

Modeling and Predicting Cancer From ToxCast Phase I Data Richard Judson, David Dix, Keith Houck, Thomas Knudsen, Imran Shah, David Reif, Matthew Martin, Holly Mortensen, Daniel Rotroff, Robert Kavlock

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Univariate Correlations With Rat Liver **Tumor Progression**

Network of genes associated with the

progression of rat liver tumor endpoints.

Associations were calculated using Fisher's

exact test, with assay AC50/LEC values ≤100

mM set to 1, and >100 mM set to 0. Only

associations with a p-value <0.01 are included.

Links between genes (yellow) and in vivo

endpoints (pink) are shown where there is a

statistical association based on the in vitro

assay results. The "Any Lesion" category

contains the "Pre-neoplastic" category, which

in turn contains the "Neoplastic" lesions

category. Disease or disorder classes (cyan)

are linked to genes.

Science Question

The goal of the ToxCast program is to link in vitro assays with in vivo toxicity in humans. One strategy to approach this goal is to look for statistical associations between the ToxCast assays and in vivo phenotypes for animal models. The questions we address here are:

- Can we find statistically significant associations between in vitro assays and cancer endpoints in rodents?
- 2. Can multi-assay models be constructed with high enough sensitivity and specificity to be useful for screening and prioritization?
- 3. Does our understanding of biology (through cancer knowledge of genes and pathways) support these models?

Research Goals

- 1. To develop a bioinformatics infrastructure that allows us to compile all ToxCast and ToxRefDB together in a way that allows high-throughput statistical analyses
- To use statistical and machine 2. learning algorithms to construct and test models linking in vitro assavs with cancer endpoints in rats and mice
- 3. To qualify these models using information from the literature on general cancer biology and mode of action information on ToxCast chemicals In particular we want to understand toxicity in terms of activation of specific genes and pathways

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Assays

A total of 687 in vitro assav endpoints were used from ToxCast. This includes multiple time points for several assays. All assays were run in concentration-response format and we extracted a characteristic concentration for each chemical-assay pair. These were either AC50 or LEC (lowest effective concentration, or the lowest concentration at which the response was significantly different than negative control). For these calculations, we did not use information on magnitude of effect (Max fold change or Emax). Assay AC50/LEC values were log transformed prior to performing statistical calculations. In addition to the in vitro assav data, we also used chemical class information, limited physico-chemical values calculated from chemical structure, and structure fragment composition fingerprints.



Gene/Pathway

Perturbation Scores

Most assays were mapped to genes (313

human and 86 rat) and gene perturbation

scores were calculated. These were the

lowest AC50/LEL observed for a chemical for

Genes were mapped to KEGG and Ingenuity

pathways and pathway perturbation scores for

a chemical were given the value of the lowest

AC50/LEC value for any assay mapped to a

gene mapped to that pathway. However, the

chemical needed to be active in at least 5

assays mapped to that pathway for a pathway

perturbation score to be calculated.

any assay mapped to that gene.

Cancer-related endpoints from rat and mouse chronic-cancer studies were extracted from ToxRefDB and dichotomized (cause / do not cause cancer)



Disease Progression



endpoint associations were calculated. Results with p<0.05 were kept. By chance, 41 such associations were expected and 58 were found. The table shows relative risk values. Green indicates there is literature support for the association.



Conazole pathway coverage. Color band indicates (red) rat liver tumoriaen

UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

Multivariate Signature For Rat Liver Proliferative Lesions

A Stepwise Logistic Regression model was created linking several assays with rat liver proliferative lesions. The signature genes are PPARG, HMGCS2 (surrogate for PPARA), CCL2 and AR, all of which are known to be associated with cancer, and all but CCL2 are linked to liver tumors in rodents and or man. The figure shows the hits of these assays against the subset of chemicals which are liver tumorigens in rats. 248/309 chemicals had rat data in ToxRefDB





Model was built for

examine the subset of

liver tumors in rat. Not

all tumorigens have

hits in the predictive

chemicals ordered by

chemical similarity) but

they do have a wide

range of activities in

addition to the model

assays (right, ordered

by activity similarity)

lesions

causino

(above.

figures

proliferative

chemicals

assays

but these





research&development



Specificity=94%

Prediction statistics for the Model Specificity is high, so few false positives are seen, but sensitivity is low, indicating that many of these chemicals act through mechanisms not adequately probed by these assays, including the need for metabolic activation / deactivation

Results/Conclusions

A simple predictive model using ToxCast assays and logistic regression produces a predictive signature for rat liver proliferative lesions. The genes showing up in the signature all have independent links to cancer. In particular, chemical-related PPAR activity is readily seen as a risk for rat liver proliferative lesions

Impact and Outcomes

- 8 chemicals not in ToxRefDB were predicted to be positive for rat liver proliferative lesions
- PFOA: Causes rat liver adenomas
- PFOS: Causes rat liver adenomas
- Diniconazole: rat liver hypertrophy Chlorothalonil: rat liver enlargement, kidnev tumors
- TCMTB: testicular and thyroid adenomas No data for Niclosamide. Methylene
- bis(thiocyanate), Phenoxyethanol

Future Directions

- We are pursuing a variety of other analysis strategies. These include:
- Focusing on particular chemical classes 1 to better control for chemical structure
- 2. Including pharmacokinetic information as it becomes available
- 3 Eccusing on specific chemicals where mode of action is well known and using these to understand the strengths and weaknesses of in vitro to in vivo mapping.

References

- M.T. Martin, R. Judson, D. Reif, D.J. Dix, R. Kavlock, "Profiling Chemicals Based on Chronic Toxicity Profiles from the U.S. EPA ToxRef Database". Environmental Health Perspectives. Vol. 117, 392-399 (2009)
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