible lifestyles. Key
 41 data include the ability of the agent or a
 42 metabolite to react with or bind to DNA,
 43 positive results in multiple test systems, or
 44 similar properties and structure-activity
 45 relationships to mutagenic carcinogens ([U.S.](#)
 46 [EPA, 2005a, §2.3.5](#)).

47 **5.5. Characterizing the overall weight**
 48 **of the evidence**

49 After evaluating the human, animal, and
 50 mechanistic evidence pertinent to an effect,
 51 the assessment answers the question: Does
 52 the agent cause the adverse effect? ([NRC,](#)
 53 [2009, 1983](#)). In doing this, the assessment
 54 develops a narrative that integrates the
 55 evidence pertinent to causation. To provide
 56 clarity and consistency, the narrative includes
 57 a standard hazard descriptor. For example,
 58 the following standard descriptors combine
 59 epidemiologic, experimental, and mechanistic
 60 evidence of carcinogenicity ([U.S. EPA, 2005a,](#)
 61 [§2.5](#)).

62 ***Carcinogenic to humans:*** There is
 63 convincing epidemiologic evidence of a
 64 causal association (that is, there is
 65 reasonable confidence that the
 66 association cannot be fully explained by
 67 chance, bias, or confounding); or there is
 68 strong human evidence of cancer or its
 69 precursors, extensive animal evidence,
 70 identification of key precursor events in
 71 animals, and strong evidence that they
 72 are anticipated to occur in humans.

73 ***Likely to be carcinogenic to humans:*** The
 74 evidence demonstrates a potential hazard
 75 to humans but does not meet the criteria
 76 for *carcinogenic*. There may be a plausible
 77 association in humans, multiple positive
 78 results in animals, or a combination of
 79 human, animal, or other experimental
 80 evidence.

81 ***Suggestive evidence of carcinogenic***
 82 ***potential:*** The evidence raises concern
 83 for effects in humans but is not sufficient
 84 for a stronger conclusion. This descriptor
 85 covers a range of evidence, from a
 86 positive result in the only available study
 87 to a single positive result in an extensive
 88 database that includes negative results in
 89 other species.

90 ***Inadequate information to assess***
 91 ***carcinogenic potential:*** No other
 92 descriptors apply. *Conflicting evidence* can
 93 be classified as *inadequate information* if

1 all positive results are opposed by
2 negative studies of equal quality in the
3 same sex and strain. *Differing results,*
4 however, can be classified as *suggestive*
5 *evidence* or as *likely to be carcinogenic*.

6 ***Not likely to be carcinogenic to humans:***

7 There is robust evidence for concluding
8 that there is no basis for concern. There
9 may be no effects in both sexes of at least
10 two appropriate animal species; positive
11 animal results and strong, consistent
12 evidence that each mode of action in
13 animals does not operate in humans; or
14 convincing evidence that effects are not
15 likely by a particular exposure route or
16 below a defined dose.

17 Multiple descriptors may be used if there
18 is evidence that carcinogenic effects differ by
19 dose range or exposure route ([U.S. EPA,](#)
20 [2005a, §2.5](#)).

21 Another example of standard descriptors
22 comes from the EPA's Integrated Science
23 Assessments, which evaluate causation for
24 the effects of the criteria pollutants in
25 ambient air ([U.S. EPA, 2010, §1.6](#)).

26 ***Causal relationship:*** Sufficient evidence to
27 conclude that there is a causal
28 relationship. Observational studies
29 cannot be explained by plausible
30 alternatives, or they are supported by
31 other lines of evidence, for example,
32 animal studies or mechanistic
33 information.

34 ***Likely to be a causal relationship:*** Sufficient
35 evidence that a causal relationship is
36 likely, but important uncertainties
37 remain. For example, observational
38 studies show an association but co-
39 exposures are difficult to address or other
40 lines of evidence are limited or
41 inconsistent; or multiple animal studies
42 from different laboratories demonstrate
43 effects and there are limited or no human
44 data.

45 ***Suggestive of a causal relationship:*** At least
46 one high-quality epidemiologic study

47 shows an association but other studies
48 are inconsistent.

49 ***Inadequate to infer a causal relationship:***

50 The studies do not permit a conclusion
51 regarding the presence or absence of an
52 association.

53 ***Not likely to be a causal relationship:***

54 Several adequate studies, covering the full
55 range of human exposure and considering
56 susceptible populations, are mutually
57 consistent in not showing an effect at any
58 level of exposure.

59 The EPA is investigating and may on a
60 trial basis use these or other standard
61 descriptors to characterize the overall weight
62 of the evidence for effects other than cancer.

6. Selecting studies for derivation of toxicity values

63 For each effect where there is credible
64 evidence of an association with the agent, the
65 assessment derives toxicity values if there are
66 suitable epidemiologic or experimental data.
67 The decision to derive toxicity values may be
68 linked to the hazard descriptor.

69 Dose-response analysis requires
70 quantitative measures of dose and response.
71 Then, other factors being equal:

- 72 - Epidemiologic studies are preferred
73 over animal studies, if quantitative
74 measures of exposure are available
75 and effects can be attributed to the
76 agent.
- 77 - Among experimental animal models,
78 those that respond most like humans
79 are preferred, if the comparability of
80 response can be determined.
- 81 - Studies by a route of human
82 environmental exposure are
83 preferred, although a validated
84 toxicokinetic model can be used to
85 extrapolate across exposure routes.
- 86 - Studies of longer exposure duration
87 and follow-up are preferred, to
88 minimize uncertainty about whether

1 effects are representative of lifetime
2 exposure.

3 - Studies with multiple exposure levels
4 are preferred for their ability to
5 provide information about the shape
6 of the exposure-response curve.

7 - Studies with adequate power to
8 detect effects at lower exposure levels
9 are preferred, to minimize the extent
10 of extrapolation to levels found in the
11 environment.

12 Studies with non-monotonic exposure-
13 response relationships are not necessarily
14 excluded from the analysis. A diminished
15 effect at higher exposure levels may be
16 satisfactorily explained by factors such as
17 competing toxicity, saturation of absorption
18 or metabolism, exposure misclassification, or
19 selection bias.

20 If a large number of studies are suitable
21 for dose-response analysis, the assessment
22 considers the study characteristics in this
23 section to focus on the most informative data.
24 The assessment explains the reasons for not
25 analyzing other groups of studies. As a check
26 on the selection of studies for dose-response
27 analysis, the EPA asks peer reviewers to
28 identify studies that were not adequately
29 considered.

7. Deriving toxicity values

30 7.1. General framework for dose- 31 response analysis

32 The EPA uses a two-step approach that
33 distinguishes analysis of the observed dose-
34 response data from inferences about lower
35 doses ([U.S. EPA, 2005a, §3](#)).

36 Within the observed range, the preferred
37 approach is to use modeling to incorporate a
38 wide range of data into the analysis. The
39 modeling yields a *point of departure* (an
40 exposure level near the lower end of the
41 observed range, without significant
42 extrapolation to lower doses) (Sections 7.2-
43 7.3).

44 Extrapolation to lower doses considers
45 what is known about the modes of action for
46 each effect (Sections 7.4-7.5). If response
47 estimates at lower doses are not required, an
48 alternative is to derive *reference values*, which
49 are calculated by applying factors to the point
50 of departure in order to account for sources
51 of uncertainty and variability (Section 7.6).

52 For a group of agents that induce an effect
53 through a common mode of action, the dose-
54 response analysis may derive a *relative*
55 *potency factor* for each agent. A full dose-
56 response analysis is conducted for one well-
57 studied *index chemical* in the group, then the
58 potencies of other members are expressed in
59 relative terms based on relative toxic effects,
60 relative absorption or metabolic rates,
61 quantitative structure-activity relationships,
62 or receptor binding characteristics ([U.S. EPA,](#)
63 [2005a, §3.2.6](#); [2000b, §4.4](#)).

64 Increasingly, the EPA is basing toxicity
65 values on combined analyses of multiple data
66 sets or multiple responses. The EPA also
67 considers multiple dose-response approaches
68 if they can be supported by robust data.

69 7.2. Modeling dose to sites of biologic 70 effects

71 The preferred approach for analysis of
72 dose is toxicokinetic modeling because of its
73 ability to incorporate a wide range of data.
74 The preferred dose metric would refer to the
75 active agent at the site of its biologic effect or
76 to a close, reliable surrogate measure. The
77 active agent may be the administered
78 chemical or a metabolite. Confidence in the
79 use of a toxicokinetic model depends on the
80 robustness of its validation process and on
81 the results of sensitivity analyses ([U.S. EPA,](#)
82 [2006a](#); [2005a, §3.1](#); [1994, §4.3](#)).

83 Because toxicokinetic modeling can
84 require many parameters and more data than
85 are typically available, the EPA has developed
86 standard approaches that can be applied to
87 typical data sets. These standard approaches
88 also facilitate comparison across exposure
89 patterns and species.

90 - Intermittent study exposures are
91 standardized to a daily average over

1 the duration of exposure. For chronic
2 effects, daily exposures are averaged
3 over the lifespan. Exposures during a
4 critical period, however, are not
5 averaged over a longer duration ([U.S.
6 EPA, 2005a, §3.1.1; 1991b, §3.2](#)).

7 - Doses are standardized to equivalent
8 human terms to facilitate comparison
9 of results from different species.

10 - Oral doses are scaled allometrically
11 using mg/kg^{3/4}-day as the equivalent
12 dose metric across species. Allometric
13 scaling pertains to equivalence across
14 species, not across lifestages, and is
15 not used to scale doses from adult
16 humans or mature animals to infants
17 or children ([U.S. EPA, 2011;
18 2005a, §3.1.3](#)).

19 - Inhalation exposures are scaled using
20 dosimetry models that apply species-
21 specific physiologic and anatomic
22 factors and consider whether the
23 effect occurs at the site of first contact
24 or after systemic circulation ([U.S. EPA,
25 2012a; 1994, §3](#)).

26 It can be informative to convert doses
27 across exposure routes. If this is done, the
28 assessment describes the underlying data,
29 algorithms, and assumptions ([U.S. EPA,
30 2005a, §3.1.4](#)).

31 In the absence of study-specific data on,
32 for example, intake rates or body weight, the
33 EPA has developed recommended values for
34 use in dose-response analysis ([U.S. EPA,
35 1988](#)).

36 **7.3. Modeling response in the range of** 37 **observation**

38 Toxicodynamic (“biologically based”)
39 modeling can incorporate data on biologic
40 processes leading to an effect. Such models
41 require sufficient data to ascertain a mode of
42 action and to quantitatively support model
43 parameters associated with its key events.
44 Because different models may provide
45 equivalent fits to the observed data but
46 diverge substantially at lower doses, critical

47 biologic parameters should be measured
48 from laboratory studies, not by model fitting.
49 Confidence in the use of a toxicodynamic
50 model depends on the robustness of its
51 validation process and on the results of
52 sensitivity analyses. Peer review of the
53 scientific basis and performance of a model is
54 essential ([U.S. EPA, 2005a, §3.2.2](#)).

55 Because toxicodynamic modeling can
56 require many parameters and more
57 knowledge and data than are typically
58 available, the EPA has developed a standard
59 set of empirical (“curve-fitting”) models
60 (<http://www.epa.gov/ncea/bmds/>) that can
61 be applied to typical data sets, including those
62 that are nonlinear. The EPA has also
63 developed guidance on modeling dose-
64 response data, assessing model fit, selecting
65 suitable models, and reporting modeling
66 results ([U.S. EPA, 2012b](#)). Additional
67 judgment or alternative analyses are used if
68 the procedure fails to yield reliable results,
69 for example, if the fit is poor, modeling may
70 be restricted to the lower doses, especially if
71 there is competing toxicity at higher doses
72 ([U.S. EPA, 2005a, §3.2.3](#)).

73 Modeling is used to derive a point of
74 departure ([U.S. EPA, 2012b; 2005a, §3.2.4](#)).
75 (See Section 7.6 for alternatives if a point of
76 departure cannot be derived by modeling.):

77 - If linear extrapolation is used,
78 selection of a response level
79 corresponding to the point of
80 departure is not highly influential, so
81 standard values near the low end of
82 the observable range are generally
83 used (for example, 10% extra risk for
84 cancer bioassay data, 1% for
85 epidemiologic data, lower for rare
86 cancers).

87 - For nonlinear approaches, both
88 statistical and biologic considerations
89 are taken into account.

90 - For dichotomous data, a response
91 level of 10% extra risk is generally
92 used for minimally adverse effects,
93 5% or lower for more severe effects.

1 - For continuous data, a response level
2 is ideally based on an established
3 definition of biologic significance. In
4 the absence of such definition, one
5 control standard deviation from the
6 control mean is often used for
7 minimally adverse effects, one-half
8 standard deviation for more severe
9 effects.

10 The point of departure is the 95% lower
11 bound on the dose associated with the
12 selected response level.

13 **7.4. Extrapolating to lower doses and** 14 **response levels**

15 The purpose of extrapolating to lower
16 doses is to estimate responses at exposures
17 below the observed data. Low-dose
18 extrapolation, typically used for cancer data,
19 considers what is known about modes of
20 action ([U.S. EPA, 2005a, §3.3.1 and §3.3.2](#)).

21 1) If a biologically based model has been
22 developed and validated for the agent,
23 extrapolation may use the fitted model
24 below the observed range if significant
25 model uncertainty can be ruled out with
26 reasonable confidence.

27 2) Linear extrapolation is used if the dose-
28 response curve is expected to have a
29 linear component below the point of
30 departure. This includes:

- 31 - Agents or their metabolites that are
32 DNA-reactive and have direct
33 mutagenic activity.
- 34 - Agents or their metabolites for which
35 human exposures or body burdens
36 are near doses associated with key
37 events leading to an effect.

38 Linear extrapolation is also used when
39 data are insufficient to establish mode of
40 action and when scientifically plausible.

41 The result of linear extrapolation is
42 described by an oral slope factor or an
43 inhalation unit risk, which is the slope of
44 the dose-response curve at lower doses
45 or concentrations, respectively.

46 3) Nonlinear models are used for
47 extrapolation if there are sufficient data
48 to ascertain the mode of action and to
49 conclude that it is not linear at lower
50 doses, and the agent does not
51 demonstrate mutagenic or other activity
52 consistent with linearity at lower doses.
53 Nonlinear approaches generally should
54 not be used in cases where mode of action
55 has not ascertained. If nonlinear
56 extrapolation is appropriate but no model
57 is developed, an alternative is to calculate
58 reference values.

59 4) Both linear and nonlinear approaches
60 may be used if there a multiple modes of
61 action. For example, modeling to a low
62 response level can be useful for
63 estimating the response at doses where a
64 high-dose mode of action would be less
65 important.

66 If linear extrapolation is used, the
67 assessment develops a candidate slope factor
68 or unit risk for each suitable data set. These
69 results are arrayed, using common dose
70 metrics, to show the distribution of relative
71 potency across various effects and
72 experimental systems. The assessment then
73 derives or selects an overall slope factor and
74 an overall unit risk for the agent, considering
75 the various dose-response analyses, the study
76 preferences discussed in Section 6, and the
77 possibility of basing a more robust result on
78 multiple data sets.

79 **7.5. Considering susceptible** 80 **populations and lifestyles**

81 The assessment analyzes the available
82 information on populations and lifestyles
83 that may be particularly susceptible to each
84 effect. A tiered approach is used ([U.S. EPA,](#)
85 [2005a, §3.5](#)).

86 1) If an epidemiologic or experimental study
87 reports quantitative results for a
88 susceptible population or lifestyle, these
89 data are analyzed to derive separate
90 toxicity values for susceptible individuals.

1 2) If data on risk-related parameters allow
2 comparison of the general population and
3 susceptible individuals, these data are
4 used to adjust the general-population
5 toxicity values for application to
6 susceptible individuals.

7 3) In the absence of chemical-specific data,
8 the EPA has developed *age-dependent*
9 *adjustment factors* for early-life exposure
10 to potential carcinogens that have a
11 mutagenic mode of action. There is
12 evidence of early-life susceptibility to
13 various carcinogenic agents, but most
14 epidemiologic studies and cancer
15 bioassays do not include early-life
16 exposure. To address the potential for
17 early-life susceptibility, the EPA
18 recommends ([U.S. EPA, 2005b, §5](#)):

- 19 - 10-fold adjustment for exposures
20 before age 2 years.
- 21 - 3-fold adjustment for exposures
22 between ages 2 and 16 years.

23 7.6. Reference values and uncertainty 24 factors

25 An *oral reference dose* or an *inhalation*
26 *reference concentration* is an estimate of an
27 exposure (including in susceptible
28 subgroups) that is likely to be without an
29 appreciable risk of adverse health effects over
30 a lifetime ([U.S. EPA, 2002, §4.2](#)). Reference
31 values are typically calculated for effects
32 other than cancer and for suspected
33 carcinogens if a well characterized mode of
34 action indicates that a necessary key event
35 does not occur below a specific dose.
36 Reference values provide no information
37 about risks at higher exposure levels.

38 The assessment characterizes effects that
39 form the basis for reference values as
40 adverse, considered to be adverse, or a
41 precursor to an adverse effect. For
42 developmental toxicity, reproductive toxicity,
43 and neurotoxicity there is guidance on
44 adverse effects and their biologic markers
45 ([U.S. EPA, 1998b, 1996, 1991b](#)).

46 To account for uncertainty and variability
47 in the derivation of a lifetime human

48 exposure where adverse effects are not
49 anticipated to occur, reference values are
50 calculated by applying a series of *uncertainty*
51 *factors* to the point of departure. If a point of
52 departure cannot be derived by modeling, a
53 no-observed-adverse-effect level or a lowest-
54 observed-adverse-effect level is used instead.
55 The assessment discusses scientific
56 considerations involving several areas of
57 variability or uncertainty.

58 **Human variation.** The assessment accounts
59 for variation in susceptibility across the
60 human population and the possibility that
61 the available data may not be
62 representative of individuals who are
63 most susceptible to the effect. A factor of
64 10 is generally used to account for this
65 variation. This factor is reduced only if
66 the point of departure is derived or
67 adjusted specifically for susceptible
68 individuals (not for a general population
69 that includes both susceptible and non-
70 susceptible individuals) ([U.S. EPA,](#)
71 [2002, §4.4.5](#); [1998b, §4.2](#); [1996, §4](#);
72 [1994, §4.3.9.1](#); [1991b, §3.4](#)).

73 **Animal-to-human extrapolation.** If animal
74 results are used to make inferences about
75 humans, the assessment adjusts for cross-
76 species differences. These may arise from
77 differences in toxicokinetics or
78 toxicodynamics. Accordingly, if the point
79 of departure is standardized to equivalent
80 human terms or is based on toxicokinetic
81 or dosimetry modeling, a factor of 10^{1/2}
82 (rounded to 3) is applied to account for
83 the remaining uncertainty involving
84 toxicokinetic and toxicodynamic
85 differences. If a biologically based model
86 adjusts fully for toxicokinetic and
87 toxicodynamic differences across species,
88 this factor is not used. In most other
89 cases, a factor of 10 is applied ([U.S. EPA,](#)
90 [2011; 2002, §4.4.5](#); [1998b, §4.2](#); [1996, §4](#);
91 [1994, §4.3.9.1](#); [1991b, §3.4](#)).

92 **Adverse-effect level to no-observed-**
93 **adverse-effect level.** If a point of
94 departure is based on a lowest-observed-
95 adverse-effect level, the assessment must

1 infer a dose where such effects are not
2 expected. This can be a matter of great
3 uncertainty, especially if there is no
4 evidence available at lower doses. A
5 factor of 10 is applied to account for the
6 uncertainty in making this inference. A
7 factor other than 10 may be used,
8 depending on the magnitude and nature
9 of the response and the shape of the dose-
10 response curve ([U.S. EPA, 2002, §4.4.5](#);
11 [1998b, §4.2](#); [1996, §4](#); [1994, §4.3.9.1](#);
12 [1991b, §3.4](#)).

13 **Subchronic-to-chronic exposure.** If a point
14 of departure is based on subchronic
15 studies, the assessment considers
16 whether lifetime exposure could have
17 effects at lower levels of exposure. A
18 factor of 10 is applied to account for the
19 uncertainty in using subchronic studies to
20 make inferences about lifetime exposure.
21 This factor may also be applied for
22 developmental or reproductive effects if
23 exposure covered less than the full critical
24 period. A factor other than 10 may be
25 used, depending on the duration of the
26 studies and the nature of the response
27 ([U.S. EPA, 2002, §4.4.5](#); [1998b, §4.2](#); [1994,](#)
28 [§4.3.9.1](#)).

29 **Incomplete database.** If an incomplete
30 database raises concern that further
31 studies might identify a more sensitive
32 effect, organ system, or lifestage, the
33 assessment may apply a database
34 uncertainty factor ([U.S. EPA, 2002, §4.4.5](#);
35 [1998b, §4.2](#); [1996, §4](#); [1994, §4.3.9.1](#);
36 [1991b, §3.4](#)). The size of the factor
37 depends on the nature of the database
38 deficiency. For example, the EPA typically
39 follows the suggestion that a factor of 10
40 be applied if both a prenatal toxicity study
41 and a two-generation reproduction study
42 are missing and a factor of 10^{1/2} if either
43 is missing ([U.S. EPA, 2002, §4.4.5](#)).

44 In this way, the assessment derives
45 candidate values for each suitable data set
46 and effect that is credibly associated with the
47 agent. These results are arrayed, using
48 common dose metrics, to show where effects

49 occur across a range of exposures ([U.S. EPA,](#)
50 [1994, §4.3.9](#)).

51 The assessment derives or selects an
52 *organ- or system-specific reference value* for
53 each organ or system affected by the agent.
54 The assessment explains the rationale for
55 each organ/system-specific reference value
56 (based on, for example, the highest quality
57 studies, the most sensitive outcome, or a
58 clustering of values). By providing these
59 organ/system-specific reference values, IRIS
60 assessments facilitate subsequent cumulative
61 risk assessments that consider the combined
62 effect of multiple agents acting at a common
63 site or through common mechanisms ([NRC,](#)
64 [2009](#)).

65 The assessment then selects an overall
66 reference dose and an overall reference
67 concentration for the agent to represent
68 lifetime human exposure levels where effects
69 are not anticipated to occur. This is generally
70 the most sensitive organ/system-specific
71 reference value, though consideration of
72 study quality and confidence in each value
73 may lead to a different selection.

74 **7.7. Confidence and uncertainty in the** 75 **reference values**

76 The assessment selects a standard
77 descriptor to characterize the level of
78 confidence in each reference value, based on
79 the likelihood that the value would change
80 with further testing. Confidence in reference
81 values is based on quality of the studies used
82 and completeness of the database, with more
83 weight given to the latter. The level of
84 confidence is increased for reference values
85 based on human data supported by animal
86 data ([U.S. EPA, 1994, §4.3.9.2](#)).

87 **High confidence:** The reference value is not
88 likely to change with further testing,
89 except for mechanistic studies that might
90 affect the interpretation of prior test
91 results.

92 **Medium confidence:** This is a matter of
93 judgment, between high and low
94 confidence.

1 **Low confidence:** The reference value is
2 especially vulnerable to change with
3 further testing.

4 These criteria are consistent with
5 guidelines for systematic reviews that
6 evaluate the quality of evidence. These also
7 focus on whether further research would be
8 likely to change confidence in the estimate of
9 effect ([Guyatt et al., 2008b](#)).

10 All assessments discuss the significant
11 uncertainties encountered in the analysis.
12 The EPA provides guidance on
13 characterization of uncertainty ([U.S. EPA,](#)

14 [2005a, §3.6](#)). For example, the discussion
15 distinguishes model uncertainty (lack of
16 knowledge about the most appropriate
17 experimental or analytic model) and
18 parameter uncertainty (lack of knowledge
19 about the parameters of a model).
20 Assessments also discuss human variation
21 (interpersonal differences in biologic
22 susceptibility or in exposures that modify the
23 effects of the agent).

24
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26 August 2013
27

28

29 EXECUTIVE SUMMARY

30

Occurrence and Health Effects

31 *tert*-Butanol does not occur naturally, but it is produced by humans for
32 multiple purposes, such as a solvent for paints, a denaturant for ethanol and several
33 other alcohols, an octane booster in gasoline, a dehydrating agent, and the
34 manufacture of flotation agents, fruit essences, and perfumes. *tert*-Butanol is also a
35 primary metabolite of methyl *tert*-butyl ether (MTBE) and ethyl *tert*-butyl ether
36 (ETBE). Exposure to *tert*-butanol primarily occurs through breathing air containing
37 *tert*-butanol vapors, as well as consuming contaminated water (or breast milk) or
38 foods. Exposure may also occur through direct skin contact.

39 Animal studies demonstrate that chronic oral exposure to *tert*-butanol is
40 associated with kidney and thyroid effects. Developmental effects (e.g., reduced fetal
41 viability) have been observed in short-term exposure to high levels of *tert*-butanol
42 (via oral or inhalation exposure) in animals. No chronic inhalation exposure studies
43 have been conducted. There is suggestive evidence that *tert*-butanol is carcinogenic
44 to humans based on predominantly benign renal tumors in male rats and benign
45 thyroid tumors in female mice.
46

47 **Effects Other Than Cancer Observed Following Oral Exposure**

48 EPA identified kidney effects as a hazard of *tert*-butanol exposure, with kidney toxicity
49 observed after oral exposure in two strains of rats, one strain of mice, and in both sexes. In mice,
50 the only kidney effect observed was an increase in kidney weight (absolute and/or relative) in both
51 sexes of mice in the 13-week study, but no treatment-related histopathological lesions were
52 reported in the kidneys of mice at 13 weeks or 2 years. Absolute and relative kidney weights were
53 increased in both male and female rats after 13 weeks and 15 months of treatment.
54 Histopathological examination also indicated kidney toxicity in both male and female rats, with
55 increased incidence of nephropathy after 13 weeks of oral exposure and transitional epithelium

1 hyperplasia observed after 2 years of oral exposure. Additionally, increased suppurative
 2 inflammation was noted in females after 2 years of oral exposure. Mode of action analysis
 3 determined that male rat kidney effects were not mediated by α_{2u} -globulin, and these effects are
 4 concluded to be relevant for human health hazard assessment.

5 **Oral Reference Dose (RfD) for Effects Other Than Cancer**

6 Kidney toxicity, represented by kidney transitional epithelial hyperplasia, was chosen as the
 7 basis for the proposed overall oral reference dose (RfD) (see Table ES-1), as it was the only
 8 noncancer endpoint for which there is credible evidence of an association with *tert*-butanol
 9 exposure. The chronic study by [NTP \(1995\)](#) and the observed kidney effects were used to derive
 10 the RfD. The endpoint of transitional epithelial hyperplasia was selected as the critical effect due to
 11 its consistency in both sexes, its specificity and its sensitivity as an indicator of kidney toxicity, and
 12 the observed dose-response relationship of effects across dose groups. Benchmark dose (BMD)
 13 modeling was utilized to derive the BMDL_{10%} of 16 mg/kg-day. The BMDL was converted to a
 14 human equivalent dose using body weight^{3/4} scaling, and this value of 3.84 mg/kg-day was used as
 15 the point of departure (POD) for RfD derivation ([U.S. EPA, 2011](#)).

16 The proposed overall RfD was calculated by dividing the POD for kidney transitional
 17 epithelial hyperplasia by a composite uncertainty factor (UF) of 30 to account for the extrapolation
 18 from animals to humans (3) and for interindividual differences in human susceptibility (10).

19 **Table ES-1. Summary of reference dose (RfD) derivation**

Effect	Basis	RfD (mg/kg-day)	Exposure description	Confidence
Kidney toxicity	Increased incidence of kidney transitional epithelial hyperplasia	1×10^{-1}	Chronic	HIGH
Proposed overall RfD	Increased incidence of kidney transitional epithelial hyperplasia	1×10^{-1}	Chronic	HIGH

20

21 **Effects Other Than Cancer Observed Following Inhalation Exposure**

22 EPA identified kidney effects as a human hazard of *tert*-butanol exposure. Both absolute
 23 and relative kidney weights were increased in male and female rats. There was an increase in
 24 nephropathy severity in male rats, which supported the increase in kidney weights. No available
 25 human studies evaluated the effects of inhalation exposure. Mode of action analysis determined
 26 that male rat kidney effects were not mediated by α_{2u} -globulin, and these effects are concluded to
 27 be relevant for human health hazard assessment.

1 **Inhalation Reference Concentration (RfC) for Effects Other Than Cancer**

2 Kidney toxicity, represented by transitional epithelial hyperplasia, was chosen as the basis
 3 for the proposed inhalation reference concentration (RfC) (see Table ES-2), as it was the only
 4 noncancer endpoint for which there is credible evidence of an association with *tert*-butanol
 5 exposure. The chronic oral exposure study in rats ([NTP, 1995](#)) was used to derive the overall RfC.
 6 A PBPK model for *tert*-butanol in rats was developed internally, and route-to-route extrapolation
 7 was used to derive equivalent inhalation PODs. The POD adjusted for the human equivalent
 8 concentration (HEC) was 26.1 mg/m³ and based on transitional epithelial hyperplasia.

9 The RfC was calculated by dividing the POD by a composite UF of 30 to account for
 10 toxicodynamic differences between animals and humans (3) and interindividual differences in
 11 human susceptibility (10).

12 **Table ES-2. Summary of reference concentration (RfC) derivation**

Effect	Basis	RfC (mg/m ³)	Exposure description	Confidence
Kidney toxicity	Increased incidence of kidney transitional epithelial hyperplasia	9 × 10 ⁻¹	Chronic	HIGH
Proposed overall RfC	Increased incidence of kidney transitional epithelial hyperplasia	9 × 10⁻¹	Chronic	HIGH

13
 14 **Evidence for Human Carcinogenicity**

15 Under EPA's *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)), the database for
 16 *tert*-butanol provides "suggestive evidence of carcinogenic potential." Human data are not available
 17 to assess the carcinogenic potential of *tert*-butanol. In 2-year studies in F344 rats and B6C3F₁ mice,
 18 male rats exhibited dose-related increases in renal tubule adenoma and combined renal tubule
 19 adenoma or carcinoma. Although data support α_{2u}-globulin deposition in the kidney of male rats,
 20 there is insufficient evidence to support this as the only or primary mechanism for renal tumor
 21 development in male rats. Therefore, the renal tumors are considered relevant to humans.
 22 However, the observed renal tumors were predominantly benign, only occurred in a single
 23 sex/species combination, and were not observed in studies that exposed the same strain of rat to
 24 ETBE, which is rapidly metabolized to *tert*-butanol. In addition, a statistically significant increase in
 25 the incidence of thyroid follicular cell adenoma was observed in a 2-year drinking water study in
 26 female mice ([NTP, 1995](#)). These tumors were all benign and only a single sex/species combination
 27 was affected. There are no studies examining the carcinogenic potential of *tert*-butanol after
 28 inhalation exposure in animals. However, internal tumors developed after oral exposure and may
 29 occur regardless of exposure route, as blood concentrations were found to be similar after oral or

1 inhalation exposures. Genotoxicity data for *tert*-butanol are inconclusive. *tert*-Butanol was negative
2 in a variety of genotoxicity assays in different cell systems including gene mutations, sister
3 chromatid exchanges, micronucleus formation and chromosomal aberrations. However, DNA
4 adducts in male Kunming mice and DNA damage in human HL-60 leukemia cells have been
5 observed. Overall, the cancer descriptor “suggestive evidence of carcinogenic potential” is
6 plausible, as some concern is raised by the positive evidence of predominantly benign renal tumors
7 in male rats and benign thyroid tumors in female mice.

8 **Quantitative Estimate of Carcinogenic Risk from Oral Exposure**

9 Lifetime oral exposure to *tert*-butanol has been associated with increased renal tubule
10 adenomas and carcinoma in male F344 rats, increased thyroid follicular cell adenomas in female
11 B6C3F₁ mice, and increased thyroid follicular cell adenomas and carcinomas in male B6C3F₁ mice.
12 The [NTP \(1995\)](#) study in rats and mice was the only available study for dose-response analysis. The
13 study included histological examinations for tumors in many different tissues, contained three
14 exposure levels and controls, contained adequate numbers of animals per dose group
15 (~50/sex/group), treated animals for up to 2 years, and included detailed reporting of methods
16 and results.

17 Although *tert*-butanol was considered to have “suggestive evidence of carcinogenic
18 potential,” EPA concluded that the main study was well-conducted and quantitative analysis may be
19 useful for providing a sense of the magnitude of potential carcinogenic risk. For renal tumors, two
20 slope factors were derived for this endpoint from the [NTP \(1995\)](#) bioassay: one based on the
21 original reported incidences and one based on the [Hard et al. \(2011\)](#) reanalysis. The two estimates
22 differed by less than 20%, and rounded to the same number at one significant figure. However, the
23 [Hard et al. \(2011\)](#) reanalysis is considered preferable, as it is based on a Pathology Working Group
24 (PWG) analysis. A slope factor was also derived for thyroid tumors in female mice. The modeled
25 *tert*-butanol PODs were scaled to HEDs according to EPA guidance by converting the BMDL₁₀ on the
26 basis of (body weight)^{3/4} scaling ([U.S. EPA, 2011, 2005a](#)). Using linear extrapolation from the
27 BMDL₁₀, a human equivalent oral slope factor was derived (slope factor = 0.1/BMDL₁₀). The more
28 sensitive endpoint of renal tumors was used because there is no data to support neither renal nor
29 thyroid tumors most relevant to humans. The oral slope factor of **1 × 10⁻² per mg/kg-day**, based
30 on the renal tubule tumor response in male F344 rats.

31 **Quantitative Estimate of Carcinogenic Risk from Inhalation Exposure**

32 No chronic inhalation exposure studies to *tert*-butanol are available. However, through the
33 oral route of exposure, lifetime exposure has been associated with increased renal tubule adenomas
34 and carcinoma in male F344 rats, increased thyroid follicular cell adenomas in female B6C3F₁ mice,
35 and increased thyroid follicular cell adenomas and carcinomas in male B6C3F₁ mice. The [NTP](#)
36 [\(1995\)](#) study in rats and mice was the only available study for dose-response analysis. The study
37 included histological examinations for tumors in many different tissues, contained three exposure

1 levels and controls, contained adequate numbers of animals per dose group (~50/sex/group),
2 treated animals for up to 2 years, and included detailed reporting of methods and results.

3 Although *tert*-butanol was considered to have “suggestive evidence of carcinogenic
4 potential,” EPA concluded that the main study was well-conducted and quantitative analysis may be
5 useful for providing a sense of the magnitude of potential carcinogenic risk. Since the available
6 evidence for *tert*-butanol carcinogenicity is from a 2 year oral exposure, route-to-route
7 extrapolation of the oral BMDL was performed to derive an inhalation equivalent BMCL. The BMCL
8 was then converted to a human equivalent concentration (HEC) according to the RfC guidelines
9 ([U.S. EPA, 1994](#)) by multiplying the BMCL by the blood:gas partition coefficient ratio. Using linear
10 extrapolation from the resulting BMCL_{10-HEC}, a human equivalent inhalation unit risk was derived
11 (inhalation unit risk = 0.1/BMCL_{10-HEC}). Extrapolation from the oral study results for renal tubule
12 adenoma or carcinoma in male F44 rats gives a unit risk of 2×10^{-3} per mg/m³, associated with
13 lifetime inhalation exposure to *tert*-butanol.

14 **Susceptible Populations and Lifestages for Cancer and Noncancer**

15 No data were identified to indicate susceptible populations or lifestages.

16 **Key Issues Addressed in Assessment**

17 Due to the observation of kidney tumors and noncancer toxicity following chronic exposure
18 to *tert*-butanol, an evaluation of whether *tert*-butanol caused α_{2u} -globulin nephropathy was
19 performed. The presence of α_{2u} -globulin in the hyaline droplets was confirmed in male rats by α_{2u}
20 immunohistochemical staining. Linear mineralization and tubular hyperplasia were reported in
21 male rats, though only in the chronic study. Other subsequent steps in the pathological sequence,
22 including necrosis, exfoliation, and granular casts, were either absent or not consistently observed
23 across subchronic or chronic studies. None of the observed effects occurred in female rats or in
24 either sex of mice. Because the available data supports the occurrence of at least two of the
25 subsequent steps in the pathological sequence, these data are sufficient to conclude that
26 α_{2u} -globulin nephropathy is occurring in the kidney of male rats following *tert*-butanol exposure.
27 Thus, the noncancer lesions associated with α_{2u} -globulin nephropathy are not considered relevant
28 to humans.

29 However, tumors develop at doses lower than some precursors of α_{2u} -globulin
30 nephropathy, such as granular casts and tubular hyperplasia ([Hard et al., 2011](#); [NTP, 1995](#)).
31 Therefore, there is insufficient evidence to support a conclusion that α_{2u} -globulin nephropathy is
32 the sole or primary contributor to renal tumor development. Because carcinogenic processes other
33 than α_{2u} -globulin nephropathy cannot be ruled out, the renal tumors are considered relevant to
34 humans.

35 In addition, some of the observed renal lesions in rats following exposure to *tert*-butanol
36 are effects commonly associated with chronic progressive nephropathy (CPN), an age-related renal
37 disease of laboratory rodents that occurs spontaneously. While it has been argued that CPN in rats

1 is not relevant to humans, it is acknowledged that the mechanism regulating CPN in rats is not
2 understood. Moreover, no key events for the exacerbation of CPN have been identified, so no mode
3 of action analysis can be performed. Therefore, kidney effects from *tert*-butanol exposure
4 associated with CPN are considered relevant to humans.

5 Sufficient data were available to develop a PBPK model in rats for both oral and inhalation
6 exposure in order to perform route-to-route extrapolation, so rat studies from both routes of
7 exposure were considered for dose-response analysis. The only endpoint available from the
8 subchronic inhalation study ([NTP, 1997](#)) was increased kidney weights, which is a less-specific
9 endpoint compared to other endpoints available for analysis from the oral study ([NTP, 1995](#)). In
10 regards to the carcinogenic effects, the 2-year oral study ([NTP, 1995](#)) was the only study to evaluate
11 lifetime carcinogenic effects and was selected for route-to-route extrapolation.

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LITERATURE SEARCH STRATEGY | STUDY SELECTION AND EVALUATION

A literature search and screening strategy were used to identify literature characterizing the health effects of *tert*-butanol. This strategy consisted of a broad search of online scientific databases and other sources in order to identify all potentially pertinent studies. In subsequent steps, references were screened to exclude papers not pertinent to an assessment of the health effects of *tert*-butanol, and remaining references were sorted into categories for further evaluation. This section describes the literature search and screening strategy in detail.

The chemical-specific search was conducted in four online scientific databases, including PubMed, Toxline, Web of Science, and TSCATS through April 2014, using the keywords and limits described in Table LS-1. The overall literature search approach is shown graphically in Figure LS-1. An additional 7 citations were obtained using additional search strategies described in Table LS-2. After electronically eliminating duplicates from the citations retrieved through these databases, 2,532 unique citations were identified.

The resulting 2,532 citations were screened into categories as presented in Figure LS-1 using the title, abstract, and/or full text for pertinence to examine the health effects of *tert*-butanol exposure.

- 12 references were identified as potential “Sources of Health Effects Data” and were considered for data extraction to evidence tables and exposure-response arrays.
- 196 references were identified as “Supporting Studies;” these included 39 studies describing physiologically-based pharmacokinetic (PBPK) models and other toxicokinetic information, 70 studies providing genotoxicity and other mechanistic information, 1 human case report, 73 not relevant exposure paradigms (including acute, dermal, eye irritation, and injection studies), 6 preliminary toxicity studies, and 7 physical dependency studies. While still considered sources of health effects information, studies investigating the effects of acute and direct chemical exposures are generally less pertinent for characterizing health hazards associated with chronic oral and inhalation exposure. Therefore, information from these studies was not considered for extraction into evidence tables. Nevertheless, these studies were still evaluated as possible sources of supporting health effects information.
- 63 references were identified as secondary sources of health effects information (e.g., reviews and other agency assessments); these references were kept as additional resources for development of the Toxicological Review.
- 2,261 references were identified as not being pertinent to an evaluation of the health effects of *tert*-butanol and were excluded from further consideration (see Figure LS-1 for exclusion categories).

The complete list of references and the sorting of these materials can be found on the HERO website at <http://hero.epa.gov>.

1 **Selection of Critical Studies for Inclusion in Evidence Tables**

2 Each study retained after the literature search and screen was evaluated for aspects of its
3 design or conduct that could affect the interpretation of results and the overall contribution to the
4 evidence for determination of hazard potential. Some general questions that were considered in
5 evaluating experimental animal studies are presented in Table LS-3. Much of the key information
6 for conducting this evaluation can generally be found in the study's methods section and in how the
7 study results are reported. Importantly, the evaluation at this stage does not consider the direction
8 or magnitude of any reported effects.

9 To facilitate this evaluation, evidence tables were constructed that systematically
10 summarize the important information from each study in a standardized tabular format as
11 recommended by the [NRC \(2011\)](#). Twelve studies identified as "Sources of Health Effects Data"
12 were considered for extraction into evidence tables for hazard identification in Chapter 1. Initial
13 review of studies found two studies to be publications of the [NTP \(1995\)](#) data prior to the release of
14 the final NTP report ([Cirvello et al., 1995](#); [Lindamood et al., 1992](#)). One publication in the
15 "Supporting Studies" category also was based on data from the NTP report ([Takahashi et al., 1993](#)).
16 There were differences between the published reports and the final NTP report; therefore, the
17 finalized [NTP \(1995\)](#) report was included in evidence tables. Data from the remaining 10 studies in
18 the "Sources of Health Effects Data" category were extracted into evidence tables.

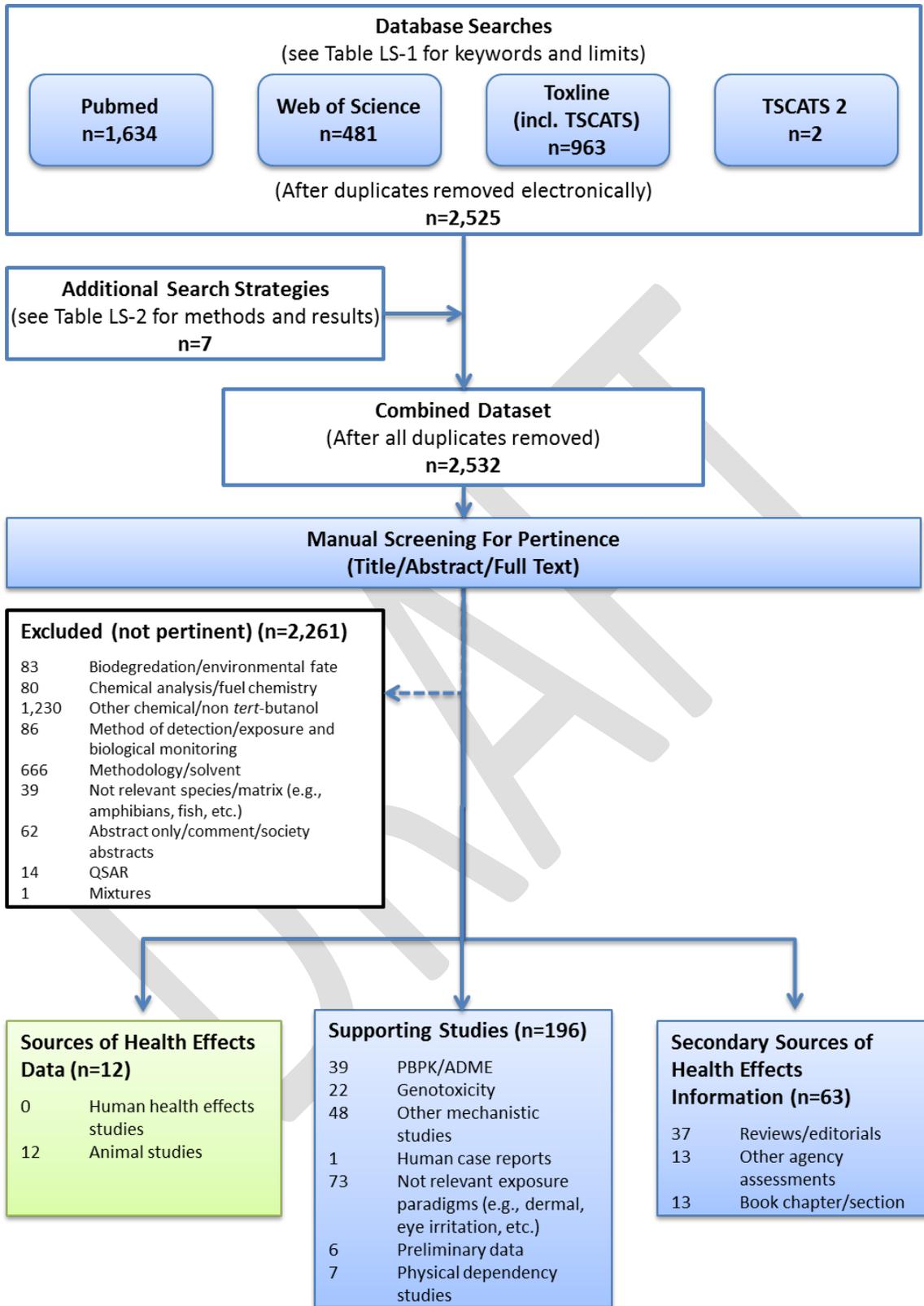
19 Supporting studies that contain pertinent information for the toxicological review and
20 augment hazard identification conclusions, such as genotoxic and mechanistic studies, studies
21 describing the kinetics and disposition of *tert*-butanol absorption and metabolism, pilot studies,
22 short term or acute studies, were not included in the evidence tables. Such supporting studies may
23 be discussed in the narrative sections of Chapter 1, or presented in Appendices, if they provide
24 additional or corroborating information.

25 **Database Evaluation**

26 The database for *tert*-butanol is comprised of animal toxicity studies containing one 2-year
27 bioassay that employs oral exposures in rats and mice; two oral subchronic studies in rats and one
28 in mice; one inhalation subchronic study in rats and mice; a re-evaluation of the [NTP \(1995\)](#) rat
29 data; two oral developmental studies; two inhalation developmental studies; and one one-
30 generation reproductive study that also evaluates other systemic effects. Several acute and short
31 term studies (including an 18-day inhalation study and a 14-day study by NTP) using oral and
32 inhalation exposures were performed mostly in rats, but were grouped as supporting studies since
33 the database of chronic and subchronic rat studies was considered sufficient. No cohort studies,
34 case-control studies, or ecological studies exist in the published literature. There was one case
35 report available. Health effect studies of gasoline and *tert*-butanol mixtures were not considered
36 pertinent to the assessment since the separate effects of the gasoline components could not be
37 determined; thus, these studies were excluded during the manual screen.

1 The “Sources of Health Effects Data” were comprised entirely of studies performed in rats
2 and mice with drinking water, oral gavage, and inhalation exposures to *tert*-butanol. These 12
3 sources were conducted according to OECD Good Laboratory Practice (GLP) guidelines, presented
4 extensive histopathological data, and/or clearly presented their methodology; thus, these are
5 considered high quality. Preliminary, acute, and short-term studies contained information that
6 supported and did not differ qualitatively from the results of the ≥ 30 day exposure studies; thus,
7 these studies are not included in the evidence tables. Some of these shorter duration studies are
8 presented in the text of the Toxicological Review and are used in sections such as “Mechanistic
9 Evidence” to augment the discussion. A more detailed discussion of methodological concerns that
10 were identified will precede each endpoint evaluated in the hazard identification section.

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Figure LS-1. Study selection strategy.

1 **Table LS-1. Details of the search strategy employed for *tert*-butanol**

Database (Search Date)	Keywords	Limits
PubMed (12/20/2012) (4/17/2014)	<i>tert</i> -butanol OR 75-65-0[<i>rn</i>] OR " <i>t</i> -butyl hydroxide" OR "2-methyl-2-propanol" OR "trimethyl carbinol" OR " <i>t</i> -butyl alcohol" OR <i>tert</i> -butanol OR " <i>tert</i> -butyl alcohol" OR <i>tert</i> -butyl alcohol[mesh]	None
Web of Science (12/20/2012) (4/17/2014)	Topic = (<i>tert</i> -butanol OR 75-65-0 OR " <i>t</i> -butyl hydroxide" OR "2-methyl-2-propanol" OR "trimethyl carbinol" OR " <i>t</i> -butyl alcohol" OR " <i>tert</i> -butanol" OR " <i>tert</i> -butyl alcohol")	Refined by: Research Areas = (cell biology OR respiratory system OR microscopy OR biochemistry molecular biology OR gastroenterology hepatology OR public environmental occupational health OR oncology OR physiology OR cardiovascular system cardiology or toxicology OR life sciences biomedicine other topics OR hematology OR pathology OR neurosciences neurology OR developmental biology)
Toxline (includes TSCATS) (1/11/2013) (4/17/2014)	<i>tert</i> -butanol OR 75-65-0 [<i>rn</i>] OR <i>t</i> -butyl hydroxide OR 2-methyl-2-propanol OR trimethyl carbinol OR <i>t</i> -butyl alcohol OR <i>tert</i> -butanol OR <i>tert</i> -butyl alcohol OR <i>tert</i> -butyl alcohol	Not PubMed
TSCATS2 (1/4/2013) (4/17/2014)	75-65-0	None

2

1 **Table LS-2. Summary of additional search strategies for *tert*-butanol**

Approach used	Source(s)	Date performed	Number of additional references identified
Manual search of citations from reviews	Review article: Mcgregor (2010) . Tertiary-butanol: A toxicological review. Crit Rev Toxicol 40(8): 697-727.	1/2013	5
	Review article: Chen (2005) . Amended final report of the safety assessment of t-butyl alcohol as used in cosmetics." Int J Toxicol 24(2): 1-20.	1/2013	2
Manual search of citations from reviews conducted by other international and federal agencies	IPCS (1987a) . Butanols: Four isomers: 1-butanol, 2-butanol, <i>tert</i> -butanol, isobutanol [WHO EHC]. Geneva, Switzerland: World Health Organization.	1/2013	None
	OSHA (1992) . Occupational safety and health guideline for <i>tert</i> -butyl alcohol. Cincinnati, OH: National Institute for Occupational Safety and Health.	1/2013	None

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1 **Table LS-3. Questions and relevant experimental information for evaluation**
 2 **of experimental animal studies**

Methodological feature	Question(s) considered	Examples of relevant information extracted
Test animal	Based on the endpoint(s) in question, are concerns raised regarding the suitability of the species, strain, or sex of the test animals on study?	Test animal species, strain, sex
Experimental setup	Are the timing, frequency and duration of exposure, as well as animal age and experimental group allocation procedures/ group size for each endpoint evaluation, appropriate for the assessed endpoint(s)?	Age/lifestage of test animals at exposure and all endpoint testing timepoints Timing and periodicity of exposure and endpoint evaluations; duration of exposure Sample size for each experimental group (e.g., animals; litters; dams) at each endpoint evaluation
Exposure	Are the exposure conditions and controls informative and reliable for the endpoint(s) in question, and are they sufficiently specific to the compound of interest?	Exposure administration techniques (e.g., route; chamber type)
Endpoint evaluation procedures	Do the procedures used to evaluate the endpoint(s) in question conform to established protocols, or are they biologically sound? Are they sensitive for examination of the outcome(s) of interest?	Specific methods for assessing the effect(s) of exposure, including related details (e.g., specific region of tissue/organ evaluated) Endpoint evaluation controls, including those put in place to minimize evaluator bias
Outcomes and data reporting	Were data reported for all pre-specified endpoint(s) and study groups, or were any data excluded from presentation/ analyses?	Data presentation for endpoint(s) of interest

Note: “Outcome” refers to findings from an evaluation (e.g., hypertrophy), whereas “endpoint” refers to the evaluation itself (e.g., liver histopathology).

3
4

1. HAZARD IDENTIFICATION

1.1. PRESENTATION AND SYNTHESIS OF EVIDENCE BY ORGAN/SYSTEM

1.1.1. Kidney Effects

Synthesis of Effects in Kidney

This section reviews the studies that investigated whether exposure to *tert*-butanol can cause kidney effects in humans or animals. The database examining kidney effects following *tert*-butanol exposure contains no human data, six studies performed in rats or mice, and one re-evaluation of the rat data from [NTP \(1995\)](#). Studies employing short-term and acute exposures that examined kidney effects are not included in the evidence tables; however, they are discussed in the text if they provide data to support mode of action or hazard identification. No methodological concerns were identified that would lead one or more studies to be considered less informative for assessing human health hazard. A pathology working group ([Hard et al., 2011](#)) re-examined kidney histopathology from the [NTP \(1995\)](#) 13-week and 2-year studies in rats to evaluate questions involving MOAs for renal tubule development. All slides were analyzed in a blinded manner. [Hard et al. \(2011\)](#) did report different incidences of adenomas or carcinomas compared with the original [NTP \(1995\)](#) study; thus, these data were presented separately. Histopathological results from both Hard and NTP will be considered for hazard identification.

tert-Butanol exposure resulted in a number of kidney effects after both oral (drinking water) and inhalation exposure in both sexes of rats and mice. Kidney effects observed after oral exposure (Table 1-1; Table 1-2; Figure 1-1) include increased kidney weight in female rats and in male and female mice (13-week exposure), and kidney inflammation, kidney transitional epithelial hyperplasia, and increased incidence and/or severity of kidney nephropathy in female rats (2-year exposure) ([NTP, 1995](#)). In a 2-year oral exposure study in male rats, increased kidney weight, increased hyaline droplets, kidney transitional epithelial hyperplasia, kidney mineralization, renal tubule hyperplasia, and increased incidence and/or severity of kidney nephropathy were observed, with some of these effects seen at earlier time periods ([NTP, 1995](#)). Other kidney effects in male rats were observed in a 10-week oral exposure study ([Acharya et al., 1997](#); [Acharya et al., 1995](#)). No changes in clinical chemistry that would typically be indicative of kidney damage have been observed with *tert*-butanol exposure. Although there were some changes in urinalysis parameters (e.g., decreased urine volume and increased specific gravity), this was accompanied by reduced water consumption and may not be related to an effect of kidney function.

The kidney is also the target organ for cancer effects (Table 1-3; Figure 1-1). Male F344 rats had an increased incidence of renal tubule adenomas and combined renal tubule adenoma or

1 carcinoma in a 2-year oral bioassay ([Hard et al., 2011](#); [NTP, 1995](#)). The highest exposure group had
2 an increase in mortality, which may in part explain the apparent non-monotonicity in the observed
3 dose-response, in which the highest exposure group had a lower incidence of tumors than the
4 middle exposure group.

5 An Independent Pathology Working Group (PWG), sponsored by Lyondell Chemical
6 Company, re-evaluated the kidney changes in the NTP 2-year study ([Hard et al., 2011](#)). The PWG
7 consisted of senior pathologists with experience in chemically-induced nephrotoxicity and renal
8 neoplasia. In all cases, PWG members were blinded to treatment groups to preclude any possible
9 bias, and used guidelines published by the Society of Toxicologic Pathology. The PWG confirmed the
10 NTP findings of atypical tubule hyperplasia and renal tubule tumors in male rats at 2-years. In
11 particular, they reported very similar overall tumor incidences in the exposed groups. However,
12 the PWG evaluation of the control groups reported fewer renal tubule adenomas and carcinomas
13 than the original NTP study. As a result, based on the PWG evaluation, all treated groups had
14 statistically significant increases in renal tubule adenomas and carcinomas (combined) as
15 compared to controls. Additionally, the PWG considered fewer of the tumors to be carcinomas as
16 compared to the original NTP study.

17 No chronic (2-year) inhalation exposure study is available, but minimal kidney effects were
18 observed in rats (mainly the males) after *tert*-butanol exposure by inhalation for 13 weeks at
19 concentrations ranging from 406–6,368 mg/m³ ([NTP, 1997](#)) (Table 1-1; Table 1-2; Figure 1-2).
20 Absolute kidney weights were elevated (9.8–11%) in male rats exposed at ≥3,274 mg/m³ (not dose-
21 dependent); relative kidney weights were statistically elevated (~9%) in males at ≥3,274 mg/m³
22 and females at 6,368 mg/m³. Male rats exhibited an increase in the severity of chronic nephropathy
23 (characterized as number of foci of regenerative tubules). Although the kidney effects were less
24 severe after inhalation exposure, a direct comparison can only be made on the basis of internal
25 dose. [ARCO \(1983\)](#) found that blood levels of *tert*-butanol and its metabolites are equivalent after a
26 single oral dose of 350 mg/kg compared to a single 6-hour inhalation exposure to 6,164 mg/m³.
27 That would indicate, based on bolus exposures, that the inhalation exposures used in the [NTP](#)
28 [\(1997\)](#) study were in the range of the lower doses used in the [NTP \(1995\)](#) oral study. On the other
29 hand, based on PBPK modeling, chronic exposure in the range of the [NTP \(1995\)](#) bioassay doses of
30 90–420 mg/kg-day lead to the same average blood concentration of *tert*-butanol as 6-hour/day, 5
31 day/week inhalation exposures to 860–4500 mg/m³, suggesting that the oral and inhalation
32 exposures in [NTP \(1995\)](#) and [NTP \(1997\)](#), respectively, overlap on the basis of internal dose.
33 Finally, the lack of either mortality or changes in body weight (both observed with oral exposure)
34 observed after the inhalation exposure suggests that a direct comparison cannot be made.
35

1
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Table 1-1. Changes in kidney weight in animals following exposure to tert-butanol

Reference and study design	Results					
<i>Kidney weight (percent change as compared to control)</i>						
Lyondell Chemical Co. (2004) Sprague-Dawley rat; 12/sex/treatment Gavage 0, 64, 160, 400, or 1,000 mg/kg-d Males: 9 weeks beginning 4 weeks prior to mating Females: 4 weeks prior to mating through PND21	Males					
	<u>Dose</u> (mg/kg-d)	<u>Left absolute</u> <u>weight</u>	<u>Left relative</u> <u>weight</u>	<u>Right absolute</u> <u>weight</u>	<u>Right relative</u> <u>weight</u>	
	0	0	0	0	0	
	64	+6	+8	+6	+8	
	160	+9	+14*	+6	+11*	
	400	+12*	+14*	+14*	+17*	
	1,000	+18*	+28*	+20*	+31*	
	Females					
	<u>Dose</u> (mg/kg-d)	<u>Left absolute</u> <u>weight</u>	<u>Left relative</u> <u>weight</u>	<u>Right absolute</u> <u>weight</u>	<u>Right relative</u> <u>weight</u>	
	0	0	0	0	0	
	64	-1	-2	+2	0	
	160	0	0	+1	0	
	400	+3	+2	+4	+2	
	1,000	+4	0	+7	+2	
NTP (1995) F344/N rat; 10/sex/treatment Drinking water 0, 2.5, 5, 10, 20, 40 mg/mL M: 0, 230, 490, 840, 1,520, 3,610 ^a mg/kg-d F: 0, 290, 590, 850, 1,560, 3,620 ^a mg/kg-d 13 weeks	Males			Females		
	<u>Dose</u> (mg/kg-d)	<u>Absolute</u> <u>weight</u>	<u>Relative</u> <u>weight</u>	<u>Dose</u> (mg/kg-d)	<u>Absolute</u> <u>weight</u>	<u>Relative</u> <u>weight</u>
	0	0	0	0	0	0
	230	+12*	+19*	290	+19*	+17*
	490	+17*	+26*	590	+16*	+15*
	840	+16*	+32*	850	+29*	+28*
	1,520	+26*	+54*	1,560	+39*	+40*
	3,610	All dead	All dead	3,620	+36*	+81*

3

Table 1-1. Changes in kidney weight in animals following exposure to tert-butanol (continued)

Reference and study design	Results					
	Males			Females		
NTP (1995) B6C3F ₁ mouse; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, 40 mg/mL) M: 0, 350, 640, 1,590, 3,940, 8,210 ^a mg/kg-d F: 0, 500, 820, 1,660, 6,430, 11,620 ^a mg/kg-d 13 weeks	<u>Dose</u> (mg/kg-d)	<u>Absolute weight</u>	<u>Relative weight</u>	<u>Dose</u> (mg/kg-d)	<u>Absolute weight</u>	<u>Relative weight</u>
	0	0	0	0	0	0
	350	+1	+1	500	0	-3
	640	+3	+2	820	-3	-1
	1,590	+2	+8	1,660	+1	0
	3,940	+6	+22*	6,430	+6	+15*
	8,210	0	+48*	11,620	+12*	+35*
NTP (1995) F344/N rat; 60/sex/treatment (10/sex/treatment evaluated at 15 months) Drinking water (0, 1.25, 2.5, 5, or 10 mg/mL) M: 0, 90, 200, or 420 ^a mg/kg-d F: 0, 180, 330, or 650 ^a mg/kg-d 2 years	<u>Dose</u> (mg/kg-d)	<u>Absolute weight</u>	<u>Relative weight</u>	<u>Dose</u> (mg/kg-d)	<u>Absolute weight</u>	<u>Relative weight</u>
	0	0	0	0	0	0
	90	+4	+8	180	+8*	+14*
	200	+11	+15*	330	+18*	+21*
	420	+7	+20*	650	+22*	+42*
	Only animals sacrificed at 15 months were evaluated for organ weights. Organs were not weighed in the 2-year mouse study					
NTP (1997) F344/N rat; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m ³) (dynamic whole-body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported	<u>Concentration</u> (mg/m ³)	<u>Absolute weight</u>	<u>Relative weight</u>	<u>Absolute weight</u>	<u>Relative weight</u>	
	0	0	0	0	0	
	406	+1	+1	-4	-1	
	824	-2	-1	0	+1	
	1,643	+3	+2	+4	+4	
	3,273	+11*	+8*	+2	+2	
	6,368	+9.8*	+9*	+4	+9*	

Table 1-1. Changes in kidney weight in animals following exposure to tert-butanol (continued)

Reference and study design	Results				
	Males			Females	
	Concentration (mg/m ³)	Absolute weight	Relative weight	Absolute weight	Relative weight
NTP (1997) B6C3F ₁ mouse; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m ³) (dynamic whole-body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported	0	0	0	0	0
	406	-6	-4	+1	-3
	824	-1	+3	+5	+9
	1,643	+4	+3	+1	-2
	3,273	-10	-3	0	+7
	6,368	+3	+6	+3	+15*

1 ^a The high-dose group had an increase in mortality.
 2 * Statistically significant $p \leq 0.05$ as determined by the study authors.
 3 Percentage change compared to control = (treated value – control value) ÷ control value × 100.
 4 Conversions from drinking water concentrations to mg/kg-d performed by study authors.
 5 Conversion from ppm to mg/m³ is 1 ppm = 3.031 mg/m³.
 6

Table 1-2. Changes in kidney histopathology in animals following exposure to tert-butanol

Reference and study design	Results
Acharya et al. (1997; 1995) Wistar rat; 5–6 males/treatment Drinking water (0 or 0.5%), 0 or 575 mg/kg-d 10 weeks	↑ tubular degeneration, degeneration of the basement membrane of the Bowman’s capsule, diffused glomeruli, and glomerular vacuolation (no incidences reported) ↓ kidney glutathione (~40%)*
Lyondell Chemical Co. (2004) Sprague-Dawley rat; 12/sex/treatment Gavage 0, 64, 160, 400, or 1,000 mg/kg-d F0 males: 9 weeks beginning 4 weeks prior to mating F0 females: 4 weeks prior to mating through PND21	There were no changes in kidney histopathology observed.

9

Table 1-2. Changes in kidney histopathology in animals following exposure to tert-butanol (continued)

Reference and study design	Results																																																					
<p>NTP (1995)</p> <p>F344/N rat; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 230, 490, 840, 1,520, 3,610^a mg/kg-d F: 0, 290, 590, 850, 1,560, 3,620^a mg/kg-d 13 weeks</p>	<p>Incidence (severity):</p> <table border="1"> <thead> <tr> <th colspan="3" data-bbox="560 415 1003 441">Males</th> <th colspan="3" data-bbox="1003 415 1408 441">Females</th> </tr> <tr> <th data-bbox="560 470 667 527"><u>Dose</u> <u>(mg/kg-d)</u></th> <th data-bbox="667 470 829 527"><u>Mineralization</u></th> <th data-bbox="829 470 1003 527"><u>Nephropathy</u></th> <th data-bbox="1003 470 1110 527"><u>Dose</u> <u>(mg/kg-d)</u></th> <th data-bbox="1110 470 1273 527"><u>Mineralization</u></th> <th data-bbox="1273 470 1408 527"><u>Nephropathy</u></th> </tr> </thead> <tbody> <tr> <td data-bbox="560 527 667 583">0</td> <td data-bbox="667 527 829 583">0/10</td> <td data-bbox="829 527 1003 583">7/10 (1.0)</td> <td data-bbox="1003 527 1110 583">0</td> <td data-bbox="1110 527 1273 583">10/10 (1.7)</td> <td data-bbox="1273 527 1408 583">2/10 (1.0)</td> </tr> <tr> <td data-bbox="560 583 667 640">230</td> <td data-bbox="667 583 829 640">0/10</td> <td data-bbox="829 583 1003 640">10/10 (1.6*)</td> <td data-bbox="1003 583 1110 640">290</td> <td data-bbox="1110 583 1273 640">10/10 (2.0)</td> <td data-bbox="1273 583 1408 640">3/10 (1.0)</td> </tr> <tr> <td data-bbox="560 640 667 697">490</td> <td data-bbox="667 640 829 697">2/10 (1.5)</td> <td data-bbox="829 640 1003 697">10/10 (2.6*)</td> <td data-bbox="1003 640 1110 697">590</td> <td data-bbox="1110 640 1273 697">10/10 (2.0)</td> <td data-bbox="1273 640 1408 697">5/10 (1.0)</td> </tr> <tr> <td data-bbox="560 697 667 753">840</td> <td data-bbox="667 697 829 753">8/10*(1.4)</td> <td data-bbox="829 697 1003 753">10/10 (2.7*)</td> <td data-bbox="1003 697 1110 753">850</td> <td data-bbox="1110 697 1273 753">10/10 (2.0)</td> <td data-bbox="1273 697 1408 753">7/10* (1.0)</td> </tr> <tr> <td data-bbox="560 753 667 810">1,520</td> <td data-bbox="667 753 829 810">4/10*(1.0)</td> <td data-bbox="829 753 1003 810">10/10 (2.6*)</td> <td data-bbox="1003 753 1110 810">1,560</td> <td data-bbox="1110 753 1273 810">10/10 (2.0)</td> <td data-bbox="1273 753 1408 810">8/10* (1.0)</td> </tr> <tr> <td data-bbox="560 810 667 867">3,610</td> <td data-bbox="667 810 829 867">4/10*(1.0)</td> <td data-bbox="829 810 1003 867">7/10 (1.1)</td> <td data-bbox="1003 810 1110 867">3,620</td> <td data-bbox="1110 810 1273 867">6/10 (1.2)</td> <td data-bbox="1273 810 1408 867">7/10* (1.0)</td> </tr> </tbody> </table>						Males			Females			<u>Dose</u> <u>(mg/kg-d)</u>	<u>Mineralization</u>	<u>Nephropathy</u>	<u>Dose</u> <u>(mg/kg-d)</u>	<u>Mineralization</u>	<u>Nephropathy</u>	0	0/10	7/10 (1.0)	0	10/10 (1.7)	2/10 (1.0)	230	0/10	10/10 (1.6*)	290	10/10 (2.0)	3/10 (1.0)	490	2/10 (1.5)	10/10 (2.6*)	590	10/10 (2.0)	5/10 (1.0)	840	8/10*(1.4)	10/10 (2.7*)	850	10/10 (2.0)	7/10* (1.0)	1,520	4/10*(1.0)	10/10 (2.6*)	1,560	10/10 (2.0)	8/10* (1.0)	3,610	4/10*(1.0)	7/10 (1.1)	3,620	6/10 (1.2)	7/10* (1.0)
Males			Females																																																			
<u>Dose</u> <u>(mg/kg-d)</u>	<u>Mineralization</u>	<u>Nephropathy</u>	<u>Dose</u> <u>(mg/kg-d)</u>	<u>Mineralization</u>	<u>Nephropathy</u>																																																	
0	0/10	7/10 (1.0)	0	10/10 (1.7)	2/10 (1.0)																																																	
230	0/10	10/10 (1.6*)	290	10/10 (2.0)	3/10 (1.0)																																																	
490	2/10 (1.5)	10/10 (2.6*)	590	10/10 (2.0)	5/10 (1.0)																																																	
840	8/10*(1.4)	10/10 (2.7*)	850	10/10 (2.0)	7/10* (1.0)																																																	
1,520	4/10*(1.0)	10/10 (2.6*)	1,560	10/10 (2.0)	8/10* (1.0)																																																	
3,610	4/10*(1.0)	7/10 (1.1)	3,620	6/10 (1.2)	7/10* (1.0)																																																	
<p>NTP (1995)</p> <p>B6C3F₁ mouse; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 350, 640, 1,590, 3,940, 8,210^a mg/kg-d F: 0, 500, 820, 1,660, 6,430, 11,620^a mg/kg-d 13 weeks</p>	<p>Histopathology data for the 13-week study were not provided, but the kidney was evaluated indicating that no changes in kidney histopathology were observed in the 13-week study.</p>																																																					

Table 1-2. Changes in kidney histopathology in animals following exposure to tert-butanol (continued)

Reference and study design	Results			
NTP (1995) F344/N rat; 60/sex/treatment (10/sex/treatment evaluated at 15 months) Drinking water (0, 1.25, 2.5, 5, 10 mg/mL) M: 0, 90, 200, 420 ^a mg/kg-d F: 0, 180, 330, 650 ^a mg/kg-d 2 years	Incidence (severity):			
	Males			
	<u>Dose (mg/kg-d)</u>	<u>Mineralization (interim)</u>	<u>Mineralization (terminal)</u>	<u>Linear mineralization (terminal)</u>
	0	1/10 (1.0)	26/50 (1.0)	0/50
	90	2/10 (1.0)	28/50 (1.1)	5/50* (1.0)
	200	5/10 (1.8)	35/50 (1.3)	24/50* (1.2)
	420	9/10* (2.3)	48/50* (2.2)	46/50* (1.7)
	<u>Dose (mg/kg-d)</u>	<u>Renal tubule hyperplasia (extended evaluation)</u>	<u>Transitional epithelium hyperplasia</u>	<u>Nephropathy severity</u>
	0	12/50 (2.3)	25/50 (1.7)	3.0
	90	16/50 (2.3)	32/50 (1.7)	3.1
	200	14/50 (2.2)	36/50* (2.0)	3.1
	420	23/50* (2.8)	40/50* (2.1)	3.3*
	Females			
	<u>Dose (mg/kg-d)</u>	<u>Mineralization^b Interim</u>	<u>Mineralization^b Terminal</u>	<u>Inflammation (suppurative) incidence</u>
	0	10/10 (2.8)	49/50 (2.6)	2/50
	180	10/10 (2.9)	50/50 (2.6)	3/50
330	10/10 (2.9)	50/50 (2.7)	13/50*	
650	10/10 (2.8)	50/50 (2.9)	17/50*	
<u>Dose (mg/kg-d)</u>	<u>Renal tubule hyperplasia</u>	<u>Transitional epithelium hyperplasia</u>	<u>Nephropathy severity</u>	
0	0/50	0/50	1.6	
180	0/50	0/50	1.9*	
330	0/50	3/50 (1.0)	2.3*	
650	1/50 (1.0)	17/50*(1.4)	2.9*	

Table 1-2. Changes in kidney histopathology in animals following exposure to tert-butanol (continued)

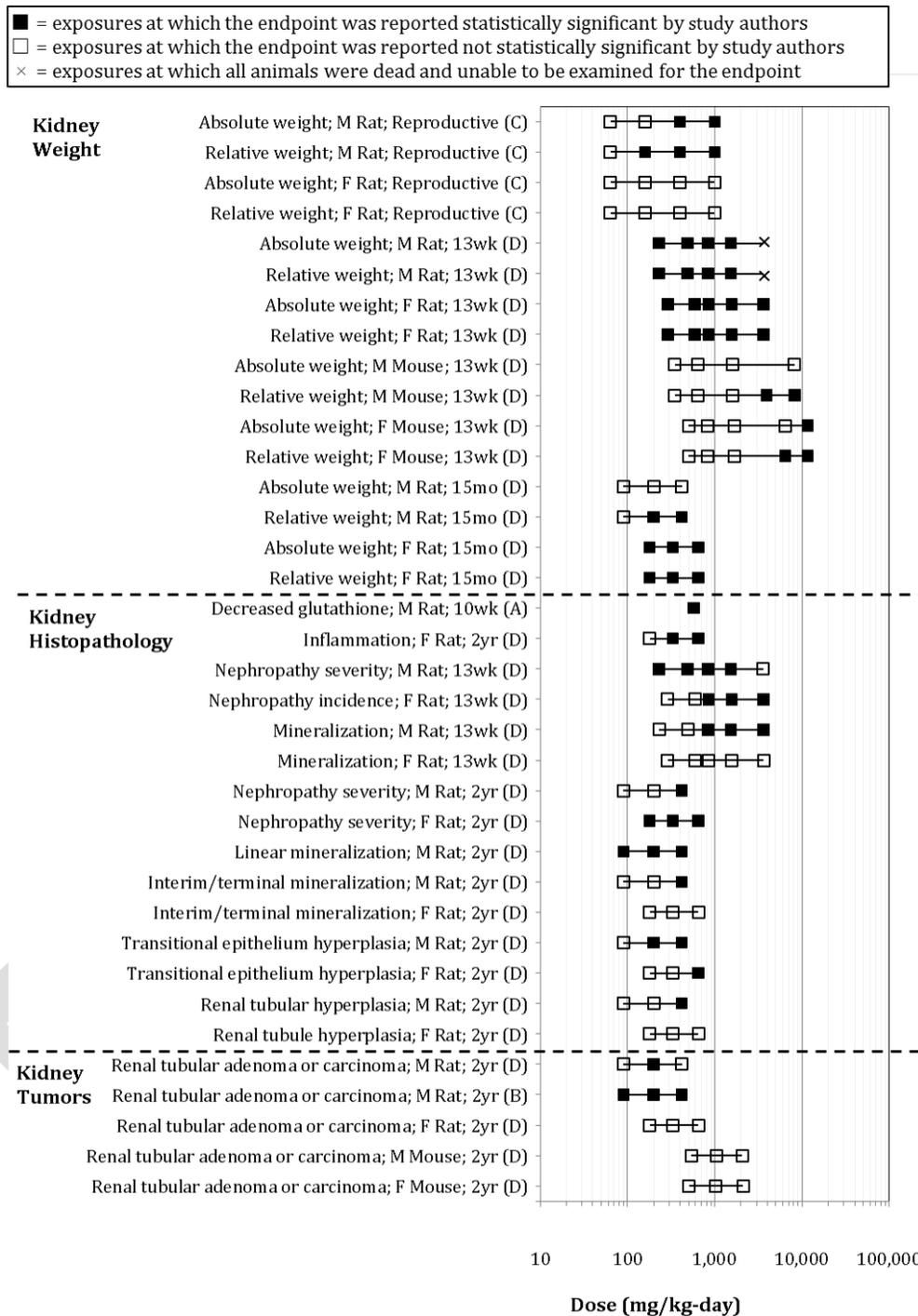
Reference and study design	Results														
<p>NTP (1995)</p> <p>B6C3F₁ mouse; 60/sex/treatment Drinking water (0, 5, 10, or 20 mg/mL) M: 0, 540, 1,040, or 2,070^a mg/kg-d F: 0, 510, 1,020, or 2,110 mg/kg-d 2 years</p>	<p>No changes in kidney related histopathology observed.^c</p>														
<p>NTP (1997)</p> <p>F344/N rat; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole-body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported</p>	<p>Male</p> <table border="1" data-bbox="597 688 987 1031"> <thead> <tr> <th>Concentration (mg/m³)</th> <th>Average severity of chronic nephropathy</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>1.0</td> </tr> <tr> <td>406</td> <td>1.4</td> </tr> <tr> <td>824</td> <td>1.4</td> </tr> <tr> <td>1,643</td> <td>1.6</td> </tr> <tr> <td>3,273</td> <td>1.9</td> </tr> <tr> <td>6,368</td> <td>2.0</td> </tr> </tbody> </table> <p>Severity categories: 1= minimal, 2= mild. No results from statistical tests reported</p>	Concentration (mg/m ³)	Average severity of chronic nephropathy	0	1.0	406	1.4	824	1.4	1,643	1.6	3,273	1.9	6,368	2.0
Concentration (mg/m ³)	Average severity of chronic nephropathy														
0	1.0														
406	1.4														
824	1.4														
1,643	1.6														
3,273	1.9														
6,368	2.0														
<p>NTP (1997)</p> <p>B6C3F₁ mouse; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole-body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported</p>	<p>There were no kidney effects observed.</p>														

1 ^a The high-dose group had an increase in mortality.
 2 ^b Linear mineralization not observed in female rats.
 3 ^c Organs were not weighed in mice during the 2-year study. Relative organ weights refer to relative to body weight
 4 * Statistically significant $p \leq 0.05$ as determined by the study authors.
 5 Conversions from drinking water concentrations to mg/kg-d performed by study authors.
 6 Conversion from ppm to mg/m³ is 1 ppm = 3.031 mg/m³.
 7
 8

1 **Table 1-3. Changes in kidney tumors in animals following exposure to**
 2 **tert-butanol**

Reference and study design	Results				
<p>NTP (1995)</p> <p>F344/N rat; 60/sex/treatment (10/sex/treatment evaluated at 15 months) Drinking water (0, 1.25, 2.5, 5, or 10 mg/mL) M: 0, 90, 200, or 420^a mg/kg-d F: 0, 180, 330, or 650^a mg/kg-d 2 years</p>	<p>Male <u>Dose</u> (mg/kg-d)</p>	<p><u>Renal tubule adenoma</u> (single)</p>	<p><u>Renal tubule adenoma</u> (multiple)</p>	<p><u>Renal tubule carcinoma</u></p>	<p><u>Renal tubule adenoma (single or multiple) or carcinoma</u></p>
	0	7/50	1/50	0/50	8/50
	90	7/50	4/50	2/50	13/50
	200	10/50	9/50*	1/50	19/50*
	420	10/50	3/50	1/50	13/50
	<p>Female <u>Dose</u> (mg/kg-d)</p>	<p><u>Renal tubule adenoma</u> (single)</p>	<p><u>Renal tubule adenoma</u> (multiple)</p>	<p><u>Renal tubule carcinoma</u></p>	<p><u>Renal tubule adenoma (single or multiple) or carcinoma</u></p>
	0	0/50	0/50	0/50	0/50
	180	0/50	0/50	0/50	0/50
	330	0/50	0/50	0/50	0/50
	650	0/50	0/50	0/50	0/50
Results do not include the animals sacrificed at 15 months.					
<p>Hard et al. (2011)</p> <p>reanalysis of the slides from male rats in the NTP (1995) study (see above)</p>	<p>Male <u>Dose</u> (mg/kg-d)</p>	<p><u>Renal tubule adenoma</u> (single)</p>	<p><u>Renal tubule adenoma</u> (multiple)</p>	<p><u>Renal tubule carcinoma</u></p>	<p><u>Renal tubule adenoma (single or multiple) or carcinoma</u></p>
	0	3/50	1/50	0/50	4/50
	90	9/50	3/50	1/50	13/50*
	200	9/50	9/50	0/50	18/50*
	420	9/50	3/50	1/50	12/50*
<p>NTP (1995)</p> <p>B6C3F₁ mouse; 60/sex/treatment Drinking water (0, 5, 10, or 20 mg/mL) M: 0, 540, 1,040, or 2,070^a mg/kg-d F: 0, 510, 1,020, or 2,110 mg/kg-d 2 years</p>	No changes in kidney-related tumors				

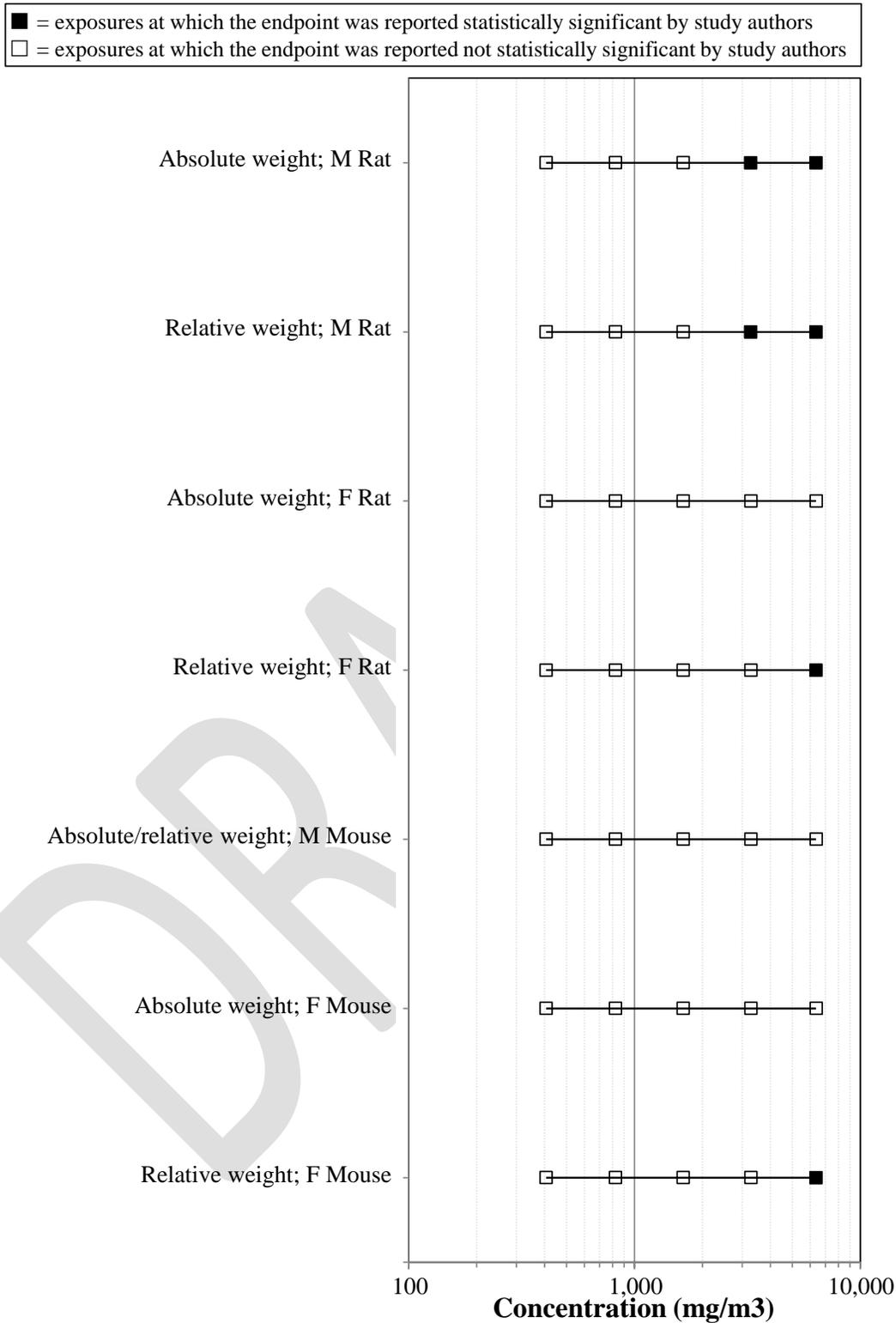
3 ^a The high-dose group had an increase in mortality.
 4 * Statistically significant $p \leq 0.05$ as determined by the study authors.
 5 Conversions from drinking water concentrations to mg/kg-d performed by study authors.
 6



Sources: (A) Acharya et al. (1997; 1995); (B) Hard et al. (2011)*; (C) Lyondell Chemical Co. (2004) (D) NTP (1995); * reanalysis of NTP (1995)

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Figure 1-1. Exposure response array for kidney effects following oral exposure to *tert*-butanol.



Source: [NTP \(1997\)](#)

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Figure 1-2. Exposure-response array of kidney effects following subchronic inhalation exposure to *tert*-butanol (no chronic studies available).

1 **Mode of Action Analysis—Kidney Effects**

2 Mode of Action Analysis for α_{2u} -globulin-associated nephropathy

3 *Description of the hypothesized MOA*

4 Several studies were identified that evaluated the role of α_{2u} -globulin in *tert*-butanol-
5 induced renal tumor development ([Borghoff et al., 2001](#); [Williams and Borghoff, 2001](#); [Takahashi et](#)
6 [al., 1993](#)). α_{2u} -Globulin is a member of a large superfamily of low-molecular-weight proteins and
7 was first characterized in male rat urine. Such proteins have been detected in various tissues and
8 fluids of most mammals (including humans), but the particular isoform of α_{2u} -globulin commonly
9 detected in male rat urine is considered specific to the male rat.

10 The hypothesized sequence of α_{2u} -globulin-associated nephropathy, as described by [U.S.](#)
11 [EPA \(1991a\)](#), is as follows. Chemicals that induce α_{2u} -globulin accumulation do so rapidly. The
12 accumulation of α_{2u} -globulin in the hyaline droplets results in hyaline droplet deposition in the P2
13 segment of the proximal tubule within 24 hours of exposure. Hyaline droplets are a normal
14 constitutive feature of the mature male rat kidney; they are particularly evident in the P2 segment
15 of the proximal tubule and contain α_{2u} -globulin ([U.S. EPA, 1991a](#)). Abnormal increases in hyaline
16 droplets have more than one etiology and can be associated with the accumulation of different
17 proteins. As hyaline droplet deposition continues, single-cell necrosis occurs in the P2 segment
18 which leads to exfoliation of these cells into the tubule lumen within 5 days of chemical exposure. In
19 response to the cell loss, cell proliferation is observed in the P2 segment after 3 weeks and
20 continues for the duration of the exposure. After 2 or 3 weeks of exposure, the cell debris
21 accumulates in the P3 segment of the proximal tubule to form granular casts. Continued chemical
22 exposure for 3 to 12 months leads to the formation of calcium hydroxyapatite in the papilla which
23 results in linear mineralization. After 1 or more years of chemical exposure, these lesions may
24 result in the induction of renal adenomas and carcinomas.

25 [U.S. EPA \(1991a\)](#) states that two questions must be addressed to determine the extent to
26 which α_{2u} -globulin mediated processes induce renal tumors and nephropathy. First, it must be
27 determined whether the α_{2u} -globulin process is occurring in male rats and therefore could be a
28 factor in renal effects. [U.S. EPA \(1991a\)](#) states that the criteria for answering this question in the
29 affirmative are as follows:

- 30 1) hyaline droplets are increased in size and number in male rats,
31 2) the protein in the hyaline droplets in male rats is α_{2u} -globulin, and
32 3) if several (but not necessarily all) additional steps in the pathological sequence are present
33 in male rats, such as:
34 (a) single-cell necrosis,
35 (b) exfoliation of epithelial cells into the tubular lumen,

- 1 (c) granular casts,
2 (d) linear mineralization, and
3 (e) tubule hyperplasia.

4 The available data relevant to this question in male rats are summarized in Table 1-5 and
5 Figure 1-3 and Figure 1-4, and will be evaluated below in accordance with the mode of action
6 (MOA) framework from the EPA cancer guidelines ([U.S. EPA, 2005a](#)).

7 If the α_{2u} -globulin process is operative, then [U.S. EPA \(1991a\)](#) states that a second question
8 must be answered as to whether the renal effects are solely due to the α_{2u} -globulin process, are a
9 combination of the α_{2u} -globulin process and other carcinogenic processes, or are due primarily to
10 other processes. [U.S. EPA \(1991a\)](#) states that the following types of data may be useful for
11 answering this question:

- 12 1) Hypothesis-testing data
13 2) Biochemical information
14 3) Sustained cell division in the proximal tubule of the male rat
15 4) Structure-activity relationships
16 5) Covalent binding to macromolecules
17 6) Genotoxicity
18 7) Nephrotoxicity
19 8) Animal bioassay data in other species-, sex-combinations
20 9) Other information not specifically listed

21 The available data relevant to this question are summarized in Table 1-6, and will be
22 evaluated below in accordance with the MOA framework from the EPA cancer guidelines ([U.S. EPA,](#)
23 [2005a](#)).

24 From these two questions, [U.S. EPA \(1991a\)](#) states that one of three possible conclusions
25 can be made:

- 26 • If renal tumors in male rats are attributable solely to the α_{2u} -globulin process, then [U.S. EPA](#)
27 [\(1991a\)](#) states that such tumors will not be used for human cancer hazard identification or
28 for dose-response extrapolations.
- 29 • If renal tumors in male rats are not linked to the α_{2u} -globulin process, then [U.S. EPA \(1991a\)](#)
30 states that such tumors are an appropriate endpoint for human hazard identification and
31 are considered, along with other appropriate endpoints, for quantitative risk estimation.

- If some renal tumors in male rats are attributable to the α_{2u} -globulin process and some attributable to other carcinogenic processes, then [U.S. EPA \(1991a\)](#) states that such tumors remain relevant for purposes of hazard identification, but a dose-response estimate based on such tumors in male rats should not be performed unless there is enough information to determine the relative contribution of each process to the overall renal tumor response.

Additionally, [U.S. EPA \(1991a\)](#) states that if the α_{2u} -globulin process is occurring in male rats, then the associated nephropathy in male rats (described above) would not be an appropriate endpoint to determine noncancer effects occurring in humans. In such a case, the characterization of human health hazard for renal toxicity would need to rely on other types of nephrotoxic effect data in male rats and/or on nephrotoxic effect data in female rats or other species.

Table 1-4. Additional kidney effects potentially relevant to mode of action in animals following exposure to *tert*-butanol

Reference and study design	Results														
Williams and Borghoff (2001) F344 rats; 4/sex Single gavage dose: 500 mg/kg	Males: ↑ binding of <i>tert</i> -butanol to α_{2u} -globulin compared to females* Females: no change in binding observed														
NTP (1995) F344/N rat; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 230, 490, 840, 1,520, 3,610 ^a mg/kg-d F: 0, 290, 590, 850, 1,560, 3,620 ^a mg/kg-d 13 weeks	Accumulation of hyaline droplets: <table border="1"> <thead> <tr> <th>Male Dose (mg/kg-d)</th> <th>Hyaline droplet accumulation</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>0/10</td> </tr> <tr> <td>230</td> <td>+^e</td> </tr> <tr> <td>490</td> <td>++</td> </tr> <tr> <td>840</td> <td>++</td> </tr> <tr> <td>1,520</td> <td>++</td> </tr> <tr> <td>3,610</td> <td>0/10</td> </tr> </tbody> </table> No information provided on females. No results from statistical tests reported.	Male Dose (mg/kg-d)	Hyaline droplet accumulation	0	0/10	230	+ ^e	490	++	840	++	1,520	++	3,610	0/10
Male Dose (mg/kg-d)	Hyaline droplet accumulation														
0	0/10														
230	+ ^e														
490	++														
840	++														
1,520	++														
3,610	0/10														
Hard et al. (2011) Reanalysis of the slides in the NTP (1995) study	Males: Confirmed accumulation of hyaline droplets increased with increasing dose-levels in 13 week study above. No incidence data available. Females: not evaluated														

13

Table 1-4. Additional kidney effects potentially relevant to mode of action in animals following exposure to tert-butanol (continued)

Reference and study design	Results
<p>Borghoff et al. (2001) F344 rat; 5/sex/treatment Analytical concentration:0, 250, 450, 1,750 ppm (0,771, 1,387 or 5,395mg/m³) 6hr/d 10 days</p>	<p>Males: positive trend for accumulation of protein droplets ($p < 0.05$), significant increase in accumulation of α_{2u}-globulin at 5,395 mg/m³ as compared to controls (no incidence data provided) Females: No positive staining for α_{2u}-globulin was observed in exposed female rats.</p>

- 1 ^a The high-dose group had an increase in mortality.
- 2 ^b Linear mineralization not observed in female rats.
- 3 ^c Organs were not weighed in mice during the 2-year study.
- 4 ^d Standard & extended evaluation combined.
- 5 ^e + or ++ indicated an increased accumulation relative to controls, as reported by the authors; no additional incidence data and
- 6 no results from statistical tests available.
- 7 * Statistically significant $p \leq 0.05$ as determined by the study authors.
- 8 Conversion from ppm to mg/m³ is 1 ppm = 3.031 mg/m³.
- 9 Percentage change compared to control = (treated value – control value) ÷ control value × 100.
- 10 Conversions from drinking water concentrations to mg/kg-d performed by study authors.
- 11

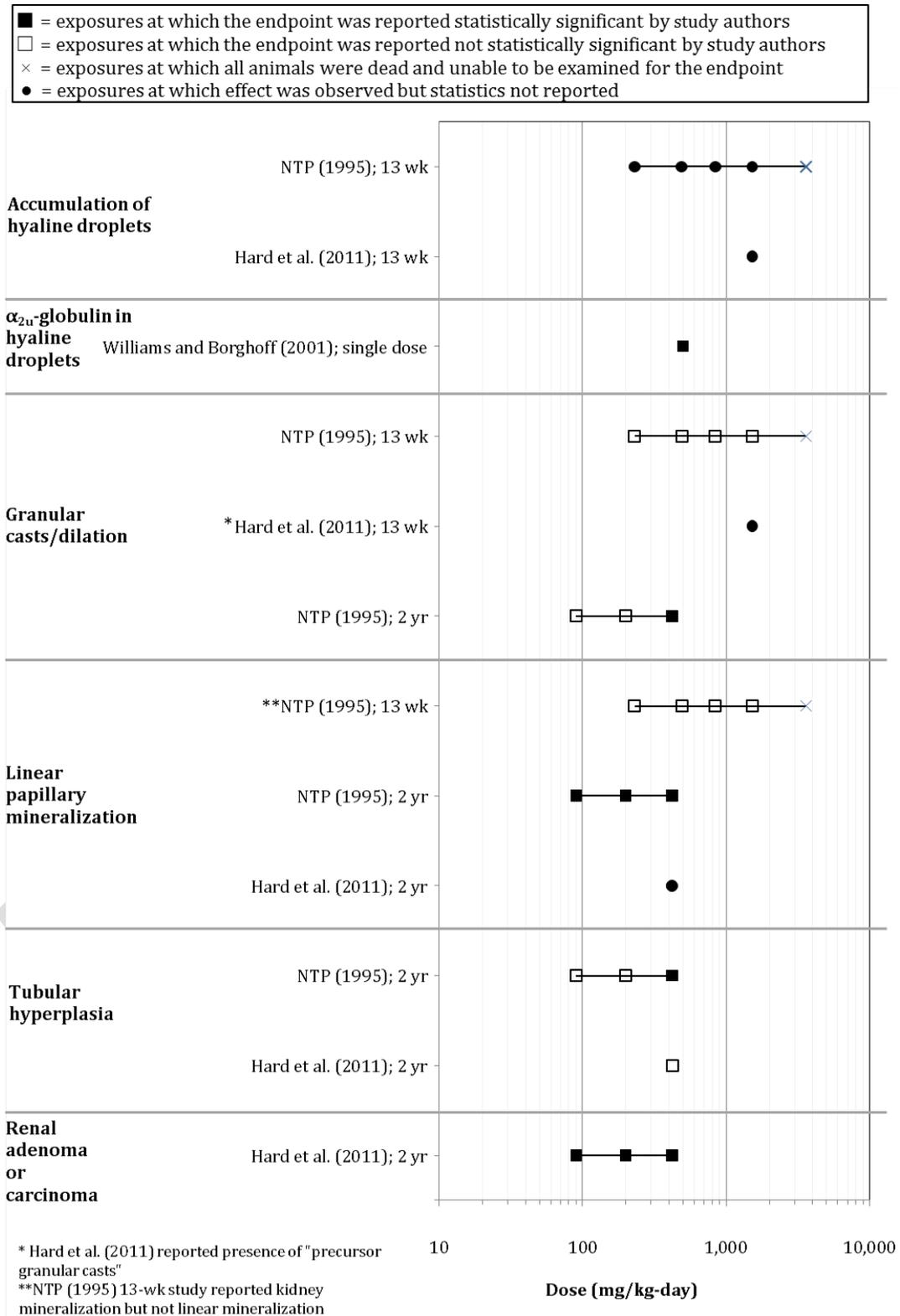
DRAFT

1 **Table 1-5. Summary of data on the α_{2u} -globulin process in male rats exposed**
 2 **to tert-butanol**

Criterion	Duration	Results	Reference
(1) hyaline droplets are increased in size and number	10 d	+	Borghoff et al. (2001)
	13 wk	(+)	NTP (1995)
	13 wk	–	NTP (1997)
	13 wk	(+) ^a	Hard et al. (2011)
(2) the protein in the hyaline droplets is α_{2u} -globulin	12 hr	+	Williams and Borghoff (2001)
	10 d	+	Borghoff et al. (2001)
(3) Several (but not necessarily all) additional steps in the pathological sequence are present in male rats, such as:			
(a) Single-cell necrosis	13 wk	–	NTP (1995)
	2 yr	–	NTP (1995)
	2 yr	–	Hard et al. (2011)
(b) exfoliation of epithelial cells into the tubular lumen	13 wk	–	NTP (1995)
	2 yr	–	NTP (1995)
	2 yr	–	Hard et al. (2011)
(c) granular casts	13 wk	–	NTP (1995)
	13 wk	(+) ^{a,c}	Hard et al. (2011)
	13 wk	–	NTP (1997)
	2 yr	– ^b	NTP (1995)
(d) linear mineralization	13 wk	–	NTP (1995)
	13 wk	–	NTP (1997)
	2 yr	+	NTP (1995)
	2 yr	(+) ^a	Hard et al. (2011)
(e) tubule hyperplasia	2 yr	+	NTP (1995)

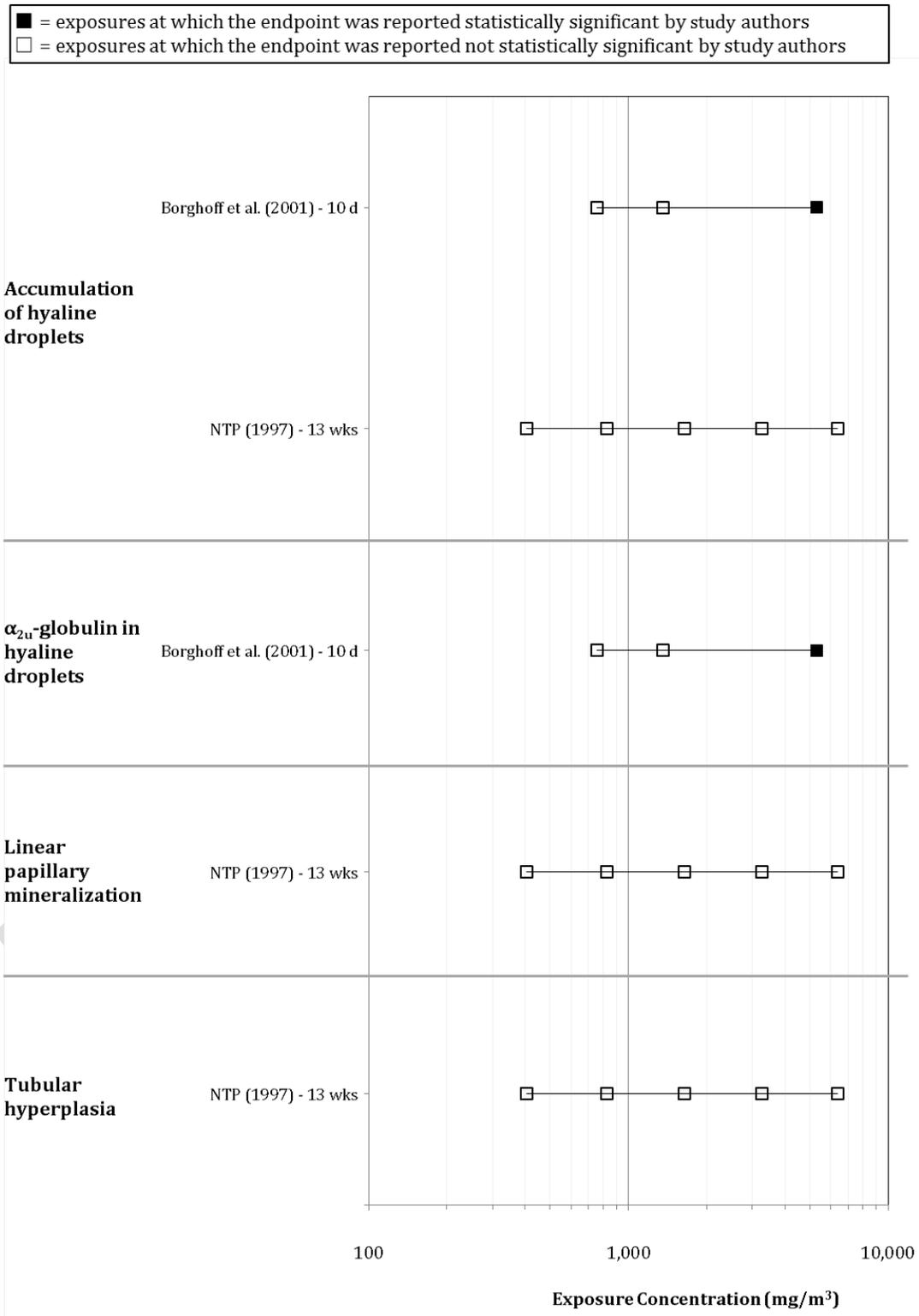
- 3 + = Statistically significant change reported in one or more treated groups.
 4 (+) = Effect was reported in one or more treated groups, but statistics not reported.
 5 – = No statistically significant change reported in any of the treated groups.
 6 ^a Re-analysis of one control and one treated group from [NTP \(1995\)](#)
 7 ^b Protein casts reported, not granular casts
 8 ^c Precursors to granular casts reported

9
 10



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Figure 1-3. Exposure-response array for components of α_{2u}-globulin nephropathy and renal tumors in male rats after oral exposure to tert-butanol.



1

2 **Figure 1-4. Exposure-response array for components of α_{2u} -globulin**
 3 **nephropathy and renal tumors in male rats after inhalation exposure to**
 4 ***tert*-butanol.**

1 **Table 1-6. Summary of additional data informing the contribution of the**
 2 **α_{2u} -globulin process on the renal tumor development in male rats exposed to**
 3 ***tert*-butanol**

Type of data Description	Reference
(1) Hypothesis-testing data	
No data	
(2) Biochemical information	
Reversible, non-covalent binding of <i>tert</i> -butanol to α_{2u} -globulin.	Williams and Borghoff (2001)
(3) Sustained cell division in the proximal tubule of the male rat	
Hyperplasia at 2 yr reported in both male and female rats, attributed to CPN.	Hard et al. (2011)
No effect on proliferation at 13 wk	NTP (1997)
Increased proliferation at 13 wk based on PCNA assay	NTP (1995)
Increased proliferation of the P2 segment at 10 days based on BrdU labeling	Borghoff et al. (2001)
(4) Structure-activity relationships	
No data	
(5) Covalent binding to macromolecules	
No data	
(6) Genotoxicity	
Limited database to conclude <i>tert</i> -butanol is genotoxic or non-genotoxic	See Appendix B.3.
(7) Nephrotoxicity	
Increased tubular regeneration and intratubule protein cast formation at 2 yr in males and females, with effects in females occurring at lower dose.	NTP (1995)
Increased severity of CPN in male rats after 13 wk inhalation exposure	NTP (1997)
Increased CPN in male and female F344/N rats following drinking water exposure for 13 wk.	NTP (1995)
Increased CPN in male and female F344/N rats following drinking water exposure for 2 yr.	NTP (1995)
(8) Animal bioassay data in other species-, sex-combinations	
Two renal tubular adenocarcinomas (not statistically significant as compared to concurrent controls) reported in male mice following drinking water exposure for 2 yr. These tumors are very rare in mice.	NTP (1995)
(9) Other data	
Dose-response and temporal concordance (see Figures 1-3 and 1-4).	

4
5

1 *Strength, consistency, specificity of association*

2 *Is the α_{2u} -globulin process occurring in male rats exposed to tert-butanol?*

3 The first criterion considered is whether hyaline droplets are increased in size and number
4 in male rats. Protein droplet accumulation was statistically significantly increased in the kidneys of
5 male rats exposed to 5,305 mg/m³ *tert*-butanol for 6 hr/day for 10 days ([Borghoff et al., 2001](#)). Data
6 from drinking water studies ([NTP, 1995](#); [Takahashi et al., 1993](#); [Lindamood et al., 1992](#))
7 demonstrated a statistically significant increase, except at the highest dose, in hyaline droplet
8 formation and severity in the proximal tubule of male rats following oral exposure to *tert*-butanol
9 for 13 weeks. Treated males had large hyaline droplets with crystal accumulation, but the controls
10 had small droplets without crystals. [NTP \(1997\)](#) stained for hyaline droplet formation in male rats
11 exposed to 0, 3,273, or 6,368 mg/m³ *tert*-butanol via inhalation for 13 weeks, and there was no
12 difference between the controls and treatment groups.

13 The second criterion considered is whether the protein in the hyaline droplets in male rats
14 is α_{2u} -globulin. Two studies measured α_{2u} -globulin immunoreactivity in the hyaline droplets of the
15 renal proximal tubular epithelium ([Borghoff et al., 2001](#); [Williams and Borghoff, 2001](#)). [Borghoff et](#)
16 [al. \(2001\)](#) observed α_{2u} -globulin immunoreactivity present in the hyaline droplets in the male rats.
17 No α_{2u} -globulin immunostaining was observed in the kidneys of the female rats. [Williams and](#)
18 [Borghoff \(2001\)](#) found the content of α_{2u} -globulin statistically significantly elevated in 72% of the
19 kidneys of male rats treated with *tert*-butanol compared with controls treated with corn oil.

20 The third criterion considered is whether several (but not necessarily all) additional steps
21 in the pathological sequence are present in male rats. Several, but not all, of the subsequent
22 histopathological lesions were observed in the available subchronic or chronic *tert*-butanol
23 exposure studies. Linear mineralization was the lesion most consistently observed in male rats and
24 was found to be statistically significantly increased in male rats after 2 years of oral exposure ([NTP,](#)
25 [1995](#)) (see Table 1-2). The 13-week study in rats ([NTP, 1995](#)) reported mineralization, but it was
26 not characterized as linear mineralization. Additionally, although the inhalation study by [NTP](#)
27 [\(1997\)](#) did not report linear mineralization at 13 weeks, this may be due to the lower internal dose
28 as compared to the oral studies. Atypical tubule hyperplasia was statistically significantly increased
29 at the highest dose following 2 years of oral exposure ([NTP, 1995](#)). Granular casts were increased at
30 the 13 week time point ([NTP, 1995](#)), though statistical significance was not determined. The
31 reanalysis of the 13-week data by [Hard et al. \(2011\)](#) concluded that the lesions were precursors to
32 granular casts and not the casts themselves. Other studies did not observe granular casts at 13
33 weeks or 2 years ([NTP, 1997, 1995](#)). The formation of protein casts were observed, but these may
34 be part of the pathology of CPN and are not thought to be related to α_{2u} -globulin nephropathy ([NTP,](#)
35 [1995](#)). Neither necrosis nor epithelial exfoliation was reported in any study.

36 In summary, the evidence supports the conclusion that *tert*-butanol causes increases in the
37 size and number of hyaline droplets, and the accumulating protein in the hyaline droplets is
38 α_{2u} -globulin. Additionally, several, but not all, of the additional steps in the pathological sequence

1 were observed, and not always consistently across studies. Therefore, the overall strength,
2 consistency, and specificity of the association between *tert*-butanol and the hypothesized key
3 events is moderate.

4 *Are the renal effects in male rats exposed to tert-butanol solely due to the α_{2u} -globulin process?*

5 As summarized in Table 1-6, there are many potential sources of additional data that may
6 inform the contribution of the α_{2u} -globulin process on renal tumor development. No hypothesis-
7 testing data, structure-activity relationships, or covalent binding data were located, so these types
8 of data are not discussed. Additional data related to dose-response concordance and temporal
9 relationships are discussed in subsequent sections.

10 In terms of biochemical information, [Williams and Borghoff \(2001\)](#) report that *tert*-butanol
11 reversibly and non-covalently binds to α_{2u} -globulin. This provides additional support to the
12 evidence that the α_{2u} -process is occurring, but it does not inform the relative contribution to renal
13 tumor development.

14 Sustained cell division in the proximal tubule of the male rat is consistent with, though not
15 specific to, the α_{2u} -process. Proliferation of the proximal tubule was significantly increased in male
16 rats after 10 days of inhalation exposure to *tert*-butanol at concentrations of 771–5,395 mg/m³
17 ([Borghoff et al., 2001](#)) and after 13 weeks of oral exposure to 1520 mg/kg-day ([NTP, 1995](#);
18 [Takahashi et al., 1993](#); [Lindamood et al., 1992](#)), but not after 13 week inhalation exposure up to
19 6368 mg/m³ ([NTP, 1997](#)). Therefore, it is unclear the extent to which increased cell division is
20 sustained. While hyperplasia was reported in chronic studies ([Hard et al., 2011](#)), it was observed in
21 both male and female rats, and attributed to CPN.

22 There are a limited number of studies available to assess the genotoxic potential of *tert*-
23 butanol (see Appendix B.3 in Supplemental Information for further details). *tert*-Butanol was
24 generally negative in a variety of genotoxicity assays and cell systems including *Salmonella*
25 *typhimurium*, *Escherichia coli* and *Neurospora crassa* ([Mcgregor et al., 2005](#); [Zeiger et al., 1987](#);
26 [Dickey et al., 1949](#)). Studies also demonstrate negative results for gene mutations, sister chromatid
27 exchanges, micronucleus formation, and chromosomal aberrations ([NTP, 1995](#); [McGregor et al.,](#)
28 [1988](#)). However, DNA adducts were found in male Kunming mice ([Yuan et al., 2007](#)), and DNA
29 damage was observed in human HL-60 leukemia cells ([Tang et al., 1997](#)). In another study by
30 [Sgambato et al. \(2009\)](#), an initial increase in DNA damage was observed as measured by nuclear
31 fragmentation, but the damage declined drastically following 4 hours of exposure and disappeared
32 entirely after 12 hours of exposure to *tert*-butanol.

33 In terms of nephrotoxicity, a number of renal effects have been reported in female rats
34 and/or in mice. Kidney transitional epithelial hyperplasia and inflammation were significantly
35 increased in both male and female F344 rats exposed for 2 years via oral exposure. F344 rats
36 exposed to *tert*-butanol at dose ranges of 230–1520 mg/kg-day and 850–3,620 mg/kg-day in males
37 and females, respectively, exhibited a statistically significant increase in the incidence of
38 nephropathy compared with controls after 13 weeks of exposure ([NTP, 1995](#)). Nephropathy

1 severity was also significantly increased at 420 mg/kg-day in males and 180-650 mg/kg-day in
2 females after 2 years of exposure ([NTP, 1995](#)). Average severity of chronic nephropathy was
3 minimal to mild in males after a 13-week inhalation exposure ([NTP, 1997](#)). Female rats also had
4 lesions associated with nephropathy ([NTP, 1995](#)), but none of the lesions were similar to those
5 observed in the male rat that are associated with α_{2u} -globulin nephropathy.

6 With respect to renal tumors, no statistically significant increases in renal tumors were
7 reported in *tert*-butanol-exposed female rats or mice compared with concurrent controls. Two
8 renal tubular adenocarcinomas were reported in male mice following drinking water exposure for
9 2 years ([NTP, 1995](#)) (one each in the low and high dose groups). Although such tumors are very
10 rare in mice, with historical control incidences of 2/1351 (0.15%) in feeding studies and 4/1093
11 (0.37%) in chamber studies ([Haseaman et al., 1998](#)), these data are not sufficient to indicate that the
12 kidney tumors observed in mice exposed to *tert*-butanol are treatment-related.

13 Overall, the strength, consistency, and specificity of the data supporting a *tert*-butanol-
14 induced α_{2u} -globulin process as the sole actor for renal effects in male rats is weak to moderate.

15 *Dose-response concordance*

16 *Is the α_{2u} -globulin process occurring in male rats exposed to tert-butanol?*

17 As shown in Figure 1-3 and Figure 1-4, the dose-response concordance among hypothesized
18 key events is mixed.

19 [Borghoff et al. \(2001\)](#) exposed male and female F344 rats to *tert*-butanol at concentrations
20 of 758, 1,364, or 5,305 mg/m³ for 6 hr/day for 10 days to assess the role of α_{2u} -globulin
21 nephropathy and renal cell proliferation. Significant tubular proliferation in males was observed at
22 all exposure levels, but accumulation of α_{2u} -globulin-positive hyaline droplets was increased only at
23 the highest dose ([Borghoff et al., 2001](#)). These data suggest that cell proliferation may be related to
24 the α_{2u} -globulin process only at the highest exposure concentration.

25 The dose-response relationships observed after 13 weeks and 2-years were also only
26 moderately concordant. Data from a drinking water study ([NTP, 1995](#); [Takahashi et al., 1993](#);
27 [Lindamood et al., 1992](#)) demonstrated hyaline droplet formation in the proximal tubule of male rats
28 at all tested doses (except at the highest dose where all rats died during weeks 5-12) following oral
29 exposure to *tert*-butanol for 13 weeks. PWG reevaluation by [Hard et al. \(2011\)](#) reported observing
30 precursors to granular casts at the only dose level evaluated (1,520 mg/kg-day). Spontaneous
31 mineralization was observed, but the linear mineralization characteristic of this MOA was not
32 observed at any dose. At the 2-year timepoint, linear mineralization was observed at all exposure
33 levels from 90-420 mg/kg-day, and renal tubular hyperplasia was observed at the highest dose of
34 420 mg/kg-day ([NTP, 1995](#)), consistent with the expected dose-response relationship. Notably,
35 however, granular casts were not observed.

36 Overall, the dose-response concordance of the association between *tert*-butanol and the
37 hypothesized key events is moderate.

1 *Are the renal effects in male rats exposed to tert-butanol solely due to the α_{2u} -globulin process?*

2 Dose-response concordance between the hypothesized key events and the occurrence of
3 renal tumors can inform whether carcinogenesis is solely due to the α_{2u} -globulin process. Male
4 F344 rats exhibited an increased incidence of renal tubule adenomas and combined renal tubule
5 adenoma or carcinoma in a 2-year oral bioassay ([Hard et al., 2011](#); [NTP, 1995](#)). Increased tumors
6 were observed at 90, 200, and 420 mg/kg-day. Although some effects related to α_{2u} -globulin
7 nephropathy, including hyaline droplets and linear mineralization, were observed at all doses,
8 tubule hyperplasia was not observed at doses lower than 420 mg/kg-day in any study and only
9 precursor granular casts were observed at a much higher dose of 1,520 mg/kg-day. Moreover, the
10 middle dose of *tert*-butanol induced the greatest incidence of tumors, so increasing the dose from
11 200 to 420 mg/kg-day led to additional markers of α_{2u} -globulin nephropathy in the form of tubule
12 hyperplasia, but without any increase in tumor burden. Thus, *tert*-butanol induced tumors at lower
13 doses than for other precursor effects such as hyperplasia and granular casts, suggesting a weak
14 dose response concordance with the incidence of tumors.

15 Therefore, on the basis of weak dose-response concordance, the data suggest that the
16 observed tumors are not solely due to α_{2u} -globulin and that other processes are primarily
17 responsible for tumors.

18 *Temporal relationship*

19 *Is the α_{2u} -globulin process occurring in male rats exposed to tert-butanol?*

20 As shown in Table 1-2 and Table 1-4, hyaline droplets and α_{2u} -globulin accumulation were
21 observed after a single dose or 10-day exposure; precursors to granular casts were observed at 13
22 weeks; and tubular hyperplasia was observed at 2 years. The observations are consistent with the
23 expected temporal relationship ([Hard et al., 2011](#); [Borghoff et al., 2001](#); [Williams and Borghoff,](#)
24 [2001](#); [NTP, 1995](#)). However, the absence of other key events such as necrosis, exfoliation, and
25 granular casts in most other studies at the anticipated time points weaken the case for α_{2u} -globulin
26 MOA. Additionally, the NTP's 13-week study in rat reported kidney mineralization, but it was not
27 characterized as linear mineralization.

28 *Are the renal effects in male rats exposed to tert-butanol solely due to the α_{2u} -globulin process?*

29 This question cannot be answered from data on temporality.

30 *Biological plausibility and coherence*

31 Both U.S. EPA and IARC have accepted the general biological plausibility and coherence of
32 role of α_{2u} -globulin-mediated nephropathy in renal tumor induction ([Swenberg and Lehman-](#)
33 [McKeeman, 1999](#); [U.S. EPA, 1991a](#)), and those rationales will not be repeated here.

34 However, a retrospective analysis has suggested that a number of α_{2u} -globulin inducing
35 chemicals fail to induce many of the pathological sequences in the α_{2u} -globulin pathway ([Doi et al.](#)

1 [2007](#)). For instance, dose-response concordance was not observed for several endpoints such as
2 linear mineralization, tubular hyperplasia, granular casts, and hyaline droplets following exposure
3 to α_{2u} -globulin-inducing chemicals such as d-limonene, decalin, propylene glycol mono-t-butyl
4 ether, and Stoddard solvent IIC (SS IIC). Although some of these chemicals induced
5 histopathological lesions that exhibited a dose response, all of them failed to induce a dose-
6 response trend for at least one of the endpoints in the sequence. Furthermore, no endpoint in the
7 pathological sequence was predictive for tumor incidence. Tumor incidence did not exhibit a dose
8 response following either d-limonene or decalin exposure. Finally, tumor incidence was not
9 predicted by the severity of a particular effect in the α_{2u} -globulin sequence as demonstrated by SS
10 IIC which induced some of the most severe nephropathy precursors relative to the other chemicals
11 but did not significantly increase kidney tumors ([Doi et al., 2007](#)). Thus, these analyses suggest that
12 another MOA may be operative for inducing tumors.

13 Moreover, renal tumors were not observed following exposure to ETBE, which is rapidly
14 metabolized to *tert*-butanol. Specifically, [Suzuki et al. \(2012\)](#) and [Saito et al. \(2013\)](#) reported no
15 increase in renal tumors in both sexes of Fischer 344 rats following 2-year oral or inhalation
16 exposures to ETBE at doses that yield similar internal concentrations (based on PBPK modeling) of
17 *tert*-butanol compared with the concentrations of the *tert*-butanol bioassays. After 13 weeks of
18 exposure to *tert*-butanol or ETBE, hyaline droplets were increased in a dose-response manner.
19 ETBE exposure increased hyaline droplets at lower internal concentrations of *tert*-butanol than by
20 direct *tert*-butanol administration. Similar to hyaline droplets, linear mineralization was increased
21 at an internal *tert*-butanol concentration approximately tenfold lower following ETBE exposure
22 than a *tert*-butanol exposure. By contrast, tubule hyperplasia and renal tumors were both observed
23 following a 2-year exposure to *tert*-butanol but not following ETBE exposure. Renal tumors and
24 tubule hyperplasia were not observed following any ETBE exposure despite achieving similar blood
25 concentrations of *tert*-butanol as the [NTP \(1995\)](#) study. The failure of internal *tert*-butanol
26 concentrations to induce histopathological lesions early in the α_{2u} -globulin pathological sequence
27 at blood levels that later induced hyperplasia and tumors suggests a lack of coherence across the
28 two data sets.

29 *Conclusions about the hypothesized MOA for α_{2u} -globulin-associated nephropathy*

30 *Is the hypothesized MOA sufficiently supported in test animals?*

31 This conclusion is divided into two sub-questions: whether the α_{2u} -globulin process is
32 occurring in male rats, and whether it is the sole contributor to renal effects in male rats.

33 With respect to the first question, *tert*-butanol induced increases in α_{2u} -globulin deposition
34 and hyaline droplet accumulation, and several of the subsequent steps in the pathological sequence
35 were observed. These data provide sufficient evidence that the α_{2u} -globulin process can be
36 operating given sufficient *tert*-butanol exposure.

1 With respect to the second question, male rats are more sensitive to the kidney effects of
2 *tert*-butanol, and the available data indicate that male rats accumulate α_{2u} -globulin in the kidney,
3 which is a specific MOA for male rats. Many of the steps in the pathological sequence of lesions
4 related to α_{2u} -globulin-associated nephropathy were observed exclusively in male rats but not in
5 female rats, or mice of either sex, and renal tumors occurred only in male rats. However, there is
6 insufficient evidence to support a conclusion that α_{2u} -globulin nephropathy is the sole or primary
7 contributor to renal tumor development. Given the inconsistencies and limitations of the
8 genotoxicity database, the effect of *tert*-butanol with respect to genotoxicity cannot be ruled out.
9 Additionally, *tert*-butanol induced tumors at lower doses than other precursor effects such as
10 hyperplasia and granular casts, with no further increase in tumor incidence coinciding with the
11 additional markers of α_{2u} -globulin nephropathy. Thus, renal tumors observed in male rats are
12 unlikely to be confounded by the presence of α_{2u} -globulin nephropathy. Therefore, on the basis of a
13 weak dose-response concordance, the data support a conclusion that processes other than
14 α_{2u} -globulin nephropathy are likely responsible for renal tumor development.

15 *Is the hypothesized MOA relevant to humans?*

16 Based on the conclusion that processes other than α_{2u} -globulin nephropathy are likely
17 responsible for renal tumor development induced by *tert*-butanol, [U.S. EPA \(1991a\)](#) states that the
18 following conclusion will be made:

- 19 • If renal tumors in male rats are not linked to the α_{2u} -globulin process, then ([U.S. EPA,](#)
20 [1991a](#)) states that such tumors are an appropriate endpoint for human hazard
21 identification and are considered, along with other appropriate endpoints, for quantitative
22 risk estimation.

23 Therefore, kidney tumors are relevant to humans for purposes of hazard identification and
24 dose-response assessment. Because female rats and both sexes of mice do not have α_{2u} -globulin
25 present, kidney effects in these animals are considered relevant to humans for both hazard
26 identification and dose-response.

27

28 *Which populations or lifestyles can be particularly susceptible to the hypothesized MOA?*

29 This question is not applicable.

30 Alternative MOA hypotheses with inadequate data for analysis

31 Other nephrotoxic responses, such as exacerbation of CPN, inflammation, transitional
32 epithelial hyperplasia, and increased kidney weight, are observed in rats and/or mice, suggesting
33 other possible processes are operative. It has been proposed that enhanced chronic progressive
34 nephropathy (CPN) is a mode of action for chemically-induced kidney tumors in male rats and that

1 renal tubule tumors induced by chemicals that concomitantly exacerbate CPN are not relevant to
2 humans ([Hard and Khan, 2004](#)).

3 CPN is an age-related renal disease of unknown etiology that occurs spontaneously in rats,
4 especially the F344, Sprague-Dawley, and Osborne-Mendel strains. Additional markers associated
5 with CPN include elevated protein and albumin in the urine and increased BUN, creatinine, and
6 cholesterol in the serum ([Hard et al., 2009](#)). CPN is often more severe in males compared with
7 females. Several of the CPN pathological effects are similar to and can obscure the lesions
8 characteristic of α_{2u} -globulin-related hyaline droplet nephropathy ([Webb et al., 1990](#)). Additionally,
9 renal effects of α_{2u} -globulin accumulation can exacerbate the effects associated with CPN ([U.S. EPA,](#)
10 [1991a](#)). However, [Webb et al. \(1990\)](#) suggested that exacerbated CPN was one component of the
11 nephropathy resulting from exposure to chemicals that induce α_{2u} -globulin nephropathy. Male rat
12 sensitivity has been noted with both CPN and α_{2u} -globulin nephropathy.

13 Increased severity of CPN occurred in both male and female rats as a result of *tert*-butanol
14 exposure. Some of the observed renal lesions in male rats following exposure to *tert*-butanol are
15 effects commonly associated with CPN. [Hard et al. \(2011\)](#) concluded that the observation of
16 transitional epithelial hyperplasia in the 2-year drinking study conducted by [NTP \(1995\)](#) was
17 associated with CPN, and not a direct effect of *tert*-butanol exposure. However, there was a strong,
18 statistically-significant, treatment-related, dose-response relationship between chronic *tert*-butanol
19 exposure and increased incidence of transitional epithelial hyperplasia in both male and female rats
20 in the [NTP \(1995\)](#) study. The severity of CPN also increased with *tert*-butanol exposure, although
21 the dose-response relationship in males was very weak (only a 10% increase in mean severity at
22 the highest dose). The very different dose-response relationships argue against a close association.
23 Moreover, even if transitional epithelial hyperplasia were associated with CPN, there is no evidence
24 to support that the effect is independent of *tert*-butanol treatment, given the robust dose-response
25 relationships. Therefore, the data are insufficient to dismiss transitional epithelial hyperplasia as
26 causally related to *tert*-butanol exposure.

27 Additionally, there have been a few research groups who have discussed the role of CPN and
28 α_{2u} -globulin accumulation on the renal tumors observed in male rats exposed to *tert*-butanol.
29 [Cruzan et al. \(2007\)](#) concluded that α_{2u} -globulin, exacerbation of CPN, or a combination of both
30 were the MOAs for the kidney tumors in males. [Hard et al. \(2011\)](#) also concluded that both α_{2u} -
31 globulin-induced nephropathy and exacerbated CPN were MOAs for the kidney tumors observed in
32 the male rats in the 2-year drinking study conducted by [NTP \(1995\)](#). However, the underlying
33 mechanisms regulating CPN and its exacerbation are not well understood, and to date, there is no
34 scientific consensus on the relevance of CPN in rats to human health hazard ([Melnick et al., 2012](#);
35 [Hard et al., 2009](#)). Moreover, no key events for the exacerbation of CPN have been identified, so no
36 MOA analysis can be performed under the EPA Cancer Guidelines MOA framework ([U.S. EPA,](#)
37 [2005a](#)). Therefore, kidney effects from *tert*-butanol exposure associated with CPN are considered
38 relevant to humans.

1 **Summary of kidney toxicity**

2 Kidney toxicity was consistently observed after oral exposure in two strains of rats and in
3 one strain of mice and in both sexes. Absolute and relative kidney weights also were increased in
4 male and female rats in both the 13-week and 2-year studies. In male and female rats,
5 histopathological examination of the kidneys indicated kidney lesions exhibiting a dose-response
6 trend, increased incidence of nephropathy after 13 weeks and 2 years, and increased transitional
7 epithelium hyperplasia and suppurative inflammation (females only) after 2 years. In mice, the
8 only kidney effect observed was an increase in kidney weight (absolute and/or relative) in both
9 sexes of mice in the 13-week study. Organs were not weighed in the 2-year mouse study, so no
10 determination can be made. Furthermore, there were no treatment-related histopathological
11 lesions in the kidneys of mice at 13 weeks or 2 years.

12 Male rats are more sensitive to the kidney effects of *tert*-butanol, and the available data
13 indicate that male rats accumulate α_{2u} -globulin in the kidney, which is a specific MOA for male rats.
14 MOA analysis determined that the renal tumors observed in male rats are mediated by other
15 processes besides α_{2u} -globulin. Therefore, in the absence of a known MOA, EPA considers the male
16 and female kidney effects observed in experimental animals to be relevant to assessing human
17 health hazard.

18 EPA identified kidney effects as a human hazard of *tert*-butanol exposure. Data on kidney
19 tumors associated with *tert*-butanol exposure are discussed as part of the overall weight of
20 evidence for carcinogenicity in Section 1.2.2.

21 **1.1.2. Thyroid Effects**

22 ***Synthesis of Effects in Thyroid***

23 This section reviews the studies that investigated whether exposure to *tert*-butanol can
24 cause thyroid effects in humans or animals. The database examining thyroid effects following
25 *tert*-butanol exposure contains no human data and two chronic studies (one in rats and one in
26 mice). Studies employing short term and acute exposures that examined thyroid effects are not
27 included in the evidence tables; however, they are discussed in the text if they provide data to
28 support mode of action or hazard identification. No methodological concerns were identified that
29 would lead one or more studies to be considered less informative for assessing human health
30 hazard.

31 A 2-year inhalation study is not available. Thyroid effects were not observed in studies in
32 rats ([NTP, 1995](#)). Thyroid toxicity was observed in mice of both sexes after 2 years of oral exposure
33 via drinking water ([NTP, 1995](#)). Follicular cell hyperplasia, as well as follicular cell adenomas, was
34 present in both male and female mice. The evidence was stronger in females due to the dose-
35 related increase in follicular cell hyperplasia reaching statistical significance in the highest two
36 doses, and the presence of a statistically significant increase in follicular cell adenomas in the high-
37 dose group. There was also a statistically significant increase in follicular cell hyperplasia at all

1 doses in male mice, but only a marginal increase in follicular cell adenomas in the mid-dose group.
2 One high-dose male mouse developed a follicular cell carcinoma. The lower tumor incidence in
3 males may be due to the increased mortality seen in the high-dose group. [NTP \(1995\)](#) noted that
4 thyroid follicular cell tumorigenesis follows a progression from hyperplasia to adenoma and
5 carcinoma, suggesting that hyperplasia is a preneoplastic lesion in the thyroid.
6

DRAFT

1 **Table 1-7. Evidence pertaining to thyroid effects in animals following oral**
 2 **exposure to *tert*-butanol**

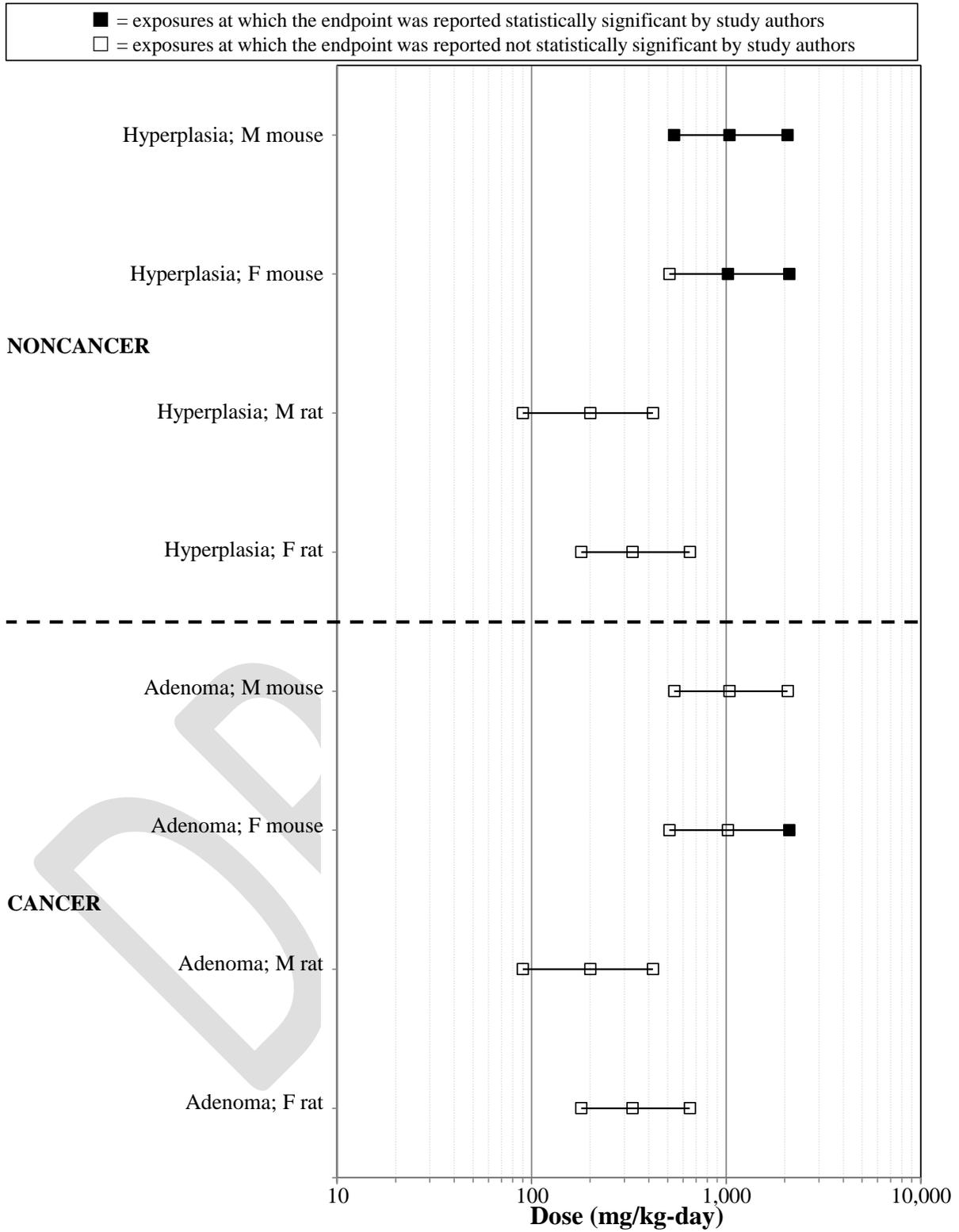
Reference and study design	Results			
<i>Follicular cell hyperplasia</i>				
NTP (1995) F344/N rat; 60/sex/treatment (10/sex/treatment evaluated at 15 months) Drinking water (0, 1.25, 2.5, 5, or 10 mg/mL) M: 0, 90, 200, or 420 ^a mg/kg-d F: 0, 180, 330, or 650 ^a mg/kg-d 2 years	Incidence ^b			
	Males		Females	
	<u>Dose (mg/kg-d)</u>	<u>Follicular cell hyperplasia</u>	<u>Dose (mg/kg-d)</u>	<u>Follicular cell hyperplasia</u>
	0	3/50	0	0/50
	90	0/49	180	0/50
NTP (1995) B6C3F ₁ mouse; 60/sex/treatment Drinking water (0, 5, 10, or 20 mg/mL) M: 0, 540, 1,040, or 2,070 ^a mg/kg-d F: 0, 510, 1,020, or 2,110 mg/kg-d 2 years	Incidence (severity)			
	Males		Females	
	<u>Dose (mg/kg-d)</u>	<u>Follicular cell hyperplasia</u>	<u>Dose (mg/kg-d)</u>	<u>Follicular cell hyperplasia</u>
	0	5/60 (1.2)	0	19/58 (1.8)
	540	18/59* (1.6)	510	28/60 (1.9)
1,040	15/59* (1.4)	1,020	33/59* (1.7)	
2,070	18/57* (2.1)	2,110	47/59* (2.2)	
<i>Tumors</i>				
NTP (1995) F344/N rat; 60/sex/treatment (10/sex/treatment evaluated at 15 months) Drinking water (0, 1.25, 2.5, 5, or 10 mg/mL) M: 0, 90, 200, or 420 ^a mg/kg-d F: 0, 180, 330, or 650 ^a mg/kg-d 2 years	Incidence ^b			
	<u>Dose (mg/kg-d)</u>	<u>Follicular cell adenoma</u>	<u>Follicular cell carcinoma</u>	
	Male			
	0	2/50	2/50	
	90	0/49	0/49	
	200	0/50	0/50	
	420	0/50	0/50	
	Female			
	0	1/50	1/50	
	180	0/50	0/50	
330	1/50	1/50		
650	0/50	0/50		

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Table 1-7. Evidence pertaining to thyroid effects in animals following oral exposure to tert-butanol (continued)

Reference and study design	Results					
<p>NTP (1995) B6C3F₁ mouse; 60/sex/treatment Drinking water (0, 5, 10, or 20 mg/mL) M: 0, 540, 1,040, or 2,070^a mg/kg-d F: 0, 510, 1,020, or 2,110 mg/kg-d 2 years</p>	Incidence					
	<u>Dose</u> (mg/kg-d)	<u>Follicular</u> <u>cell</u> <u>adenoma</u>	<u>Mortality-</u> <u>adjusted</u> <u>rate (%)</u>	<u>Follicular</u> <u>cell</u> <u>carcinoma</u> <u>or adenoma</u>	<u>Mortality-</u> <u>adjusted</u> <u>rate</u> <u>(%)</u>	<u>Follicular</u> <u>cell</u> <u>carcinoma</u> ^c
	Male					
	0	1/60	3.6	1/60	3.6	0/60
	540	0/59	0.0	0/59	0.0	0/59
	1,040	4/59	10.1	4/59	10.1	0/59
	2,070	1/57	5.9	2/57	8.7	1/57
	Female					
	0	2/58	5.6	2/58	5.6	0/58
	510	3/60	8.6	3/60	8.6	0/60
	1,020	2/59	4.9	2/59	4.9	0/59
	2,110	9/59*	19.6	9/59*	19.6	0/59

1 ^aThere was a significant decrease in survival in the high-dose group.
 2 ^bResults do not include the animals sacrificed at 15 months.
 3 ^cMortality-adjusted rates were not calculated by study authors for follicular cell carcinoma.
 4 * Statistically significant $p \leq 0.05$ as determined by the study authors.
 5 Conversions from drinking water concentrations to mg/kg-d performed by study authors.
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Source: [NTP \(1995\)](#)

Figure 1-5. Exposure-response array of thyroid follicular cell effects following chronic oral exposure to tert-butanol.

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1 **Mode of Action Analysis—Thyroid Effects**

2 There are inadequate data to determine the MOA for *tert*-butanol-induced thyroid follicular
3 cell lesions. The mechanism of formation of these lesions resulting from *tert*-butanol exposure has
4 not been specifically studied; however, [Blanck et al. \(2010\)](#) conducted a short-term study
5 examining the hepatic and thyroid effects of *tert*-butanol exposure to provide additional data on the
6 thyroid tumors observed in the chronic [NTP \(1995\)](#) study. *tert*-Butanol did not have any effect on
7 liver weight when compared to the control group, but the livers were visibly enlarged, in some
8 cases accompanied by centrilobular hepatocellular hypertrophy, in some treated groups. There
9 were no treatment-related histological alterations in the thyroid in *tert*-butanol treated mice. Only a
10 slight statistically nonsignificant increase in thyroid stimulating hormone (TSH) was observed after
11 3 days of exposure, but both thyroxine (T₄) and triiodothyronine (T₃) levels were decreased.
12 Sustained increases in TSH, resulting in sustained thyroid follicular cell proliferation, may
13 eventually result in progression of hyperplasia to adenoma and carcinoma ([U.S. EPA, 1998a](#)). Based
14 on alterations in hepatic phase I and phase II enzyme activities and gene expression, the data from
15 [Blanck et al. \(2010\)](#) suggest a possible role for increased thyroid hormone clearance in the liver in
16 *tert*-butanol-induced thyroid tumors. The available support for this hypothesis, however, is weak.
17 In particular, [Blanck et al. \(2010\)](#) did not find any significant changes in TSH levels, though the
18 study duration was short (≤14 days), and there are no data regarding thyroid cell proliferation after
19 exposure to *tert*-butanol.

20 **Summary of thyroid toxicity**

21 EPA identified thyroid effects as a potential human hazard of *tert*-butanol exposure. The
22 thyroid endpoints reported following chronic exposure to *tert*-butanol include follicular cell
23 hyperplasia, follicular cell adenoma, and follicular cell carcinoma. Together with the evidence of
24 significantly increased incidence of thyroid follicular cell adenomas in high-dose females, these
25 observations support the finding that the increased hyperplasia in male mice is a preneoplastic
26 effect rather than an adaptive response. There is no conclusive MOA for the development of thyroid
27 follicular cell adenomas, although there is some evidence supporting greater clearance of thyroid
28 hormones by the liver causing continual secretion of TSH by the pituitary leading to follicular cell
29 hyperplasia and tumors. Data on thyroid tumors associated with *tert*-butanol exposure are
30 discussed further as part of the overall weight of evidence for carcinogenicity in Section 1.2.2.

31 **1.1.3. Reproductive and Developmental Effects**

32 **Synthesis of reproductive and developmental toxicity**

33 This section reviews the studies that investigated whether exposure to *tert*-butanol can
34 cause reproductive and developmental effects in humans or animals. This section contains
35 information on reproductive effects, systemic developmental effects, as well as
36 neurodevelopmental effects following *tert*-butanol exposure. The database examining reproductive

1 or developmental effects following *tert*-butanol exposure contains no human data and 7 studies
2 performed in rats and mice. Three studies evaluating reproductive effects included changes in
3 reproductive organs (one one-generation reproductive study and two subchronic studies). In
4 addition, there was one two-year oral study in rats and mice that evaluated reproductive
5 histopathology and did not find any treatment-related effects. The studies selected for discussion
6 below exposed animals via oral gavage, drinking water, and inhalation for ≥ 63 days. The collection
7 of reproductive studies on *tert*-butanol is limited by the absence of any two-generation
8 reproductive oral or inhalation studies. Four studies evaluated developmental effects (three
9 developmental studies and one one-generation reproductive study). As mentioned in the Study
10 Selection, the studies selected for discussion below exposed animals to *tert*-butanol via inhalation,
11 gavage, and drinking water. No methodological concerns were identified that would lead one or
12 more studies to be considered less informative for assessing human health hazard, but the [Faulkner
13 et al. \(1989\)](#) study did not provide results in the dam that could be used to adequately determine if
14 fetal effects occurred due to maternal toxicity. Three studies evaluated neurodevelopmental effects
15 following *tert*-butanol exposure in rats and mice. These studies selected for discussion below
16 exposed animals via liquid diet and inhalation. The collection of neurodevelopmental studies on
17 *tert*-butanol is limited in that all studies were conducted prior to Developmental Toxicity
18 Guidelines being available from the [U.S. EPA \(1991b\)](#) and OECD; as such, there are study design
19 considerations for each of the studies. [Daniel and Evans \(1982\)](#) had a small number of animals per
20 treatment group, presentation of results provided limited use of the data with no comparisons to
21 controls, and there was no long term neurodevelopmental testing. [Nelson et al. \(1991\)](#) evaluated
22 neurodevelopmental effects after either paternal or maternal exposure. Although the study authors
23 used two different exposure concentrations, the exposures were not run concurrently nor was
24 there information provided on exposure methods to indicate the studies were conducted similarly.

25 Reproductive effects

26 Reproductive endpoints, such as sex organ weights, estrous cycle length, and sperm effects
27 were examined following either oral or inhalation exposure in three subchronic studies ([Lyondell
28 Chemical Co., 2004](#); [NTP, 1997, 1995](#))(Table 1-8;Figure 1-6;Figure 1-7). The only reproductive
29 effect noted was an increase in estrous cycle length. Estrous cycle length was increased (28%
30 increase, $p < 0.01$) in female mice orally exposed to 11,620 mg/kg-day. No significant changes in
31 estrous cycle length were observed following oral exposure in rats, or inhalation exposure in mice
32 or rats.

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Table 1-8. Evidence pertaining to reproductive effects in animals following exposure to *tert*-butanol

Reference and study design	Results
<i>Male reproductive effects</i>	
<p>Lyondell Chemical Co. (2004)</p> <p>Sprague-Dawley rat; 12/sex/treatment Gavage 0, 64, 160, 400, or 1,000 mg/kg-d F0 males: 9 weeks beginning 4 weeks prior to mating F0 females: 4 weeks prior to mating through PND21</p>	<p>F0 reproductive effects No significant effect on weights of male reproductive organs or sperm observed</p>
<p>NTP (1995)</p> <p>F344/N rat; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 230, 490, 840, 1,520, 3,610^a mg/kg-d F: 0, 290, 590, 850, 1,560, 3,620^a mg/kg-d 13 weeks</p>	<p>No significant effect on weights of male reproductive organs or sperm observed</p>
<p>NTP (1995)</p> <p>B6C3F₁ mouse; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 350, 640, 1,590, 3,940, 8,210^a mg/kg-d F: 0, 500, 820, 1,660, 6,430, 11,620^a mg/kg-d 13 weeks</p>	<p>No significant effect on weights of male reproductive organs or sperm observed</p>
<p>NTP (1997)</p> <p>F344/N rat; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported</p>	<p>No significant effect on weights of male reproductive organs or sperm observed</p> <p>Evaluations were performed only for concentrations ≥542 ppm (1,643 mg/m³)</p>
<p>NTP (1997)</p> <p>B6C3F₁ mouse; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported</p>	<p>No significant effect on weights of male reproductive organs or sperm observed</p> <p>Evaluations were performed only for concentrations ≥542 ppm (1,643 mg/m³)</p>

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Table 1-8. Evidence pertaining to reproductive effects in animals following exposure to tert-butanol (continued)

Reference and study design	Results
<i>Female reproductive effects</i>	
<p>Lyondell Chemical Co. (2004)</p> <p>Sprague-Dawley rat; 12/sex/treatment Gavage 0, 64, 160, 400, or 1,000 mg/kg-d F0 males: 9 weeks beginning 4 weeks prior to mating F0 females: 4 weeks prior to mating through PND21</p>	<p>Pregnancy index</p> <p>91.7% 91.7% 100% 100% 91.7%</p>
<p>NTP (1995)</p> <p>F344/N rat; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 230, 490, 840, 1,520, 3,610^a mg/kg-d F: 0, 290, 590, 850, 1,560, 3,620^a mg/kg-d 13 weeks</p>	<p>No significant effect on female estrous cycle (0, -2, -4, 0, +8 % change relative to control)</p>
<p>NTP (1995)</p> <p>B6C3F₁ mouse; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 350, 640, 1,590, 3,940, 8,210^a mg/kg-d F: 0, 500, 820, 1,660, 6,430, 11,620^a mg/kg-d 13 weeks</p>	<p>↑ length of estrous cycle <i>Response relative to control:</i> 0, +5, +5, +5, +6, +28*%</p>
<p>NTP (1997)</p> <p>F344/N rat; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported</p>	<p>No significant effect on female estrous cycle (0, -4, +2, +4 % change relative to control)</p> <p>Evaluations were performed only for concentrations ≥542 ppm (1,643 mg/m³)</p>
<p>NTP (1997)</p> <p>B6C3F₁ mouse; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported</p>	<p>No significant effect on female estrous cycle (0, -3, -9, -5 % change relative to control)</p> <p>Evaluations were only performed for concentrations ≥542 ppm (1,643 mg/m³)</p>

- 1 * Statistically significant $p \leq 0.05$ as determined by the study authors.
- 2 Conversions from drinking water concentrations to mg/kg-d performed by study authors.
- 3 Conversion from ppm to mg/m³ is 1 ppm = 3.031 mg/m³.
- 4 Percentage change compared to control = (treated value – control value) ÷ control value × 100.
- 5

1 Developmental effects

2 Data from three developmental studies (two oral, one inhalation) suggest that fetal effects
3 are generally observed at doses that cause toxicity in the dams as measured by clinical signs (e.g.,
4 decreased body weight gain, and/or food consumption) (Table 1-9; Figure 1-6; Figure 1-7).

5 Developmental effects of *tert*-butanol observed after oral exposure (liquid diets or gavage)
6 in several mouse strains and one rat strain include measures of fetal loss or viability (e.g., increased
7 number of resorptions, decreased numbers of neonates per litter) and decreased fetal body weight
8 ([Lyondell Chemical Co., 2004](#); [Faulkner et al., 1989](#); [Daniel and Evans, 1982](#)). [Daniel and Evans](#)
9 [\(1982\)](#) also observed decreases in body weight gain during PND 2–10; however, data suggest that
10 the effect may be due to maternal behavior or nutritional status. In addition, one study reported
11 increased incidence of variations of the skull or sternebrae in two mouse strains, but the difference
12 was not statistically significant ([Faulkner et al., 1989](#)). Similar developmental effects were also
13 observed after whole-body inhalation exposure in Sprague-Dawley rats for 7 hours/day on GDs 1–
14 19 ([Nelson et al., 1989](#)). Fetal effects included concentration-related reductions in body weight in
15 male and female fetuses and higher incidence of skeletal variations when analyzed on the basis of
16 individual fetuses (but not on a per litter basis). In contrast to the oral exposure studies in mice
17 and rats, however, there was no effect on measures of fetal loss.

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1 **Table 1-9. Evidence pertaining to developmental effects in animals following**
 2 **exposure to *tert*-butanol**

Reference and study design	Results																																			
<p>Daniel and Evans (1982)</p> <p>Swiss Webster (Cox) mouse; 15 pregnant dams/treatment Liquid diet (0, 0.5, 0.75, 1.0%, w/v) 0 (isocaloric amounts of maltose/dextrin), 3,324, 4,879, 6,677 mg/kg-d GD 6–20</p>	<p>No statistical analysis was conducted on any of these data</p> <p>Maternal</p> <p>Percent change compared to control:</p> <table border="1" data-bbox="690 499 1421 808"> <thead> <tr> <th><u>Dose</u> (mg/kg-d)</th> <th><u>Food consumption</u> (mean g/animal/day)</th> <th><u>Body weight</u> gain</th> <th><u>Number of litters</u> (% pregnant dams)</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>0</td> <td>0</td> <td>11 (77%)</td> </tr> <tr> <td>3,324</td> <td>+2</td> <td>-3</td> <td>12 (80%)</td> </tr> <tr> <td>4,879</td> <td>-3</td> <td>-19</td> <td>8 (53%)</td> </tr> <tr> <td>6,677</td> <td>-4</td> <td>-20</td> <td>7 (47%)</td> </tr> </tbody> </table> <p>Authors note that lower food consumption in higher <i>tert</i>-butanol dose groups reflects problems with pair feeding and maternal sedation.</p> <p>Fetal</p> <p>Percent change compared to control:</p> <table border="1" data-bbox="690 1024 1230 1281"> <thead> <tr> <th><u>Dose</u> (mg/kg-d)</th> <th><u>Number of neonates/litter</u></th> <th><u>Fetal body weight</u> on PND 2</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>0</td> <td>0</td> </tr> <tr> <td>3,324</td> <td>-1</td> <td>-7</td> </tr> <tr> <td>4,879</td> <td>-29</td> <td>-19</td> </tr> <tr> <td>6,677</td> <td>-49</td> <td>-38</td> </tr> </tbody> </table> <p>Number of stillborn also increased with dose (3, 6, 14, and 20, respectively), but the number of stillborn per litter was not provided. The high dose also caused a delay in eye opening and a lag in weight gain during PND 2–10 (information was only provided in text or figures)</p>	<u>Dose</u> (mg/kg-d)	<u>Food consumption</u> (mean g/animal/day)	<u>Body weight</u> gain	<u>Number of litters</u> (% pregnant dams)	0	0	0	11 (77%)	3,324	+2	-3	12 (80%)	4,879	-3	-19	8 (53%)	6,677	-4	-20	7 (47%)	<u>Dose</u> (mg/kg-d)	<u>Number of neonates/litter</u>	<u>Fetal body weight</u> on PND 2	0	0	0	3,324	-1	-7	4,879	-29	-19	6,677	-49	-38
<u>Dose</u> (mg/kg-d)	<u>Food consumption</u> (mean g/animal/day)	<u>Body weight</u> gain	<u>Number of litters</u> (% pregnant dams)																																	
0	0	0	11 (77%)																																	
3,324	+2	-3	12 (80%)																																	
4,879	-3	-19	8 (53%)																																	
6,677	-4	-20	7 (47%)																																	
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0	0	0																																		
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4,879	-29	-19																																		
6,677	-49	-38																																		

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Table 1-9. Evidence pertaining to developmental effects in animals following exposure to *tert*-butanol (continued)

Reference and study design	Results																				
<p>Faulkner et al. (1989)</p> <p>CBA/J mouse; 7 pregnant females in control, 12 pregnant females in treated Gavage (10.5 mmoles/kg twice a day); 0 (tap water), 1,556 mg/kg-d GD 6–18</p>	<p>Maternal results not reported.</p> <p>Fetal</p> <table border="0"> <tr> <td></td> <td colspan="2">Percent change compared to control:</td> <td colspan="2">Incidence:</td> </tr> <tr> <td><u>Dose</u> (mg/kg-d)</td> <td><u>Live fetuses/litter</u></td> <td><u>Fetal weight</u></td> <td><u>Sternebral variations</u></td> <td><u>Skull variations</u></td> </tr> <tr> <td>0</td> <td>0</td> <td>0</td> <td>4/28</td> <td>1/28</td> </tr> <tr> <td>1,556</td> <td>-41*</td> <td>-4</td> <td>7/30</td> <td>3/30</td> </tr> </table> <p>Sternal variations: misaligned or unossified sternebrae Skull variations: moderate reduction in ossification of supraoccipital bone</p> <p>Number of total resorptions (10 resorptions/66 implants in controls, 37/94 implants in treated) and resorptions per litter (+118%) increased ($p < 0.05$)</p>		Percent change compared to control:		Incidence:		<u>Dose</u> (mg/kg-d)	<u>Live fetuses/litter</u>	<u>Fetal weight</u>	<u>Sternebral variations</u>	<u>Skull variations</u>	0	0	0	4/28	1/28	1,556	-41*	-4	7/30	3/30
	Percent change compared to control:		Incidence:																		
<u>Dose</u> (mg/kg-d)	<u>Live fetuses/litter</u>	<u>Fetal weight</u>	<u>Sternebral variations</u>	<u>Skull variations</u>																	
0	0	0	4/28	1/28																	
1,556	-41*	-4	7/30	3/30																	
<p>Faulkner et al. (1989)</p> <p>C57BL/6J mouse; 5 pregnant females in controls, 9 pregnant females treated Gavage (10.5 mmoles/kg twice a day) 0 (tap water), 1,556 mg/kg-d GD 6–18</p>	<p>Maternal results not reported.</p> <p>Fetal</p> <table border="0"> <tr> <td></td> <td colspan="2">Percent change compared to control:</td> <td colspan="2">Incidence:</td> </tr> <tr> <td><u>Dose</u> (mg/kg-d)</td> <td><u>Live fetuses/litter</u></td> <td><u>Fetal weight</u></td> <td><u>Sternal variations</u></td> <td><u>Skull variations</u></td> </tr> <tr> <td>0</td> <td>0</td> <td>0</td> <td>5/21</td> <td>1/21</td> </tr> <tr> <td>1,556</td> <td>-58%*</td> <td>-4</td> <td>9/16</td> <td>7/16</td> </tr> </table> <p>Sternal variations: misaligned or unossified sternebrae Skull variations: moderate reduction in ossification of supraoccipital bone</p> <p>Number of total resorptions (4 resorptions/44 implants in controls, 38/68 implants in treated) and resorptions per litter (+428%) increased ($p < 0.05$)</p>		Percent change compared to control:		Incidence:		<u>Dose</u> (mg/kg-d)	<u>Live fetuses/litter</u>	<u>Fetal weight</u>	<u>Sternal variations</u>	<u>Skull variations</u>	0	0	0	5/21	1/21	1,556	-58%*	-4	9/16	7/16
	Percent change compared to control:		Incidence:																		
<u>Dose</u> (mg/kg-d)	<u>Live fetuses/litter</u>	<u>Fetal weight</u>	<u>Sternal variations</u>	<u>Skull variations</u>																	
0	0	0	5/21	1/21																	
1,556	-58%*	-4	9/16	7/16																	

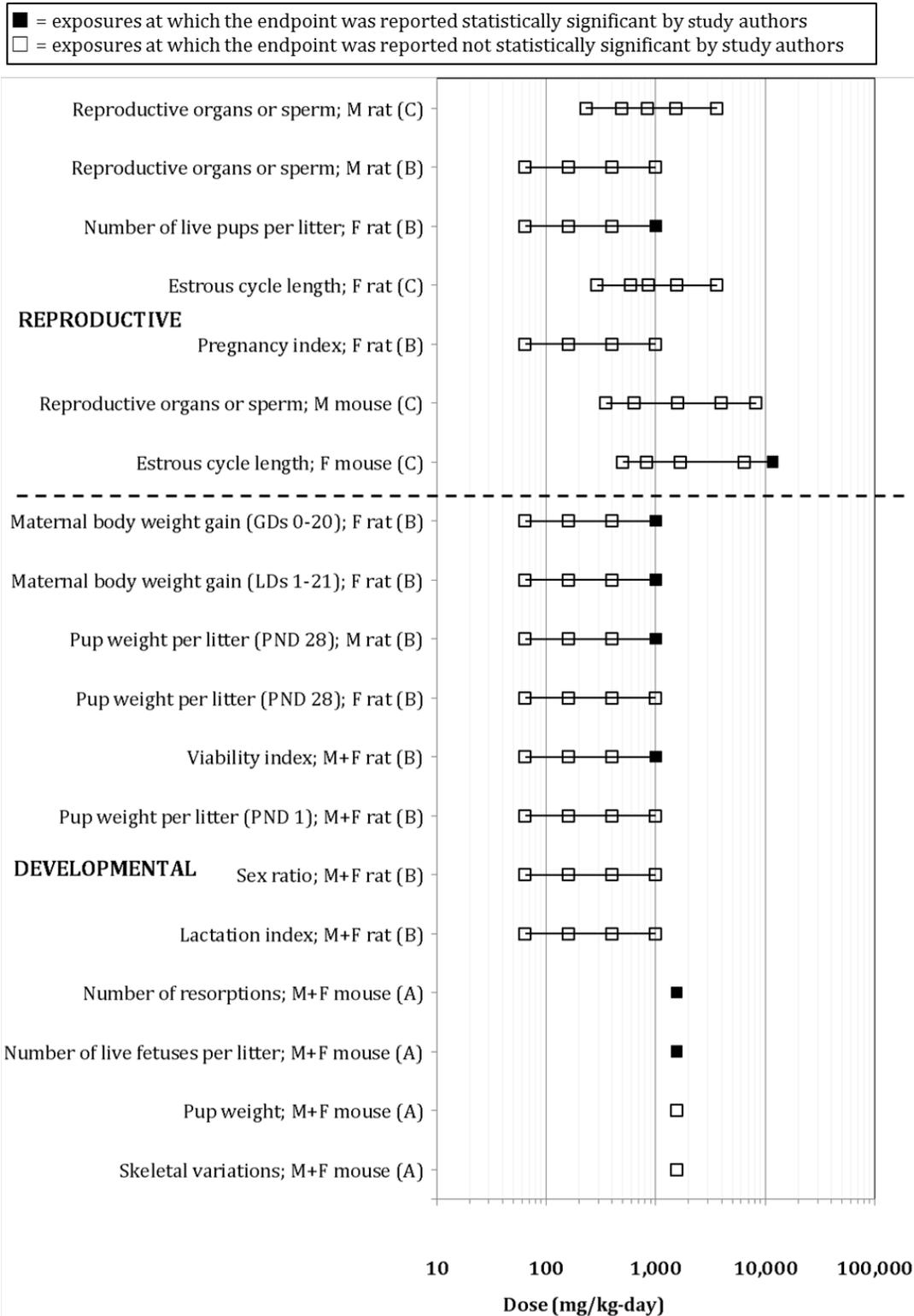
Table 1-9. Evidence pertaining to developmental effects in animals following exposure to tert-butanol (continued)

Reference and study design	Results				
<p>Lyondell Chemical Co. (2004)</p> <p>OECD guideline 421 study:</p> <p>Sprague-Dawley rat; 12/sex/treatment Gavage 0, 64, 160, 400, or 1,000 mg/kg-d F0 males: 9 weeks beginning 4 weeks prior to mating F0 females: 4 weeks prior to mating through PND21 F1 Males and Females: 7 weeks (throughout gestation and lactation; 1 male and 1 female from each litter was dosed directly from PND 21-28)</p>	<p>Response relative to control</p> <p><u>Dose</u> (mg/kg-d) 0 64 160 400 1000</p> <p>Maternal effects</p> <p>Body weight gain GD 0-20</p> <p>0 -3 -4 0 -16*</p> <p>Food consumption GD 0-20</p> <p>0 0 0 +4 0</p> <p>Body weight gain PND 1-21</p> <p>0 +3 -10 +3 +100*</p> <p>Food consumption LD1-14</p> <p>0 -2 -6 0 -16</p> <p>Live pups/litter <i>response relative to control</i></p> <p>0 -9 -11 -7 -33*</p> <p><u>Dams dosed with 1000 or 400 mg/kg/d showed CNS effects (e.g., ataxia, lethargy) which became undetectable by 4-weeks of exposure in animals exposed to 400 mg/kg/d but not those in the higher dose group.</u></p> <p>F1 effects</p> <p>Viability index (pup survival to PND4)</p> <p>96.4% 98.7% 98.2% 99.4% 74.1%*</p> <p>Lactation index (pup survival to PND21)</p> <p>100% 100% 100% 99.2% 98.8%</p> <p>Sex ratio (% males)</p> <p>54.4 52.3 50.9 53.4 52.1</p> <p>Pup weight/litter PND 1 relative to control (%)</p> <p>0 +6 +4 +7 -10</p> <p>Pup weight PND 28 relative to control (%)</p> <p>M: 0 +2 0 0 -12*</p> <p>F: 0 0 -4 -2 -8</p>				
<p>Nelson et al. (1989)</p> <p>Sprague-Dawley rat; 15 pregnant dams/treatment</p>	<p>Maternal: Unsteady gait (no statistical tests reported), dose-dependent ↓ in body weight gain (results presented in figure only), dose-dependent ↓ in food consumption ranging from 7–36% depending on dose and time</p> <p>Fetal</p>				

Table 1-9. Evidence pertaining to developmental effects in animals following exposure to tert-butanol (continued)

Reference and study design	Results				
Analytical concentration: 0, 2,200, 3,510, 5,030 ppm (0, 6,669, 10,640, 15,248 mg/m ³), (dynamic whole body chamber) 7 hr/d GD 1–19 Generation method, analytical concentration and method were reported	Percent change compared to control:				
	<u>Dose</u> (mg/m ³)	<u>Number of live fetuses/litter</u>	<u>Resorptions per litter</u>		
	0	0	0		
	6,669	0	+9		
	10,640	+15	-18		
	15,248	+8	0		
	Percent change compared to control:				
				Incidence:	
	<u>Dose</u> (mg/m ³)	<u>Fetal weight (males)</u>	<u>Fetal weight (females)</u>	<u>Skeletal variation by litter</u>	<u>Skeletal variation by fetus</u>
	0	0	0	10/15	18/96
6,669	-9*	-9*	14/17	35/104	
10,640	-12*	-13*	14/14	53/103*	
15,248	-32*	-31*	12/12	76/83*	
Skeletal variation by litter refers to the number of variations observed in the number of litters examined. Skeletal variation by fetus refers to the number of variations observed in the total number of fetuses examined. Fetuses are not categorized by litter.					

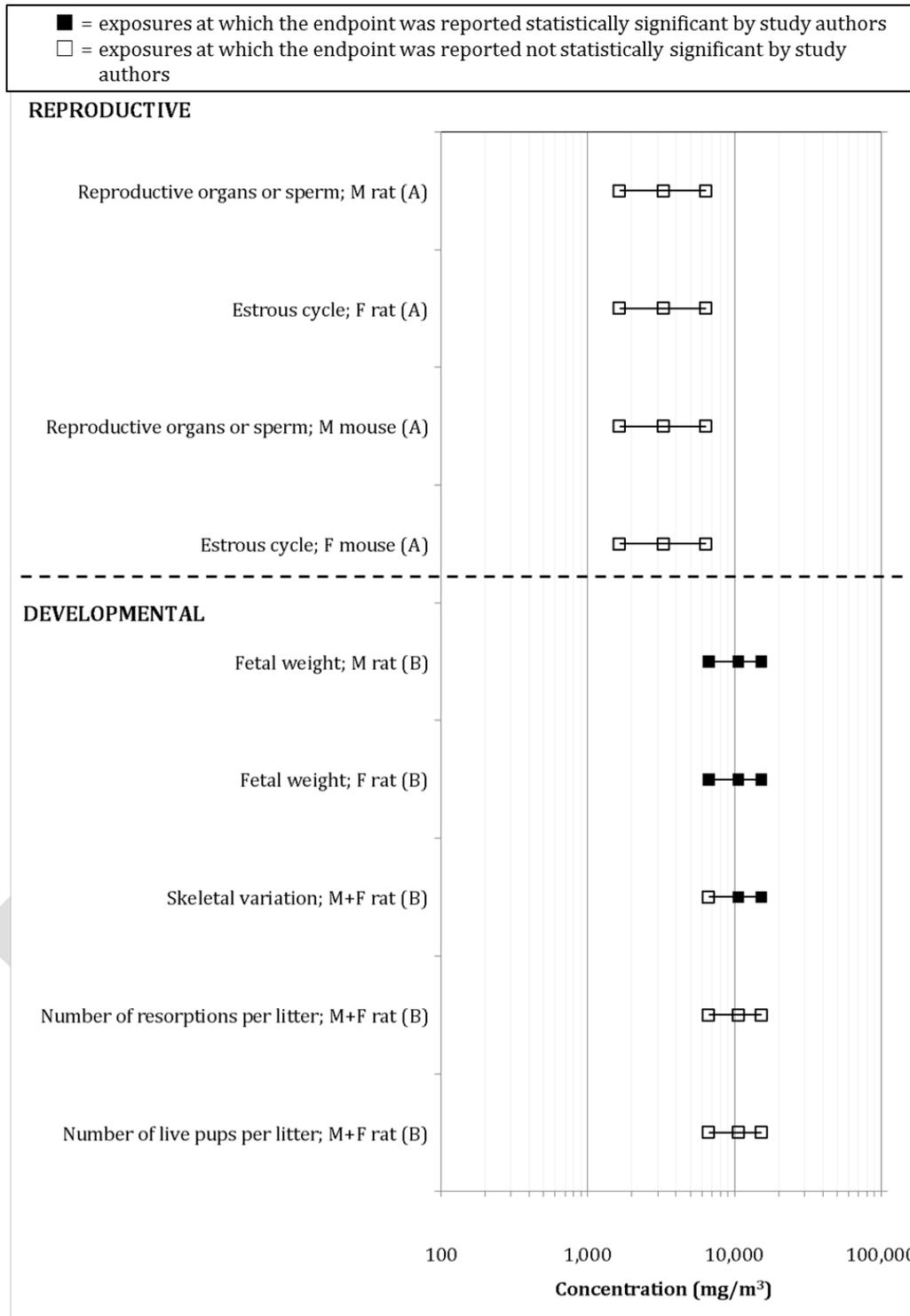
1 * Statistically significant $p \leq 0.05$ as determined by study authors.
 2 Conversions from diet concentrations to mg/kg-d performed by study authors.
 3 Conversion from ppm to mg/m³ is 1 ppm = 3.031 mg/m³.
 4 Percentage change compared to control = (treated value – control value) ÷ control value × 100.
 5
 6
 7
 8
 9



Sources: (A) [Faulkner et al. \(1989\)](#); (B) [Lyondell Chemical Co. \(2004\)](#); (C) [NTP \(1995\)](#)

Figure 1-6. Exposure-response array of reproductive and developmental effects following oral exposure to *tert*-butanol.

1



Sources: (A) [NTP \(1997\)](#); (B) [Nelson et al. \(1989\)](#)

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Figure 1-7. Exposure-response array of reproductive and developmental effects following inhalation exposure to *tert*-butanol.

1 Neurodevelopmental Effects

2 In addition to the developmental effects noted above, neurodevelopmental effects also have
3 been observed. This includes changes in rotarod performance following oral or inhalation
4 exposures, as well as decreases in open field behavior and cliff avoidance following oral exposure,
5 and reduced time hanging on wire after inhalation exposure during gestation (Table 1-10; Figure
6 1-6; Figure 1-7).

7 *Rotarod performance*

8 Looking across studies, not all the findings were consistent. While [Daniel and Evans \(1982\)](#)
9 found decreased rotarod performance in mouse pups of dams orally exposed during gestation,
10 [Nelson et al. \(1991\)](#) observed an increase in rotarod performance in rat pups of dams exposed via
11 inhalation during gestation.

12 *Neurochemical measurements*

13 In addition to behavioral effects, one study evaluated biochemical or physiological changes
14 in the brain of offspring exposed during gestation or early in the postnatal period. [Nelson et al.](#)
15 [\(1991\)](#) found statistically significant changes in neurochemical measurements in the brain in
16 offspring of dams exposed via inhalation during gestation; however, the two concentrations tested
17 were not run concurrently, and very little data were provided.

18 *Physiological and psychomotor development*

19 Data also suggest that neurodevelopmental effects were not solely due to in utero exposure
20 ([Daniel and Evans, 1982](#)). [Daniel and Evans \(1982\)](#) cross-fostered half of the mouse pups born to
21 treated mother with untreated surrogate females to test the effects of maternal nutrition and
22 behavioral factors on the pups' physiological and psychomotor development. Results indicated that
23 pups fostered with control dams performed significantly better than those maintained with treated
24 dams (Table 1-10) ([Daniel and Evans, 1982](#)). Results were only presented in figures and were not
25 compared with controls.
26

1 **Table 1-10. Evidence pertaining to neurodevelopmental effects in animals**
 2 **following exposure to *tert*-butanol**

Reference and study design	Results
<p>Daniel and Evans (1982)</p> <p>Liquid diet (0, 0.5, 0.75, or 1.0%, w/v); GD6–20; Swiss Webster (Cox) mouse; 15 pregnant dams/treatment; after birth half the pups were nursed with their treated dams and the other half were fostered by untreated dams who recently gave birth</p> <p>0 (isocaloric amounts of maltose/dextrin), 3,324, 4,879, or 6,677 mg/kg-d</p>	<ul style="list-style-type: none"> • a dose-dependent increase righting reflex time, with more time needed in animals maintained with maternal dams • a dose-dependent decrease in open field behavior, with less activity in pups maintained with maternal dams • a dose-dependent decrease in rotarod performance with the pups from maternal dams having lower performances • a dose-dependent decrease in the amount of time the pups were able to avoid a cliff, with animals maintained with their maternal dams having less avoidance time
<p>Nelson et al. (1991)</p> <p>Sprague-Dawley rat; 15 pregnant dams/treatment</p> <p>Analytical concentration: 0, 6,000, or 12,000 mg/m³; (dynamic whole body chamber) 7 hr/d</p> <p>GD 1–19</p> <p>Generation method, analytical concentration and method were reported</p>	<p>Data were not presented specifically by dose nor were any tables or figures of the data provided</p> <p>Maternal toxicity was noted by decreased food consumption and body weight gains</p> <p>Results in offspring</p> <ul style="list-style-type: none"> • increase in rotarod performance in high-dose group (16 versus 26 revolutions/min for controls and 12,000 mg/m³ animals, respectively) • decreased time held on wire in the performance ascent test in the low-dose group (16 sec versus 10 sec for controls and 1,750 mg/m³ animals, respectively) <p>The following differences in neurochemical measurements in the brain between control and treated offspring were observed,</p> <ul style="list-style-type: none"> • 53% decrease in norepinephrine in the cerebellum at 12,000 mg/m³ • 57% decrease in met-enkephalin in the cerebrum at 12,000 mg/m³ and 83% decrease at 6,000 mg/m³ • 61% decrease in β-endorphin in the cerebellum at 12,000 mg/m³ • 67% decrease in serotonin in the midbrain at 6,000 mg/m³

3

Table 1-10. Evidence pertaining to neurodevelopmental effects in animals following exposure to *tert*-butanol (*continued*)

Reference and study design	Results
<p>Nelson et al. (1991)</p> <p>adult male Sprague-Dawley rats (18/treatment) mated to untreated females</p> <p>Analytical concentration: 0, 6,000, or 12,000 mg/m³; (dynamic whole body chamber) 7 hr/d for 6 wk</p> <p>Generation method, analytical concentration and method were reported</p>	<p>Data were not presented specifically by dose nor were any tables or figures of the data provided</p> <p>Results (generally only specified as paternally treated versus controls) in offspring indicate</p> <ul style="list-style-type: none"> • increase in rotarod performance (16 versus 20 revolutions/min for controls and 12,000 mg/m³ animals, respectively) • decreased time in open field (less time to reach the outer circle of the field, 210 sec versus 115 seconds for controls and 12,000 mg/m³ animals, respectively) <p>The following differences in neurochemical measurements in the brain between control and treated offspring were observed</p> <ul style="list-style-type: none"> • 39% decrease in norepinephrine in the cerebellum at 12,000 mg/m³ • 40% decrease in met-enkephalin in the cerebrum at 12,000 mg/m³ and 75% decrease at 6,000 mg/m³ • 71% decrease in β-endorphin in the cerebellum at 12,000 mg/m³ • 47% decrease in serotonin in the midbrain at 6,000 mg/m³

1 * Statistically significant $p \leq 0.05$ as determined by study authors.
 2 Conversions from diet concentrations to mg/kg-d performed by study authors.
 3 Percentage change compared to control = (treated value – control value) ÷ control value × 100.
 4

5 **Mechanistic Evidence**

6 No mechanistic evidence is available for reproductive or developmental effects, including
 7 neurodevelopmental effects.

8 **Summary of Reproductive and Developmental Toxicity**

9 EPA concluded that the evidence does not support reproductive effects as a potential
 10 human hazard of *tert*-butanol exposure. There are no two-generation reproductive studies
 11 available by oral or inhalation exposure. Two oral exposure studies ([Lyondell Chemical Co., 2004](#);
 12 [NTP, 1995](#)) and one subchronic inhalation study ([NTP, 1997](#)) are available. Overall, reproductive
 13 effects observed due to exposure to *tert*-butanol were limited to altered length of estrous cycle
 14 ([NTP, 1995](#)), but there is no available information to infer how this effect may influence
 15 reproductive ability.

16 EPA identified suggestive evidence of developmental effects as a potential human hazard of
 17 *tert*-butanol exposure. Exposure during gestation resulted in increased fetal loss, decreased fetal
 18 body weight, and possible increases in skeletal variations in exposed offspring or pups, although
 19 effects were not always consistent across exposure routes (oral and inhalation). Dams had
 20 decreased body weight and/or body weight gains, decreased food consumption, and/or clinical
 21 signs of intoxication at the same doses that *tert*-butanol caused fetal effects. Neurodevelopmental

1 effects including decreased brain weight, changes in brain biochemistry, and changes in behavioral
2 performances have also been observed. Each of the neurodevelopmental studies, however, had
3 limitations in the study design and/or reporting. In addition, results from the neurodevelopmental
4 studies were not always consistent between studies or across dose.

5 **1.1.4. Carcinogenicity (other than in the kidney or thyroid)**

6 ***Synthesis of Carcinogenicity Data (Other than in the Kidney or Thyroid)***

7 This section reviews the studies that investigated whether exposure to *tert*-butanol can
8 cause cancers (other than in the kidney or thyroid) in humans or animals. The database examining
9 carcinogenicity following *tert*-butanol exposure contains no human data and two chronic studies,
10 one in rats and one in mice. As mentioned in the Study Selection, the studies providing data on
11 carcinogenicity exposed animals via drinking water for ≥ 30 days. Shorter duration studies do not
12 generally evaluate carcinogenicity, but any shorter duration studies that examined carcinogenicity
13 are discussed in the text if they provide data to support mode of action or hazard identification. No
14 methodological concerns were identified that would lead one or more studies to be considered less
15 informative for assessing human health hazard.

16 Kidney and thyroid tumors are presented above with the specific organ hazard
17 identification. No other treatment-related changes in tumors in other organs were noted in the 2-
18 year oral rat or mouse studies conducted by [NTP \(1995\)](#), which evaluated a comprehensive set of
19 tissues/organs. There is no 2-year inhalation study.

20 ***Mechanistic Evidence***

21 Available mechanistic evidence was previously discussed in the context of kidney and
22 thyroid tumors (Sections 1.1.1 and 1.1.2).

23 ***Summary of Carcinogenicity Evidence***

24 There are limited data available on the carcinogenicity of *tert*-butanol. There are 2-year
25 oral studies in one strain of rats and one strain of mice, but no 2-year inhalation studies. EPA
26 identified suggestive evidence of kidney and thyroid tumors as a potential human hazard.

27 **1.1.5. Other Toxicological Effects**

28 ***Synthesis of Other Toxicity Data***

29 The database for effects other than kidney, thyroid, reproductive, developmental (including
30 neurodevelopmental) and cancer contain only 14 rodent studies. As previously mentioned in the
31 Study Selection, all selected studies employed inhalation, oral gavage, or drinking water exposures
32 for ≥ 30 days. Studies employing short term and acute exposures that examined other toxicological
33 effects are not included in the evidence tables; however, they are discussed in the text if they

1 provide data to support mode of action or hazard identification. No studies were removed for
2 methodological concerns.

3 *tert*-Butanol also has been found to have CNS effects similar to ethanol in terms of animals
4 appearing intoxicated and having withdrawal symptoms after cessation of oral or inhalation
5 exposure. Severity of CNS symptoms such as withdrawal increased with dose and duration of
6 exposure. However, study quality concerns (e.g., short exposure durations, lack of data reporting,
7 small number of animals per treatment group) associated with all of the studies in the database
8 preclude a clear understanding of potential neurotoxicity following *tert*-butanol exposure, and
9 therefore, CNS studies are not presented in evidence tables.

10 Effects in other tissues were observed with less consistency. These include decreased body
11 weight, liver effects, and urinary bladder effects.

12 Body weight

13 Body weight was decreased by >10% in both rats and mice with subchronic and chronic
14 exposure, with males generally more affected than females (Table 1-11). The concentrations used
15 in the subchronic inhalation study did not decrease body weights. However, a short-term (i.e., 18-
16 day) inhalation study in rats observed a >10% decrease in body weight at concentrations about
17 threefold higher (in mg/m³) than the highest concentration used in the subchronic study. The same
18 concentrations did not have any effect on the body weight in mice with short-term inhalation
19 exposure.

20 Liver effects

21 Although some rodent studies observed statistically significant changes in relative liver
22 weight with *tert*-butanol exposure, absolute liver weight was significantly increased only in female
23 rats after subchronic oral exposure (Table 1-12). The results pertaining to histopathology changes
24 were inconsistent (Table 1-13). The oral [NTP \(1995\)](#) subchronic and chronic studies did not
25 observe treatment-related effects on liver histopathology in both sexes of F344 rats, but in a 10-
26 week study in a different rat strain (Wistar rats), several liver lesions (including necrosis) and
27 increased liver glycogen were seen in male rats (no females were included in the study) with the
28 only dose used ([Acharya et al., 1997](#); [Acharya et al., 1995](#)). The study did not provide any incidence
29 or severity data. The dose used in this study was in the range of the lower doses used in the [NTP](#)
30 [\(1995\)](#) study. In the developmental study by [Lyondell Chemical Co. \(2004\)](#), the F1 SD rats treated
31 by *tert*-butanol for at least 9 weeks did not show any liver effects. An increased incidence of fatty
32 liver was observed in the male mice of the highest dose group in the 2-year mouse bioassay, but no
33 histopathologic changes were seen in the subchronic mouse study. No changes in liver
34 histopathology were observed in the [NTP \(1997\)](#) subchronic inhalation study.

1 Urinary bladder effects

2 Several studies also reported effects in the urinary bladder (Table 1-14). Transitional
3 epithelial hyperplasia was observed in male rats and mice after 13 weeks of exposure at doses of
4 3,610 mg/kg-day (male rats) and ≥ 3940 mg/kg-day (male mice). Male mice exposed at doses of
5 2,070 mg/kg-day for 2 years of also exhibited transitional epithelial hyperplasia. Neither female
6 rats nor female mice showed increased incidences of transitional epithelial hyperplasia. Both sexes
7 of mice demonstrated incidence of inflammation in the urinary bladder after both subchronic and
8 chronic exposures, with a greater incidence in males compared to females.

9 An exposure-response array of these effects in body weight, liver, and urinary bladder is
10 provided in Figure 1-8 and Figure 1-9 for oral and inhalation studies, respectively.

11 ***Mechanistic Evidence***

12 No mechanistic evidence is available for these other toxicological effects.

13 ***Summary of Other Toxicity Data***

14 EPA concluded that the evidence does not support body weight changes, liver effects, and
15 urinary bladder effects as potential human hazards of *tert*-butanol exposure.
16

1
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Table 1-11. Evidence pertaining to effects on body weight in animals following exposure to *tert*-butanol

Reference and study design	Results																																
<p>Acharya et al. (1995)</p> <p>Wistar rat; 5–6 males/treatment Drinking water (0 or 0.5%), 0 or 575 mg/kg-d 10 weeks</p>	<p>Body weight in treated animals lower than controls by ~7% (p< 0.05); (results only provided in a figure)</p>																																
<p>Lyondell Chemical Co. (2004)</p> <p>Sprague-Dawley rat; 12/sex/treatment Gavage 0, 64, 160, 400, or 1,000 mg/kg-d F0 males: 9 weeks beginning 4 weeks prior to mating F0 females: 4 weeks prior to mating through PND21</p>	<p>Percent change compared to control:</p> <table border="1"> <thead> <tr> <th colspan="2">F0 Males</th> <th colspan="2">F0 Females</th> </tr> <tr> <th>Dose (mg/kg-d)</th> <th>Body weight</th> <th>Dose (mg/kg-d)</th> <th>Body weight</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>0</td> <td>0</td> <td>0</td> </tr> <tr> <td>64</td> <td>-2</td> <td>64</td> <td>0</td> </tr> <tr> <td>160</td> <td>-4</td> <td>160</td> <td>-2</td> </tr> <tr> <td>400</td> <td>+2</td> <td>400</td> <td>+1</td> </tr> <tr> <td>1,000</td> <td>-7</td> <td>1,000</td> <td>+4</td> </tr> </tbody> </table>	F0 Males		F0 Females		Dose (mg/kg-d)	Body weight	Dose (mg/kg-d)	Body weight	0	0	0	0	64	-2	64	0	160	-4	160	-2	400	+2	400	+1	1,000	-7	1,000	+4				
F0 Males		F0 Females																															
Dose (mg/kg-d)	Body weight	Dose (mg/kg-d)	Body weight																														
0	0	0	0																														
64	-2	64	0																														
160	-4	160	-2																														
400	+2	400	+1																														
1,000	-7	1,000	+4																														
<p>NTP (1995)</p> <p>F344/N rat; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 230, 490, 840, 1,520, 3,610^a mg/kg-d F: 0, 290, 590, 850, 1,560, 3,620^a mg/kg-d 13 weeks</p>	<p>Percent change compared to control:</p> <table border="1"> <thead> <tr> <th colspan="2">Males</th> <th colspan="2">Females</th> </tr> <tr> <th>Dose (mg/kg-d)</th> <th>Body weight</th> <th>Dose (mg/kg-d)</th> <th>Body weight</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>0</td> <td>0</td> <td>0</td> </tr> <tr> <td>230</td> <td>-4</td> <td>290</td> <td>+2</td> </tr> <tr> <td>490</td> <td>-5*</td> <td>590</td> <td>+1</td> </tr> <tr> <td>840</td> <td>-12*</td> <td>850</td> <td>+1</td> </tr> <tr> <td>1,520</td> <td>-17*</td> <td>1,560</td> <td>-2</td> </tr> <tr> <td>3,610</td> <td>All dead</td> <td>3,620</td> <td>-21*</td> </tr> </tbody> </table>	Males		Females		Dose (mg/kg-d)	Body weight	Dose (mg/kg-d)	Body weight	0	0	0	0	230	-4	290	+2	490	-5*	590	+1	840	-12*	850	+1	1,520	-17*	1,560	-2	3,610	All dead	3,620	-21*
Males		Females																															
Dose (mg/kg-d)	Body weight	Dose (mg/kg-d)	Body weight																														
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1,520	-17*	1,560	-2																														
3,610	All dead	3,620	-21*																														
<p>NTP (1995)</p> <p>B6C3F₁ mouse; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 350, 640, 1,590, 3,940, 8,210^a mg/kg-d F: 0, 500, 820, 1,660, 6,430, 11,620^a mg/kg-d 13 weeks</p>	<p>Percent change compared to control:</p> <table border="1"> <thead> <tr> <th colspan="2">Males</th> <th colspan="2">Females</th> </tr> <tr> <th>Dose (mg/kg-d)</th> <th>Body weight</th> <th>Dose (mg/kg-d)</th> <th>Body weight</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>0</td> <td>0</td> <td>0</td> </tr> <tr> <td>350</td> <td>-1</td> <td>500</td> <td>+3</td> </tr> <tr> <td>640</td> <td>+1</td> <td>820</td> <td>-1</td> </tr> <tr> <td>1,590</td> <td>-4</td> <td>1,660</td> <td>+4</td> </tr> <tr> <td>3,940</td> <td>-14*</td> <td>6,430</td> <td>-6</td> </tr> <tr> <td>8,210</td> <td>-24*</td> <td>11,620</td> <td>-15*</td> </tr> </tbody> </table> <p>High-dose females had a significantly lower initial weight, but also had a significantly lower body weight gain indicating that there was some effect of treatment</p>	Males		Females		Dose (mg/kg-d)	Body weight	Dose (mg/kg-d)	Body weight	0	0	0	0	350	-1	500	+3	640	+1	820	-1	1,590	-4	1,660	+4	3,940	-14*	6,430	-6	8,210	-24*	11,620	-15*
Males		Females																															
Dose (mg/kg-d)	Body weight	Dose (mg/kg-d)	Body weight																														
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1,590	-4	1,660	+4																														
3,940	-14*	6,430	-6																														
8,210	-24*	11,620	-15*																														

3

Table 1-11. Evidence pertaining to effects on body weight in animals following exposure to tert-butanol (continued)

Reference and study design	Results			
<p>NTP (1995)</p> <p>F344/N rat; 60/sex/treatment (10/sex/treatment evaluated at 15 months) Drinking water (0, 1.25, 2.5, 5, 10 mg/mL) M: 0, 90, 200, 420^a mg/kg-d F: 0, 180, 330, 650^a mg/kg-d 2 years</p>	Percent change compared to control:			
	Males		Females	
	<u>Dose</u> (mg/kg-d)	<u>Body weight</u>	<u>Dose</u> (mg/kg-d)	<u>Body weight</u>
	0	0	0	0
	90	-15	180	-2
	200	-18	330	-5
	420	-24	650	-21
	Only animals that survived at the end of 2 years were evaluated for body weight. Note: statistical significance not determined by study authors.			
<p>NTP (1995)</p> <p>B6C3F₁ mouse; 60/sex/treatment Drinking water (0, 5, 10, 20 mg/mL); M: 0, 540, 1,040, 2,070^a mg/kg-d F: 0, 510, 1,020, 2,110 mg/kg-d 2 years</p>	Percent change compared to control:			
	Males		Females	
	<u>Dose</u> (mg/kg-d)	<u>Body weight</u>	<u>Dose</u> (mg/kg-d)	<u>Body weight</u>
	0	0	0	0
	540	+1	510	-2
	1,040	-2	1,020	-3
	2,070	-1	2,110	-12
	Only animals that survived at the end of 2 years were evaluated for body weight. Note: statistical significance not determined by study authors.			
<p>NTP (1997)</p> <p>F344/N rat; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported</p>	Percent change compared to control:			
	<u>Concentration</u> (mg/m ³)	Males <u>Body weight</u>		Females <u>Body weight</u>
	0	0		0
	406	-1		-5
	824	-2		-1
	1,643	+2		0
	3,273	+3		0
	6,368	+2		-3
<p>NTP (1997)</p> <p>B6C3F₁ mouse; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and</p>	Percent change compared to control:			
	<u>Concentration</u> (mg/m ³)	Males <u>Body weight</u>		Females <u>Body weight</u>
	0	0		0
	406	+4		+3
	824	-2		-3
	1,643	0		+3
	3,273	-4		-6

Table 1-11. Evidence pertaining to effects on body weight in animals following exposure to *tert*-butanol (continued)

Reference and study design	Results		
method were reported	6,368	0	-8

- 1 ^aThere was a significant decrease in survival in the high-dose group.
- 2 * Statistically significant $p \leq 0.05$ as determined by study authors.
- 3 Conversions from drinking water concentrations to mg/kg-d performed by study authors.
- 4 Percentage change compared to control = (treated value – control value) ÷ control value × 100.
- 5

DRAFT

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Table 1-12. Changes in liver weight in animals following exposure to tert-butanol

Reference and study design	Results																																																
<p>Acharya et al. (1995) Wistar rat; 5–6 males/treatment Drinking water (0 or 0.5%), 0 or 575 mg/kg-d 10 weeks</p>	<p>No significant treatment-related effects (results were only provided in a figure)</p>																																																
<p>Lyondell Chemical Co. (2004) Sprague-Dawley rat; 12/sex/treatment Gavage 0, 64, 160, 400, or 1,000 mg/kg-d Males: 9 weeks beginning 4 weeks prior to mating Females: 4 weeks prior to mating through PND21</p>	<p>Percent change compared to control:</p> <table border="1"> <thead> <tr> <th colspan="3">Males</th> <th colspan="3">Females</th> </tr> <tr> <th>Dose (mg/kg-d)</th> <th>Absolute weight</th> <th>Relative weight</th> <th>Dose (mg/kg-d)</th> <th>Absolute weight</th> <th>Relative weight</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> </tr> <tr> <td>64</td> <td>-1</td> <td>0</td> <td>64</td> <td>-4</td> <td>-4</td> </tr> <tr> <td>160</td> <td>-3</td> <td>+1</td> <td>160</td> <td>-7</td> <td>-5</td> </tr> <tr> <td>400</td> <td>-2</td> <td>-1</td> <td>400</td> <td>+2</td> <td>+1</td> </tr> <tr> <td>1,000</td> <td>+8</td> <td>+16*</td> <td>1,000</td> <td>+8</td> <td>+3</td> </tr> </tbody> </table>	Males			Females			Dose (mg/kg-d)	Absolute weight	Relative weight	Dose (mg/kg-d)	Absolute weight	Relative weight	0	0	0	0	0	0	64	-1	0	64	-4	-4	160	-3	+1	160	-7	-5	400	-2	-1	400	+2	+1	1,000	+8	+16*	1,000	+8	+3						
Males			Females																																														
Dose (mg/kg-d)	Absolute weight	Relative weight	Dose (mg/kg-d)	Absolute weight	Relative weight																																												
0	0	0	0	0	0																																												
64	-1	0	64	-4	-4																																												
160	-3	+1	160	-7	-5																																												
400	-2	-1	400	+2	+1																																												
1,000	+8	+16*	1,000	+8	+3																																												
<p>NTP (1995) F344/N rat; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 230, 490, 840, 1,520, 3,610^a mg/kg-d F: 0, 290, 590, 850, 1,560, 3,620^a mg/kg-d 13 weeks</p>	<p>Percent change compared to control:</p> <table border="1"> <thead> <tr> <th colspan="3">Males</th> <th colspan="3">Females</th> </tr> <tr> <th>Dose (mg/kg-d)</th> <th>Absolute weight</th> <th>Relative weight</th> <th>Dose (mg/kg-d)</th> <th>Absolute weight</th> <th>Relative weight</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> <td>0</td> </tr> <tr> <td>230</td> <td>-2</td> <td>+4</td> <td>290</td> <td>+11*</td> <td>+9*</td> </tr> <tr> <td>490</td> <td>+1</td> <td>+8*</td> <td>590</td> <td>+10*</td> <td>+9*</td> </tr> <tr> <td>840</td> <td>+5</td> <td>+20*</td> <td>850</td> <td>+12*</td> <td>+11*</td> </tr> <tr> <td>1,520</td> <td>+8</td> <td>+31*</td> <td>1,560</td> <td>+15*</td> <td>+16*</td> </tr> <tr> <td>3,610</td> <td>All dead</td> <td>All dead</td> <td>3,620</td> <td>+9*</td> <td>+41*</td> </tr> </tbody> </table>	Males			Females			Dose (mg/kg-d)	Absolute weight	Relative weight	Dose (mg/kg-d)	Absolute weight	Relative weight	0	0	0	0	0	0	230	-2	+4	290	+11*	+9*	490	+1	+8*	590	+10*	+9*	840	+5	+20*	850	+12*	+11*	1,520	+8	+31*	1,560	+15*	+16*	3,610	All dead	All dead	3,620	+9*	+41*
Males			Females																																														
Dose (mg/kg-d)	Absolute weight	Relative weight	Dose (mg/kg-d)	Absolute weight	Relative weight																																												
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3,940	0	+14*	6,430	-2	+6																																												
8,210	-16	+22*	11,620	-6	+13*																																												

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Table 1-12. Changes in liver weight in animals following exposure to tert-butanol (continued)

Reference and study design	Results					
<p>NTP (1995)</p> <p>F344/N rat; 60/sex/treatment (10/sex/treatment evaluated at 15 months) Drinking water (0, 1.25, 2.5, 5 or 10 mg/mL) M: 0, 90, 200, or 420^a mg/kg-d F: 0, 180, 330, or 650^a mg/kg-d 2 years</p>	Percent change compared to control:					
	Males			Females		
	<u>Dose</u> (mg/kg-d)	<u>Absolute</u> <u>weight</u>	<u>Relative</u> <u>weight</u>	<u>Dose</u> (mg/kg-d)	<u>Absolute</u> <u>weight</u>	<u>Relative</u> <u>weight</u>
	0	0	0	0	0	0
	90	+2	+7	180	-14*	-8
	200	+8	+11	330	-3	-1
	420	+1	+14*	650	-6	+9*
	Only animals sacrificed at 15 months were evaluated for organ weights. Organ weights were not measured in the 2-year mouse study					
<p>NTP (1997)</p> <p>F344/N rat; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported</p>	Percent change compared to control:					
	Males			Females		
	<u>Concentration</u> (mg/m ³)	<u>Absolute</u> <u>weight</u>	<u>Relative</u> <u>weight</u>	<u>Absolute</u> <u>weight</u>	<u>Relative</u> <u>weight</u>	
	0	0	0	0	0	
	406	-8	-8	0	+3	
	824	-2	-1	0	0	
	1,643	+1	-1	+3	+2	
	3,273	+10	+7	+9	+9*	
	6,368	+5	+5	+4	+8*	
<p>NTP (1997)</p> <p>B6C3F₁ mouse; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported</p>	Percent change compared to control:					
	Males			Females		
	<u>Concentration</u> <u>on</u> (mg/m ³)	<u>Absolute</u> <u>weight</u>	<u>Relative</u> <u>weight</u>	<u>Absolute</u> <u>weight</u>	<u>Relative</u> <u>weight</u>	
	0	0	0	0	0	
	406	-1	0	+1	-4	
	824	+4	+9	+1	+5	
	1,643	+7	+5	+5	+1	
	3,273	-8	-2	+2	+9*	
	6,368	+5	+7	+8	+21*	

1 ^aThe high-dose group had an increase in mortality.
 2 * Statistically significant $p \leq 0.05$ as determined by study authors.
 3 Conversions from drinking water concentrations to mg/kg-d performed by study authors.

- 1 Conversion from ppm to mg/m³ is 1 ppm = 3.031 mg/m³.
 2 Percentage change compared to control = (treated value – control value) ÷ control value × 100.
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4 **Table 1-13. Changes in liver histopathology in animals following exposure to**
 5 ***tert*-butanol**

Reference and study design	Results
<p>Acharya et al. (1997; 1995)</p> <p>Wistar rat; 5–6 males/treatment Drinking water (0, 0.5%), 0, 575 mg/kg-d 10 weeks</p>	<p>↑ liver glycogen (~ 7 fold)*</p> <p>↑ incidence of centrilobular necrosis, vacuolation of hepatocytes, loss of hepatocyte architecture, peripheral proliferation, and lymphocyte infiltration (incidences and results of statistical tests not reported)</p>
<p>Lyondell Chemical Co. (2004)</p> <p>Sprague-Dawley rat; 12/sex/treatment Gavage 0, 64, 160, 400, or 1,000 mg/kg-d</p> <p>Males: 9 weeks beginning 4 weeks prior to mating Females: 4 weeks prior to mating through PND21</p>	<p>No treatment-related effects observed.</p>
<p>NTP (1995)</p> <p>F344/N rat; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, or 40 mg/mL) M: 0, 230, 490, 840, 1,520, 3,610^a mg/kg-d F: 0, 290, 590, 850, 1,560, 3,620^a mg/kg-d 13 weeks</p>	<p>Histopathology data for the 13-week study were not provided, but the liver was evaluated indicating that no changes in liver histopathology were observed in the 13-week study.</p>
<p>NTP (1995)</p> <p>B6C3F₁ mouse; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, 40 mg/mL) M: 0, 350, 640, 1,590, 3,940, 8,210^a mg/kg-d F: 0, 500, 820, 1,660, 6,430, 11,620^a mg/kg-d 13 weeks</p>	<p>Histopathology data for the 13-week study were not provided, but the liver was evaluated indicating that no changes in liver histopathology were observed in the 13-week study.</p>
<p>NTP NTP (1995)</p> <p>F344/N rat; 60/sex/treatment (10/sex/treatment evaluated at 15 months) Drinking water (0, 1.25, 2.5, 5, 10 mg/mL) M: 0, 90, 200, or 420^a mg/kg-d F: 0, 180, 330, or 650^a mg/kg-d 2 years</p>	<p>No treatment-related effects observed.</p>

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Table 1-13. Changes in liver histopathology in animals following exposure to tert-butanol (continued)

Reference and study design	Results			
	Males		Females	
	<u>Dose</u> (mg/kg-d)	<u>Incidence of fatty</u> <u>change</u>	<u>Dose</u> (mg/kg-d)	<u>Incidence of fatty</u> <u>change</u>
NTP (1995) B6C3F ₁ mouse; 60/sex/treatment Drinking water (0, 5, 10, 20 mg/mL) M: 0, 540, 1,040, or 2,070 ^a mg/kg-d F: 0, 510, 1,020, or 2,110 mg/kg-d 2 years	0	12/59	0	11/60
	540	5/60	510	8/60
	1,040	8/59	1,020	8/60
	2,070	29/59*	2,110	6/60
NTP (1997) F344/N rat; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m ³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported	Histopathology data for the 13-week study were not provided, but the liver was evaluated in control and high-dose group indicating that no changes in liver histopathology were observed in the 13-week study.			
NTP (1997) B6C3F ₁ mouse; 10/sex/treatment Analytical concentration: 0, 134, 272, 542, 1,080, or 2,101 ppm (0, 406, 824, 1,643, 3,273 or 6,368 mg/m ³) (dynamic whole body chamber) 6 hr/d, 5 d/wk 13 weeks Generation method (Sonimist Ultrasonic spray nozzle nebulizer), analytical concentration and method were reported	Authors stated that there were no treatment-related microscopic observations, but data were not provided.			

^aThe high-dose group had an increase in mortality.

* Statistically significant $p \leq 0.05$ as determined by study authors.

Conversions from drinking water concentrations to mg/kg-d performed by study authors.

Conversion from ppm to mg/m³ is 1 ppm = 3.031 mg/m³.

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1 **Table 1-14. Changes in urinary bladder histopathology in animals following**
 2 **oral exposure to *tert*-butanol**

Reference and study design	Results																																																											
<p>NTP (1995)</p> <p>F344/N rat; 10/sex/treatment Drinking water (0, 2.5, 5, 10, 20, 40 mg/mL) M: 0, 230, 490, 840, 1,520, 3,610^a mg/kg-d F: 0, 290, 590, 850, 1,560, 3,620^a mg/kg-d 13 weeks</p>	<p>Incidence (severity):</p> <table border="1" data-bbox="586 401 1424 814"> <thead> <tr> <th colspan="3" data-bbox="586 401 1040 443">Males</th> <th colspan="3" data-bbox="1040 401 1424 443">Females</th> </tr> <tr> <th data-bbox="586 443 781 485"></th> <th data-bbox="781 443 1040 485"><u>Transitional epithelial hyperplasia</u></th> <th data-bbox="1040 443 1424 485"><u>Transitional epithelial hyperplasia</u></th> <th colspan="3"></th> </tr> <tr> <th data-bbox="586 485 781 527"><u>Dose (mg/kg-d)</u></th> <th data-bbox="781 485 1040 527"></th> <th data-bbox="1040 485 1424 527"><u>Dose (mg/kg-d)</u></th> <th colspan="3"></th> </tr> </thead> <tbody> <tr> <td data-bbox="586 527 781 569">0</td> <td data-bbox="781 527 1040 569">0/10</td> <td data-bbox="1040 527 1424 569">0</td> <td colspan="3" data-bbox="1040 527 1424 569">0/10</td> </tr> <tr> <td data-bbox="586 569 781 611">230</td> <td data-bbox="781 569 1040 611">not evaluated</td> <td data-bbox="1040 569 1424 611">290</td> <td colspan="3" data-bbox="1040 569 1424 611">not evaluated</td> </tr> <tr> <td data-bbox="586 611 781 653">490</td> <td data-bbox="781 611 1040 653">not evaluated</td> <td data-bbox="1040 611 1424 653">590</td> <td colspan="3" data-bbox="1040 611 1424 653">not evaluated</td> </tr> <tr> <td data-bbox="586 653 781 695">840</td> <td data-bbox="781 653 1040 695">0/10</td> <td data-bbox="1040 653 1424 695">850</td> <td colspan="3" data-bbox="1040 653 1424 695">not evaluated</td> </tr> <tr> <td data-bbox="586 695 781 737">1,520</td> <td data-bbox="781 695 1040 737">1/10 (3.0)</td> <td data-bbox="1040 695 1424 737">1,560</td> <td colspan="3" data-bbox="1040 695 1424 737">0/10</td> </tr> <tr> <td data-bbox="586 737 781 779">3,610</td> <td data-bbox="781 737 1040 779">7/10* (2.9)</td> <td data-bbox="1040 737 1424 779">3,620</td> <td colspan="3" data-bbox="1040 737 1424 779">3/10 (2.0)</td> </tr> </tbody> </table> <p>Severity: 1 = minimal, 2 = mild, 3 = moderate, 4 = marked</p>						Males			Females				<u>Transitional epithelial hyperplasia</u>	<u>Transitional epithelial hyperplasia</u>				<u>Dose (mg/kg-d)</u>		<u>Dose (mg/kg-d)</u>				0	0/10	0	0/10			230	not evaluated	290	not evaluated			490	not evaluated	590	not evaluated			840	0/10	850	not evaluated			1,520	1/10 (3.0)	1,560	0/10			3,610	7/10* (2.9)	3,620	3/10 (2.0)		
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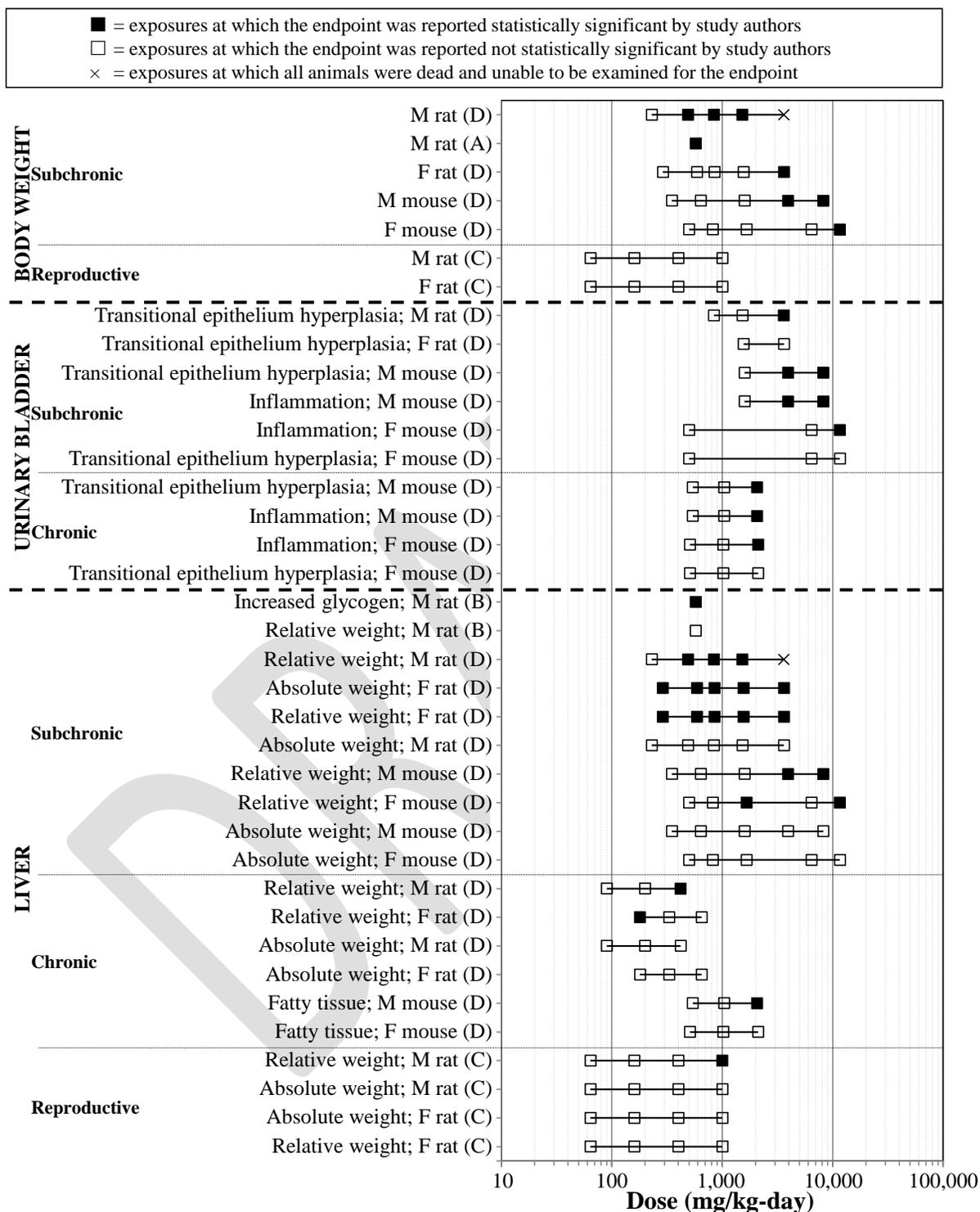
Table 1-14. Evidence pertaining to urinary bladder effects in animals following oral exposure to *tert*-butanol (continued)

Reference and study design	Results					
<p>NTP (1995)</p> <p>B6C3F₁ mouse; 60/sex/treatment Drinking water (0, 5, 10, or 20 mg/mL) M: 0, 540, 1,040, 2,070^a mg/kg-d F: 0, 510, 1,020, 2,110 mg/kg-d 2 years</p>	Incidence (severity):					
	Males			Females		
	<u>Dose</u> (mg/kg-d)	<u>Transitional</u> <u>epithelial</u> <u>hyperplasia</u>	<u>Inflam-</u> <u>mation</u>	<u>Dose</u> (mg/kg-d)	<u>Transitional</u> <u>epithelial</u> <u>hyperplasia</u>	<u>Inflam-</u> <u>mation</u>
	0	1/59 (2.0)	0/59	0	0/59	0/59
	540	3/59 (1.7)	3/59 (1.7)	510	0/60	0/60
	1,040	1/58 (1.0)	1/58 (1.0)	1,020	0/59	0/59
	2,070	17/59* (1.8)	37/59* (2.0)	2,110	3/57 (1.0)	4/57* (2.0)
	Severity: 1 = minimal, 2 = mild, 3 = moderate, 4 = marked					

- 1 ^aThe high-dose group had an increase in mortality.
- 2 * Statistically significant $p \leq 0.05$ as determined by study authors.
- 3 Conversions from drinking water concentrations to mg/kg-d performed by study authors.

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Sources: (A) Acharya et al. (1995); (B) Acharya et al. (1997; 1995); (C) Lyondell Chemical Co. (2004); (D) NTP (1995)

Figure 1-8. Exposure-response array of other effects following oral exposure to tert-butanol.

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1.2. INTEGRATION AND EVALUATION

1.2.1. Effects Other Than Cancer

The strongest evidence following *tert*-butanol exposure is for kidney, with toxicity observed after oral exposure in two strains of rats and in one strain of mice and in both sexes. In mice, the only kidney effect observed was an increase in kidney weight (absolute and/or relative) in both sexes of mice in the 13-week study, but no treatment-related histopathological lesions were reported in the kidneys of mice at the 13-week or 2-year time points (NTP, 1995). In male rats, effects related to the accumulation of α_{2u} -globulin in the kidney have been reported, including precursors to granular casts, linear mineralization, and tubular hyperplasia, but these are not considered relevant to humans (Hard et al., 2011; Cirvello et al., 1995; NTP, 1995; Lindamood et al., 1992). However, several other effects in the kidney unrelated to α_{2u} -globulin were observed in female and/or male rats. Absolute and relative kidney weights were increased in both male and female rats after both 13 weeks and 15 months of treatment (NTP, 1995). Histopathological examination also indicated kidney toxicity in both male and female rats, with increased incidence of nephropathy after 13 weeks of oral exposure and transitional epithelium hyperplasia observed after 2 years of oral exposure (NTP, 1995). Additionally, increased inflammation (suppurative) was noted in females after 2 years oral exposure (NTP, 1995). EPA identified kidney effects as a human hazard of *tert*-butanol oral exposure.

Fewer and less severe kidney effects were observed via inhalation than via oral exposure, likely due to the differing levels of internal doses achieved via the different routes. Specifically, available inhalation studies (NTP, 1997) were conducted at concentrations that are comparable, in terms of *tert*-butanol blood concentration, to the lower range of doses in oral studies. Moreover, there is convincing toxicokinetic data to indicate that *tert*-butanol is absorbed by both routes, and kidney effects are remote from the site of absorption. EPA identified kidney effects as a human hazard of *tert*-butanol inhalation exposure.

Thyroid follicular cell hyperplasia was observed in the mice after 2 years of exposure via drinking water (NTP, 1995); and EPA identified thyroid effects as a potential human hazard of *tert*-butanol exposure. However, this endpoint most likely reflects early events in the neoplastic progression of thyroid follicular cell tumors following *tert*-butanol exposure (see Section 1.1.2) and was not considered further for dose-response analysis and derivation of noncancer reference values.

EPA identified suggestive evidence of developmental effects as a potential human hazard of *tert*-butanol exposure. Exposure to high doses of *tert*-butanol during gestation resulted in some effects in exposed offspring or pups, although the effects were not always consistent across exposure routes (oral and inhalation). Dams exhibit effects at the same doses as fetal effects. Neurodevelopmental effects have also been observed; however, the neurodevelopmental studies had limitations in the study design and/or reporting and results were inconsistent between studies

1 or across dose. Thus, these effects were not considered further for dose-response analysis and
2 derivation of reference values.

3 EPA concluded that the evidence does not support reproductive effects, body weight
4 changes, liver effects, and urinary bladder effects as potential human hazards of *tert*-butanol
5 exposure. Thus, these effects were not considered further for dose-response analysis and the
6 derivation of reference values.

7 **1.2.2. Carcinogenicity**

8 Under EPA's *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)), the database for
9 *tert*-butanol provides "suggestive evidence of carcinogenic potential," based on a statistically
10 significant increase in renal tumors (renal tubule adenomas and carcinomas) in male F344 rats and
11 a statistically significant increase in thyroid follicular cell adenomas in female B6C3F₁ mice, all
12 exposed to *tert*-butanol in drinking water for 2 years ([Cirvello et al., 1995](#); [NTP, 1995](#)). There are
13 no available studies of cancer in humans associated with exposure to *tert*-butanol.

14 In the [NTP \(1995\)](#) rodent bioassay, *tert*-butanol-exposed male rats had a significant
15 increase in renal tumors compared to controls, a result confirmed by a PWG reevaluation ([Hard et
16 al., 2011](#)). Although mechanistic data show that α_{2u} -globulin-related processes occur with *tert*-
17 butanol exposure, there is insufficient evidence to support a conclusion that α_{2u} -globulin
18 nephropathy is the sole or primary contributor to renal tumor development. Specifically,
19 *tert*-butanol induced tumors at lower doses than those for other precursor effects such as
20 hyperplasia and granular casts, with no further increase in tumor incidence coinciding with the
21 induction of additional markers of α_{2u} -globulin nephropathy. Based on analysis of available mode
22 of action data, these tumors are not attributed to α_{2u} -globulin and are considered relevant in
23 humans ([U.S. EPA, 1991a](#)). *tert*-Butanol was negative in a variety of genotoxicity assays in different
24 cell systems including gene mutations, sister chromatid exchanges, micronucleus formation, and
25 chromosomal aberrations. However, DNA adducts in male Kunming mice and DNA damage in
26 human HL-60 leukemia cells have been observed. Overall, the mode(s) of carcinogenic action for
27 *tert*-butanol in the kidney and the thyroid are not known, and these tumor data are considered
28 relevant to humans.

29 As emphasized in the Cancer Guidelines ([U.S. EPA, 2005a](#)), selection of the cancer descriptor
30 follows a full evaluation of the available evidence. The carcinogenicity evidence for *tert*-butanol
31 could be considered a borderline case between two cancer descriptors—"suggestive evidence of
32 carcinogenic potential" and "likely to be carcinogenic to humans." The descriptor of "suggestive
33 evidence of carcinogenic potential" is appropriate when a concern for potential carcinogenic effects
34 in humans is raised, but the data are judged not sufficient for a stronger conclusion. Exposure to
35 *tert*-butanol produced a positive tumor response at more than one site (kidney and thyroid) and in
36 more than one species (rat and mouse). These data appear to correspond closely to one of the
37 examples in the Cancer Guidelines ([U.S. EPA, 2005a](#)) for the descriptor of "likely to be carcinogenic
38 to humans;" i.e., "an agent that has tested positive in animal experiments in more than one species,

1 sex, strain, site, or exposure route, with or without evidence of carcinogenicity in humans.” Several
2 aspects of the data support the conclusion that these data are not sufficient to characterize *tert*-
3 butanol as “likely to be carcinogenic to humans.”

4 First, the renal tumors associated with *tert*-butanol exposure in the [NTP \(1995\)](#) rodent
5 bioassay were predominantly benign. Based on the PWG reevaluation ([Hard et al., 2011](#)), among
6 the three treated groups, only two of the 43 animals with tumors has carcinomas (there were no
7 carcinomas among the 4 control animals with tumors). Additionally, no kidney tumors were
8 observed in female rats or in either sex of mice. Furthermore, ETBE, which is rapidly metabolized
9 to *tert*-butanol, did not induce kidney tumors in the same strain of rats at doses that resulted in
10 similar internal concentrations of *tert*-butanol. Therefore, the level of concern raised by renal
11 tumors associated with *tert*-butanol exposure is reduced based on the predominance of benign
12 tumors, an increase in renal tumors in a single sex/species combination only, and the lack of
13 coherence with the metabolically-related compound ETBE.

14 The thyroid tumors associated with *tert*-butanol exposure were also predominantly benign.
15 In the [NTP \(1995\)](#) rodent bioassay, only female mice had a statistically significant increase in
16 thyroid tumors; none of these were carcinomas. In males, decreased survival complicates the
17 interpretation of thyroid tumors because male mice had an increased incidence of thyroid follicular
18 cell hyperplasia at all exposure levels, but there was no significant increase in thyroid tumors at any
19 exposure. Interestingly, one thyroid follicular cell carcinoma occurred in a high-dose male, but
20 limited conclusions can be drawn from this single observation. Thyroid tumors were not observed
21 in either sex of the rat exposed chronically to *tert*-butanol. Additionally, ETBE did not induce
22 thyroid tumors, although only rats and not mice were tested. Therefore, the level of concern raised
23 by thyroid tumors associated with *tert*-butanol exposure is reduced based on the predominance of
24 benign tumors and an increase in thyroid tumors in a single sex/species combination only.

25 Overall, the cancer descriptor “suggestive evidence of carcinogenic potential” was selected,
26 as some concern is raised by the positive evidence of predominantly benign renal tumors in male
27 rats and thyroid tumors in female mice.

28 The Cancer Guidelines ([U.S. EPA, 2005a](#)) indicate that for tumors occurring at a site other
29 than the initial point of contact, the weight of evidence for carcinogenic potential may apply to all
30 routes of exposure that have not been adequately tested at sufficient doses. An exception occurs
31 when there is convincing toxicokinetic data that absorption does not occur by other routes.
32 Information available on the carcinogenic effects of *tert*-butanol via the oral route demonstrates
33 that tumors occur in tissues remote from the site of absorption. Information on the carcinogenic
34 effects of *tert*-butanol via the inhalation and dermal routes in humans or animals is not available.
35 Based on the observation of systemic tumors following oral ingestion, and in the absence of
36 information to indicate otherwise, it is assumed that an internal dose will be achieved regardless of
37 the route of exposure. Therefore, there is “suggestive evidence of carcinogenic potential” from
38 exposure to *tert*-butanol by all routes of exposure.

- 1 **1.2.3. Susceptible Populations and Lifestages for Cancer and Noncancer Outcomes**
- 2 No data were identified to indicate any possible susceptible populations or lifestages.

DRAFT

2. DOSE-RESPONSE ANALYSIS

2.1. ORAL REFERENCE DOSE FOR EFFECTS OTHER THAN CANCER

The RfD (expressed in units of mg/kg-day) is defined as an estimate (with uncertainty spanning perhaps an order of magnitude) of a daily exposure to the human population (including sensitive subgroups) that is likely to be without an appreciable risk of deleterious effects during a lifetime. It can be derived from a no-observed-adverse-effect level (NOAEL), lowest-observed-adverse-effect level (LOAEL), or the 95% lower bound on the benchmark dose (BMDL), with uncertainty factors (UFs) generally applied to reflect limitations of the data used.

2.1.1. Identification of Studies and Effects for Dose-Response Analysis

EPA identified kidney effects as a human hazard of *tert*-butanol exposure. Studies within this effect category were evaluated using general study quality characteristics (as discussed in Section 6 of the Preamble) to help inform the selection of studies from which to derive toxicity values. Rationales for selecting the studies and effects to represent each of these hazards are summarized below.

Human studies are preferred over animal studies when quantitative measures of exposure are reported and the reported effects are determined to be associated with exposure. However, there are no available human occupational or epidemiological studies of oral exposure to *tert*-butanol.

Animal studies were evaluated to determine which studies provided: (a) the most relevant routes and durations of exposure; (b) multiple exposure levels to provide information about the shape of the dose-response curve; and (c) power to detect effects at low exposure levels ([U.S. EPA, 2002](#)). Sufficient data were available to develop a PBPK model in rats for both oral and inhalation exposure in order to perform route-to-route extrapolation, so rat studies from both routes of exposure were considered for dose-response analysis. The database for *tert*-butanol includes several studies and data sets that are potentially suitable for use in deriving reference values. Specifically, effects associated with *tert*-butanol exposure in animals include observations of organ weight and/or histological changes in the kidney observed in several chronic and subchronic studies.

Kidney Toxicity

EPA identified kidney effects as a human hazard of *tert*-butanol-induced toxicity based on findings of organ weight changes in rats and mice, as well as histopathology in rats. These findings were consistent across multiple chronic, subchronic, and short-term studies following oral and

1 inhalation exposure. Acharya et al. (1997; 1995) used a single exposure group and did not provide
2 incidence or severity data, and thus was not considered for dose-response assessment. Lyondell
3 Chemical Co. (2004) and NTP (1997) were of subchronic or shorter duration, and so were set aside
4 given the availability of a longer duration study. Therefore, the NTP 2-year drinking water study
5 (NTP, 1995) was identified most suitable for dose-response assessment considering the study
6 duration, comprehensive reporting of outcomes, multiple species tested, and multiple doses tested.

7 In the NTP (1995) drinking water study, male F344 rats were exposed to approximate
8 doses of 0, 90, 200, or 420 mg/kg-day; female F344 rats were exposed to approximate doses of 0,
9 180, 330, or 650 mg/kg-day; male B6C3F₁ mice were exposed to approximate doses of 0, 540,
10 1,040, or 2,070 mg/kg-day; and female B6C3F₁ mice were exposed to approximate doses of 0, 510,
11 1,020, or 2,110 mg/kg-day. Reduced body weights and survival were observed and reflected in
12 some of the effects. Kidney effects including changes in organ weight and/or histopathology were
13 observed in both sexes in rats and mice. Effects were also observed after 13 weeks, 15 months, and
14 2 years of treatment (NTP, 1995). Effects were more consistent and occurred at lower doses in rats
15 as compared to mice, so as a result, only data in the more sensitive species of rats were used for
16 dose-response assessment. Endpoints potentially confounded by the presence of α_{2u} -globulin
17 nephropathy in male rats, such as linear mineralization and renal tubule hyperplasia, were not used
18 for dose-response analysis. Specific endpoints chosen for analysis were absolute and relative
19 kidney weight (observed in males and females), kidney inflammation (observed only in females),
20 and kidney transitional epithelial hyperplasia (observed in males and females). For most
21 endpoints, the data at the longest duration of 2 years were selected. However, as discussed in
22 Section 1.1.1, 2-year kidney weight data were not considered because organs were only weighed at
23 15 months.

24 **2.1.2. Methods of Analysis**

25 No biologically based dose-response models are available for *tert*-butanol. In this situation,
26 EPA evaluates a range of dose-response models thought to be consistent with underlying biological
27 processes to determine how to best empirically model the dose-response relationship in the range
28 of the observed data. Consistent with this approach, all models available in EPA's Benchmark Dose
29 Software (BMDS) were evaluated. Consistent with EPA's *Benchmark Dose Technical Guidance*
30 *Document* (U.S. EPA, 2012b), the benchmark dose (BMD) and the 95% lower confidence limit on the
31 BMD (BMDL) were estimated using a benchmark response (BMR) of 10% change from the control
32 mean for organ weight data in the absence of information regarding the level of change that is
33 considered biologically significant. Furthermore, the BMD and BMDL were estimated to facilitate a
34 consistent basis of comparison across endpoints, studies, and assessments. A benchmark response
35 (BMR) of 10% extra risk was considered appropriate for the quantal data on incidences of kidney
36 inflammation and kidney transitional epithelial hyperplasia. For each endpoint, the BMDL estimate
37 (95% lower confidence limit on the BMD, as estimated by the profile likelihood method) and Akaike
38 Information Criterion (AIC) value were used to select a best-fit model among models exhibiting

1 adequate fit. If the BMDL estimates were “sufficiently close,” that is, differed by at most 3-fold, the
2 model selected was the one that yielded the lowest AIC value. If the BMDL estimates were not
3 sufficiently close, the lowest BMDL was selected as the POD. The estimated BMDLs were used as
4 points of departure (PODs). Further details including the modeling output and graphical results for
5 the best-fit model for each endpoint can be found in Appendix C of the Supplemental Information.

6 In general, absolute and relative kidney weight data may both be considered appropriate
7 endpoints for analysis ([Bailey et al., 2004](#)). However, in the [NTP \(1995\)](#) 2-year drinking water
8 study, there was a noticeable decrease in body weight in exposed animals relative to controls at the
9 15 month interim sacrifice (see Table 1-1). In such a case, relative kidney weights are the
10 preferred, so changes in absolute kidney weights were not analyzed.

11 Human equivalent doses (HEDs) for oral exposures were derived from the PODs estimated
12 from the laboratory animal data as described in EPA’s *Recommended Use of Body Weight^{3/4} as the*
13 *Default Method in Derivation of the Oral Reference Dose* ([U.S. EPA, 2011](#)). In this guidance, EPA
14 advocates a hierarchy of approaches for deriving HEDs from data in laboratory animals, with the
15 preferred approach being physiologically-based toxicokinetic modeling. Other approaches can
16 include using chemical-specific information in the absence of a complete physiologically-based
17 toxicokinetic model. As discussed in Appendix B of the Supplemental Information, several rat
18 physiologically based pharmacokinetic (PBPK) models for *tert*-butanol have been developed and
19 published, but a validated human PBPK model for *tert*-butanol for extrapolating doses from animals
20 to humans is not available. In lieu of either chemical-specific models or data to inform the
21 derivation of human equivalent oral exposures, a body weight scaling to the ³/₄ power (i.e., BW^{3/4})
22 approach is applied to extrapolate toxicologically equivalent doses of orally administered agents
23 from adult laboratory animals to adult humans for the purpose of deriving an oral RfD. BW^{3/4}
24 scaling was not employed for deriving HEDs from studies in which doses were administered
25 directly to early postnatal animals because of the absence of information on whether allometric
26 (i.e., body weight) scaling holds when extrapolating doses from neonatal animals to adult humans
27 due to presumed toxicokinetic and/or toxicodynamic differences between lifestages ([U.S. EPA,](#)
28 [2011](#); [Hattis et al., 2004](#)).

29 Consistent with EPA guidance ([U.S. EPA, 2011](#)), the PODs estimated based on effects in adult
30 animals are converted to HEDs employing a standard dosimetric adjustment factor (DAF) derived
31 as follows:

$$DAF = (BW_a^{1/4} / BW_h^{1/4}),$$

34 where

35 BW_a = animal body weight

36 BW_h = human body weight

37

Using a standard BW_a of 0.25 kg for rats and a BW_h of 70 kg for humans ([U.S. EPA, 1988](#)), the resulting DAFs for rats is 0.24. Applying this DAF to the POD identified for effects in adult rats yields a POD_{HED} as follows (see Table 2-1):

$$\text{POD}_{\text{HED}} = \text{Laboratory animal dose (mg/kg-day)} \times \text{DAF}$$

Table 2-1 summarizes the sequence of calculations leading to the derivation of a human-equivalent POD for each data set discussed above.

Table 2-1. Summary of derivations of points of departure

Endpoint and Reference	Species/sex	Model ^a	BMR	BMD (mg/kg-d)	BMDL (mg/kg-d)	POD _{ADJ} ^b (mg/kg-d)	POD _{HED} ^c (mg/kg-d)
<i>Kidney</i>							
Increased relative kidney weight NTP (1995)	Rat/M	Exponential (M4)	10%	117	48	48	11.5
Increased relative kidney weight NTP (1995)	Rat/F	Linear	10%	158	133	133	31.9
Kidney inflammation NTP (1995)	Rat/F	Log-probit	10%	254	200	200	48
Kidney transitional epithelial hyperplasia NTP (1995)	Rat/M	Log-logistic	10%	30	16	16	3.84
Kidney transitional epithelial hyperplasia NTP (1995)	Rat/F	Multistage, 3-degree	10%	412	339	339	81.4

^aFor modeling details, see Appendix C in Supplemental Information.

^bFor studies in which animals were not dosed daily, administered doses were adjusted to calculate the TWA daily doses prior to BMD modeling.

^cHED PODs were calculated using BW^{3/4} scaling ([U.S. EPA, 2011](#)).

2.1.3. Derivation of Candidate Values

Under EPA's *A Review of the Reference Dose and Reference Concentration Processes* ([U.S. EPA, 2002; Section 4.4.5](#)), also described in the Preamble, five possible areas of uncertainty and variability were considered. An explanation follows:

1 An intraspecies uncertainty factor, UF_H , of 10 was applied to all PODs to account for
2 potential differences in toxicokinetics and toxicodynamics in the absence of information on the
3 variability of response in the human population following oral exposure to *tert*-butanol.

4 An interspecies uncertainty factor, UF_A , of 3 ($10^{1/2} = 3.16$, rounded to 3) was applied to all
5 PODs because $BW^{3/4}$ scaling is used to extrapolate oral doses from laboratory animals to humans.
6 Although $BW^{3/4}$ scaling addresses some aspects of cross-species extrapolation of toxicokinetic and
7 toxicodynamic processes, some residual uncertainty remains. In the absence of chemical-specific
8 data to quantify this uncertainty, EPA's $BW^{3/4}$ guidance ([U.S. EPA, 2011](#)) recommends use of an
9 uncertainty factor of 3.

10 A subchronic to chronic uncertainty factor, UF_S , of 1 was applied to all PODs since the
11 endpoints examined were all observed following chronic exposure.

12 A LOAEL to NOAEL uncertainty factor, UF_L , of 1 was applied to all PODs because the current
13 approach is to address this factor as one of the considerations in selecting a BMR for benchmark
14 dose modeling. In this case, BMRs of a 10% change in relative kidney weight, a 10% extra risk of
15 kidney inflammation, and a 10% extra risk of transitional cell hyperplasia were selected under an
16 assumption that they represent minimal biologically significant changes.

17 A database uncertainty factor, UF_D , of 1 was applied to all PODs. The *tert*-butanol toxicity
18 database includes a chronic toxicity study in rats and mice ([NTP, 1995](#)), a subchronic toxicity study
19 in rats and mice ([NTP, 1997](#)), and developmental toxicity studies in rats and mice ([Lyondell
20 Chemical Co., 2004](#); [Faulkner et al., 1989](#); [Daniel and Evans, 1982](#)). In the developmental studies,
21 no effects were observed at exposure levels below 1000 mg/kg-day, and effects observed at
22 ≥ 1000 mg/kg-day were accompanied by evidence of maternal toxicity. These exposure levels are
23 much higher than the PODs for kidney effects, suggesting developmental toxicity is not a sensitive
24 endpoint. The *tert*-butanol database contains a one-generation reproductive toxicity study in rats
25 ([Lyondell Chemical Co., 2004](#)), though no multigenerational reproductive study has been
26 performed. There are no immunotoxicity studies for *tert*-butanol. Information provided by studies
27 on ETBE, which is rapidly metabolized to systemically-available *tert*-butanol, can help in
28 considering the lack of a *tert*-butanol multigenerational reproductive study or an immunotoxicity
29 study. No adverse effects were reported in one- and two-generation reproductive/developmental
30 studies on ETBE ([Gaoua, 2004a, b](#)), and the database for ETBE does not indicate immunotoxicity
31 ([Banton et al., 2011](#); [Li et al., 2011](#)). Thus, although there are some gaps in the toxicity database for
32 *tert*-butanol, the available data on *tert*-butanol, informed by the data on ETBE, do not suggest that
33 additional studies would lead to identification of a more sensitive endpoint or a lower POD.
34 Therefore, a database UF_D of 1 was applied.

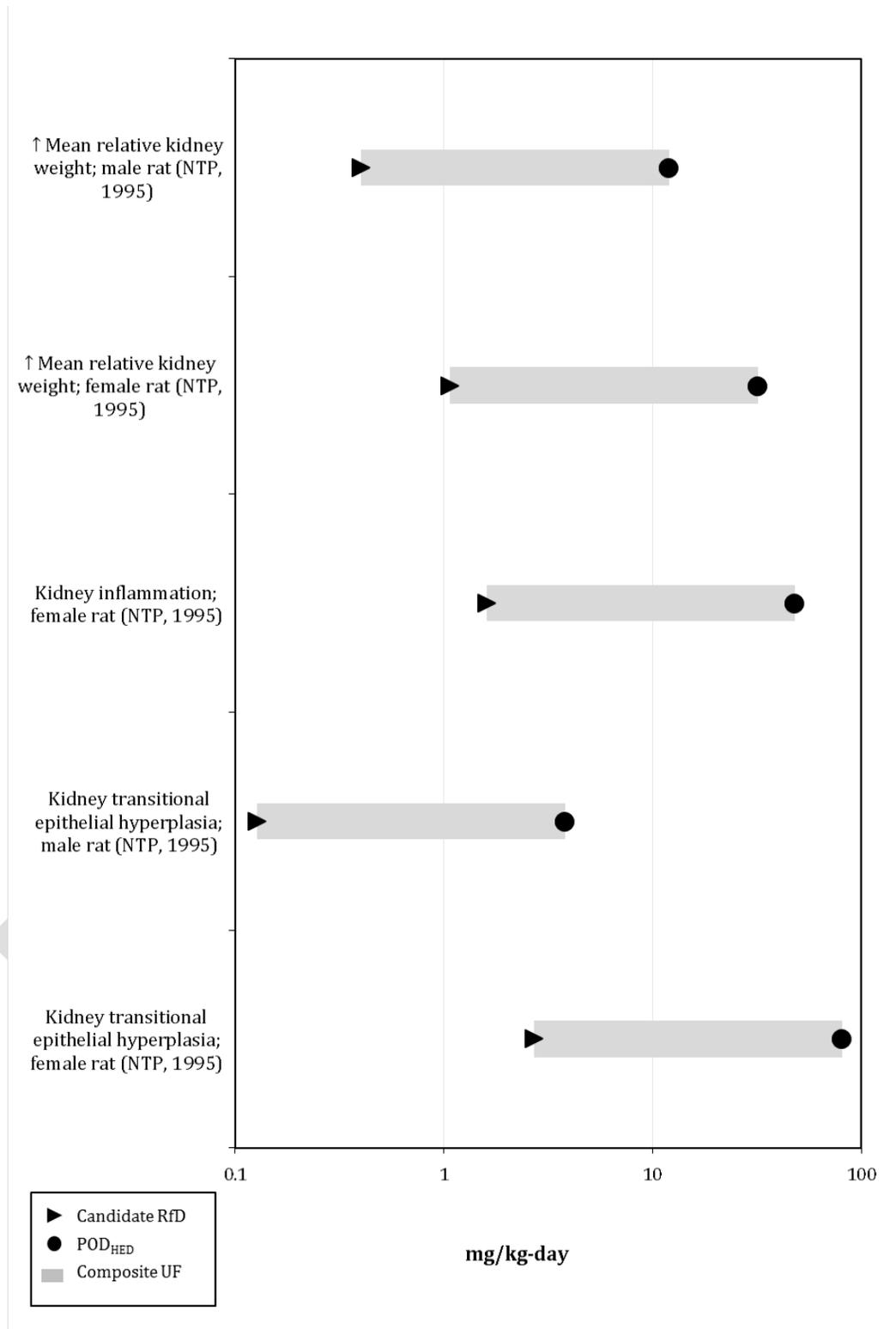
35 Table 2-2 is a continuation of Table 2-1 and summarizes the application of UFs to each POD
36 to derive a candidate value for each data set. The candidate values presented in the table below are
37 preliminary to the derivation of the organ/system-specific reference values. These candidate values
38 are considered individually in the selection of a representative oral reference value for a specific

- 1 hazard and subsequent overall RfD for *tert*-butanol.
 2 Figure 2-1 presents graphically the candidate values, UFs, and PODs, with each bar
 3 corresponding to one data set described in Table 2-1 and Table 2-2.
 4

5 **Table 2-2. Effects and corresponding derivation of candidate RfDs**

Endpoint and Reference	POD _{HED} (mg/kg-d)	POD type	UF _A	UF _H	UF _L	UF _S	UF _D	Composite UF	Candidate value (mg/kg-d)
<i>Kidney</i>									
Increased relative kidney weight; male rat NTP (1995)	12	BMDL _{10%}	3	10	1	1	1	30	4 × 10 ⁻¹
Increased relative kidney weight; female rat NTP (1995)	32	BMDL _{10%}	3	10	1	1	1	30	1 × 10 ⁰
Kidney inflammation; female rat NTP (1995)	48	BMDL _{10%}	3	10	1	1	1	30	2 × 10 ⁰
Kidney transitional epithelial hyperplasia; male rat NTP (1995)	3.8	BMDL _{10%}	3	10	1	1	1	30	1 × 10 ⁻¹
Kidney transitional epithelial hyperplasia; female rat NTP (1995)	81	BMDL _{10%}	3	10	1	1	1	30	3 × 10 ⁰

6
7



1

2

Figure 2-1. Candidate RfD values with corresponding POD and composite UF.

1 **2.1.4. Derivation of Organ/System-Specific Reference Doses**

2 Table 2-3 distills the candidate values from Table 2-2 into a single value for the kidney.
 3 Organ or system-specific reference values may be useful for subsequent cumulative risk
 4 assessments that consider the combined effect of multiple agents acting at a common site.

5 ***Kidney Toxicity***

6 For *tert*-butanol, candidate values were for several different effects in both sexes, spanning
 7 a range from 1×10^{-1} to 3×10^0 mg/kg-day, for an overall thirtyfold range. To estimate an exposure
 8 level below which kidney toxicity from *tert*-butanol exposure is not expected to occur, the RfD for
 9 increased incidence of transitional epithelial hyperplasia in male rats (**1×10^{-1} mg/kg-day**) is
 10 proposed as the kidney-specific reference dose for *tert*-butanol. Unlike kidney inflammation, this
 11 effect was observed in both sexes, with males appearing to be more sensitive than females.
 12 Additionally, it is a more specific and more sensitive indicator of kidney toxicity than the relatively
 13 non-specific endpoint of kidney weight changes. Confidence in this kidney-specific RfD is high. The
 14 PODs are based on modeled benchmark dose estimates, and the candidate values are derived from
 15 a well-conducted study, involving a sufficient number of animals per group, including both sexes,
 16 and assessing a wide range of kidney endpoints.

17 **Table 2-3. Organ/system-specific RfDs and proposed overall RfD for *tert*-**
 18 **butanol**

Effect	Basis	RfD (mg/kg-day)	Exposure description	Confidence
Kidney toxicity	Increased incidence of transitional epithelial hyperplasia	1×10^{-1}	Chronic	HIGH
Proposed overall RfD	Increased incidence of transitional epithelial hyperplasia	1×10^{-1}	Chronic	HIGH

19

20 **2.1.5. Selection of the Proposed Overall Reference Dose**

21 For *tert*-butanol, only kidney effects were identified as a hazard; thus a single
 22 organ/system-specific reference dose was derived. Therefore, the kidney-specific RfD of
 23 **1×10^{-1} mg/kg-day** is also proposed as an estimated exposure level below which deleterious
 24 effects from *tert*-butanol exposure are not expected to occur.

25 The overall reference dose is derived to be protective of all types of effects for a given
 26 duration of exposure and is intended to protect the population as a whole including potentially
 27 susceptible subgroups ([U.S. EPA, 2002](#)). Decisions concerning averaging exposures over time for
 28 comparison with the RfD should consider the types of toxicological effects and specific lifestages of
 29 concern. Fluctuations in exposure levels that result in elevated exposures during these lifestages

1 could potentially lead to an appreciable risk, even if average levels over the full exposure duration
2 were less than or equal to the RfD. In the case of *tert*-butanol, no specific lifestages have been
3 identified as a potentially susceptible subgroup.

4 **2.1.6. Confidence Statement**

5 A confidence level of high, medium, or low is assigned to the study used to derive the RfD,
6 the overall database, and the RfD itself, as described in Section 4.3.9.2 of EPA's *Methods for*
7 *Derivation of Inhalation Reference Concentrations and Application of Inhalation Dosimetry* ([U.S. EPA](#)
8 [1994](#)). The overall confidence in this RfD is high. Confidence in the principal study ([NTP, 1995](#)) is
9 high. This study was well-conducted, complied with FDA GLP regulations, involved a sufficient
10 number of animals per group (including both sexes), and assessed a wide range of tissues and
11 endpoints. Although there are some gaps in the toxicity database for *tert*-butanol, these areas are
12 informed by the data on ETBE, a parent compound of *tert*-butanol. Therefore, the confidence in the
13 database is high. Reflecting high confidence in the principal study and high confidence in the
14 database, confidence in the RfD is high.

15 **2.1.7. Previous IRIS Assessment**

16 An oral assessment for *tert*-butanol was not previously available on IRIS.

17 **2.2. INHALATION REFERENCE CONCENTRATION FOR EFFECTS OTHER** 18 **THAN CANCER**

19 The inhalation reference concentration (RfC) (expressed in units of mg/m³) is defined as an
20 estimate (with uncertainty spanning perhaps an order of magnitude) of a continuous inhalation
21 exposure to the human population (including sensitive subgroups) that is likely to be without an
22 appreciable risk of deleterious effects during a lifetime. It can be derived from a NOAEL, LOAEL, or
23 the 95% lower bound on the benchmark concentration (BMCL), with UFs generally applied to
24 reflect limitations of the data used.

25 **2.2.1. Identification of Studies and Effects for Dose-Response Analysis**

26 EPA identified kidney effects as a human hazard of *tert*-butanol exposure. Studies within
27 this effect category were evaluated using general study quality characteristics (as discussed in
28 Section 6 of the Preamble) to help inform the selection of studies from which to derive toxicity
29 values. Rationales for selecting the studies and effects to represent this hazard are summarized
30 below.

31 Human studies are preferred over animal studies when quantitative measures of exposure
32 are reported and the reported effects are determined to be associated with exposure. However,
33 there are no available human occupational or epidemiological studies of inhalation exposure to
34 *tert*-butanol.

1 Animal studies were evaluated to determine which study provided: (a) the most relevant
2 routes and durations of exposure; (b) multiple exposure levels to provide information about the
3 shape of the dose-response curve; and (c) power to detect effects at low exposure levels ([U.S. EPA,
4 2002](#)). Sufficient data were available to develop a PBPK model in rats for both oral and inhalation
5 exposure in order to perform route-to-route extrapolation, so rat studies from both routes of
6 exposure were considered for dose-response analysis. The database for *tert*-butanol includes a
7 several studies and data sets that are potentially suitable for use in deriving reference values.
8 Specifically, effects associated with *tert*-butanol exposure in animals include observations of organ
9 weight and histological changes in the kidney in several chronic and subchronic studies.

10 ***Kidney Toxicity***

11 EPA identified kidney effects as a human hazard of *tert*-butanol exposure based on findings
12 of organ weight changes in rats and mice and histopathology in rats. These findings were
13 consistent across multiple chronic, subchronic, and short-term studies following oral and inhalation
14 exposure. Acharya et al. ([1997; 1995](#)) used a single exposure group and did not provide incidence
15 or severity data, so was not considered for dose-response assessment. [Lyondell Chemical Co.
16 \(2004\)](#) was of shorter than subchronic duration, and so was set aside given the availability of a
17 longer duration studies. Given the availability of a chronic study, the subchronic studies of [NTP
18 \(1995\)](#) and [NTP \(1997\)](#) would normally also be set aside for dose-response analysis. [NTP \(1997\)](#) is
19 the longest duration study via the inhalation route, not requiring route-to-route extrapolation, so
20 was kept for comparison purposes. Overall, the NTP 2-year drinking water study [NTP \(1995\)](#) was
21 identified as the study most suitable for dose-response assessment, given the study duration,
22 comprehensive reporting of outcomes, use of multiple species tested, multiple doses tested, and
23 availability of a PBPK model for route-to-route extrapolation. This study was discussed previously
24 in Section 2.1.1 as part of the derivation of the oral reference dose, so will not be reviewed here
25 again. The [NTP \(1997\)](#) subchronic inhalation study is described in more detail below.

26 [NTP \(1997\)](#) was a well-designed subchronic study that evaluated the effect of *tert*-butanol
27 exposure on multiple species at multiple inhalation doses. Briefly, groups of F344 rats and B6C3F₁
28 mice (10 per sex per species) were exposed to *tert*-butanol (>99% pure) at concentrations of 0,
29 409, 819, 1,637, 3,274 or 6,366 mg/m³ by inhalation for 6 hours per day, 5 days per week, for 13
30 weeks ([NTP, 1997](#)). Absolute kidney weights were elevated (10–11%) in male rats exposed at
31 ≥3,274 mg/m³; relative kidney weights were statistically significantly elevated (~9%) in males at
32 ≥3,274 mg/m³ and females at 6,366 mg/m³. Male rats exhibited an increase in the severity of
33 chronic nephropathy (characterized as number of foci of regenerative tubules). There were few
34 endpoints available for consideration in the subchronic study, but changes in kidney weights were
35 also observed in the oral studies, such as the [NTP \(1995\)](#) 2-year drinking water study.

1 **2.2.2. Methods of Analysis**

2 No biologically based dose-response models are available for *tert*-butanol. In this situation,
 3 EPA evaluates a range of dose-response models thought to be consistent with underlying biological
 4 processes to determine how best to empirically model the dose-response relationship in the range
 5 of the observed data. Consistent with this approach, all models available in EPA’s Benchmark Dose
 6 Software (BMDS) were evaluated. Consistent with EPA’s *Benchmark Dose Technical Guidance*
 7 *Document* ([U.S. EPA, 2012b](#)), the benchmark dose (BMD) and the 95% lower confidence limit on the
 8 BMD (BMDL) were estimated using a benchmark response (BMR) of 10% change from the control
 9 mean for organ weight data in the absence of information regarding what level of change is
 10 considered biologically significant, and also to facilitate a consistent basis of comparison across
 11 endpoints, studies, and assessments. A benchmark response (BMR) of 10% extra risk was
 12 considered appropriate for the quantal data on incidences of kidney inflammation and kidney
 13 transitional epithelial hyperplasia. The estimated BMDLs were used as points of departure (PODs).
 14 Further details including the modeling output and graphical results for the best-fit model for each
 15 endpoint can be found in Appendix C of the Supplemental Information.

16 In general, absolute and relative kidney weight data may both be considered appropriate
 17 endpoints for analysis ([Bailey et al., 2004](#)). However, in the [NTP \(1995\)](#) 2-year drinking water
 18 study, there was a noticeable decrease in body weight in exposed animals relative to controls at the
 19 15 month interim sacrifice (see Table 1-1). In such a case, relative kidney weights are preferred, so
 20 changes in absolute kidney weights from [NTP \(1995\)](#) were not analyzed. However, body weights
 21 were not impacted in the [NTP \(1997\)](#) subchronic inhalation study. Based on a historical review of
 22 26 studies of control rats from 1-month bioassays, [Bailey et al. \(2004\)](#) concluded that neither
 23 absolute kidney weight nor relative kidney:body (or kidney:brain) weight are optimal for
 24 evaluating organ weight changes. Since neither approach is preferred, both were considered to be
 25 appropriate for BMD analysis of the [NTP \(1997\)](#) data set.

26 ***PODs from Inhalation Studies***

27 Because the RfC is applicable to a continuous lifetime human exposure but derived from
 28 animal studies featuring intermittent exposure, EPA guidance ([U.S. EPA, 1994](#)) provides
 29 mechanisms for: (1) adjusting experimental exposure concentrations to a value reflecting
 30 continuous exposure duration (ADJ) and (2) determining a human equivalent concentration (HEC)
 31 from the animal exposure data. The former employs an inverse concentration-time relationship to
 32 derive a health-protective duration adjustment to time-weight the intermittent exposures used in
 33 the studies. The modeled benchmark concentration from the inhalation study ([NTP, 1997](#)) was
 34 adjusted to reflect a continuous exposure by multiplying it by (6 hours per day) ÷ (24 hours per
 35 day) and (5 days per week) ÷ (7 days per week) as follows:

36
$$\text{BMCL}_{\text{ADJ}} = \text{BMCL (mg/m}^3\text{)} \times (6 \div 24) \times (5 \div 7)$$

 37
$$= \text{BMCL (mg/m}^3\text{)} \times (0.1786)$$

1 The RfC methodology provides a mechanism for deriving a HEC from the duration-adjusted
2 POD (BMCL_{ADJ}) determined from the animal data. The approach takes into account the extra-
3 respiratory nature of the toxicological responses and accommodates species differences by
4 considering blood:air partition coefficients for *tert*-butanol in the laboratory animal (rat or mouse)
5 and humans. According to the RfC guidelines ([U.S. EPA, 1994](#)), *tert*-butanol is a Category 3 gas
6 because extra-respiratory effects were observed. [Kaneko et al. \(2000\)](#) measured a blood:gas
7 partition coefficient of 531 ± 102 for *tert*-butanol in the male Wistar rat, while [Borghoff et al.](#)
8 [\(1996\)](#) measured a value of 481 ± 29 in male F344 rats. A blood:gas partition coefficient of 462 was
9 reported for *tert*-butanol in humans ([Nihlén et al., 1995](#)). The calculation $(H_{b/g})_A \div (H_{b/g})_H$ was used
10 to calculate a blood:gas partition coefficient ratio to apply to the delivered concentration. Because
11 F344 rats were used in the study, the blood:gas partition coefficient for F344 rats was used. Thus,
12 the calculation was: $481 \div 462 = 1.04$. Therefore, a ratio of 1.04 was used to calculate the HEC. This
13 allowed a BMCL_{HEC} to be derived as follows:

14

$$\begin{aligned} \text{BMCL}_{\text{HEC}} &= \text{BMCL}_{\text{ADJ}} (\text{mg}/\text{m}^3) \times (\text{interspecies conversion}) \\ &= \text{BMCL}_{\text{ADJ}} (\text{mg}/\text{m}^3) \times (481 \div 462) \\ &= \text{BMCL}_{\text{ADJ}} (\text{mg}/\text{m}^3) \times (1.04) \end{aligned}$$

18 Table 2-4 summarizes the sequence of calculations leading to the derivation of a human-
19 equivalent point of departure for each inhalation data set discussed above.

20

1 **Table 2-4. Summary of derivation of PODs following inhalation exposure**

Endpoint and Reference	Species/ Sex	Model ^a	BMR	BMC ^b (mg/m ³)	BMCL ^b (mg/m ³)	POD _{ADJ} ^b (mg/m ³)	POD _{HEC} ^c (mg/m ³)
<i>Kidney</i>							
Increased relative kidney weight NTP (1997)	Male F344 rats	Linear	10%	6309	4821	861	861
Increased absolute kidney weight NTP (1997)	Male F344 rats	Hill	10%	1931	1705	304	304
Increased relative kidney weight NTP (1997)	Female F344 rats	No model selected	10%	--	--	--	--
Increased absolute kidney weight NTP (1997)	Female F344 rats	No model selected	10%	--	--	--	--

2 ^aFor modeling details, see Appendix C in Supplemental Information.

3 ^bBMCs, BMCLs, and PODs were adjusted for continuous daily exposure by multiplying by (hours exposed per day /
4 24 hrs) × (days exposed per week / 7 days).

5 ^cPOD_{HEC} calculated by adjusting the POD_{ADJ} by the DAF (=1.0) for a category 3 gas ([U.S. EPA, 1994](#)).

6 ^dBMD modeling failed to successfully calculate a BMD value (see Appendix C).

8 ***PODs from oral studies – use of PBPK model for route-to-route extrapolation***

9 A PBPK model for *tert*-butanol in rats has been developed, as described in Appendix B.
10 Using this model, route-to-route extrapolation of the oral BMDLs to derive inhalation PODs was
11 performed as follows. First, the internal dose in the rat at each oral BMDL (assuming continuous
12 exposure) was estimated using the PBPK model, to derive an “internal dose BMDL.” Then, the
13 inhalation air concentration (again, assuming continuous exposure) that led to the same internal
14 dose in the rat was estimated using the PBPK model. The resulting BMCL was then converted to a
15 human equivalent concentration POD using the methodology previously described in “PODs from
16 inhalation studies”:

17
$$\text{BMCL}_{\text{HEC}} = \text{BMCL}_{\text{ADJ}} (\text{mg}/\text{m}^3) \times (\text{interspecies conversion})$$

18
$$= \text{BMCL}_{\text{ADJ}} (\text{mg}/\text{m}^3) \times (481 \div 462)$$

19
$$= \text{BMCL}_{\text{ADJ}} (\text{mg}/\text{m}^3) \times (1.04)$$

20 A critical decision in the route-to-route extrapolation is the selection of the internal dose
21 metric that establishes “equivalent” oral and inhalation exposures. For *tert*-butanol-induced kidney
22 effects, the two options are the concentration of *tert*-butanol in blood and rate of *tert*-butanol
23 metabolism. Note that using the kidney concentration of *tert*-butanol will lead to the same route-
24 to-route extrapolation relationship as *tert*-butanol in blood, since the distribution from blood to
25 kidney is independent of route. There are no data to suggest that metabolites of *tert*-butanol

1 mediate its renal toxicity. In the absence of evidence that would suggest otherwise, it is assumed
 2 that *tert*-butanol itself is the active toxicological agent. Therefore, the concentration of *tert*-butanol
 3 in blood was selected as the dose metric.

4 Table 2-5 summarizes the sequence of calculations leading to the derivation of a human-
 5 equivalent point of departure for each oral data set discussed above.

6 **Table 2-5. Summary of derivation of inhalation points of departure derived**
 7 **from route-to-route extrapolation from oral exposures**

Endpoint and reference	Species/sex	BMR	BMDL (mg/kg-d)	Internal dose ^a (mg/L)	Equivalent POD _{HEC} ^b (mg/m ³)
<i>Kidney</i>					
Mean relative kidney weight NTP (1995)	Rat/M	10%	48	2.34	79.6
Mean relative kidney weight NTP (1995)	Rat/F	10%	133	7.46	231
Kidney inflammation NTP (1995)	Rat/F	10%	200	12.6	359
Kidney transitional epithelial hyperplasia NTP (1995)	Rat/M	10%	16	0.745	26.1
Kidney transitional epithelial hyperplasia NTP (1995)	Rat/F	10%	339	27.9	638

8 ^a Average blood concentration of *tert*-butanol under continuous oral exposure at the BMDL.

9 ^b Continuous inhalation human equivalent concentration that leads to the same average blood concentration of
 10 *tert*-butanol as continuous oral exposure at the BMDL.
 11

12 ***PODs carried forth to derivation of candidate values***

13 For the derivation of candidate values, it must be considered whether PODs from the
 14 inhalation study of [NTP \(1997\)](#) would provide a better basis than the route-to-route extrapolated
 15 PODs based on the oral study of [NTP \(1995\)](#). The only endpoint available from [NTP \(1997\)](#) is
 16 increased kidney weights. The corresponding PODs from this subchronic inhalation study are
 17 substantially higher than those for the same endpoint derived by route-to-route extrapolation from
 18 the chronic study ([NTP, 1995](#)), consistent with longer duration requiring a lower dose to elicit an
 19 effect. Additionally, as discussed in Section 2.1.3, kidney weight is a less-specific endpoint
 20 compared to some of the other endpoints available for analysis from the oral study ([NTP, 1995](#)).
 21 Therefore, the PODs derived from PBPK model-based route-to-route extrapolation are the
 22 preferred basis for deriving kidney-specific candidate RfCs, as they are based on a longer (chronic)
 23 duration and a more specific endpoint.

1 **2.2.3. Derivation of Candidate Values**

2 Under EPA's *A Review of the Reference Dose and Reference Concentration Processes* ([U.S. EPA, 2002; Section 4.4.5](#)), also described in the Preamble, five possible areas of uncertainty and
3 variability were considered. An explanation follows:
4

5 An intraspecies uncertainty factor, UF_H , of 10 was applied to all PODs to account for
6 potential differences in toxicokinetics and toxicodynamics in the absence of information on the
7 variability of response in the human population following inhalation exposure to *tert*-butanol.

8 An interspecies uncertainty factor, UF_A , of 3 ($10^{1/2} = 3.16$, rounded to 3) was applied to all
9 PODs to account for residual uncertainty in the extrapolation from laboratory animals to humans in
10 the absence of information to characterize toxicodynamic differences between rodents and humans
11 after inhalation exposure to *tert*-butanol. This value is adopted by convention where an adjustment
12 from animal to a human equivalent concentration has been performed as described in EPA's
13 *Methods for Derivation of Inhalation reference Concentrations and Application of Inhalation*
14 *Dosimetry* ([U.S. EPA, 1994](#)).

15 A subchronic to chronic uncertainty factor, UF_S , of 1 was applied to the PODs derived from
16 the [NTP \(1995\)](#) study, as the endpoints were observed following chronic exposure. For the PODs
17 derived from the subchronic [NTP \(1997\)](#) study, a UF_S of 10 was applied to account for extrapolation
18 from subchronic to chronic duration.

19 A LOAEL to NOAEL uncertainty factor, UF_L , of 1 was applied to all PODs because the current
20 approach is to address this factor as one of the considerations in selecting a BMR for benchmark
21 dose modeling. In this case, BMRs of a 10% change in kidney weight, a 10% extra risk of kidney
22 inflammation, and a 10% extra risk of transitional cell hyperplasia were selected under an
23 assumption that they represent minimal biologically significant changes.

24 A database uncertainty factor, UF_D , of 1 was applied to all PODs. The *tert*-butanol toxicity
25 database includes a chronic toxicity study in rats and mice ([NTP, 1995](#)), a subchronic toxicity study
26 in rats and mice ([NTP, 1997](#)), and developmental toxicity studies in rats and mice ([Lyondell
27 Chemical Co., 2004; Faulkner et al., 1989; Daniel and Evans, 1982](#)). In the developmental studies,
28 no effects were observed at exposure levels below 1000 mg/kg-day, and effects observed at
29 ≥ 1000 mg/kg-day were accompanied by evidence of maternal toxicity. These exposure levels are
30 much higher than the PODs for kidney effects, suggesting developmental toxicity is not a sensitive
31 endpoint. The *tert*-butanol database contains a one-generation reproductive toxicity study in rats
32 ([Lyondell Chemical Co., 2004](#)), though no multigenerational reproductive study has been
33 performed. There are no immunotoxicity studies for *tert*-butanol. Information provided by studies
34 on ETBE, which is rapidly metabolized to systemically-available *tert*-butanol, can help in
35 considering the lack of a *tert*-butanol multigenerational reproductive study or an immunotoxicity
36 study. No adverse effects were reported in one- and two-generation reproductive/developmental
37 studies on ETBE ([Gaoua, 2004a, b](#)), and the database for ETBE does not indicate immunotoxicity
38 ([Banton et al., 2011; Li et al., 2011](#)). Thus, although there are some gaps in the toxicity database for

1 *tert*-butanol, the available data on *tert*-butanol, informed by the data on ETBE, do not suggest that
 2 additional studies would lead to identification of a more sensitive endpoint or a lower POD.
 3 Therefore, a database UF_D of 1 was applied.

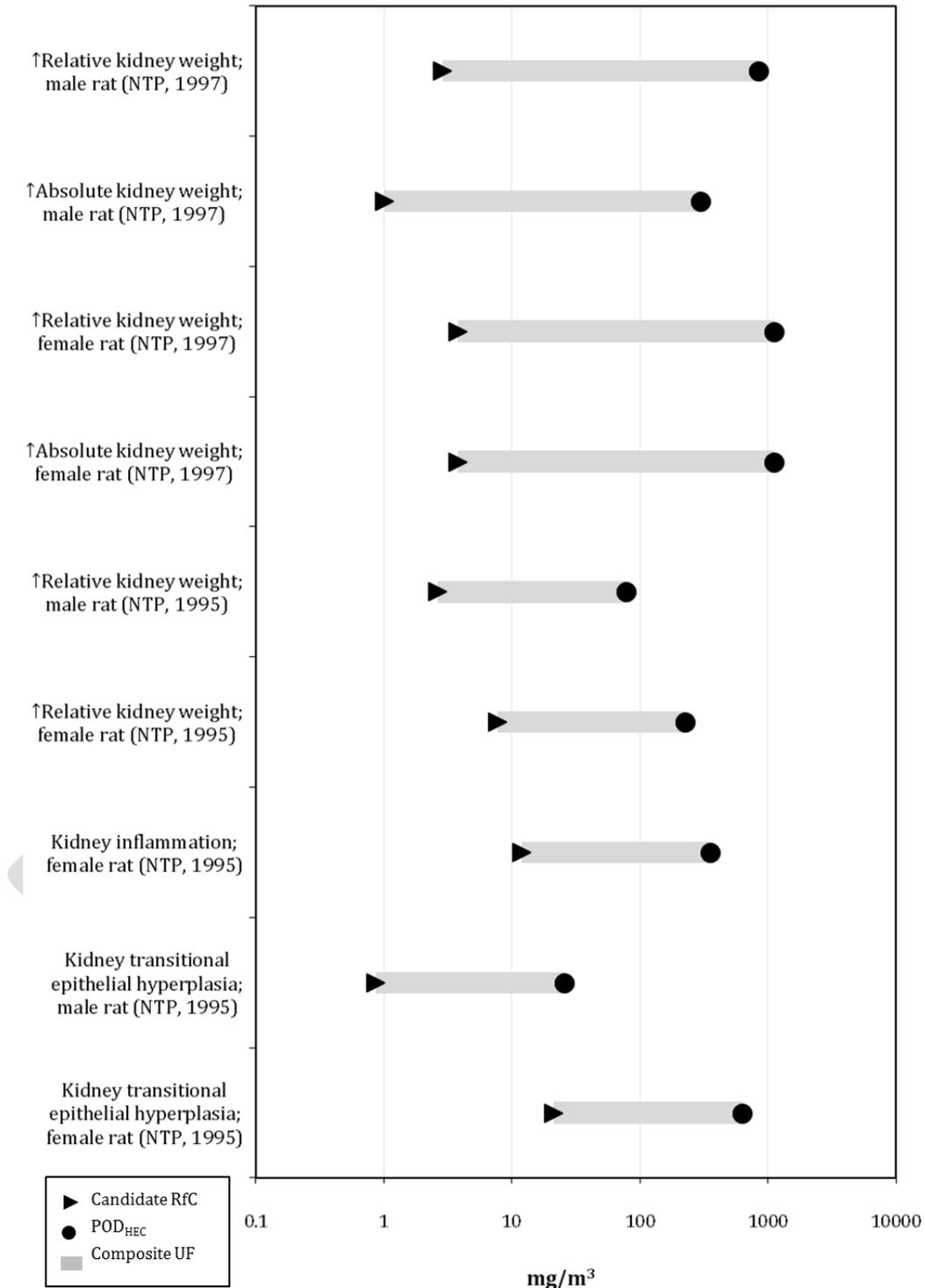
4 Table 2-6 is a continuation of Table 2-4 and Table 2-5, and summarizes the application of
 5 UFs to each POD to derive a candidate value for each data set. The candidate values presented in
 6 the table below are preliminary to the derivation of the organ/system-specific reference values.
 7 These candidate values are considered individually in the selection of a representative inhalation
 8 reference value for a specific hazard and subsequent overall RfC for *tert*-butanol.

9 **Table 2-6. Effects and corresponding derivation of candidate values**

Endpoint (Sex and species) and Reference	POD _{HEC} ^a (mg/m ³)	POD type	UF _A	UF _H	UF _L	UF _S	UF _D	Composite UF	Candidate value (mg/m ³)
<i>Kidney</i>									
Increased relative kidney weight; male rat NTP (1997)	861	BMCL _{10%}	3	10	1	10	1	300	3 × 10 ⁰
Increased absolute kidney weight; male rat NTP (1997)	304	BMCL _{10%}	3	10	1	10	1	300	1 × 10 ⁰
Increased relative kidney weight; female rat NTP (1997)	1137	NOAEL	3	10	1	10	1	300	4 × 10 ⁰
Increased absolute kidney weight; female rat NTP (1997)	1137	NOAEL	3	10	1	10	1	300	4 × 10 ⁰
Increased relative kidney weight; male rat NTP (1995)	79.6	BMCL _{10%}	3	10	1	1	1	30	3 × 10 ⁰
Increased relative kidney weight; female rat NTP (1995)	231	BMCL _{10%}	3	10	1	1	1	30	8 × 10 ⁰
Kidney inflammation; female rat NTP (1995)	359	BMCL _{10%}	3	10	1	1	1	30	1 × 10 ¹
Kidney transitional epithelial hyperplasia; male rat NTP (1995)	26.1	BMCL _{10%}	3	10	1	1	1	30	9 × 10 ⁻¹
Kidney transitional epithelial hyperplasia; female rat NTP (1995)	638	BMCL _{10%}	3	10	1	1	1	30	2 × 10 ¹

10

1



2

3

4

Figure 2-2. Candidate RfC values with corresponding POD and composite UF.

1 **2.2.4. Derivation of Organ/System-Specific Reference Concentrations**

2 Table 2-7 distills the candidate values from Table 2-6 into a single value for the kidney.
 3 Organ or system-specific reference values may be useful for subsequent cumulative risk
 4 assessments that consider the combined effect of multiple agents acting at a common site.

5 ***Kidney Toxicity***

6 For *tert*-butanol, candidate values were for several different effects in both sexes, spanning
 7 a range from 9×10^{-1} to 2×10^1 mg/m³, for an overall twenty-fold range. To estimate an exposure
 8 level below which kidney toxicity from *tert*-butanol exposure is not expected to occur, the RfC for
 9 increased incidence of transitional epithelial hyperplasia in male rats (9×10^{-1} mg/m³) is proposed
 10 as the kidney-specific reference concentration for *tert*-butanol, consistent with the selection of the
 11 kidney-specific RfD (see Section 2.1.4). As discussed previously, unlike kidney inflammation, this
 12 effect was observed in both sexes, with males appearing to be more sensitive than females.
 13 Additionally, it is a more specific and more sensitive indicator of kidney toxicity than the relatively
 14 non-specific endpoint of kidney weight changes. Confidence in this kidney-specific RfC is medium.
 15 The PODs are based on modeled benchmark dose estimates, and the candidate values are derived
 16 from a well-conducted study, involving a sufficient number of animals per group, including both
 17 sexes, assessing a wide range of kidney endpoints, and availability of a PBPK model for route-to-
 18 route extrapolation.

19

20 **Table 2-7. Organ/system-specific RfCs and proposed overall RfC for**
 21 ***tert*-butanol**

Effect	Basis	RfC (mg/m ³)	Exposure description	Confidence
Kidney toxicity	Increased incidence of transitional epithelial hyperplasia	9×10^{-1}	Chronic	HIGH
Proposed overall RfC	Increased incidence of transitional epithelial hyperplasia	9×10^{-1}	Chronic	HIGH

22

23 **2.2.5. Selection of the Proposed Overall Reference Concentration**

24 For *tert*-butanol, only kidney effects were identified as a hazard; thus, a single
 25 organ/system-specific reference concentration was derived. Therefore, the kidney-specific RfC of
 26 9×10^{-1} mg/m³ is also proposed as an estimated exposure level below which deleterious effects
 27 from *tert*-butanol exposure are not expected to occur.

28 The overall reference concentration is derived to be protective of all types of effects for a
 29 given duration of exposure and is intended to protect the population as a whole including

1 potentially susceptible subgroups ([U.S. EPA, 2002](#)). Decisions concerning averaging exposures over
2 time for comparison with the RfC should consider the types of toxicological effects and specific
3 lifestages of concern. Fluctuations in exposure levels that result in elevated exposures during these
4 lifestages could potentially lead to an appreciable risk, even if average levels over the full exposure
5 duration were less than or equal to the RfC. In the case of *tert*-butanol, no specific lifestages have
6 been identified has a potentially susceptible subgroup.

7 **2.2.6. Confidence Statement**

8 A confidence level of high, medium, or low is assigned to the study used to derive the RfC,
9 the overall database, and the RfC itself, as described in Section 4.3.9.2 of EPA's *Methods for*
10 *Derivation of Inhalation Reference Concentrations and Application of Inhalation Dosimetry* ([U.S. EPA,](#)
11 [1994](#)). The overall confidence in this RfC is high. Confidence in the principal study ([NTP, 1995](#)) is
12 high. This study was well-conducted, compiled with FDA GLP regulations, involved a sufficient
13 number of animals per group (including both sexes), and assessed a wide range of tissues and
14 endpoints. Although there are some gaps in the toxicity database for *tert*-butanol, these areas are
15 informed by the data on ETBE, a parent compound of *tert*-butanol. Therefore, the confidence in the
16 database is high. Reflecting high confidence in the principal study and high confidence in the
17 database, confidence in the RfC is high.

18 **2.2.7. Previous IRIS Assessment**

19 An inhalation assessment for *tert*-butanol was not previously available on IRIS.

20 **2.2.8. Uncertainties in the Derivation of the Reference Dose and Reference Concentration**

21 The following discussion identifies uncertainties associated with the RfD and RfC for
22 *tert*-butanol. To derive the RfD, the UF approach ([U.S. EPA, 2000a, 1994](#)) was applied to a POD
23 based on kidney toxicity in rats treated chronically. To derive the RfC, this same approach was
24 applied, but a PBPK model was used to extrapolate from oral to inhalation exposure. UFs were
25 applied to the POD to account for extrapolating from an animal bioassay to human exposure, the
26 likely existence of a diverse population of varying susceptibilities, and database deficiencies. These
27 extrapolations are carried out with default approaches given the lack of data to inform individual
28 steps.

29 The database for *tert*-butanol contains no human data on adverse health effects from
30 subchronic or chronic exposure. Data on the effects of *tert*-butanol are derived from a small
31 database of studies in rats and mice. The database for *tert*-butanol exposure includes one lifetime
32 bioassay, several reproductive/developmental studies, and several subchronic studies.

33 Although the database is adequate for reference value derivation, there is uncertainty
34 associated with the lack of a comprehensive multigeneration reproductive toxicity study.
35 Additionally, only subchronic and short-term inhalation studies have been conducted, and no
36 chronic inhalation studies are available. Developmental studies identified significant increases in

1 fetal loss, decreases in fetal body weight, and possible increases in skeletal variations in exposed
2 offspring or pups. However, effects were not always consistent across exposure routes, and
3 significant material toxicity was present whenever developmental effects were observed.

4 The toxicokinetic and toxicodynamic differences for *tert*-butanol between the animal
5 species in which the POD was derived and humans are unknown. Although sufficient information is
6 available to develop a PBPK model in rats to evaluate difference across routes of exposure, the
7 *tert*-butanol database lacks an adequate model that would inform potential interspecies differences.
8 Generally, it was found that rats appear more susceptible than mice, and males appear more
9 susceptible than females to *tert*-butanol toxicity. However, the underlying mechanistic basis of
10 these apparent differences is not understood. Most importantly, it is unknown which animal
11 species and/or sexes may be more comparable to humans.

12 **2.3. ORAL SLOPE FACTOR FOR CANCER**

13 The carcinogenicity assessment provides information on the carcinogenic hazard potential
14 of the substance in question and quantitative estimates of risk from oral and inhalation exposure
15 that may be derived. Quantitative risk estimates may be derived from the application of a low-dose
16 extrapolation procedure. If derived, the oral slope factor is a plausible upper bound on the estimate
17 of risk per mg/kg-day of oral exposure.

18 **2.3.1. Analysis of Carcinogenicity Data**

19 As noted in Section 1.2.2, EPA concluded that there is “suggestive evidence of carcinogenic
20 potential” for *tert*-butanol. The *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)) state:

21
22 When there is suggestive evidence, the Agency generally would not attempt a dose-
23 response assessment, as the nature of the data generally would not support one; however
24 when the evidence includes a well-conducted study, quantitative analysis may be useful for
25 some purposes, for example, providing a sense of the magnitude and uncertainty of
26 potential risks, ranking potential hazards, or setting research priorities.
27

28 The only data available on potential carcinogenicity was derived from the 2-year drinking
29 water study in rats and mice by ([NTP, 1995](#)). This study was considered suitable for dose-response
30 analysis. It was conducted in accordance with Food and Drug Administration (FDA) Good
31 Laboratory Practice (GLP) Regulations, and all aspects were subjected to retrospective quality
32 assurance audits. The study included histological examinations for tumors in many different
33 tissues, contained three exposure levels and controls, contained adequate numbers of animals per
34 dose group (~50/sex/group), treated animals for up to 2 years, and included detailed reporting of
35 methods and results. Additionally, the renal tumors were re-examined by a Pathology Working
36 Group ([Hard et al., 2011](#)).

37 Dose-related increasing trends in tumors were noted at the following sites:

- Renal tubule adenomas and carcinomas in male rats; and
- Thyroid follicular adenomas in female mice.

These tumors were statistically significantly increased by pairwise comparison (Fisher exact test, $p \leq 0.05$) and by trend test (Cochran-Armitage trend test, $p \leq 0.05$). Based on analysis of mode of action data, it was concluded that processes other than α_{2u} -globulin nephropathy are likely responsible for the male rat renal tumors, so these tumors may be suitable for quantitative analysis (U.S. EPA, 1991a). Additionally, a thyroid follicular carcinoma was observed in male mice, so it is possible that the thyroid follicular adenomas in female mice could progress to malignant form. Therefore, the thyroid follicular adenomas in female mice may also be considered suitable for quantitative analysis. Considering these data along with the uncertainty associated with the suggestive nature of the weight of evidence, EPA concluded that quantitative analyses may be useful for providing a sense of the magnitude of potential carcinogenic risk.

2.3.2. Dose-Response Analysis—Adjustments and Extrapolations Methods

The U.S. EPA *Guidelines for Carcinogen Risk Assessment* (U.S. EPA, 2005a) recommends that the method used to characterize and quantify cancer risk from a chemical is determined by what is known about the MOA of the carcinogen and the shape of the cancer dose-response curve. The linear approach is recommended if the MOA of carcinogenicity has not been established (U.S. EPA, 2005a). In the case of *tert*-butanol, the modes of carcinogenic action for renal tubule and thyroid follicular tumors are not fully understood (see Section 1.2.2). Therefore, a linear low-dose extrapolation approach was used to estimate human carcinogenic risk associated with *tert*-butanol exposure.

The modeled *tert*-butanol PODs were scaled to HEDs according to EPA guidance (U.S. EPA, 2011, 2005a). In particular, the BMDL was converted to an HED by assuming that doses in animals and humans are toxicologically equivalent when scaled by body weight raised to the $3/4$ power. Standard body weights of 0.025 for mice, 0.25 kg for rats, and 70 kg for humans was used (U.S. EPA, 1988). The following formula was used for the conversion of oral BMDL to oral HED for rat endpoints:

$$\begin{aligned} \text{Scaled HED in mg/kg-day} &= (\text{BMDL in mg/kg-day}) \times (\text{animal body weight}/70)^{1/4} \\ &= (\text{BMDL in mg/kg-day}) \times 0.24 \end{aligned}$$

Details of the modeling and the model selection process can be found in Appendix C of the Supplemental Information. PODs for estimating low-dose risk were identified at doses at the lower end of the observed data, generally corresponding to 10% extra risk. Because initial modeling overestimated the control, due to the non-monotonicity of the observed dose-response, the POD

1 was derived after dropping the highest exposure group ([U.S. EPA, 2012b](#)). The highest exposure
2 group also had increased mortality, which may in part explain the observed non-monotonicity.

3 **2.3.3. Derivation of the Oral Slope Factor**

4 The PODs estimated for each tumor site are summarized in Table 2-8. The lifetime oral
5 cancer slope factor for humans is defined as the slope of the line from the lower 95% bound on the
6 exposure at the POD to the control response (slope factor = 0.1/BMDL₁₀). This slope, a 95% upper
7 confidence limit, represents a plausible upper bound on the true risk. Using linear extrapolation
8 from the BMDL₁₀, human equivalent oral slope factors were derived for each species/tumor site
9 combination and are listed in Table 2-8.

10 The oral slope factors derived from the [NTP \(1995\)](#) bioassay differ by twenty-fold,
11 depending on the species and tumor site. The most sensitive endpoint of renal tumors was used to
12 derive the oral slope factor because there are no data to support any one result as most relevant for
13 extrapolating to humans. Two slope factors were derived for this endpoint from the [NTP \(1995\)](#)
14 bioassay, one based on the original reported incidences and the other based on the [Hard et al.](#)
15 [\(2011\)](#) reanalysis. The two estimates differed by less than 20%, and rounded to the same number
16 at one significant figure. However, the [Hard et al. \(2011\)](#) reanalysis is considered preferable, as it is
17 based on a PWG analysis. Therefore, the recommended slope factor for providing a sense of the
18 magnitude of potential carcinogenic risk associated with lifetime oral exposure to *tert*-butanol is
19 **1 × 10⁻² per mg/kg-day**, based on the renal tubule tumor response in male F344 rats.

20 **Table 2-8. Summary of the oral slope factor derivations**

Tumor	Species/Sex	Selected Model	BMR	BMD (mg/kg-d)	POD= BMDL (mg/kg-d)	BMDL _{HED} ^a (mg/kg-d)	Slope factor ^b (mg/kg-day) ⁻¹
Renal tubule adenoma or carcinoma	Male F344 rat; dose as administered	1° Multistage (high dose dropped)	10%	70	42	10.1	1 × 10 ⁻²
Renal tubule adenoma or carcinoma [Hard et al. (2011) reanalysis]	Male F344 rat; dose as administered	1° Multistage (high dose dropped)	10%	54	36	8.88	1 × 10 ⁻²
Thyroid follicular cell adenoma	Female B6C3F1 mouse	3° Multistage	10%	2002	1437	201	5 × 10 ⁻⁴

21
22 ^aHED PODs were calculated using BW^{3/4} scaling ([U.S. EPA, 2011](#)).

23 ^bHuman equivalent slope factor = 0.1/BMDL_{10HED}; see Appendix C of the Supplemental Information for details of
24 modeling results.

1 **2.3.4. Uncertainties in the Derivation of the Oral Slope Factor**

2 There is uncertainty when extrapolating data from animals to estimate potential cancer
 3 risks to human populations from exposure to *tert*-butanol (see Table 2-9). Uncertainty in the
 4 magnitude of the recommended oral slope factor is reflected to some extent in the range of slope
 5 factors; the oral slope factor based on the male rat data was about twenty-fold higher than the oral
 6 slope factor based on female mouse data (Table 2-9). These comparisons show that the selection of
 7 target organ, animal species, and interspecies extrapolation can impact the oral cancer risk
 8 estimate. Although the thyroid follicular cell tumors occurred in male and female mice, high
 9 mortality in high-dose male mice limited the usefulness of the data. Renal tubule tumors occurred
 10 in male rats, but not female rats. Therefore, only the data in male rats and female mice were
 11 available for deriving the oral slope factor. There are no other chronic studies to replicate these
 12 findings or that examined other animal models. There are no data in humans to support the tumors
 13 observed in animals. Although changing the methods used to derive the oral slope factor could
 14 change the results, standard practices were used due to the lack of a mouse or human PBPK model
 15 or specific MOA to indicate other methods would be preferable. Additionally, considering the
 16 uncertainty associated with the suggestive nature of the weight of evidence, the oral slope factor is
 17 recommended only for providing a sense of the magnitude of potential carcinogenic risk.

18 **Table 2-9. Summary of uncertainties in the derivation of cancer risk values for**
 19 ***tert*-butanol**

Consideration and Impact on Cancer Risk Value	Decision	Justification and Discussion
Selection of target organ ↓ oral slope factor, up to twenty-fold, if renal tumors not selected.	The kidney was selected as the target organ.	As there are no data to support any one result as most relevant for extrapolating to humans, the most sensitive result for kidney renal tubular adenomas and carcinomas was used to derive the oral slope factor. However, the overall evidence for carcinogenicity was considered “suggestive.”
Selection of data set Unknown change in oral slope factor, since no other studies are available.	NTP (1995) as principal oral (drinking water) study to derive cancer risks for humans.	NTP (1995) was a well-conducted study. It was also the only bioassay available. Additional bioassays might add support to the findings or provide results for different (possibly lower) doses, which may affect the oral slope factor.
Selection of extrapolation approach (Selection of extrapolation approach could change the recommended cancer risk values.)	Oral data used for OSF.	No extrapolation methods were used.
Selection of dose metric Alternatives could ↓ or ↑ slope factor	Used administered dose converted to HED units.	Additional runs using the administered dose without conversion to HED units were also conducted, resulting in a similar oral slope

Consideration and Impact on Cancer Risk Value	Decision	Justification and Discussion
		factor. For rats, a PBPK model of internal dose was available, but the POD changed by less than 1.2-fold when modeling was based on internal doses. For mice, no PBPK model was available, so using a PBPK model for determining internal doses could have an unknown effect on the estimated OSF value.
Interspecies extrapolation of dosimetry and risk Alternatives could ↓ or ↑ slope factor (e.g., 3.5-fold ↓ [scaling by body weight] or ↑ 2-fold [scaling by BW ^{2/3}])	The default approach of body weight ^{3/4} was used.	There are no data to suggest an alternative approach. Because the dose metric was not an area under the curve, BW ^{3/4} scaling was used to calculate equivalent cumulative exposures for estimating equivalent human risks. While the true human correspondence is unknown, this overall approach is expected to neither over- nor underestimate human equivalent risks.
Dose-response modeling Alternatives could ↓ or ↑ slope factor	Used multistage dose-response model to derive a BMD and BMDL.	No biologically based models for <i>tert</i> -butanol were available. The multistage model has biological support and is the model most consistently used in EPA cancer assessments.
Low-dose extrapolation ↓ cancer risk estimate would be expected with the application of nonlinear low-dose extrapolation	Linear extrapolation of risk in low-dose region used.	Linear low-dose extrapolation for agents without a known MOA is supported.
Statistical uncertainty at POD ↓ oral slope factor 1.7-fold if BMD used as the POD rather than BMDL	BMDL (preferred approach for calculating plausible upper bound slope factor)	Limited size of bioassay results in sampling variability; lower bound is 95% CI on administered exposure at 10% extra risk of renal tumors.
Sensitive subpopulations ↑ oral slope factor to unknown extent	No sensitive populations have been identified.	No chemical-specific data are available to determine the range of human toxicodynamic variability or sensitivity, including the susceptibility of children. Because determination of a mutagenic MOA has not been made, an age-specific adjustment factor is not applied.

1

2 **2.3.5. Previous IRIS Assessment: Oral Slope Factor**

3 A cancer assessment for *tert*-butanol was not previously available on IRIS.

4 **2.4. INHALATION UNIT RISK FOR CANCER**

5 The carcinogenicity assessment provides information on the carcinogenic hazard potential
6 of the substance in question and quantitative estimates of risk from oral and inhalation exposure
7 may be derived. Quantitative risk estimates may be derived from the application of a low-dose

1 extrapolation procedure. If derived, the inhalation unit risk is a plausible upper bound on the
2 estimate of risk per $\mu\text{g}/\text{m}^3$ air breathed.

3 **2.4.1. Analysis of Carcinogenicity Data**

4 As noted in Section 1.2.2, EPA concluded that there is “suggestive evidence of carcinogenic
5 potential” for *tert*-butanol. The *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)) state:

6
7 When there is suggestive evidence, the Agency generally would not attempt a dose-
8 response assessment, as the nature of the data generally would not support one; however,
9 when the evidence includes a well-conducted study, quantitative analysis may be useful for
10 some purposes. For example, it could provide a sense of the magnitude and uncertainty of
11 potential risks, rank potential hazards, or set research priorities.
12

13 The only data available on potential carcinogenicity were from the 2-year drinking water
14 study in rats and mice by [NTP \(1995\)](#), discussed previously in Section 2.3.1. Because a PBPK model
15 for the rat is available to conduct route-to-route extrapolation (discussed below), the male rat renal
16 tubule adenoma and carcinoma data are suitable for quantitative analysis to support an inhalation
17 unit risk. Considering these data and uncertainty associated with the suggestive nature of the
18 weight of evidence, EPA concluded that quantitative analyses may be useful for providing a sense of
19 the magnitude of potential carcinogenic risk.

20 **2.4.2. Dose Response Analysis – Adjustments and Extrapolation Methods**

21 Details of the modeling and the model selection process can be found in Appendix C of the
22 Supplemental Information. A POD for estimating low-dose risk was identified at doses at the lower
23 end of the observed data corresponding to 10% extra risk.

24 A PBPK model for *tert*-butanol in rats has been developed, as described in Appendix B.
25 Using this model, route-to-route extrapolation of the oral BMDL to derive an inhalation POD was
26 performed as follows. First, the internal dose in the rat at the oral BMDL (assuming continuous
27 exposure) was estimated using the PBPK model, to derive an “internal dose BMDL.” Then, the
28 inhalation air concentration (again assuming continuous exposure) that led to the same internal
29 dose in the rat was estimated using the PBPK model, resulting in a route-to-route extrapolated
30 BMCL.

31 A critical decision in the route-to-route extrapolation is the selection of the internal dose
32 metric to use that established “equivalent” oral and inhalation exposures. For *tert*-butanol-induced
33 kidney effects, the two options are the concentration of *tert*-butanol in blood and rate of *tert*-
34 butanol metabolism. Note that using the kidney concentration of *tert*-butanol will lead to the same
35 route-to-route extrapolation relationship as *tert*-butanol in blood, since the distribution from blood
36 to kidney is independent of route. There are no data that suggest metabolites of *tert*-butanol
37 mediate its renal toxicity. In the absence of evidence that would suggest otherwise, it is assumed

1 that *tert*-butanol itself is the active toxicological agent. Therefore, the concentration of *tert*-butanol
 2 in blood was selected as the dose metric to derive the BMCL.

3 The RfC methodology provides a mechanism for deriving a HEC from the BMCL determined
 4 from the animal data. The approach takes into account the extra-respiratory nature of the
 5 toxicological responses and accommodates species differences by considering blood:air partition
 6 coefficients for *tert*-butanol in the laboratory animal (rat or mouse) and humans. According to the
 7 RfC guidelines ([U.S. EPA, 1994](#)), *tert*-butanol is a Category 3 gas because extra-respiratory effects
 8 were observed. [Kaneko et al. \(2000\)](#) measured a blood:gas partition coefficient of 531 ± 102 for *tert*-
 9 butanol in the male Wistar rat, while [Borghoff et al. \(1996\)](#) measured a value of 481 ± 29 in male
 10 F344 rats. A blood:gas partition coefficient of 462 was reported for *tert*-butanol in humans ([Nihlén](#)
 11 [et al., 1995](#)). The calculation $(H_{b/g})_A \div (H_{b/g})_H$ was used to calculate a blood:gas partition coefficient
 12 ratio to apply to the delivered concentration. Because F344 rats were used in the study, the
 13 blood:gas partition coefficient for F344 rats was used. Thus, the calculation was: $481 \div 462 = 1.04$.
 14 Therefore, a ratio of 1.04 was used to calculate the HEC. This allowed a $BMCL_{HEC}$ to be derived as
 15 follows:

$$\begin{aligned} 16 \quad BMCL_{HEC} &= BMCL_{ADJ} \text{ (mg/m}^3\text{)} \times \text{(interspecies conversion)} \\ 17 &= BMCL_{ADJ} \text{ (mg/m}^3\text{)} \times (481 \div 462) \\ 18 &= BMCL_{ADJ} \text{ (mg/m}^3\text{)} \times (1.04) \end{aligned}$$

19
 20
 21 The U.S. EPA *Guidelines for Carcinogen Risk Assessment* ([U.S. EPA, 2005a](#)) recommend that
 22 the method used to characterize and quantify cancer risk from a chemical is determined by what is
 23 known about the MOA of the carcinogen and the shape of the cancer dose-response curve. The
 24 linear approach is recommended if the MOA of carcinogenicity has not been established ([U.S. EPA,](#)
 25 [2005a](#)). In the case of *tert*-butanol, the mode of carcinogenic action for renal tubule tumors is not
 26 fully understood (see Section 1.2.2). Therefore, a linear low-dose extrapolation approach was used
 27 to estimate human carcinogenic risk associated with *tert*-butanol exposure.

28 **2.4.3. Inhalation Unit Risk Derivation**

29 The results from route-to-route extrapolation of the male rat renal tubule tumor data are
 30 summarized in Table 2-10. The lifetime inhalation unit risk for humans is defined as the slope of
 31 the line from the lower 95% bound on the exposure at the POD to the control response (inhalation
 32 unit risk = $0.1/BMCL_{10}$). This slope, a 95% upper confidence limit represents a plausible upper
 33 bound on the true risk. Using linear extrapolation from the $BMCL_{10}$, a human equivalent inhalation
 34 unit risk was derived, as listed in Table 2-10.

35 Two inhalation unit risks were derived from the [NTP \(1995\)](#) bioassay: one based on the
 36 original reported incidences and one based on the [Hard et al. \(2011\)](#) reanalysis. The two estimates
 37 differ by less than 20%, but the [Hard et al. \(2011\)](#) reanalysis is considered preferable, as it is based

1 on a PWG analysis. Therefore, the recommended inhalation unit risk for providing a sense of the
 2 magnitude of potential carcinogenic risk associated with lifetime inhalation exposure to
 3 *tert*-butanol is 2×10^{-3} per mg/m^3 , or 2×10^{-6} per $\mu\text{g}/\text{m}^3$, based on the renal tubule tumor
 4 response in male F344 rats.

5 **Table 2-10. Summary of the inhalation unit risk derivation**

Tumor	Species/Sex	BMR	BMDL (mg/kg-d)	Internal Dose ^a (mg/L)	POD= BMCL _{HEC} ^c (mg/m ³)	Unit Risk ^b (mg/m ³) ⁻¹
Renal tubule adenoma or carcinoma	Male F344 rat	10%	41.6	2.01	68.7	1×10^{-3}
Renal tubule adenoma or carcinoma [Hard et al. (2011) reanalysis]	Male F344 rat	10%	36.3	1.74	59.8	2×10^{-3}

6 ^a Average blood concentration of *tert*-butanol under continuous oral exposure at the BMDL.
 7 ^b Continuous inhalation human equivalent concentration that leads to the same average blood concentration of
 8 *tert*-butanol as continuous oral exposure at the BMDL.
 9 ^c Human equivalent inhalation unit risk = 0.1/BMCL_{HEC}.
 10

11 **2.4.4. Uncertainties in the Derivation of the Inhalation Unit Risk**

12 There is uncertainty when extrapolating data from animals to estimate potential cancer
 13 risks to human populations from exposure to *tert*-butanol (see Table 2-11). Uncertainty in the
 14 magnitude of the recommended inhalation unit risk can be inferred to some extent from the range
 15 of oral slope factors; the oral slope factor based on the male rat data was about twenty-fold higher
 16 than the oral slope factor based on female mouse data (Table 2-9). These comparisons show that
 17 the selection of target organ, animal species, and interspecies extrapolation can impact the
 18 inhalation unit risk estimate. Although the thyroid follicular cell tumors occurred in male and
 19 female mice, high mortality in high-dose male mice limited the usefulness of the data. Additionally,
 20 no PBPK model was available in mice for use in route-to-route extrapolation, so these data could
 21 not be used to estimate an inhalation unit risk. Renal tubule tumors occurred in male rats, but not
 22 female rats. Therefore, only the data in male rats were available for deriving the inhalation unit
 23 risk. There are no other chronic studies to replicate these findings or that examined other animal
 24 models. There are no data in humans to support the tumors observed in animals. Although
 25 changing the methods used to derive the inhalation unit risk could change the results, standard
 26 practices were used due to the lack of a mouse or human PBPK model or specific MOA to indicate
 27 other methods which would be preferable. Additionally, considering the uncertainty associated
 28 with the suggestive nature of the weight of evidence, the inhalation unit risk is recommended only
 29 for providing a sense of the magnitude of potential carcinogenic risk.

1 **Table 2-11. Summary of uncertainties in the derivation of cancer risk values**
 2 **for *tert*-butanol**

Consideration and Impact on Cancer Risk Value	Decision	Justification and Discussion
Selection of target organ Inhalation unit risk may change by an unknown amount if a PBPK model to extrapolate mouse thyroid tumors to the inhalation route were available.	The kidney was selected as the target organ.	No PBPK model to extrapolation mouse thyroid tumors was available. Additionally, the overall evidence for carcinogenicity was considered “suggestive.”
Selection of data set Unknown change in inhalation unit risk, since no other studies are available.	NTP (1995) as principal oral (drinking water) study to derive cancer risks for humans.	NTP (1995) was a well-conducted study. It was also the only bioassay available. Additional bioassays might add support to the findings or provide results for different (possibly lower) doses, which may affect the inhalation unit risk.
Selection of extrapolation approach Different PBPK model could ↓ or ↑ inhalation unit risk.	PBPK model-based extrapolation of oral data used for inhalation unit risk.	PBPK model accurately predicted <i>tert</i> -butanol toxicokinetics. Data and model predictions were within 2-fold of each other.
Selection of dose metric Alternatives could ↓ or ↑ inhalation unit risk.	Used <i>tert</i> -butanol concentration in blood as the dose metric for route-to-route extrapolation, converted to HEC.	In the absence of evidence that would suggest that metabolites of <i>tert</i> -butanol are responsible for carcinogenicity, it is assumed that <i>tert</i> -butanol itself is the active toxicological agent. An alternative dose metric of <i>tert</i> -butanol metabolism would result in a 1.2-fold decrease in the inhalation unit risk.
Interspecies extrapolation of dosimetry and risk Alternatives could ↓ or ↑ inhalation unit risk..	The default approach for a Category 3 gas was used.	There are no data to suggest an alternative approach. While the true human correspondence is unknown, this overall approach is expected to neither over- nor underestimate human equivalent risks.
Dose-response modeling Alternatives could ↓ or ↑ inhalation unit risk.	Used multistage dose-response model to derive a BMD and BMDL.	No biologically based models for <i>tert</i> -butanol were available. The multistage model has biological support and is the model most consistently used in EPA cancer assessments.
Low-dose extrapolation ↓ cancer risk estimate would be expected with the application of nonlinear low-dose extrapolation .	Linear extrapolation of risk in low-dose region used.	Linear low-dose extrapolation for agents without a known MOA is supported.
Statistical uncertainty at POD ↓ inhalation unit risk 1.7-fold if the BMD used to derive the inhalation POD rather than BMDL.	BMDL (preferred approach for calculating plausible upper bound)	Limited size of bioassay results in sampling variability; lower bound is 95% CI on administered exposure at 10% extra risk of renal tumors.
Sensitive subpopulations	No sensitive populations	No chemical-specific data are available to

Consideration and Impact on Cancer Risk Value	Decision	Justification and Discussion
↑ inhalation unit risk to unknown extent.	have been identified.	determine the range of human toxicodynamic variability or sensitivity, including the susceptibility of children. Because determination of a mutagenic MOA has not been made, an age-specific adjustment factor is not applied.

1

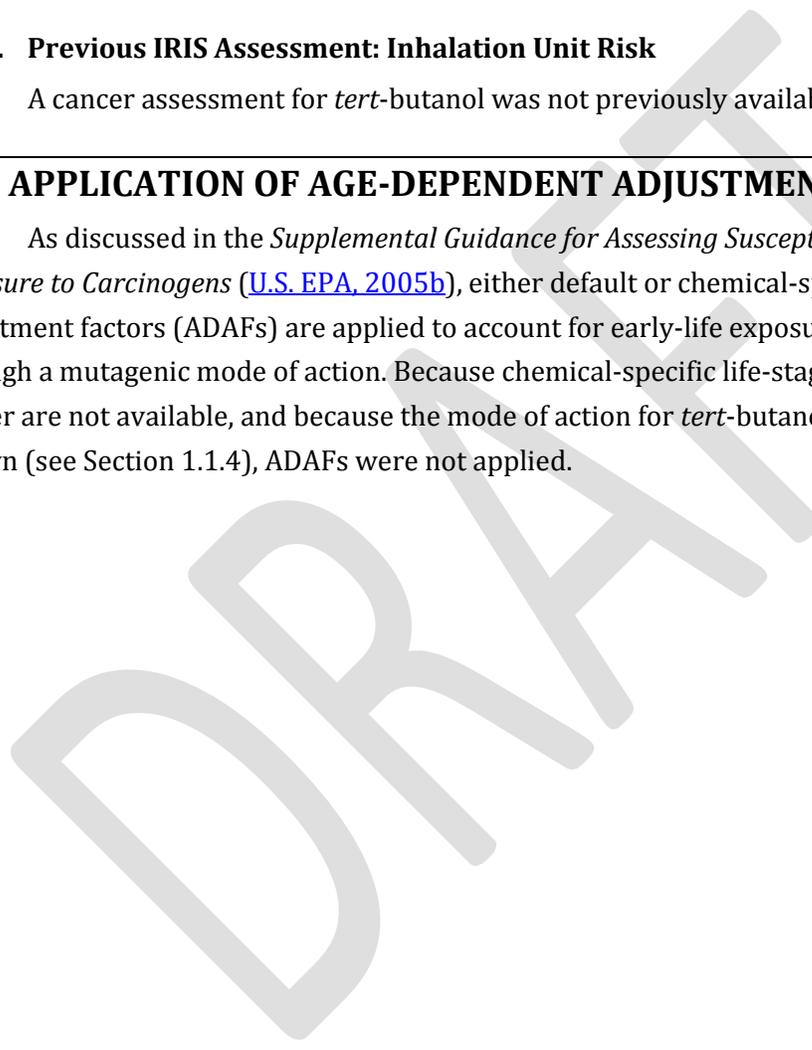
2 **2.4.5. Previous IRIS Assessment: Inhalation Unit Risk**

3 A cancer assessment for *tert*-butanol was not previously available on IRIS.

4 **2.5. APPLICATION OF AGE-DEPENDENT ADJUSTMENT FACTORS**

5 As discussed in the *Supplemental Guidance for Assessing Susceptibility from Early-Life*
 6 *Exposure to Carcinogens* ([U.S. EPA, 2005b](#)), either default or chemical-specific age-dependent
 7 adjustment factors (ADAFs) are applied to account for early-life exposure to carcinogens that act
 8 through a mutagenic mode of action. Because chemical-specific life-stage susceptibility data for
 9 cancer are not available, and because the mode of action for *tert*-butanol carcinogenicity is not
 10 known (see Section 1.1.4), ADAFs were not applied.

11



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