Dose response from high throughput gene expression studies and the influence of time and cell line on inferred mode of action by ontologic enrichment.

Introduction

Gene expression with ontologic enrichment and connectivity mapping tools is widely used to infer modes of action (MOA) for therapeutic drugs. Despite progress in high-throughput (HT) genomic systems, strategies suitable to identify industrial chemical MOA are needed. The L1000 is a HT genomics platform that measures 1000 landmark genes to computationally predict expression across an equivalent whole human genome array. We used the L1000 system with visualization tools to assess predicted gene expression changes and ontologic enrichment for 9 agrochemicals at 9 concentrations (0.001, 0.003, 0.01, 0.03, 0.1, 0.3, 1, 3, 10 µM) across 5 cell lines (Table 1 & 2) at 6 and 24 hr after treatment. Genes significant for a monotonic dose response (log likelihood ratio test with permutations) were used for analyses.

Table 1. Cell lines selected for exposure screens			
Cell Line	Replicates	Characteristics	
HepaRG	5	Metabolically competent hepatic cells	
A549	3	Lung cancer	
A673	3	Bone cancer	
HT29	3	Colorectal cancer	
MCF7	3	Breast cancer	

 Table 2. Agrochemicals selected for gene expression analysis with the
L1000 platforms. Each compound was tested across 9 treatment concentrations plus controls, and sampled at 6 hours and 24 hours.

Agricultural Use	CAS No.
Chlorophenoxy herbicide	94-75-7
Amide Herbicide	87674-68-8
Amine fungicide/antioxidant	122-39-4
unclassified pesticide	153233-91-1
Triazole Fungicide	114369-43-6
Azole Pesticide	35554-44-0
Chlorophenoxy Herbicide	94-74-6
Sulfonylurea, Herbicide	122931-48-0
Urea herbicide	51707-55-2
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Methods



Figure 1. Cells were exposed in 384-well plates for scanning of landmark gene expression. The 1000 measured landmark genes were used to computationally predict the equivalent of an Affymetrix U133A full genome microarray (>20,000 probes). Each plate batch used independent vehicle controls, allowing for batch-specific normalization by Z-scores. Normalized whole array equivalent data was analyzed for differential expression by Likelihood ratio test of monotonic dose response, with 100 permutations using the R/Bioconductor package IsoGene.¹ Genes significantly up or down expressed relative to vehicle controls for a dose dependent monotonic response (FDR<0.05) were selected for ontology enrichment. Ontologic enrichment of genes from the predicted expression profiles was performed against the public Reactome ontology and patterns of functional category enrichment were compared using visualization software developed at the Hamner Institutes.²

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no significant enrichment with down-regulated genes.

Figure 3. Differences in ontologic enrichment across time: 2,4-D at 6Hr (A549 cells) and at 24Hrs (A673 cells) All colored nodes significant at FDR<0.05 and with a minimum of 5 query elements found in category elements. White nodes are non-significant. Color coding in **3a** is relative to enrichment FDR. Color coding in **3b** is relative to the Venn diagram, with mustard indicating enrichment with both up- and down-regulated genes. Node size is proportional to the number of query elements found in the ontology category.



Differential gene expression levels varied significantly across cell lines and times, with no one cell line consistently responsive for any chemical at both times. A broad assessment of the 9 chemicals showed that A549 and HT29 cells (at 6 hr) and HepaRG and A673 cells (at 24hr) were typically the most responsive. At 6hr, A549 and HT29 cells tended to show significant gene enrichment of cell cycle mitotic processes. At 24hr, A673 cells tended to show gene enrichment of immune response, disease and cellular metabolic pathways.

The herbicide 2,4-D has been linked *in vitro* (HepG2 cells) to gene expression changes related to cell cycle processes, DNA repair mechanisms and immune response pathways³. At 6hr, the gene response observed with A549 cells showed ontologic enrichment for many of these same process categories. However, by 24hrs., the greatest number of gene expression changes were seen with A673 cells. These too demonstrated significant enrichment for immune processes but there was an absence of the cell cycle and mitotic process signals seen earlier.

Comparison of HepaRG and A673 cell responses (at 24hr) for Fenbuconazole, a fungicide associated with liver effects, showed enrichment of common parent pathways (DNA repair, cell signaling) in both cell types, but several child categories of these were identified only in HepaRGs. Fenbuconazole exposure is linked with increased liver weight, hepatocellular hypertrophy and proliferation, and clinical chemistry changes (changes in cholesterol and triglyceride levels, and increases in serum activity of hepatic enzymes) were observed in rat, dog and mouse exposures. This fungicide has also been seen to induce mouse liver cytochrome P450 activity in a manner similar to phenobarbital.⁴ Active ToxCast assays for Fenbuconazole are associated with cell cycle pathways, cell proliferation and cell death.

Both A673 and HepaRG cells exposed to Fenbuconazole detected significant upregulation in numerous cytochrome P450 genes (12 with A673 cells, and 25 with HepaRG cells), resulting in a strong enrichment signal for general metabolism processes. However, only HepaRG cells showed wide ranging enrichment in cell cycle and mitotic processes and cell signaling processes consistent with cell proliferation and hypertrophy.

toxicological studies. MOA.

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Conclusions

• The L1000 platform has promise for HT genomic testing in human cells at reasonable costs after better validation efforts.

• Differential expression response varied widely across cell lines and time points. While L1000-based methods worked for predictions from expression responses, our sampling of compounds and time points in this poster remians limited.

• Where response was robust for multiple cell lines at a given time point, differences were seen between cell lines in ontologic enrichment. Future efforts with this platform might benefit from using compounds with more complete data bases on modes of action, including previous studies that include other gene expression data and

• These findings suggest selection of appropriate time points and the use of multiple cell models should be considered in HT genomics strategies designed to inform chemical

References

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