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Effects of ToxCast Phase I Chemicals on Steroidogenesis in H295R Human Adrenocortical Carcinoma Cells

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Abstract No.
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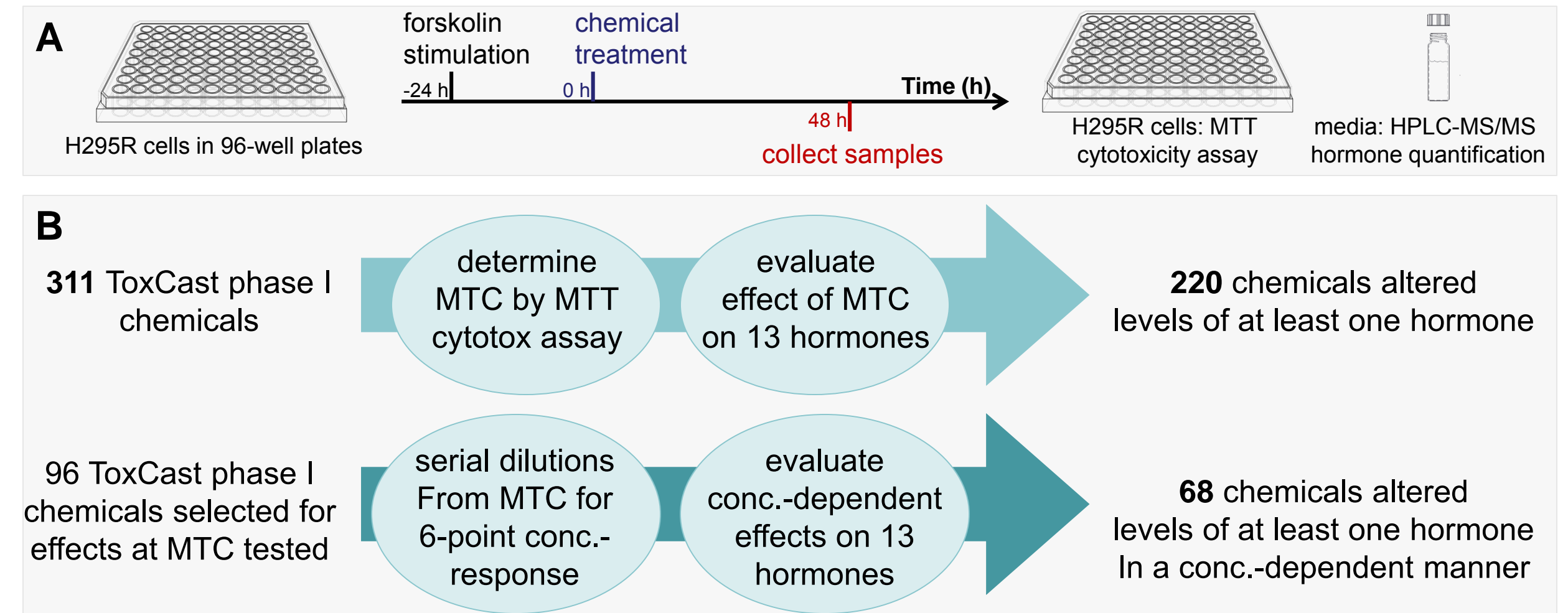
Abstract & Objectives

A delicate balance of steroid hormones is essential for proper development and reproduction. Disruption of steroidogenesis by environmental toxicants results in altered hormone levels causing adverse reproductive and developmental effects. H295R human adrenocortical carcinoma cells were used to evaluate the effect of chemicals on steroidogenesis. Using a 96-well format, cells were pre-stimulated with 10 μ M forskolin for 48 hr to induce steroidogenesis followed by chemical exposure for 48 hr. Media were removed and 13 hormone analytes were quantified by HPLC-MS/MS including progestagens (pregnenolone [PREG], progesterone [PROG], and their hydroxylated metabolites), glucocorticoids (corticosterone, cortisol, and their deoxy-precursors), androgens (dehydroepiandrosterone, androstenedione, and testosterone), and estrogens (estrone and estradiol). Initially, 311 unique ToxCast Phase I chemicals (primarily pesticides) were tested at a single non-cytotoxic maximum tolerated concentration (MTC). 220 chemicals were found to alter the levels of at least one hormone analyte. Based on the single concentration analysis, 96 chemicals disrupting 4s hormones were selected for six-point concentration-response evaluation (0.003 – 100 μ M). Concentration-dependent disruption of at least one hormone was observed with 68 of the selected chemicals. By evaluating the effects of chemicals on 13 hormones this assay provides valuable mechanistic insight into the possible targets for chemical perturbation in the steroidogenic pathway. For example, <10 chemicals altered PREG or 17 α OH-PREG while 27 and 35 chemicals had an effect on PROG and 17 α OH-PROG levels, respectively. These results demonstrate that the chemicals evaluated likely do not target CYP17a hydroxylase activity. However, 33 chemicals altered testosterone levels and 38 chemicals concentration-dependently altered estradiol levels revealing significant disruption of subsequent dehydrogenation and aromatization steps. Cumulatively, these results suggest CYP17a lyase and hydroxysteroid dehydrogenase activity are the likely targets for the disruption of steroidogenesis by the subset of ToxCast Phase I chemicals evaluated. *This abstract does not necessarily reflect US EPA policy.*

OBJECTIVES

- Develop high-throughput protocol for testing disruption of steroidogenesis capable of measuring many steroid analytes in the pathway
- Develop a cost-effective screening pipeline for testing effects of chemicals on H295R cells
- Characterize the profile of effects on steroidogenesis across a diverse library of chemicals

Study Design & Workflow



Steroidogenesis Pathway

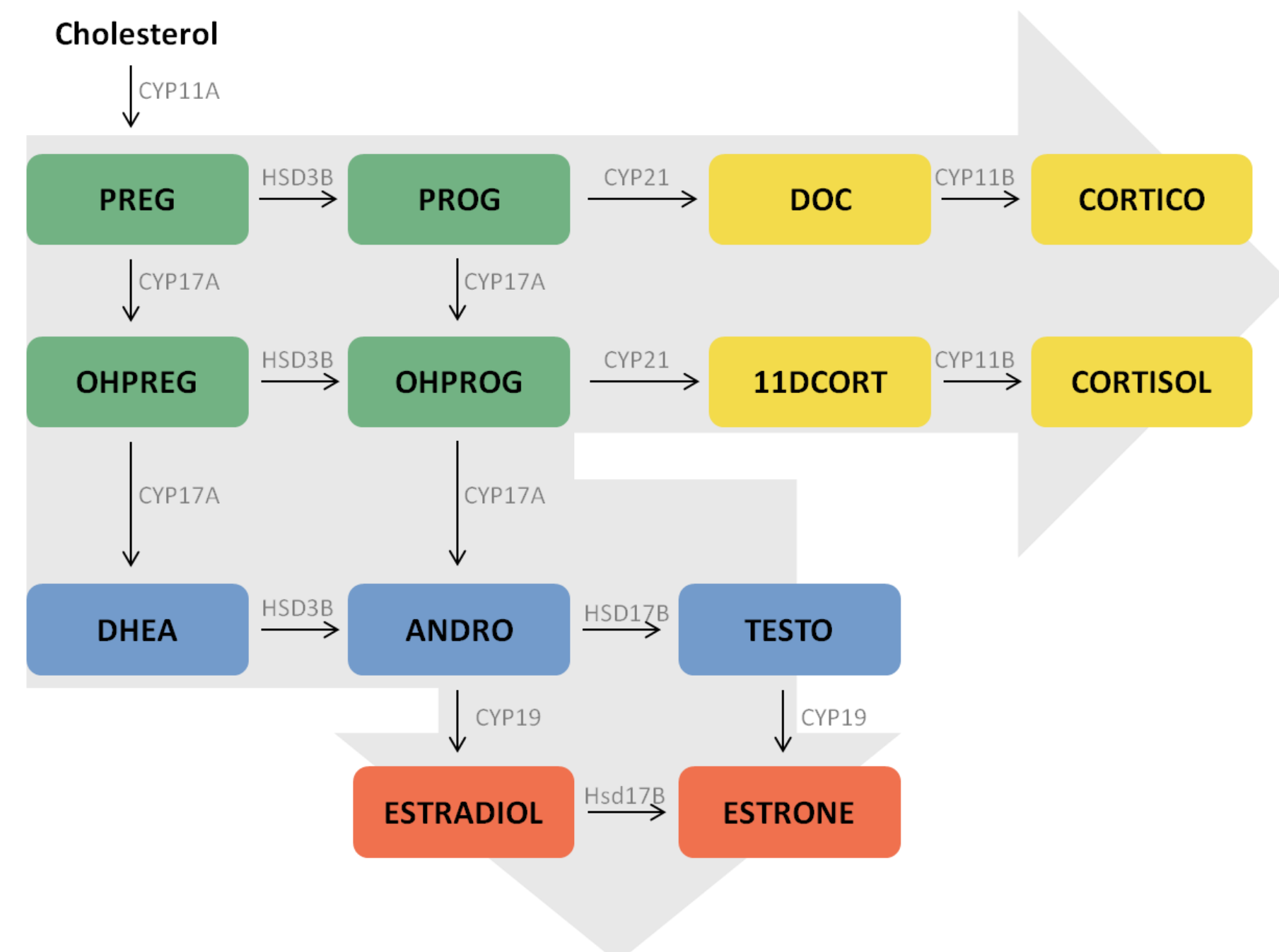


Figure 3. Steroidogenesis Pathway: Stimulation of steroidogenesis in immortalized H295R human adrenocortical carcinoma cells by 10 μ M forskolin results in detectable levels of steroid hormone production. The steroid hormones evaluated in this study can be categorized into four classes: progestagens (represented in green), glucocorticoid/mineralocorticoid (yellow), androgens (blue) and estrogens (red). Gene symbols for the enzymes carrying out each step in steroidogenesis are indicated in grey. Briefly, cholesterol is committed to steroid hormone by CYP11A. Progestagens can be metabolized by CYP21 to form glucocorticoids and mineralocorticoids or by CYP17A to form androgens and subsequently estrogens via CYP19.

Stimulation of Steroidogenesis

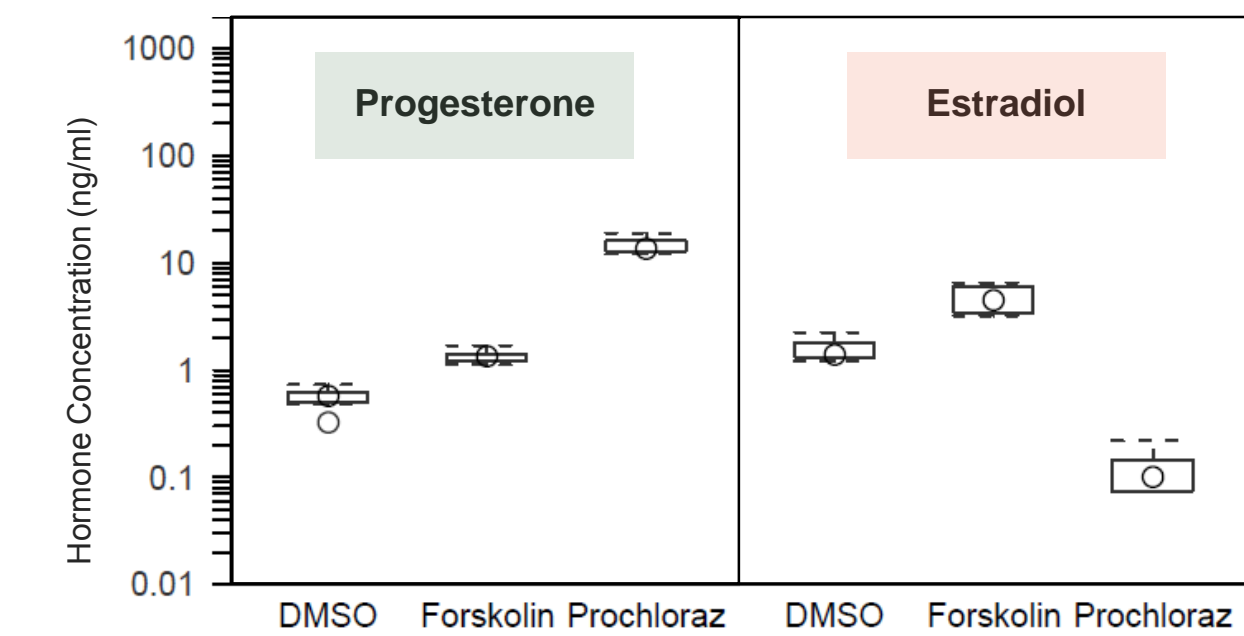


Figure 4. Stimulation of H295R Cells: Stimulation of steroidogenesis with 10 μ M Forskolin induced hormone production. Conversely, treatment with 3 μ M Prochloraz, inhibiting Cyp17 activity, increased progesterone levels while inhibiting estradiol. All treatments were conducted with a 24 hr pre-stimulus using 10 μ M forskolin followed by 48 hr treatment in 0.1% DMSO.

Profiling Chemical Effects on Steroidogenesis

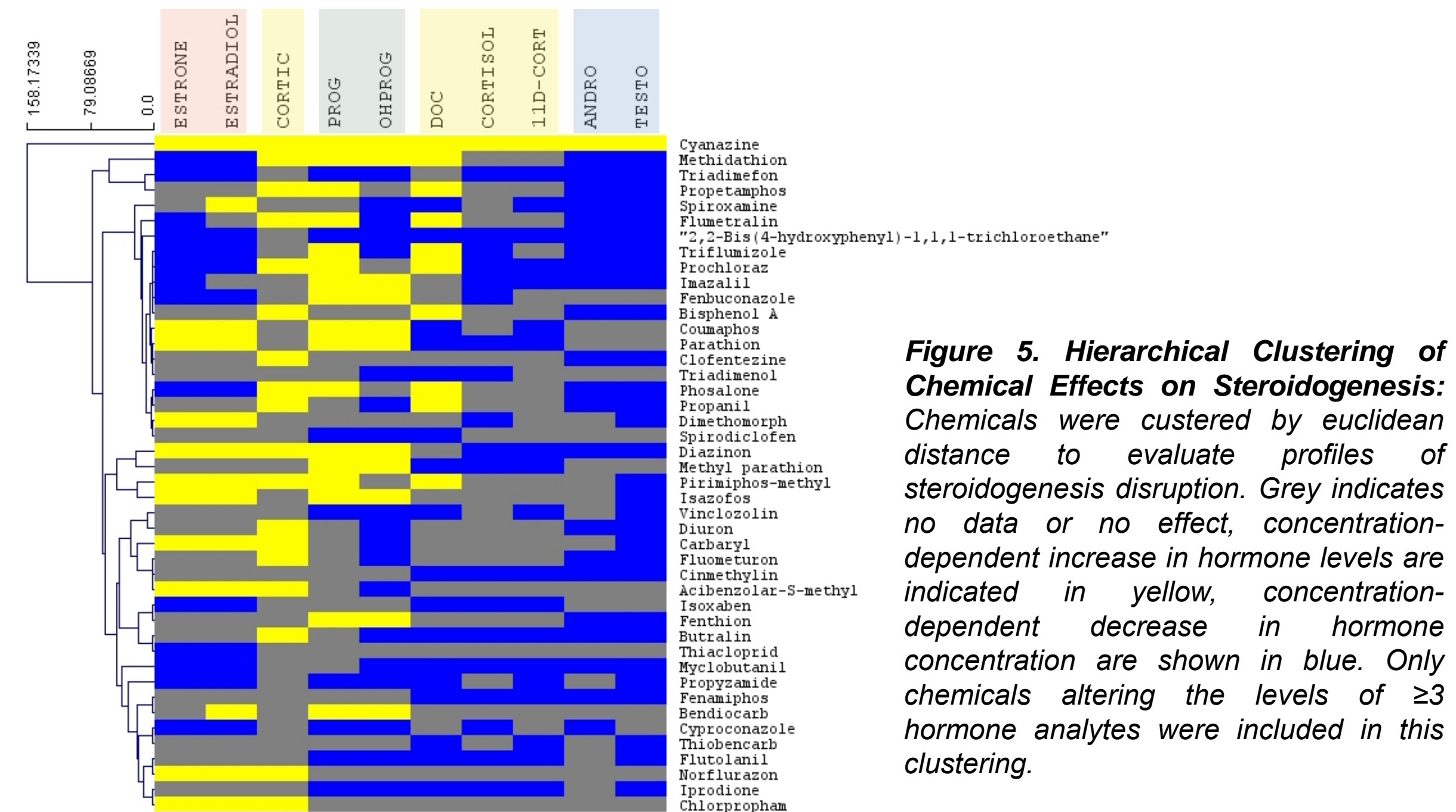


Figure 5. Hierarchical Clustering of Chemical Effects on Steroidogenesis: Chemicals were clustered by euclidean distance to evaluate profiles of steroidogenesis disruption. Grey indicates no data or no effect, concentration-dependent increase in hormone levels are indicated in yellow, concentration-dependent decrease in hormone concentration are shown in blue. Only chemicals altering the levels of ≥ 3 hormone analytes were included in this clustering.

Chemical Effects on Steroidogenesis

Table 1. Summary of Concentration-Response Evaluation of Hormone Concentrations for 96 ToxCast Phase I Chemicals

Hormone	Acronym	Quantification			Chemicals with Conc.-Dependent Effects
		Range (ng/ml)	Average Concentration (ng/ml)		
pregnenolone	PREG	2-400	3.3		9 ($\uparrow 3$, $\downarrow 6$)
17 α -OH pregnenolone	OH-PREG	5-1000	8.4		3 ($\uparrow 1$, $\downarrow 3$)
progesterone	PROG	0.2-40	0.4		27 ($\uparrow 8$, $\downarrow 19$)
17 α -OH progesterone	OH-PROG	0.2-40	15.2		35 ($\uparrow 21$, $\downarrow 14$)
deoxycorticosterone	DOC	0.5-100	3.9		29 ($\uparrow 18$, $\downarrow 11$)
corticosterone	CORTICO	0.5-100	0.6		19 ($\uparrow 0$, $\downarrow 19$)
11-deoxycortisol	11DCORT	5-1000	246.7		20 ($\uparrow 19$, $\downarrow 1$)
cortisol	CORTISOL	0.5-100	17.4		21 ($\uparrow 20$, $\downarrow 1$)
dehydroepiandrosterone	DHEA	3-600	3.9		3 ($\uparrow 2$, $\downarrow 1$)
androstenedione	ANDRO	1-200	111.9		24 ($\uparrow 23$, $\downarrow 1$)
testosterone	TESTO	0.1-20	2.9		33 ($\uparrow 32$, $\downarrow 1$)
estrone	ESTRONE	0.03-6	2.1		38 ($\uparrow 17$, $\downarrow 21$)
estradiol	ESTRADIOL	0.03-6	0.2		36 ($\uparrow 13$, $\downarrow 23$)

Summary

- ▶ H295R cells are a suitable model for high-throughput screening for chemical effects on steroidogenesis
- ▶ 24 hr stimulus with forskolin prior to chemical treatment allows for the detection of both increases and decreases in hormone levels
- ▶ 68 of the 96 chemicals selected for concentration-response evaluation elicited concentration-dependent effects on at least one hormone analyte
- ▶ Distinct profiles of steroidogenesis disruption can be observed among chemicals, demonstrating the utility of this model to not only identify chemicals that perturb steroidogenesis, but also the ability to evaluate possible mechanisms underlying altered steroidogenesis